

Effect of Formulated Artocarpus champeden Extract on Parasite Growth and Immune Response of Plasmodium berghei-Infected Mice

by Aty Widyawaruyanti

Submission date: 23-Mar-2020 11:11AM (UTC+0800)

Submission ID: 1280089166

File name: owth_and_Immune_Response_of_Plasmodium_berghei-Infected_Mice.pdf (1.19M)

Word count: 4950

Character count: 25596

Research Article

Effect of Formulated *Artocarpus champeden* Extract on Parasite Growth and Immune Response of *Plasmodium berghei*-Infected Mice

A. Widyawaruyanti^{1,2}, N. Harwiningtias,³ L. Tumewu,² A.F. Hafid,^{1,2} and Soetjipto⁴

¹Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy, Universitas Airlangga, Surabaya 60115, Indonesia

²Natural Product Medicine Research and Development, Institute of Tropical Disease, Universitas Airlangga, Surabaya 60115, Indonesia

³Faculty of Pharmacy, Universitas Airlangga, Surabaya 60115, Indonesia

⁴Department of Biochemistry, Faculty of Medicine, Universitas Airlangga, Surabaya 60132, Indonesia

Correspondence should be addressed to A. Widyawaruyanti; aty-w@fl.unair.ac.id

Received 28 June 2019; Revised 4 January 2020; Accepted 8 February 2020; Published 29 February 2020

Guest Editor: Chukwuma O. Agubata

Copyright © 2020 A. Widyawaruyanti et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. The ethanol extract of *Artocarpus champeden* stem bark (ACEE) has been proven to exhibit antimalarial activity. Despite the antimalarial effects observed, mechanisms of immune response to explain the antimalarial activity of ACEE remain poorly characterized. Here, we show the production of pro- and anti-inflammatory cytokines T helper 1 (Th1: IFN- γ , TNF- α) and T helper 2 (Th2: IL-10) from *Plasmodium berghei*-infected mice treated with formulated ACEE in order to better characterize the mechanism behind ACEE antimalarial activity. In addition, we have also determined the effect of formulated ACEE on parasite growth and liver function. **Methods.** Balb/c mice were infected with *P. berghei* strain ANKA and then administered daily doses of ACEE at a dose of 20, 50, and 100 mg/kg BW, and survival time was recorded. We determined the presence of *P. berghei* in the blood of the infected mice and inhibition of *P. berghei* growth. In order to assess the liver function of infected mice, we determined aspartate transaminase (AST) and alanine transaminase (ALT) levels. Determination of cytokines Th1 (IFN- γ , TNF- α) and Th2 (IL-10) levels was conducted using enzyme-linked immunosorbent assay (ELISA). **Results.** We found that formulated ACEE inhibited parasite growth and showed the highest antimalarial activity at 100 mg/kg BW. AST and ALT levels were found to be in the normal range, and there was no significant difference among control and treatment groups ($P > 0.05$). Infected mice treated with formulated ACEE showed a significant increase in the production of IFN- γ on day 7 and an increased production of TNF- α on day 4 and day 7. No effect on IL-10 production was observed. Decreased parasitemia and longer survival time were observed when compared with untreated infected mice. **Conclusion.** This study suggests that the administration of ACEE was effective in inhibiting *P. berghei* growth in infected mice and extending survival time. No effect on liver function was observed based on AST and ALT levels. The antimalarial effects of ACEE could be explained in part by the enhanced production of the proinflammatory cytokines IFN- γ and TNF- α . ACEE treatment may provide novel therapeutic strategies for malaria in future.

1. Introduction

In 2018, there were an estimated 228 million new cases of malaria worldwide, in which 405,000 were estimated to have resulted in death [1]. Between 2010 and 2015, the malaria mortality rate among children under five years of age fell by an estimated 35%. Nevertheless, malaria remains a significant

threat to under-fives, claiming the life of one child every two minutes [1]. Moreover, increasing resistance to current antimalarial drugs remains a threat to global efforts to control and eliminate malaria [2]. Therefore, safe and effective alternative antimalarial drugs are urgently needed.

Plants have long been potential sources of antimalarial drugs. *Cinchona* species are well known for their antimalarial

properties. Quinine, isolated from *Cinchona* species, is one of the most common drugs used to treat malaria. Artemisinin, its active compound, has a higher chemotherapeutic index than chloroquine, a common antimalarial drug [3]. Plants used to treat malaria traditionally are potential sources of new antimalarial drugs [4, 5].

Indonesia's high biodiversity makes it an excellent bioprospecting source for new antimalarial drugs. *Artocarpus champeden* (Moraceae family), which in Indonesia is commonly known as *cempedak*, has been used as a traditional medicine to cure malaria [6]. In our previous studies, several prenylated flavones were isolated from *A. champeden* stem bark extract and showed potent antimalarial activity against *Plasmodium falciparum* [7]. Heteroflavanone C was the most potent compound among the compounds isolated, which has inhibitory activity against the growth of *P. falciparum* 3D7 strain with an IC_{50} value of 0.0006 $\mu\text{g}/\text{mL}$ [7]. Furthermore, artopeden A, an isoprenylated flavone, was also isolated from the bark of *A. champeden*. This compound showed potent antimalarial activity with an IC_{50} value of 0.045 $\mu\text{g}/\text{mL}$ [8]. Hafid et al. isolated another antimalarial active compound from *A. champeden* stem bark, which identified as morachalcone A with an IC_{50} value of 0.18 $\mu\text{g}/\text{mL}$ [9].

It is known that the most substantial content of *A. champeden* is flavonoids [10]. Flavonoids can affect immune responses and act as an anti-inflammatory agent. This has been demonstrated *in vitro* by inhibiting TNF- α (proinflammatory cytokines) as well as through upregulation of Th1 and downregulation mechanisms such as Th2 [11–13]. Another study has been done by Chang (2007), who stated that flavonoids modulate IFN- γ expression [14]. Flavonoids also have the ability to increase the production of NO and tumor necrosis factor (TNF- α) [15]. Flavonoids isolated from *Bidens pilosa*, centaaurin, and centaureidin can increase IFN- γ production in T cells of the mice spleen [14]. Therefore, we assumed that flavonoids from *A. champeden* were possible to have activity in modulating TNF- α and IFN- γ expression as well.

In malaria infection, cytokines are mediators involved in pathogenesis and the elimination of the parasite [16]. Clark stated that TNF- α is a mediator of severe malaria. Proinflammatory cytokines, including TNF- α and IFN- γ , are thought to be responsible for cerebral symptoms [16, 17]. Cytokines control *Plasmodium* infection intracellularly through Th1 cells. Later, this requires counterbalance through the Th2 response to eliminate the pathogen extracellularly by Th2 cells by producing antibodies. IL-10 is a cytokine of Th2 that can decrease parasitemia in the late phase [18].

In order to develop the herbal product of *A. champeden*, its stem bark extract was further formulated by adding several excipients as capsule dosage form (formulated ACEE: *Artocarpus champeden* ethanol extract). The formulation of the extract could affect the activity; therefore, the antimalarial activity of the formulated extract needed to be evaluated. In this study, there is evidence that the administration of formulated ACEE was effective in inhibiting *P. berghei* growth in infected mice and extending survival time. No effect on liver function was observed based on AST and ALT levels.

Currently, the immune response to formulated ACEE administration is not well known, especially in relation to malaria. Herein, we demonstrated the effect of formulated ACEE on host immune response, especially on the expression of Th1/Th2 cytokine when infected by *P. berghei*. Treatment of formulated ACEE resulted in an increased production of cytokines Th1 (IFN- γ , TNF- α) when compared with untreated mice.

2. Materials and Methods

2.1. Material. An 80% ethanol extract of *A. champeden* stem bark (ACEE) was formulated as a capsule dosage form containing 15 mg extract per 300 mg granule. A wet granulation method was conducted to produce formulated ACEE by adding several excipients, including PEG 6000, PVP K25, lactose, Avicel PH-101, and Primogel. The formula is proprietary. The formulation was produced at the Faculty of Pharmacy, Universitas Airlangga, Surabaya, Indonesia.

2.2. Parasite. *P. berghei* ANKA strain was obtained from the Eijkman Institute for Molecular Biology, Jakarta. This parasite has been maintained at the Institute of Tropical Disease, Universitas Airlangga, Surabaya, Indonesia.

2.3. Animals. Female mice Balb/c strain at the age of 6–8 weeks and 20–30 g of weight was obtained from the Animal Experimental Development Unit-Gadjah Mada University (Yogyakarta, Indonesia). Animals were maintained at the Animal Laboratory of the Institute of Tropical Disease, Universitas Airlangga. Permission and approval for animal studies were obtained from the Faculty of Veterinary Medicine, Universitas Airlangga, with ethical clearance No: 194-KE.

2.4. In Vivo Antimalarial Activity and Effect on Liver Function. *In vivo* antimalarial testing was performed based on Peter's test (a 4-day suppressive test) with modifications [19]. The test was performed using 6 mice per group. Mice were infected intraperitoneally with 0.2 mL (1×10^7) of *P. berghei* on day 0. The treatment was initiated after parasite infection occurred (day 2). *P. berghei* infected mice were treated orally with formulated ACEE (granule form suspended in CMC-Na 0.5%) twice a day for five days (day 2 until day 6) at a dose equal to ACEE 20, 50, and 100 mg/kg body weight. Giemsa-stained thin blood smears were made for 7 days (day 2 until day 8). The infected red blood cells were counted under the microscope to determine the parasitemia and the inhibition of parasite growth. Mice blood was taken intracardially at day 8 to determine the effect on liver function by the observed AST and ALT levels.

The percentage of growth inhibition of *P. falciparum* was calculated using the following formula:

$$100 - \left(\frac{X_c}{X_k} \times 100\% \right), \quad (1)$$

where X_e is the % parasitemia growth of an experimental group and X_k is the % parasitemia growth of the negative control.

2.5. Determination of Immune Response (IFN- γ , TNF- α , and IL-10) on *P. berghei*-Infected Mice. Female mice Balb/c strain was divided into uninfected and *P. berghei*-infected groups. Each group was further divided into untreated and treated groups. Each group consisted of 6 mice. For the infected group, mice were infected intraperitoneally with 0.2 mL (1×10^7) of *P. berghei* on day 0. For the treated group, mice were given ACEE orally (granule form suspended in CMC-Na 0.5%) at a dose of 100 mg/kg BW or CMC-Na 5% twice a day for four days (day 2 until day 5). ACEE at a dose of 100 mg/kg BW was chosen based on previous *in vivo* antimalarial test results (Table 1). The uninfected and untreated group was used as controls to determine the normal levels of IFN- γ , TNF- α , and IL-10. The infected and untreated group was used as a natural malaria pathogenesis model. Parasitemia was observed every day (day 2 until day 12). On days 2, 4 and 7 after infection, mice were euthanized, and blood was collected to determine IFN- γ , TNF- α , and IL-10 by ELISA. Indirect sandwich ELISA TNF- α , IFN- γ , IL-10 cytokine of TNF- α , IFN- γ , and IL-10 was conducted using pairs of capture and detection Abs (BioLegend: Mouse cytokine ELISA MAXTM Deluxe Sets).

2.6. Statistical Analysis. The numerical data are presented as mean \pm standard deviation. Data were analyzed by SPSS 20.0. The significance of the mean difference between independent groups was determined using a one-way analysis of variance (ANOVA) and least significant difference (LSD) analysis. The P value <0.05 is considered significant.

3. Result

3.1. In Vivo Antimalarial Activity and Toxicity Effect on Liver Function. Formulated ACEE at a dose equal to ACEE 20, 50, and 100 mg/kg inhibited parasite growth by 57.36% (± 9.18), 64.48% (± 5.38), and 72.24% (± 4.30), respectively. The best inhibition shown was at a dose equal to ACEE 100 mg/kg. There was a significant difference in parasite growth inhibition among groups ($P > 0.05$). The parasite growth and inhibition data are described in Table 1 and Figure 1.

The effect of ACEE on liver function was observed by determining the AST and ALT levels of the treated mice groups compared with the control group. The AST and ALT levels of the treated groups ranged from 264.33–278.33 U/L and 79.16–115.66 U/L, respectively. Meanwhile, AST and ALT levels of the control group were 266.28 U/L and 89.57 U/L, respectively.

The normal AST and ALT levels of mice are 70–100 U/L and 25–200 U/L, respectively. We observed that AST and ALT of all groups were in the normal range. Statistical analysis results showed that there were no significant differences in AST and ALT levels among groups ($P > 0.05$). The AST and ALT levels are described in Table 2.

TABLE 1: Percentage of parasite growth and inhibition of *P. berghei* at day 6 and day 8.

Group	Parasite growth (%) ^a		Parasite growth inhibition (%) ^a	
	Day 6	Day 8	Day 6	Day 8
ACEE 20 mg/kg	1.28 \pm 0.26	1.48 \pm 0.32 ^a	62.67 \pm 7.49	57.36 \pm 9.18 ^a
ACEE 50 mg/kg	1.39 \pm 0.24	1.24 \pm 0.18 ^b	59.34 \pm 6.98	64.48 \pm 5.38 ^a
ACEE 100 mg/kg	1.00 \pm 0.26	0.97 \pm 0.15 ^a	70.66 \pm 7.58	72.24 \pm 4.30 ^a
Untreated	3.42 \pm 0.61	3.48 \pm 0.33 ^{ab}	—	—

^aData represent mean \pm SD ($n = 6$); ^{ab}significant difference among groups ($P < 0.05$); untreated group as control group.

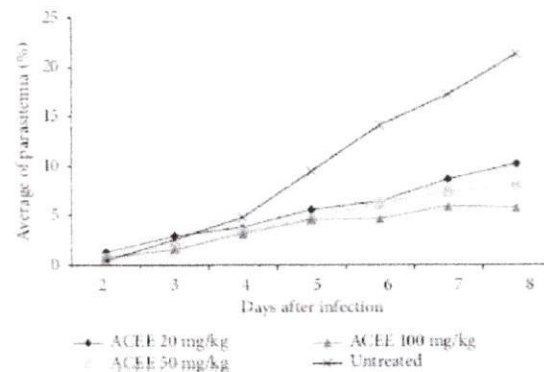


FIGURE 1: The mean of parasitemia of treated and untreated groups on day 2 until day 8 after parasite infection.

TABLE 2: AST and ALT levels of the treated and control groups.

Group	AST (U/L) ^a	ALT (U/L) ^a
ACEE 20 mg/kg	278.33 \pm 31.05	85.66 \pm 29.14
ACEE 50 mg/kg	264.33 \pm 25.53	115.66 \pm 34.91
ACEE 100 mg/kg	269.00 \pm 70.01	79.16 \pm 17.87
Untreated	266.28 \pm 67.48	89.57 \pm 40.22

^aData represent mean \pm SD ($n = 6$); no significant different among groups ($P > 0.05$).

ACEE showed the highest antimalarial activity at a dose equal to ACEE 100 mg/kg; no effect on liver function based on AST and ALT levels was observed. Therefore, further investigation of the immune response of *P. berghei*-infected mice treated with ACEE was done by administering ACEE at a dose equal to 100 mg/kg.

3.2. Determination of Immune Response (IFN- γ , TNF- α , and IL-10) on *P. berghei*-Infected Mice

3.2.1. Parasitemia Observation. The antimalarial activity of ACEE on *P. berghei*-infected mice was determined by the percentage of parasitemia (observation on day 2 until day 12), parasitemia growth, and inhibition of growth (observation on day 4, day 7, and day 12). The results showed that

TABLE 3: Parasite growth and inhibition on *P. berghei*-infected mice.

Groups	Parasitemia growth (%) ^a			Inhibition (%) ^a			Mean of survival days
	Day 4	Day 7	Day 12	Day 4	Day 7	Day 12	
Infected/untreated	1.38 ± 0.08 ^a	4.55 ± 0.92 ^c	4.48 ± 0.29 ^d	—	—	—	11.00 ± 0.00
Infected/CMC-Na	1.37 ± 0.14 ^b	6.10 ± 1.87 ^c	5.92 ± 1.77 ^d	NI	NI	NI	10.33 ± 1.75
Infected/ACEE	0.36 ± 0.22 ^{ab}	1.91 ± 0.57 ^c	3.75 ± 0.89 ^d	74.03 ± 16.16	55.53 ± 12.65	36.53 ± 15.09	11.83 ± 1.60

^aData represent mean ± SD (n = 6); infected/untreated group as control group; NI: no inhibition; ^{b,c,d}significant difference among groups (P < 0.05).

on the infected and untreated groups, the parasites began to grow on day 2 and increased until day 12. ACEE demonstrated the highest inhibitory effect on parasite growth on day 4 (Table 3).

3.2.2. Survival Rate Observation. There was increased survival for infected mice given ACEE, in which mean of survival days was 11.83 (±1.60) days. Infected and untreated mice survived until day 11. Meanwhile, the administration of CMC-Na 0.5% resulted in decreased mean survival days by 10.33 (±1.75) days. It was possibly caused by a high parasitemia level of mice in infected/untreated and infected/CMC-Na groups (Table 3).

3.2.3. Cytokine Determination. Immune responses of mice were conducted through the determination of the production of cytokine Th-1 (IFN- γ and TNF- α) and cytokine Th-2 (IL-10) (Table 4).

Statistical analysis showed that there was no significant difference in cytokine production between uninfected groups on day 4 and day 7. However, the production of TNF- α was significantly different between infected/ACEE group and the other groups on day 4. There was no significant difference between the infected/ACEE group and the infected/CMC-Na group on day 7. The production of IFN- γ was significantly different between uninfected and infected groups on day 4 and day 7. Infected groups had increased TNF- α and IFN- γ levels compared with uninfected groups. IL-10 production was not different among groups on day 4 and day 7.

4. Discussion

4.1. In Vivo Antimalarial Activity and Toxicity Effect on Liver Function. *Artocarpus* (Moraceae family) contains approximately 50 species; many of these are used in traditional medicine [20]. Several *Artocarpus* species are reported to exhibit antimalarial activities against *P. falciparum* and *P. berghei*. A prenylated stilbene compound isolated from aerial parts of *A. integer* was reported to have antimalarial activity with an IC₅₀ value of 1.7 μ g/mL [21]. *A. altilis* leaf extract showed antimalarial activity with an IC₅₀ value of 1.32 μ g/mL against *P. falciparum* and ED₅₀ value of 0.82 mg/kg body weight against *P. berghei* [22]. Several compounds isolated from *A. champeden* extract showed outstanding activity against *P. falciparum* [7–9]. Therefore, *A. champeden* stem bark extract was formulated as a capsule dosage form (formulated ACEE) and further developed as an antimalarial drug.

This study showed that formulated ACEE exhibited antimalarial activity against *P. berghei* in a dose-dependent manner. Formulated ACEE at a dose equal to ACEE 20, 50, and 100 mg/kg body weight given twice a day for five days inhibited *P. berghei* growth by more than 50% by day 8 of observation.

Antiplasmodial activity of an extract that performs percentage parasitemia suppression $\geq 50\%$ at a dose of 500, 250, and 100 mg/kg body weight per day in vivo can be classified as moderate, good, and very good, respectively [23]. Based on this classification, we can suggest that ACEE showed very good activity at doses of 20, 50, and 100 mg/kg body weight. The highest inhibition resulted from ACEE at a dose of 100 mg/kg body weight. This result was in accordance with previous study as stated above [7–9, 21, 22].

The results of liver function observation showed that AST and ALT levels of treated groups and the untreated group were in a normal range. It is therefore likely that ACEE treatment, even at the highest dose, does not affect the liver function.

4.2. Determination of Immune Response (IFN- γ , TNF- α , and IL-10) on *P. berghei*-Infected Mice. The results showed that TNF- α levels were higher with lower parasitemia in the infected/ACEE-treated group compared with infected/CMC-Na treated and infected/untreated group on day 4 and day 7. An increased TNF- α level was in accordance with decreased parasitemia in the infected group. ACEE treatment increased cytokine production that resulted in lower parasitemia. This is shown by lower parasitemia in the infected/ACEE group compared with infected/CMC-Na and infected/untreated group on day 4 and day 7. Increased TNF- α at the beginning of the infection is likely to have inhibited parasite multiplication and provided immunity protection through plasmodium [24, 25]. TNF- α is induced in monocytes and macrophages when infected erythrocytes rupture, releasing parasite antigens into the bloodstream. TNF- α also acts on lymphocytes and monocytes by increasing *P. falciparum* inhibition via mechanisms without phagocytosis [26]. Previous studies have reported that TNF- α increases within 24 hours after *P. berghei* infection. The response of TNF- α at the beginning of the infection indicates immunity protection against plasmodium [25]. Macrophage activation as an initial response after the infection was likely to have increased TNF- α production and stimulate NK cells to produce IFN- γ [27].

IFN- γ levels were significantly increased on day 7, and the infected/ACEE group had the highest IFN- γ of 290.32 ± 100.64 pg/mL compared with infected/untreated

TABLE 4: Cytokines productions (IFN- γ , TNF- α and IL-10) of mice

Groups	TNF- α (pg/mL)			IFN- γ (pg/mL)			IL-10 (pg/mL)		
	Day 2	Day 4	Day 7	Day 2	Day 4	Day 7	Day 2	Day 4	Day 7
Uninfected/CMC-Na	0.04 \pm 0.00 ^a	1.46 \pm 0.76 ^{ab}	1.21 \pm 0.62 ^{ab}	18.09 \pm 6.97	18.74 \pm 0.29 ^{abc}	18.56 \pm 0.00 ^{abc}	49.76 \pm 8.30	46.19 \pm 2.15	45.24 \pm 1.73
Uninfected/ACEE	0.04 \pm 0.00	1.12 \pm 0.48 ^{cd}	0.66 \pm 0.19 ^{cd}	18.09 \pm 6.97	19.4 \pm 2.01 ^{def}	73.09 \pm 38.32 ^{def}	49.76 \pm 8.30	45.95 \pm 2.29	46.67 \pm 2.15
Infected/untreated	1.34 \pm 0.50	3.39 \pm 1.25 ^{abc}	3.77 \pm 2.59 ^{abc}	210.50 \pm 96.50	256.52 \pm 56.57 ^{ad}	185.22 \pm 33.27 ^{ad}	74.52 \pm 48.15	51.19 \pm 13.44	77.14 \pm 50.32
Infected/CMC-Na	1.34 \pm 0.50	3.74 \pm 0.84 ^{bd}	7.94 \pm 3.30 ^{bc}	210.50 \pm 96.50	189.11 \pm 103.99 ^{bc}	225.130 \pm 61.13 ^{bc}	74.52 \pm 48.15	60.00 \pm 19.65	49.52 \pm 3.69
Infected/ACEE	1.34 \pm 0.50	5.99 \pm 2.08 ^{abc,de}	9.19 \pm 3.58 ^{bd}	210.50 \pm 96.50	208.74 \pm 78.39 ^d	290.32 \pm 100.64 ^d	74.52 \pm 48.15	51.67 \pm 12.69	51.19 \pm 11.55

Data represent mean \pm SD (n = 6); ^{a,b,c,d,e,f} significant difference among groups (P < 0.05).

and infected/CMC-Na groups with IFN- γ of 185.22 ± 33.27 pg/mL and 225.13 ± 61.13 pg/mL, respectively. IFN- γ is a Th1 cytokine, which activates macrophage. Active macrophage will release NO to eliminate the parasites present [28]. This condition was in accordance with the result in which the IFN- γ level was higher in the group with lower parasitemia (infected/ACEE) compared with the group with higher parasitemia (infected/untreated and infected/CMC-Na) on day 4 and day 7.

The TNF- α and IFN- γ levels of the infected/ACEE group were increased on day 7, even though inhibition of parasite growth considered high compared with the infected/untreated group. Prolonged induction of proinflammatory cytokines might be associated with the risk of developing severe disease. Therefore, IL-10 was necessary for proinflammatory suppression in the host during infection [29]. Meanwhile, this study result showed that the production of IL-10 on infected groups was not significantly different among groups on day 4 and day 7. ACEE treatment has not demonstrated the ability to balance the Th1 (TNF- α , IFN- γ) and Th2 (IL-10) response. Additional study is needed to improve antimalarial outcomes of ACEE treatment. Observation of the toxic effect regarding the modulating impact of proinflammatory cytokines needs to be addressed.

The IFN- γ levels in the uninfected/ACEE group were higher than those in the uninfected/CMC-Na group on day 7. This result demonstrates that ACEE could stimulate IFN- γ production in normal conditions. Therefore, it is possible to use ACEE for preventative malaria treatment.

5. Conclusions

This study suggests that the administration of formulated ACEE was effective in inhibiting parasite growth in *P. berghei*-infected mice and there was no effect on liver function based on AST and ALT levels. Furthermore, ACEE treatment was associated with the enhanced production of the proinflammatory cytokines IFN- γ and TNF- α . ACEE treatment also decreased the parasitemia level and extended survival time of *P. berghei*-infected mice. The formulated *Artocarpus champeden* ethanol extract (ACEE) treatment may provide novel therapeutic strategies for malaria.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare no conflicts of interest.

Acknowledgments

The authors wish to express their gratitude to Prof. Dr. Dt. Tahir of Tahir Foundations for the article submission support funding. This research was funded by project research grant IM-HERE Faculty of Pharmacy, Universitas Airlangga, Surabaya, Indonesia.

References

- [1] World Health Organization, *World Malaria Report 2019*, World Health Organisation, Geneva, Switzerland, 2019, <https://www.who.int/publications-detail/world-malaria-report-2019>.
- [2] World Health Organization, "Responding to antimalarial drug resistance," 2017, http://www.who.int/malaria/areas/drug_resistance/overview/en/.
- [3] F. Mojab, "Antimalarial natural product: a review," *Avicenna Journal of Phytomedicine*, vol. 2, no. 2, pp. 52–62, 2011.
- [4] J. Bero and J. Quetin-Leclercq, "Natural products published in 2009 from plants traditionally used to treat malaria," *Planta Medica*, vol. 77, no. 6, pp. 631–640, 2011.
- [5] C. R. Nogueira and L. M. X. Lopes, "Antiplasmodial natural products," *Molecules*, vol. 16, no. 3, pp. 2146–2190, 2011.
- [6] E. H. Hakim, S. A. Achmad, L. D. Juliawaty et al., "Prenylated flavonoids and related compounds of the Indonesian *Artocarpus* (Moraceae)," *Journal of Natural Medicines*, vol. 60, no. 3, pp. 161–184, 2006.
- [7] A. Widyawaruyanti, Subehan, S. K. Kalaumi et al., "New prenylated flavones from *Artocarpus champeden*, and their antimalarial activity in vitro," *Journal of Natural Medicines*, vol. 61, no. 4, pp. 410–413, 2007.
- [8] H. Morita, N. C. Zaini, T. S. Wahyuni, W. Ekasari, A. Widyawaruyanti, and Y. Hirasawa, "Artopeden A, a new antiplasmodial isoprenylated flavone from *Artocarpus champeden*," *Heterocycles*, vol. 79, no. 1, pp. 1121–1126, 2009.
- [9] A. F. Hafid, N. P. Ariantari, L. Tumewu et al., "The active marker compound identification of *Artocarpus champeden* spreng stem bark extract, morachacone A as antimalarial," *International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 4, no. 5, pp. 246–249, 2012.
- [10] E. H. Hakim, L. D. Juliawaty, Y. M. Syah, and S. A. Achmad, "Molecular diversity of *Artocarpus champeden* (Moraceae): a species endemic to Indonesia," *Molecular Diversity*, vol. 9, no. 1–3, pp. 149–158, 2005.
- [11] S.-Y. Lyu and W.-B. Park, "Production of cytokine and NO by RAW 264.7 macrophages and PBMC in vitro incubation with flavonoids," *Archives of Pharmacological Research*, vol. 28, no. 5, pp. 573–581, 2005.
- [12] M. Comalada, I. Ballester, E. Bailón et al., "Inhibition of pro-inflammatory markers in primary bone marrow-derived mouse macrophages by naturally occurring flavonoids: analysis of the structure-activity relationship," *Biochemical Pharmacology*, vol. 72, no. 8, pp. 1010–1021, 2006.
- [13] D. Messaoudeno, H. Belguendouz, M. L. Ahmedi et al., "Ex vivo effect of flavonoids extracted from *Artemisia herba alba* on cytokines and nitric oxide production in Algerian patients with Admantiades-Behcets's disease," *Journal of Inflammation*, vol. 8, no. 1, p. 35, 2011.
- [14] S.-L. Chang, Y.-M. Chiang, C. L.-T. Chang et al., "Flavonoids, centaurein and centaureidin, from *Bidens pilosa*, stimulate IFN- γ expression," *Journal of Ethnopharmacology*, vol. 112, no. 2, pp. 232–236, 2007.
- [15] R. Gomez-Flores, C. L. Calderon, L. W. Scheibel et al., "Immunoenhancing properties of *Plantago major* leaf extract," *Phytotherapy Research*, vol. 14, no. 8, pp. 617–622, 2000.
- [16] M. English and C. R. I. C. Newton, "Malaria: pathogenicity and disease," *Chemical Immunology and Allergy*, vol. 80, pp. 50–69, 2002.
- [17] I. A. Clark, W. B. Cowden, G. A. Butcher, and N. H. Hunt, "Possible roles for tumor necrosis factor in the pathology of

- malaria." *The American Journal of Pathology*, vol. 127, no. 1, pp. 192-198, 1987.
- [18] I. Angulo and M. Fresno, "Cytokines in the pathogenesis of and protection against malaria," *Clinical and Vaccine Immunology*, vol. 9, no. 6, pp. 1145-1152, 2002.
- [19] W. Peters, J. H. Portus, and B. L. Robinson, "The chemotherapy of rodent malaria, XXII," *Annals of Tropical Medicine & Parasitology*, vol. 69, no. 2, pp. 155-171, 1975.
- [20] U. B. Jagtap and V. A. Bapat, "Artocarpus: a review of its traditional uses, phytochemistry and pharmacology," *Journal of Ethnopharmacology*, vol. 129, no. 2, pp. 142-166, 2010.
- [21] C. Boonlaksiri, W. Oonnanant, P. Kongsacree, P. Kittakoop, M. Tanticharoen, and Y. Thebtaranonth, "An antimalarial stilbene from *Artocarpus integer*," *Phytochemistry*, vol. 54, no. 4, pp. 415-417, 2000.
- [22] A. F. Hafid, R. P. Septiani, L. H. Fabriana, N. Febrianty, D. Ranggaditya, and A. Widyawaruyanti, "Antimalarial activity of crude extracts of *Artocarpus heterophyllus*, *Artocarpus altilis* and *Artocarpus camansi*," *Asian Journal of Pharmaceutical and Clinical Research*, vol. 9, no. 1, pp. 279-281, 2016.
- [23] L. Bantie, S. Assefa, T. Teklehaimanot, and E. Engidawork, "In vivo antimalarial activity of the crude leaf extract and solvent fractions of *Croton macrostachyus* Hoehst. (Euphorbiaceae) against *Plasmodium berghei* in mice," *BMC Complementary and Alternative Medicine*, vol. 14, no. 1, p. 79, 2014.
- [24] I. A. Clark, J. L. Virelizier, E. A. Carawell, and P. R. Wood, "Possible importance of macrophage-derived mediators in acute malaria," *Infect Immunology*, vol. 981, no. 32, pp. 1058-1066, 1981.
- [25] J. B. Souza, K. H. Williamson, T. Otani, and J. H. Playfair, "Early gamma interferon responses in lethal and nonlethal murine blood-stage malaria," *Infect Immunology*, vol. 65, no. 5, pp. 1593-1599, 1997.
- [26] A. L. Richard, "Tumour necrosis factor and associated cytokines in the host's response to malaria," *International Journal for Parasitology*, vol. 27, no. 10, pp. 1251-1263, 1997.
- [27] G. I. Bancroft and J. P. Kelly, "Macrophage activation and innate resistance to infection in SCID mice," *Immunology*, vol. 191, no. 4-5, pp. 424-431, 1993.
- [28] L. Malaguarrera and S. Musumeci, "The immune response to *Plasmodium falciparum* malaria," *The Lancet Infectious Disease*, vol. 2, no. 8, pp. 472-478, 2002.
- [29] M. Niikura, S.-I. Inoue, and F. Kobayashi, "Role of Interleukin-10 in malaria: focusing on coinfection with lethal and nonlethal murine malaria parasites," *Journal of Biomedicine and Biotechnology*, vol. 2011, Article ID 383962, 8 pages, 2011.

Effect of Formulated Artocarpus champeden Extract on Parasite Growth and Immune Response of Plasmodium berghei-Infected Mice

ORIGINALITY REPORT

13%

SIMILARITY INDEX

5%

INTERNET SOURCES

12%

PUBLICATIONS

0%

STUDENT PAPERS

PRIMARY SOURCES

- 1 Rajesh Kumar. "Evaluation of localized and systemic immune responses in cutaneous leishmaniasis caused by Leishmania tropica: interleukin-8, monocyte chemotactic protein-1 and nitric oxide are major regulatory factors : Immune profile of CL patients", Immunology, 01/22/2010
Publication 1%
- 2 T. K. Lim. "Edible Medicinal And Non Medicinal Plants", Springer Science and Business Media LLC, 2012
Publication 1%
- 3 Kenny Ang, Michael Holmes, Ursula Kara. "Immune-mediated parasite clearance in mice infected with Plasmodium berghei following treatment with manzamine A", Parasitology Research, 2001
Publication 1%

4

Internet Source

1%

5

Fakhereldin M. Omer, Eleanor M. Riley.
"Transforming Growth Factor β Production Is
Inversely Correlated with Severity of Murine
Malaria Infection", The Journal of Experimental
Medicine, 1998

Publication

<1%

6

Nagib, P.R.A.. "Trypanosoma cruzi: Populations
bearing opposite virulence induce differential
expansion of circulating $CD3^+CD4^-CD8^-$ T
cells and cytokine serum levels in young and
adult rats", Experimental Parasitology, 200708

Publication

<1%

7

Waranya Lakornwong, Kwanjai Kanokmedhakul,
Somdej Kanokmedhakul, Palangpon
Kongsaeree et al. " Triterpene Lactones from
Cultures of sp. KM01 ", Journal of Natural
Products, 2014

Publication

<1%

8

Huseyin Fidan. "Influence of Different Atropine
Therapy Strategies on Fenthion-Induced Organ
Dysfunction in Rats", Basic & Clinical
Pharmacology & Toxicology, 5/2007

Publication

<1%

9

C. Chiyaka, W. Garira, S. Dube. "Modelling
Immune Response and Drug Therapy in Human

<1%

Malaria Infection", Computational and
Mathematical Methods in Medicine, 2008

Publication

10

Fakhreldin M. Omer, J. Brian de Souza, Eleanor M. Riley. " Differential Induction of TGF- β Regulates Proinflammatory Cytokine Production and Determines the Outcome of Lethal and Nonlethal Infections ", The Journal of Immunology, 2003

Publication

<1%

11

Laychiluh Bantie Mekonnen. "In vivo antimalarial activity of the crude root and fruit extracts of *Croton macrostachyus* (Euphorbiaceae) against *Plasmodium berghei* in mice", Journal of Traditional and Complementary Medicine, 2015

Publication

<1%

12

Matalaka, K.Z.. "Prolactin enhances production of interferon- γ , interleukin-12, and interleukin-10, but not of tumor necrosis factor- α , in a stimulus-specific manner", Cytokine, 20030221

Publication

<1%

13

Wills, P.J.. "Protective effect of *Lygodium flexuosum* (L.) Sw. extract against carbon tetrachloride-induced acute liver injury in rats", Journal of Ethnopharmacology, 20061206

Publication

<1%

14

bmcmusculoskeletdisord.biomedcentral.com

Internet Source

<1 %

15

malariajournal.biomedcentral.com

Internet Source

<1 %

16

www.thieme-connect.de

Internet Source

<1 %

17

Mônica M. de Almeida Lopes, Kellina O. de Souza, Ebenezer de Oliveira Silva. "Cempedak — *Artocarpus champeden*", Elsevier BV, 2018

Publication

<1 %

18

Navneet Kishore, Pradeep Kumar, Karuna Shanker, Akhilesh Kumar Verma. "Human disorders associated with inflammation and the evolving role of natural products to overcome", *European Journal of Medicinal Chemistry*, 2019

Publication

<1 %

19

Kou, Wei. "Central catecholamine depletion inhibits peripheral lymphocyte proliferation and splenocyte cytokine production", DuEPublico: University of Duisburg-Essen Publications Online, 2004.

Publication

<1 %

20

socalip.org

Internet Source

<1 %

21

www.jimmunol.org

Internet Source

<1 %

22 academic.oup.com
Internet Source <1%

23 T Fujioka. "Effect of methylprednisolone on the kinetics of cytokines and liver function of regenerating liver in rats", *Hepatology Research*, 2001
Publication <1%

24 www.brazjurol.com.br
Internet Source <1%

25 Petteri Nieminen, Pirjo Lindström-Seppä, Minna Juntunen, Juha Asikainen et al. "IN VIVO EFFECTS OF BISPHENOL A ON THE POLECAT (MUSTELA PUTORIUS)", *Journal of Toxicology and Environmental Health, Part A*, 2002
Publication <1%

26 www.tandfonline.com
Internet Source <1%

27 Mary Gawthrop. "Guidelines for malaria prevention in travellers: what is new?", *Practice Nursing*, 2020
Publication <1%

28 iai.asm.org
Internet Source <1%

29 scielo.conicyt.cl
Internet Source <1%

30 Hort, G.. "Delayed type hypersensitivity-associated disruption of splenic periarteriolar lymphatic sheaths coincides with temporary loss of IFN- γ production and impaired eradication of bacteria in *Brucella abortus*-infected mice", *Microbes and Infection*, 200302

Publication

<1%

31 A. Ishih, T. Nagata, F. Kobayashi, T. Miyase, M. Terada. "Cytokine and antibody production during the course of resolution in *Plasmodium yoelii* 17XL-infected BALB/c mice treated with febrifugine and isofebrifugine mixture from leaves of *Hydrangea macrophylla* var. Otaksa", *Parasitology Research*, 2004

Publication

<1%

32 www.iosrjournals.org

Internet Source

<1%

33 Mohamed Balaha, Samah Kandeel, Ahmed Kabel. "Phloretin either alone or in combination with duloxetine alleviates the STZ-induced diabetic neuropathy in rats", *Biomedicine & Pharmacotherapy*, 2018

Publication

<1%

34 pt.scribd.com

Internet Source

<1%

35 Shefalee K. Bhavsar, Paulomi Joshi, Mamta B.

Shah, D.D. Santani. " Investigation into Hepatoprotective Activity of . ", Pharmaceutical Biology, 2008

Publication

<1%

36

Wang, G.J.. "Flavonoids with iNOS inhibitory activity from Pogonatherum crinitum", Journal of Ethnopharmacology, 20080619

Publication

<1%

37

V. Muñoz, M. Sauvain, G. Bourdy, J. Callapa, S. Bergeron, I. Rojas, J.A. Bravo, L. Balderrama, B. Ortiz, A. Gimenez, E. Deharo. "A search for natural bioactive compounds in Bolivia through a multidisciplinary approach", Journal of Ethnopharmacology, 2000

Publication

<1%

38

Fumiaki Tanaka, Hiroshi Yamaguchi, Mitsuhiko Ohta, Kohjiro Mashino, Hideto Sonoda, Noriaki Sadanaga, Hiroshi Inoue, Masaki Mori. "Intratumoral injection of dendritic cells after treatment of anticancer drugs induces tumor-specific antitumor effect in vivo", International Journal of Cancer, 2002

Publication

<1%

39

Biology of C Reactive Protein in Health and Disease, 2016.

Publication

<1%

40

D. Klayman. "Qinghaosu (artemisinin): an

antimalarial drug from China", Science, 1985

Publication

<1%

41

www.iss.it

Internet Source

<1%

42

"Abstracts", Fundamental and Clinical
Pharmacology, 04/2011

Publication

<1%

43

"Progress in Immunology", Springer Science
and Business Media LLC, 1989

Publication

<1%

Exclude quotes Off

Exclude matches Off

Exclude bibliography On

Effect of Formulated Artocarpus champeden Extract on Parasite Growth and Immune Response of Plasmodium berghei-Infected Mice

GRADEMARK REPORT

FINAL GRADE

/0

GENERAL COMMENTS

Instructor

PAGE 1

PAGE 2

PAGE 3

PAGE 4

PAGE 5

PAGE 6

PAGE 7