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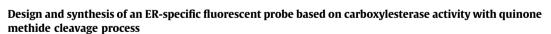
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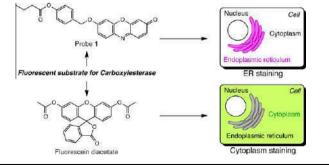
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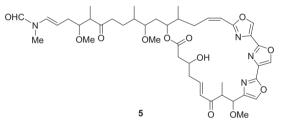
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Wataru Hakamata*, Aki Machida, Tadatake Oku, Toshiyuki Nishio





Compd

3i

3j

3k

3n

R

3-CI

3-Br

3-1

3-Me

EC50 (µM)

14.0

3.6

7.3

2.0

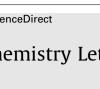
CC50 (µM)

>100

>100

>100

>100





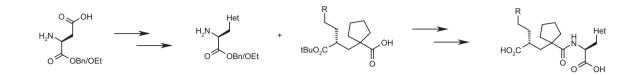
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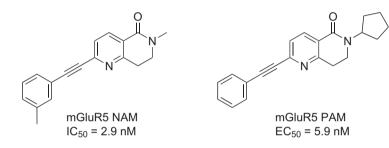
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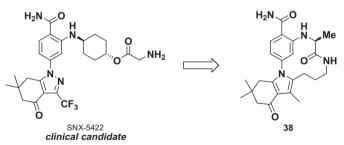
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Macrocyclic lactams as potent Hsp90 inhibitors with excellent tumor exposure and extended biomarker activity

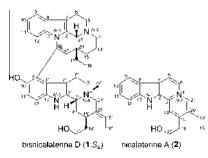
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New antiplasmodial indole alkaloids from Hunteria zeylanica

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Two new indole alkaloids, bisnicalaterine D (1), consisting of an eburnane and a corynanthe type of skeletons, and nicalaterine A (2) were isolated from the bark of *Hunteria zeylanica*. Their structures were elucidated by various spectroscopic data such as NMR and CD spectra. A series of bisnicalaterines and nicalaterine A showed potent antiplasmodial activity against *Plasmodium falciparum* 3D7.



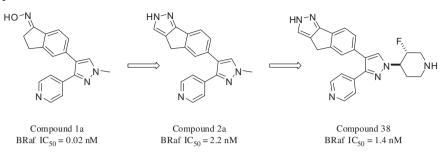
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Non-oxime pyrazole based inhibitors of B-Raf kinase

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*Corresponding author

O⁺ Supplementary data available via ScienceDirect

COVER

Botulinum neurotoxins are the most deadly toxins known to man, approximately 10 million times more deadly than cyanide. Botulinum neurotoxins are classified by the US Centers for Disease Control (CDC) as bioterrorism agents. The etiological agent responsible for botulinum intoxication is a metalloprotease; as such this is a key therapeutic target. Currently, there are no approved pharmacological treatments for botulinum intoxication. Discovering molecules that could be used as a path forward for therapeutic development as botulinum protease inhibitors is tantamount. A benzylidene cyclopentenedione-based inhibitor was found to be the first affinity reagent to covalently modify the active site of botulinum neurotoxin A light chain metalloprotease. Its kinetic parameters are reported and such an approach for inhibition of this deadly neurotoxin. [Capková, K.; Hixon, M. S.; Pellett, S.; Barbieri, J. T.; Johnson, E. A.; Janda, K. D. *Bioorg, Med. Chem. Lett.* **2010**, *20*, 206.]

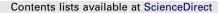
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New antiplasmodial indole alkaloids from Hunteria zeylanica

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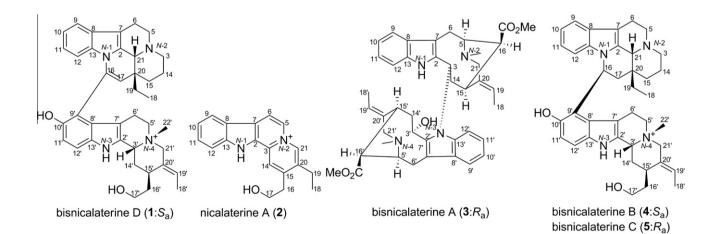
ABSTRACT

Two new indole alkaloids, bisnicalaterine D (1), consisting of an eburnane and a corynanthe type of skeletons, and nicalaterine A ($\mathbf{2}$) were isolated from the bark of *Hunteria zeylanica*. Their structures were elucidated by various spectroscopic data such as NMR and CD spectra. A series of bisnicalaterines and nicalaterine A showed potent antiplasmodial activity against *Plasmodium falciparum* 3D7.

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In our search for new bioactive alkaloids from tropical plants in Malaysia and Indonesia, we have previously reported a series of cassiarins with potent antiplasmodial activity.^{1,2} *Hunteria zeylanica* (Retz.) Gardner ex Thwaites is a member of the Apocynaceae family in Malaysia, found mostly in Pahang and Selangor,³ and the bark and leaves have been known to produce various skeletal alkaloids depending on the area where the plants were distributed.^{4–9} In our

previous paper,^{10,11} we have reported the isolation of new bisindole alkaloids, bisnicalaterines A–C (**3–5**) from *H. zeylanica*. In this Letter, we report the isolation and structure elucidation of bisnicalaterine D (**1**), a new bisindole alkaloid consisting of an eburnane and a corynanthe type of skeletons, and nicalaterine A (**2**) as well as the antimalarial activity of **1–5** against *Plasmodium falciparum* 3D7.



* Corresponding author. E-mail address: moritah@hoshi.ac.jp (H. Morita).

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter © 2011 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2011.03.104

The bark of *H. zeylanica*¹² was extracted with MeOH, and part (33 g) of the extract was treated with 3% tartaric acid (pH 2) and then partitioned with EtOAc. The aqueous layer was treated with saturated Na₂CO₃ (aq) to pH 10 and extracted with CHCl₃ and *n*-BuOH subsequently. The *n*-BuOH fraction was subjected to an HP-20 column (H₂O/MeOH 0:1 to 1:0), and the 80% MeOH fraction (3.5 g) was further separated by using a Sephadex LH-20 column. Fractions containing **1** was then separated by a silica gel column (CHCl₃/MeOH, 9:1 to 0:1), followed by an ODS Sep-Pak (MeOH/H₂O 1:9 to 1:0) to give bisnicalaterine D (**1**, 8.1 mg, 0.003%), while purification of fractions containing **2** by an amino silica gel column (CHCl₃/MeOH, 9:1 to 0:1) and a silica gel column (CHCl₃/MeOH, 8:2) yielded nicalaterine A (**2**, 2.0 mg, 0.0008%).

Bisnicalaterine D (1),¹³ a yellowish amorphous solid, $[\alpha]_D^{20}-72$ (*c* 1.0, MeOH), showed a molecular formula, $C_{39}H_{47}N_4O_2$, which was determined by HRESIMS [*m*/*z* 603.3689 (M)⁺, Δ -0.5 mmu]. IR absorption band (3430 cm⁻¹) was characteristic of amino or

Table 1

 1 H and 13 C NMR data of bisnicalaterine D (1) and nicalaterine A (2) in CD₃OD at 300 K^a

$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	Position	1		2		
3a $3.37 (1H, m)$ 46.7 132.4 3b $3.37 (1H, m)$ 53.1 $8.69 (1H, br s)$ 127.3 5a $3.87 (1H, m)$ 16.9 $8.44 (1H, br s)$ 117.0 6a $3.13 (1H, m)$ 16.9 $8.44 (1H, br s)$ 117.0 7 108.2 123.0 8 128.7 122.3 9 7.48 (1H, d, 7.6) 119.7 $8.21 (1H, d, 7.6)$ 120.6 10 $6.97 (1H, m)$ 121.7 $7.39 (1H, t, 7.6)$ 130.4 11 $6.69 (1H, t, 8.3)$ 113.4 $7.70 (1H, br s)$ 113.7 13 136.2 $7.64 (1H, t, 7.6)$ 130.4 14a $1.83 (1H, m)$ 19.4 $8.58 (1H, s)$ 112.1 14b $1.98 (1H, m)$ 19.4 $8.58 (1H, s)$ 151.1 15b $1.86 (1H, m)$ 29.1 151.1 151.1 15b $1.86 (1H, m)$ 29.1 151.1 151.1 15b $1.86 (1H, m)$ 29.1 150.1 36.2 17 $5.26 (1H, s)$ 11		[δ _H (J, Hz)]	$[\delta_c]$	[δ _H (J, Hz)]	$[\delta_c]$	
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$\begin{array}{cccccccccccccccccccccccccccccccccccc$	6b	3.30 (1H, m)				
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hydroxyl group. ¹H and ¹³C NMR data (Table 1) suggested the presence of one sp³ quaternary carbon, 12 sp³ methylenes, three sp³ methines, three methyls, eight sp² methines, and 12 sp² quaternary carbons. The ¹³C NMR spectrum of **1** is very similar to those of bisnicalaterines B (**4**).¹¹ Compared to spectroscopic data of **4**, **1** has two less sp³ carbons, two additional sp² carbons (δ_c 131.9 and δ_c 117.7) and also 2 amu smaller which suggests the existence of an additional double bond in **1**. The existence of a double bond between C-16 and C-17 was confirmed by the HMBC correlations of H-17 to C-16, C-19, C-20, C-21 and C-9', H₃-18 to C-20, and H-21 to C-15. Further analysis of the two-dimensional NMR data (¹H–¹H COSY, HMQC, and HMBC spectra) revealed the gross structure of **1** as shown in Figure 1.

The stereochemistry of each monoterpeneindole unit in **1** was assigned by NOESY correlations as shown in computer-generated 3D drawing (Fig. 2). In unit A, the NOESY correlations of H₂-19/H-21 suggested that H-21 and an ethyl group (C-18–C-19) were β -oriented. While in unit B, the NOESY correlations of H-3'/H₃-22', H-21'b, and H₂-16', H₃-22'/H-21'b and H-21'b/H-16'b suggested that H-3', C-16' and C-22' were β -oriented, while the correlations of H-19'/H-21'a and H-15'/H₃-18' established the *E*-configuration of the ethylidene side chain. Thus the relative stereochemistry of units A and B was assigned as shown in Figure 2.

In the case of bisnicalaterines B (**4**) and C (**5**), there were two possible conformations around C-16–C-9' bond, the twisted and extended conformations.¹¹ In bisnicalaterine D (**1**), the NOESY correlation of H-6'b/H-21 and the highly shifted chemical shift ($\delta_{\rm H}$ 1.53 and 2.50, respectively) of H-6'a and H₃-22' at ammonium nitrogen atom suggested that **1** possessed the twisted conformation observed in bisnicalaterine B (**4**).¹¹

The absolute structure of **1** was deduced by comparing its CD spectrum to that of bisnicalaterine B (**4**). The CD spectrum¹³ of **1** showed a similar CD pattern to that of **4**,¹¹ thus the absolute structure of **1** was determined to be of 20*R*, 21*R*, 4'*R*, 3'*R*, 15'*R*.

Nicalaterine A (**2**),¹⁴ a yellowish amorphous solid, showed a molecular formula, $C_{19}H_{19}N_2O$, which was determined by HRESIMS $[m/z \ 291.1496 \ (M)^+, \Delta -0.1 \ mmu]$. IR absorption band (3430 cm⁻¹) was characteristic of amino or hydroxyl group. The UV spectrum suggested the presence of a highly conjugated ring system [λ_{max} 386 (ε 8000), 346 (8300), 291 (6900), 240 (16,100), and 235 (sh, 15,000)] as in flavopereirine.¹⁵ ¹H and ¹³C NMR data (Table 1) suggested the presence of three sp³ methylenes, one methyl, eight sp²

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Figure 1. Selected 2D NMR correlations for bisnicalaterine D (1).

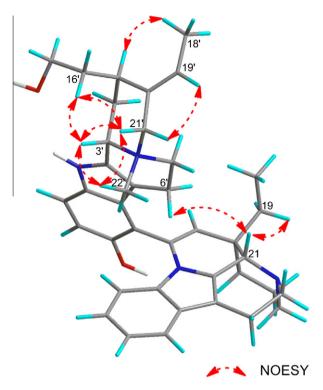


Figure 2. Selected NOESY correlations for bisnicalaterine D (1).

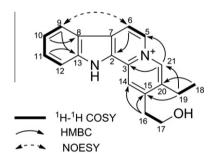


Figure 3. Selected 2D NMR correlations for nicalaterine A (2).

Table 2				
Antiplasmodial activity	of 1-5	against P.	falciparum	3D7

Antiplasmodial activity (IC 50, μ M)	Cytotoxic activity ^a (IC ₅₀ , μ M)	SI
1 >50	>50	_
2 0.11	>50	>450
3 4.36	16.2	3.7
4 1.13	>50	>44
5 0.05	>50	>1000

^a Against HL-60.

methines, and seven sp² quaternary carbons. The HMBC correlations of H₂-19 to C-15 and C-21, H₃-18 to C-20, and H₂-16 to C-15 and C-14 allowed the attachment of the ethyl side chain (C-18 and C-19) to C-20 and the 2-hydroxyethyl side chain to C-15. Further analysis of the two-dimensional NMR data (¹H-¹H COSY, HSQC, and HMBC spectra in CD₃OD) revealed the gross structure of **2** as shown in Figure 3.

Antimalarial activity¹⁶⁻¹⁸ for **1–5** against *P. falciparum* 3D7 was evaluated (Table 2). Nicalaterine A (**2**) and bisnicalaterine C (**5**)

showed potent antimalarial activity (IC_{50} 0.11 and 0.05 µM, respectively) with a good selectivity (SI > 450 and >1000, respectively). Bisnicalaterine C (**5**) with an extended conformation showed 20 times more effective than that of bisnicalaterine B (**4**) with a twisted conformation. On the other hand, bisnicalaterine D (**1**), which also possessed a twisted conformation, showed practically no antimalarial activity. It is interesting to note that the conformation around the C-16 - C-9' bond may play important roles to show antimalarial activity.

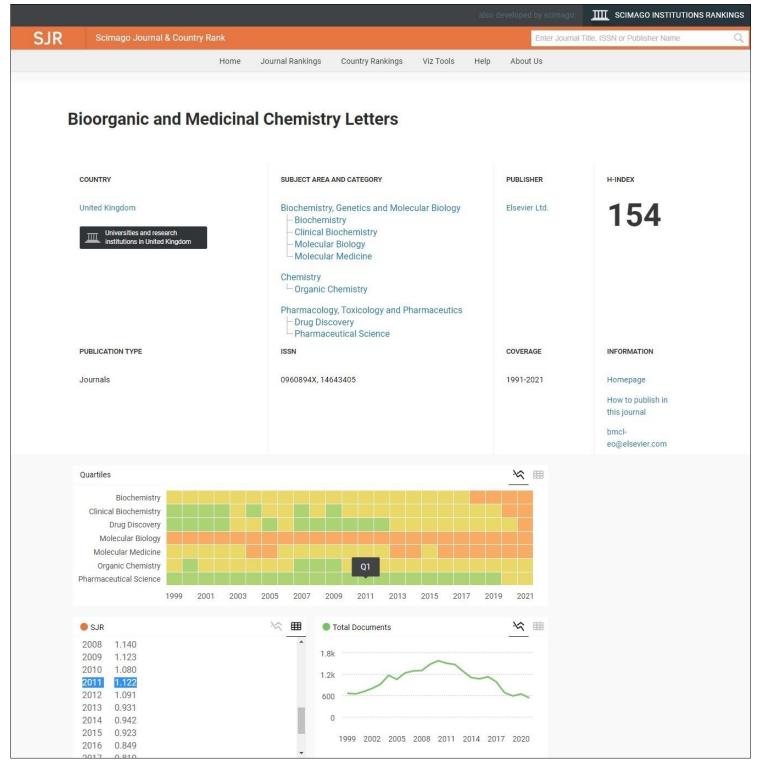
Acknowledgments

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References and notes

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- 12. Bark of *H. zeylanica* were collected in Kampung Padang, Malaysia, in 1994. The botanical identification was made by Mr. Teo Leong Eng, Faculty of Science, University of Malaya. A voucher specimen (Herbarium No. KL 4345) is deposited at the Herbarium of the Department of Chemistry, University of Malaya, Kuala Lumpur, Malaysia.
- 13. Bisnicalaterine D, yellowish amorphous solid, $[\alpha]_D^{20} 72$ (*c* 1.0, MeOH), UV (MeOH) λ_{max} 217 (ε 16800), 249 (6100), 277.5 (5300), 302 (4100); CD (MeOH) λ_{max} 205 ($\Delta \varepsilon$ 5.95), 218 (-6.09), 243 (6.54), 280 (-4.22), 365 (3.07); IR (KBr) v_{max} 3430 cm⁻¹; ¹H and ¹³C NMR data see Table 1; EI-MS *m*/*z* 603 M⁺; HRESIMS [*m*/*z* 603.3689 (M)⁺, calcd for C₃₉H₄₇N₄O₂, 603.3694].
- Nicalaterine A (2), a yellowish amorphous solid, UV (MeOH) λ_{max}, 235 (sh, 15,000), 240 (16,100), 291 (6900), 346 (8300), and 386 (ε 8000); IR (KBr) ν_{max} 3430 cm⁻¹, ¹H and ¹³C NMR data see Table 1; EI-MS *m/z* 291 M⁺; HRESIMS [*m/z* 291.1496 (M)⁺, calcd for C₁₉H₁₉N₂O, 291.1497].
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- 16. Human malaria parasites were cultured according to the method of Trager and Jensen.¹⁷ The antimalarial activity of the isolated compounds was determined by the procedure described by Budimulja et al.¹⁸ In brief, stock solutions of the samples were prepared in DMSO (final DMSO concentrations of <0.5%) and were diluted to the required concentration with complete medium (RPMI 1640 supplemented with 10% human plasma, 25 mM HEPES and 25 mM NaHCO₃) until the final concentrations of samples in culture plate wells were 10; 1; 0,1; 0,001 µ/mL. The malarial parasite *P. falciparum* 3D7 clone was propagated in a 24-well culture plates. Growth of the parasite was monitored by making a blood smear fixed with MeOH and stained with Geimsa stain. The antimalarial activity of each compound was expressed as an IC₅₀ value, defined as the concentration of the compound causing 50% inhibition of parasite growth relative to an untreated control. The percentage of growth inhibition % = 100–[(test parasitemia/control parasitemia) × 100]. Chloroqine: IC₅₀ 0.011 µM.
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Biochemistry, Genetics and Molecular Biology #59/115 Clinical Biochemistry	49th				
Biochemistry, Genetics and Molecular Biology #96/167 Molecular Medicine	42nd				
Biochemistry, Genetics and Molecular Biology #227/386 Molecular Biology	41st				