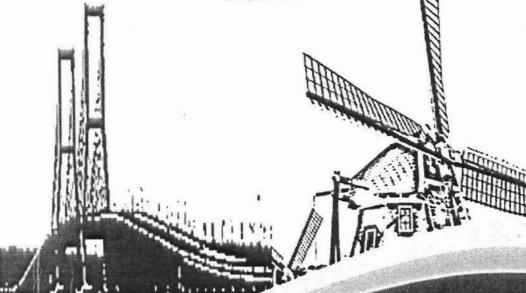


The Book of Pediatric Neuro-developmental

Analyze and overcome pediatric neuro-developmental problem in novel perspective view



Editor: Darto Saharso Prastiya Indra Gunawan

Department of Child Health Faculty of Medicine Airlangga University - Dr. Soetomo Hospital, Surabaya-Indonesia in Collaboration with Dutch Foundation for Postgraduate Medical Courses in Indonesia

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Stem Cell Research In Cerebral Palsy Option of Treatment in Future?

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Childhood cerebral palsy is a non-progressive brain disease that results from various cerebral insults that can occur before birth and 1 month after birth. Patient primarily present with motor developmental delay or motor dysfunction and possible mental retardation, epilepsy, behavioural disorders and sensory and perceptual disturbances. Conventional therapies for treating cerebral palsy include physical therapy, motor function training, language training, surgery and intramuscular injections of botulinum toxin. However these methods hav not improved cerebral injuries in patients with moderate-to-severe cerebral palsy (An, 2006).

Stem cell transplantation is a novel and promising treatment for cerebral palsy. However, this procedure is still in the initial stages of investigation (Caroll, 2011). Several preclinical experiments on animal models of cerebral palsy have been carried out to demonstrate the potential of cell transplantation to minimize damage and promote recovery. However, limited clinical trials have been initiated to study the effect cell therapy in humans.

Human umbilical cord blood cells (hUCBCs) have been explored to a great extent in cerebral palsy. hUCBCs have been administered in rat models of neonatal hypoxia/ ischemia. I alleviate spastic paresis in neonatal rat models resulting in norma walking (Meier, 2006). It is also observed that these cells exhibit neuroprotective effect in the striatum, and decrease the number of activated microglial cells in the cerebral cortex of treated animals further resulting in better functional recovery (Pimentel-Coelho 2010). They protect the mature neurons in the neocortex from injury, bring about near-normalization of brain damage in th subventricular zone (SVZ) leading to significant improvement in behavioral functions. The long lasting effect of these cells is du to the paracrine effects of hUCBCs which stimulate recovery in the injured brain and protect against further brain damage (Bas 2012). On transplantation, hUCBCs have shown to ameliorat neurological and motor deficits in CP model by reducing th levels of pro-inflammatory cytokines (Interleukin-1a (IL-1a Interleukin-1 β (IL-1 β), and Tumor necrosis factor α (TNFc (Rosenkranz, 2013). Human umbilical cord blood (hUCB) cel have also shown to reduce sensorimotor deficits after hypox ischemic brain injury in neonatal rats. The dimensions of cortic maps and receptive fields, which are significantly altered after injury, are largely restored. Additionally, the lesion induce hyperexcitability is no longer observed in treated animal compared to control animals. The results demonstrate that hUC cells reinstall the way central neurons process information by normalizing inhibitory and excitatory processes (Geissler, 2013).

Various preclinical studies have shown that transplantation of stem cells in the CP models lead to survival, homing and differentiation of these cells into neurons, oligodendrocytes, astrocytes etc.

A Chinese study, wherein neural stem cells derived from human fetal brain (hNSCs) were transplanted into cerebral ventricle of HI injury neonatal rat, too demonstrated the survival, migration and differentiation capacity of these cells in rat brain (Qu, 2005). Similarly, Zheng et al showed that Multipotent astrocytic stem cells (MASCs) from mice transplanted into a rat model of hypoxia-ischemia (HI) survive, migrate and differentiate into neurons and astrocytes (Zheng, 2006). Park et al reported clonal neural stem cells (NSCs) when transplanted into brains of postnatal hypoxic-ischemic (HI) injury mice, home preferentially to and integrate extensively within the ischemic areas. They differentiate into neurons and oligodendrocytes, the cell types damaged due to HI (Park, 2006). Yasuhara et al, investigated the efficacy of intrahippocampal transplantation of bone marrow derived multipotent progenitor cells (MPCs) in HI injury. They found that transplanted MPCs ameliorated motor deficits associated with HI injury (Yasuhara, 2006). Webber et al in their study highlighted the protective effects of oligodendrocyte precursor cell transplantation in neonatal inflammation-induced rat model of periventricular leukomalacia (Webber, 2009). Cher et al, transplanted magnetically labeled mesenchymal stem cells in a model of perinatal brain injury. They found that these cell migrate to lesion sites and proliferate. They are neuroprotective and indirectly contribute to brain repair (Chen, 2010). Study b Titomanlio et al, implanted neurosphere-derived precursors in neonatal mouse model of cerebral palsy induced by excitotoxicity They observed that cells migrated to the lesion site, remained undifferentiated at day 10, and differentiated into oligodendrocyt and neurons at day 42. Although grafted cells finally die there few weeks later, this procedure triggered a reduction in lesion size and an improvement in memory performance compared with untreated animals (Titomanlio, 2011). All the above preclinica studies have been carried out in animal models of acute hypoxic injury, hence showing significant results. But, similar results an difficult to replicate in human cases since the intervention always cannot be carried out immediately post injury. Thus, more studie should be carried out in chronic injury models. Based on this observation, it can also be concluded that earlier the intervention better is the outcome.

Below are few of the published studies carried out in human cases of cerebral palsy.

Papadopoulos et al, administered autologous umbilical com blood cells in 2 children diagnosed with spastic diplegic CP. The found that this therapy was safe, feasible and led to functiona improvements in children which was seen by the change in GMFCS (Papadopoulos, 2011).

Luan et al, carried out a study on 45 patients diagnosed with severe CP. They underwent transplantation of neural progenitor cell (NPC) derived from aborted fetal tissue. After 1 year, the developmental level in gross motor, fine motor, and cognition of the treatment group was significantly higher compared to the control group. These results suggested that NPC transplantation is a safe and effective therapeutic method for treating children with severe CP (Luan, 2012).

Li et al, transplanted autologous bone marrow mesenchymal cells in an 11 year old CP case with visual impairment. On six month follow up, he could walk better and his vision had improved significantly. These findings were supported by the electrophysiological examinations (Li, 2012).

Purandare et al, reported a case of cerebral palsy who was administered with autologous bone marrow mononuclear cells. On follow up, they recorded a significant improvement in motor, sensory, cognitive, and speech. Bowel and bladder control was also achieved. On the GMFCSE& R level, the patient was promoted from grade III to I. Hence, concluding that intrathecal infusion of autologous BMMNCs is feasible, effective, and safe in CP patients. (Purandare, 2012). Chen et al, injected neural stem cell-like (NSC-like) cell derived from autologous marrow mesenchymal stem cells in 30 cases of cerebral palsy. The control group only received rehabilitation treatment. On follow up, they observed an increasin the GMFM scores and language quotients compared to the control group. No adverse events were recorded indicating tha NSC-like cells are safe and effective for the treatment of moto deficits related to cerebral palsy (Chen, 2013).

Minet al, carried out a double blind, randomized, controlled trial in which the researcger administered allogeneic umbilical conblood cells potentiated with recombinant human erythropoietin (rhEPO) in CP patients. Compared with the EPO and control the pUCB group had signifanctly higher scores on the GMPM and BSID II Mental and motor scales at 6 months. They observed improvementin motor and cognitive dysfunction in children with CP, accompanied by structural andmetabolic changes in the brain. (Min, 2013).

Wang et al, reported a case of a 5-year old girl with CI who underwent umbilical cord mesenchymal stem cells (MSCs transplantation. She was treated with multiple times of intravenour and intrathecal administration of MSCs derived from her youn sister and was followed up for 28 months. The gross moto dysfunction was improved. Immunity was enhanced, physica strength improved along with speech and comprehension (Wang 2013). Jensen et al, recently published a study wherein a 2 ¹/₂ year old boy received autologousumbilical cord blood mononuclear cells intravenously. At 2-months follow-up the boy's motorcontrol improved, spastic paresis was largely reduced, and eyesight was recovered, as did the EEG. He smiled when played with, was able to sit and to speak simple words. At 40 months, independent eating, walking in gait trainer, crawling, and moving from prone position to freesitting were possible, and there was significantly improved receptive and expressive speech competence (fourword sentences, 200 words). This suggested that autologous cord bloodtransplantation could be a treatment alternative for cerebral palsy (Jensen, 2013).

Stem cell therapy for cerebral palsy still remains in its infant stage. Although cellular therapy for cerebral palsy has moved from the preclinical studies to bedside therapy; evidence remains inconclusive regarding multitude of variables. These variables are pertaining to cerebral palsyand cellular therapy. Pre-clinical models of effects of cellular therapy in cerebral palsy are farfrom the ideal state and show benefits only in acute injury. Majority of the human application of stem cells in cerebral palsy is for individuals who already have established pathology, hence at a chronic stage. Animal models of chronic injury are therefore required to study the efficacy and mechanism of action of stem cells. The individuals suffering from cerebral palsy are from various age groups, and present with varied kinds and severitie of clinical manifestations; there is only a limited evidence about which of these groups will benefit the most from cellular therapy

Stem cell therapy has been extensively studied but stineeds to be standardized before itbecomes a definitive treatmer modality. Autologous BMMNCs are safe and feasible option but heir effectiveness needs more clinical trials. Other types of ster cells need to establishsafety and efficacy. Though not a cure, ster cell therapy has emerged as a novel therapeuticoption to improve the quality of life.

References

- An T, Guo XQ, Pu XH. 2006. Study progress in ear intervention of high-risk infants for the prevention an treatment of cerebral palsy. Clin Pediatr, 24:696-8.
- Bae SH, Kong TH, Lee HS, Kim KS, Hong KS, Chopp M, Kan MS, Moon J. 2012. Long-lasting paracrine effects of huma cord blood cells on damaged neocortex in an animal model cerebral palsy. Cell Transplant, 21(11):2497-515
- 3. Caroll JE, Mays RW. 2011. Update on stem cell therapy for cerebral palsy. Expert Opin Biol Ther. 9:52.

- Chen A, Siow B, Blamire AM, Lako M, Clowry GJ. 2010. Transplantation of magnetically labeled mesenchymal stem cells in a model of perinatal brain injury. Stem Cell Res, 5(3):255-66.
- 5. Chen G, Wang Y, Xu Z, Fang F, Xu R, Wang Y, et al. 2013. Neural stem cell-likecells derived from autologous bone mesenchymal stem cells for the treatment ofpatients with cerebral palsy. J Transl Med, 11:21.
- Geissler M, Dinse HR, Neuhoff S, Kreikemeier K, Meier C.
 2011. Human umbilical cord blood cells restore brain damage induced changes in rat somatosensory cortex. PLoS One, 6(6):e20194.
- 7. Jensen A, Hamelmann E. 2013. First autologous cell therapy of cerebral palsy caused byhypoxic-ischemic brain damage in a child after cardiac arrest-individual treatmentwith cord blood. Case Rep Transplant, 951827.
- 8. Li M, Yu A, Zhang F, Dai G, Cheng H, Wang X, et al. 2012. Treatment of one case of cerebralpalsy combined with posterior visual pathway injury using autologous bonemarrow mesenchymal stem cells. J Transl Med, 10:100.
- 9. Luan Z, Liu W, Qu S, Du K, He S, Wang Z, et al. 2012. Effects of neuralprogenitor cell transplantation in children with severe cerebral palsy. Cell Transplant, 21 Suppl 1:S91-8.

- Meier C, Middelanis J, Wasielewski B, Neuhoff S, Roth Haerer A, Gantert M, Dinse HR, Dermietzel R, Jensen A 2006. Spastic paresis after perinatal brain damage in rat is reduced by human cord blood mononuclear cells. Pediat Res, 59(2):244-9.
- Min K, Song J, Kang JY, Ko J, Ryu JS, Kang MS, et al. 2013 Umbilical cord blood therapy potentiated with erythropoietic forchildren with cerebral palsy: a double-blind, randomized placebo-controlled trial.Stem Cells, 31(3):581-91.
- 12. Pimentel-Coelho PM, Magalhães ES, Lopes LM, deAzeved LC, Santiago MF, Mendez-Otero R. 2010. Human cord bloo transplantation in a neonatal rat model of hypoxicischemi brain damage: functional outcome related to neuroprotectio in the striatum. Stem Cells Dev, 19(3):351-8.
- 13. Papadopoulos KI, Low SS, Aw TC, Chantarojanasiri T. 201 Safety and feasibility of autologousumbilical cord bloo transfusion in 2 toddlers with cerebral palsy and the role low dose granulocyte-colony stimulating factor injections Restor Neurol Neurosci, 29(1): 17-22.
- 14. Park KI, Himes BT, Stieg PE, Tessler A, Fischer I, Snyde EY. 2006. Neural stem cells may be uniquely suited for combined gene therapy and cell replacement: Evidence from engraftment of Neurotrophin-3-expressing stem cells in hypoxic-ischemic brain injury. Exp Neurol, 199(1):179-90.

- Purandare C, Shitole DG, Belle V, Kedari A, Bora N, Joshi M. 2012. Therapeutic potentialof autologous stem cell transplantation for cerebral palsy. Case Rep Transplant, 2012:825289.
- 16. Qu SQ, Luan Z, Yin GC, Guo WL, Hu XH, Wu NH. 2005. Transplantation of human fetal neural stem cells into cerebral ventricle of the neonatal rat following hypoxic-ischemic injury: survival, migration and differentiation. Zhonghua Er Ke Za Zhi, 43(8):576-9.
- 17. Rosenkranz K, Tenbusch M, May C, Marcus K, Meier C. 2013. Changes in Interleukin-1 alpha serum levels after transplantation of umbilical cord blood cells in a model of perinatal hypoxic-ischemic brain damage. Ann Anat, 195(2):122-7.
- 18. Tanaka N, Kamei N, Nakamae T, Yamamoto R, Ishikawa M, Fujiwara H, et al. 2010. CD133+ cells from human umbilical cord blood reduce cortical damage and promote axonal growth in neonatal rat organ co-cultures exposed to hypoxia. Int J Dev Neurosci, 28(7):581-7.
- 19. Titomanlio L, Bouslama M, Le Verche V, Dalous J, Kaindl AM, Tsenkina Y, et al. 2011. Implanted neurosphere-derived precursors promote recovery after neonatal excitotoxic brain injury. Stem Cells Dev, 20(5):865-79.

- 20. Wang L, Ji H, Zhou J, Xie J, Zhong Z, et al. 2013. Therapeutic potential of umbilical cord mesenchymal stromal cellstransplantation for cerebral palsy: a case report. Case Rep Transplant, 2013:146347.
- 21. Wasielewski B, Jensen A, Roth-Härer A, Dermietzel R Meier C. 2012. Neuroglial activation and Cx43 expression are reduced upon transplantation of human umbilical cord blood cells after perinatal hypoxic-ischemic injury. Brain Res. 1487:39-53.
- 22. Webber DJ, van Blitterswijk M, Chandran S. 2009 Neuroprotective effect of oligodendrocyte precursor cell transplantation in a long-term model of periventricular leukomalacia. Am J Pathol, 175(6):2332-42
- 23. Yasuhara T, Matsukawa N, Yu G, Xu L, Mays RW, Kovach J. 2006. Behavioral and histological characterization of intrahippocampal grafts of human bone marrow-derived multipotent progenitor cells in neonatal rats with hypoxic ischemic injury. Cell Transplant, 15(3):231-8.
- Zheng T, Rossignol C, Leibovici A, Anderson KJ, Steindler DA Weiss MD. 2006. Transplantation of multipotent astrocytic stem cells into a rat model of neonatal hypoxicischemic encephalopathy. Brain Res, 1112(1):99-105.

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