THE INDIAN VETERINARY JOURNAL SINCE - 1924

Journal of the INDIAN VETERINARY ASSOCIATION ESTD - 1922 Regd. No. SI. No. 96/1967 (CHENNAI)



No. 11, Pasumpon Muthuramalinga Thevar Salai (Chamiers Road), Nandanam, Chennai - 600 035, Tamil Nadu, India Tel. : +91 44 2435 1006 Email : ivj83@yahoo.com ONLINE : www.ivj.org.in Vol. 96

DECEMBER 2019

THE INDIAN VETERINARY JOURNAL

(Official Organ of the Indian Veterinary Association)

EDITORIAL COMMITTEE

Dr A.V. KRISHNAN, Chief Editor B.V.Sc., M.V.Sc (Path.) Retd. Additional Director of A.H., TN

Dr S. SUKUMAR, Managing Editor

B.V.Sc., M.V.Sc (Vet. Micro), Ph.D. (Biotech) Retd. Professor of Biotechnology, TANUVAS, TN

Dr V. Titus George, Editor

B.V.Sc., M.V.Sc., Ph.D. (Patho) Retd. Prof. of Pathology, TANUVAS Dr I. Ponnu Pandian, Editor B.V.Sc., PGDAEM Dr K. Venukopalan, Editor B.V.Sc., M.V.Sc., Ph.D. (Poul.) Retd. Prof. of Poultry Science, TANUVAS

EDITORIAL BOARD

CHAIRMAN

Dr. Chirantan Kadian B.V.Sc & A.H. President, Indian Veterinary Association

MEMBERS

Prof. Dr C. Balachandran M.V.Sc, Ph.D., PGDAJ, PGDEVP, DICVP FAO Fellow, FIAVP, FNAVS, FASAW Vice-Chancellor Tamilnadu Veterinary and Animal Sciences University Madhavaram Milk Colony, Chennai - 600 051.

Dr. P.K. Panwar BVSc & AH, MVSc, Department of Animal Husbandry Uttar Pradesh Dr. Subhash Chander Ahlawat BVSc & AH. Department of Animal Husbandry Haryana

MANAGING COMMITTEE OF INDIAN VETERINARY ASSOCIATION

President : Dr. Chirantan Kadian

Secretary General : Dr. R.S. Patel

Treasurer : Dr. I. Ponnu Pandian

Joint Secretary : Dr. M.K. Srikanth

Vice Presidents : Dr. L. Chandrasekaran, HQ, Tamilnadu Dr. Arun Sirkeck, Himachal Pradesh

Dr. S.G. Yalagod, Karnataka Dr. Ilang, Nagaland

Dr. Ramji Lal Meena, Rajasthan Dr. Sudhir Kumar, Uttar Pradesh

Zonal Secretaries :

Dr. Amit Nain, Punjab Dr. R. Vasantha Rayalu, Andhra Pradesh Dr. Bhuban Ch. Sarmah, Assam Dr. Umesh Balaji Sontakke, Maharashtra Dr. Pankaj Kumar, Jharkhand

Managing Editor, IVJ :

Dr. S. Sukumar, Tamil Nadu

No. 12

3

THE INDIAN VETERINARY JOURNAL

(Official organ of the Indian Veterinary Association)

CONTENTS

Prevalence of Two Zoonotic Intestinal Parasites Strongyloides stercoralis and Hymenolepis nana in

house rat from a slum area in Surabaya District, East Java, Indonesia R. Heru Prasetyo 09 Antidiabetic Activity of Ketapang (Terminalia catappa L.) Leaves Extract in **Streptozotocin-Induced Diabetic Mice** Suhailah Hayaza, Siti Istigomah, Raden Joko Kuncoroningrat Susilo, Bilgis Inayatillah, Arif Nur Muhammad Ansori, Dwi Winarni, Saikhu Akhmad Husen, and Win Darmanto 11 Ferula Hermonis Roots Extract on Testicular Biometry and Reproductive Hormones in Local Ducks K.C.K. Al-Salhie and S.K.M. Al-Hummod 14 Identification of Legionella Pneumophila Serogroups as Zoonotic Disease Agent Distributed in Water Sources of East Java Eduardus Bimo Aksono, Kadek Rachmawati, Retno Bijanti and Herinda Pertiwi 17 Management of Feline Idiopathic Cystitis (FIC) Using Probiotic Combination Treatment Miyayu Soneta Sofyan, Nabiha Rosman, Bayu Krisnu, Juriah Binti Kamaludeen, Tri Bhawono Dadi and Herinda Pertiwi 20 . . . A Study on the Effect of Meniran (Phyllanthus Niruri Linn) Extract to Improve Infundibulum and Egg Production of Laying Chicken Infected with Escherichia Coli Emy K. Sabdoningrum, Sri Hidanah, Sri Chusniati, Adinda Rizky and Erma Safitri 22 Therapy of Rat Bone Marrow Mesenchymal Stem Cell (RBM-MSC) at White Rattusnorvegicus Induced Carbon Black against VEGF Expression Bodhi Agustono, Sri PantjaMadyawati, Rimayanti and Widjiati 25 Progesterone Profile of Dairy Cows which Experienced the Failure of Pregnancy to Artificial Insemination (AI) Sri P.Madyawati, IsnainiFadilah, Trilas Sardjito, Mas'ud Hariadi, Pudji Srianto, Suherni Susilowati and Erma Safitri 27 Characterization and Production of Polyclonal Antibody Anti Excretory Secretory Protein of Blastocystis sp Briantono Willy Rendragraha, Lucia Tri Suwanti, Rahadju Ernawati, Mufasirin Mufasirin, Setiawan Koesdarto, Wiwiek Tyasningsih, Soelih Estoe Pangestie, Heni Puspitasari and Septian Hakim Susantoputro 30

Vol. 96

GENERAL ARTICLES :

December 2019

5

Comparative Performance of TANUVAS Aseel, Gramapriya and Indigenous Desi Bird under Backyard Condition in Dharmapuri District		
R.Thangadurai and P.S.Shanmugam		33
Antigenotoxic Effect of Nano-quercetin on 7,12-dimethylbenz[a]anthracene Induced Genotoxocity Sprague Dawley Rats	' in	
S.Shahana, R.Madheswaran, P.Balachandran, A.Arivuchelvan and G.A.Balasubramaniam		35
Improving the Cell Wall Estimation in Starch Rich Cereal Grains		
Sonali Prusty, S.S. Kundu, K.S. Bisitha, U.B. Sontakke and Vijay Kumar Sharma		38
CLINICAL AND FIELD ARTICLES :		
Management of Cutaneous Papilloma in a Labrador Dog – A Case Report D.Sumathi, P.Ramesh, N.Pazhanivel, M.Sandya Bhavani, K.Amirinder Singh and M.G.Jayathangaraj		42
Silicone Resin Plastination of Helminth Parasites for Preservation		
S.T. Bino Sundar, S.Sivagnanam and Bhaskaran Ravi Latha		44
A Rare Case of Cutaneous Angiofibroma in a Cow and its Surgical Treatment R. Uma Rani and N. Pazhanivel		46
Cutaneous Streptothricosis in a Jersey Cross Bred Cow and its Therapeutic Management S.Saravanan and K.M. Palanivel		48
Preslaughter Stress in Banyuwangi Cattle During Transport Muhammad Thohawi Elziyad Purnama, Winda Kusuma Dewi, Shabrina Fauzia Prayoga, Nadia Marva Triana, Bondan Sigit Purnomo Aji, Faisal Fikri and Iwan Sahrial Hamid		50
Trauma induced Primary Hypovolemic Shock in a Deer K.P. Prabhakaran and R. Madheswaran		52
The Protective effect of <i>Swietenia macrophylla</i> Extract Nanoparticles on Pulmonary Damage in Streptozotocin-Induced Diabetic Rats		52
S.A. Sudjarwo and G. Wardani		54
A Case Report of Babesia gibsoni in Dog and its Therapeutic Management		
Prabhavathy Harikrishnan, C.Jayanthi and M.Ranjith Kumar	•••	57
FLASH BACK - A Golden article from the collection of digitalised archival of IVJ		59
Author and Subject Index	69 8	§ 70

Rana, C. S., Tiwari, J. K., Dangwal, L. R. and Sundriya, R. C. (2012) Herbal remedies for sexual capability. *Indian J. Traditional Knowledge*, **11** (4): 646-651

SPSS, Statistical Package for the Social Sciences (2015) Quantitative Data Analysis with IBM SPSS version 23: A Guide for Social Scientists. New York: Routledge. ISBN 978-0-415-57918-6.

Sturkie, P.D. (1986) Avian Physiology . 4th Ed. Springer-Verlag. New York, Berlin Heidelberg Tokyo.P:504.

Vatsalya, V. and Arora, K.L. (2012) Allometric growth of testes in relation to age ,body weight and selected blood parameters in male Japanese quail (*coturnixcoturnix Japonica*).*Int. J. Poult. Sci.*, **11**(4): 251-258.

Wilson, J.L., Krista, M., McDaniel, G.R. and Sutton, C.D. (1988) Correlation of broiler breeder male semen and testes morphology. *Poult.Sci.*, **67**: 660-668.

Zanoli, P., Benelli, A., Rivasi, M., Baraldi, C., Vezzalini, F and Barald, M. (2003) Opposite effect of acute and subchronic treatments with Ferula hermonis on copulatory behavior of male rats. *Int. J.Impot. Res.*, **15**:450–455.

Indian Vet. J., December 2019, 96 (12) : 17 - 19

Identification of *Legionella Pneumophila* Serogroups as Zoonotic Disease Agent Distributed in Water Sources of East Java

Eduardus Bimo Aksono¹, Kadek Rachmawati, Retno Bijanti and Herinda Pertiwi

Institute of Tropical Disease, Faculty of Veterinary Medicine, Faculty of Vocational Studies Universitas Airlangga - Kampus C Mulyorejo, Surabaya, Indonesia. 60115.

(Received : June, 2019 207/19 Accepted : August, 2019)

Abstract

The aims of this study was to understand the differences of *L.pneumophila* serogroups distribution in well water, tap water, ice cubes, hospital water and hotel water in East Java-Indonesia. A total of 60 water samples were tested by polymerase chain reaction and then it was analyzed by phylogenetic tree. Out of the 60 water samples collected, 12% of the samples were contaminated with *L. pneumophila*. The phylogenetic tree revealed *L.pneumophila* contamination in well water from Surabaya and tap water from Sidoarjo and the ice cubes from Sidoarjo, while the bacterial contamination in 2 well water isolate from Surabaya classified into their own group.

Key words: *L.pneumophila*, serogroup contamination distribution.

Legionella spp. are gram-negative bacteria, found in nature, and spreads by air upto 300 m radius. These bacteria are zoonotic and, infect the lungs leading to pneumonia which can be life-threatening, by inhalation of contaminated droplets of water (Ghotaslou *et al*, 2013). These bacteria thrives well in any environments; 56% in domestic hot water system, 44% in water cooling system and their ideal conditions are at the pH 6.9 and temperature 35-37°C with availability of ferric oxide as nutrients in the water. The bacteria are prevalent in any buildings with cooling and hot water system such as in office building and hotels. That is why the illness caused by the bacteria is also called "sick building syndrome".

Materials and Methods

60 samples of water (200ml) were collected from different environments in Surabaya-Sidoarjo-Kediri East Java such as well water (25), tap water (5), ice cubes (5), hospital water (16), and hotel water (9). Samples were put in a sterile vials then filtered using a Millipore membrane 0.22 µm, and they were put in a 50 ml conical flask and rinsed with 1 ml Phasphate Buffer solution (PBS) vortex for 10 minutes. 1 ml of this was transferred into into

¹Corresponding author : Email : eduardus-b-a-h@fkh.unair.ac.id

Origin of Samples	Results		Species
	Negative	Positive	Species
Well water	23/25	2/25	L. pneumophila
Tap water	4/5	1/5	L. pneumophila
Ice cubes	4/5	1/5	L. pneumophila
Hospital water	16/16	0/16	L. pneumophila
Hotel water	6/9	3/9	L. pneumophila
TOTAL	53/60	7/60	L. pneumophila

Table I. Presence of Legionella bacteria in water samples from East Java by PCR

eppendorf tubes, and centrifuged at 13000 rpm for 3 minutes. The supernatant was discarded and the sediment was used for DNA extraction using DNA extraction kit (QIAamp®DNA mini kit Qiagen) as per the manufacturer's instructions. Phylogenetic analysis was done by using software Genetix Mac Ver. 10.0

Results and Discussion

Table I showed that using PCR method for the 60 water samples collected in East Java, 12% of the samples (7/60) were contaminated by L. pneumophila. In details, 8% of the well water samples (2/25), 2% of the tap water samples (1/5), 2% of the ice cubes samples(1/5), 0% of the hospital water samples (0/16) and 33.33% of the hotel water samples (3/9) were positive for L. pnuemophila.

Results of Phylogenetic analysis showed that Legionella pneumophila contamination from well water was 1 from Surabaya and in tap water from Sidoarjo which were closer to L.pneumophila serogroup 2, 3, 4, 6, 9, 10, 12, isolates reported from Brazil, China, Spain and Australia. L.pneumophila contamination in the ice cubes from Sidoarjo were closer to serogroup1, 7, 8, 11, 13, 14, while the bacteria contaminating well water were 2 and water from hotels of Surabaya (hotel water isolate 1, 2 and 3).

The Legionella bacteria contamination was found in little quantity (>1,000 CFU/liter) (Qin *et al*, 2012) in tap water and it is not dangerous, (Aksono and Hermadi, 2017). However, they can multiply fast in flood waters.

The infection is transmitted through water spray or aerosol contaminated with

Legionella microbes. The infected person will fall ill after five or six days from the exposure. Legionella infection risk is less in small houses than the big building. In family housing, the infection was found only 6-30%. Prevention of L.pneumophila transmission could be contained by added chlorine >2 mg/liter of water (Moran-Gilad *et al*, 2014;Sánchez-Busó*et al*, 2015).

In this research, *L.pneumophila* distribution of serogroups from the natural water (such as well water) was different from different water source (such as water tank and drinking water system in hotels). These results are in line with the previous research conducted in Japan and Canada (Amemura-Maekawa *et al*, 2012; Reimer *et al*, 2009), they may correlated with water source characteristic differences (such as,

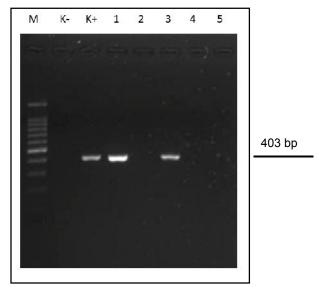


Fig 1. Results of PCR *L.pneumophila* in water samples from East Java by electrophoresis gel in 2% (403 bp). M (marker); K+ (positive control); K-(negative control); 1 (positive from tap water); 3 (positive from ice cubes).

Eduardus Bimo Aksono et al.

temperature and pH level). There are various types of water sources, while cooling tower and drinking water system tend to have similar characteristics due to same water treatments. *Legionella* has high prevalence, due to their faster intracellular growth, and in diverse hot water sources (Qin *et al*, 2013).

L. pneumophilla isolated from hot water tank in hotels had higher genetic diversity, compared with natural water and drinking water. This result may be related to growth in amoeba which adapts in different environment. It has been reported that the growth of *L. Pneumophila* in amoebae host depends on the bacterial genetics (Buse *et al*, 2012; Dey *et al*, 2009).

Summary

L.pneumophila serogroups isolated from environment water in East Java had a closer corelation with serogroups from Brazil, China, Spain and Australia and some samples were comparable to the local serogroups.

Acknowledgement

We would like to express our gratitude to Rector of Universitas Airlangga and the Ministry of Research, Technology and Higher Education for PDUPTresearch grants given in 2018.

References

Aksono, E.B., and Hermadi, H.A. (2017) Rapid detection of *Legionella* pneumophila in the water environment in Surabaya-Indonesia. *International J. of Environ. Biology*.**7(1)** : 5-10.

Amemura-Maekawa, J., Kikukawa, K., Helbig, J.H., Kaneko, S., Suzuki-Hashimoto, A., Furuhata, K., Chang, B., Murai, M., Ichinose, M., Ohnishi, M., and Kura, F.(2012) Distribution of

monoclonal antibody subgroups and sequence-based types among *Legionella* pneumophila serogroup 1 isolates derived from cooling tower water, bathwater, and soil in Japan. *Appl. Environ. Microbiol.* **78**:4263–4270.

Buse, H.Y., and Ashbolt, N.J. (2011) Differential growth of *Legionella* pneumophila strains within a range of amoebae at various temperatures associated with in-premise plumbing. *Lett. Appl. Microbiol.* **53**:217–224.

Dey, R., Bodennec J., Mameri, M.O., and Pernin, P (2009) Free-living freshwater amoebae differ in their susceptibility to the pathogenic bacterium *Legionella* pneumophila. *FEMS Microbiol. Lett.*290:10–17.

Ghotaslou, R., Sefidan, F.Y., Akhi, M.T., Soroush, M.H., and Hejazi, M.S. (2013) Detection of *Legionella* Contamination in Tabriz Hospitals by PCR Assay. *Adv Pharm Bull.* **3(1)**: 131–134.

Moran-Gilad, J., Mentasti, M., Lazarovitch, T., Huberman, Z., Stocki, T., Sadik, C., Shahar, T., Anis, E., Valinsky, L., Harrison, TG., and Grotto I., (2014) ESCMID Study Group for *Legionella* Infections (ESGLI). Molecular epidemiology of Legionnaires' disease in Israel. *Clin Microbiol Infect* **20**:690–696.

Reimer, A.R., Au, S., Schindle, S., and Bernard, K.A. (2010) *Legionella* pneumophila monoclonal antibody subgroups and DNA sequence types isolated in Canada between 1981 and 2009: laboratory component of national surveillance. *Eur. J. Clin. Microbiol. Infect. Dis.***29**:191–205.

Sánchez-Busó, L., Coscollà, M., Palero, F., Camaró, M.L., Gimeno, A., Moreno, P., Escribano, I., López Perezagua, M.M., Colomina, J., Vanaclocha, H., and González-Candelas, F. (2015) Geographical and temporal structures of *Legionella* pneumophila sequence types in Comunitat Valenciana (Spain), 1998 to 2013. *Appl Environ Microbiol* **81**:7106–7113.

Qin,T.,Tian,Z., Ren,H., Hu,G., Zhou,H., Lu,J., Luo,C., Liu,Z., and Shao, Z. (2012) Application of EMA-qPCR as a complementary tool for the detection and monitoring of *Legionella* in different water systems. *World J. Microbiol. Biotechnol.***28**:1881–1890.

Qin, T., Yan, G., Ren, H., Zhou, H., Wang, H., , Y., Zhao, M., Guan, H., Li, M., and Shao, Z. (2013) High prevalence, genetic diversity and intracellular growth ability of *Legionella* in hot spring environments. *PLoS One* **8(3)**: e59018.