

# Associations between P53, Transforming Growth Factor Beta-1, and Interleukin-10 Serum Levels with Advanced Liver Disease and Hepatitis B Virus Infection

Citrawatidyahkencono Wungu<sup>1,2</sup>, Mochamad Amin<sup>2</sup>, Ulfa Kholili<sup>3</sup>, Gwennyichsan Prabowo<sup>1</sup>,  
Poernomoboedi Setiawan<sup>3</sup>, Soetjipto<sup>4,5</sup>, Retno Handajani<sup>4,5</sup>

<sup>1</sup>Researcher, Department of Physiology and Medical Biochemistry, Medical Faculty of Universitas Airlangga, Jl. Prof. Dr. Moestopo 47, Surabaya, Indonesia, <sup>2</sup>Researcher, Institute of Tropical Disease, Universitasairlangga Campus C, Surabaya, Indonesia, <sup>3</sup>Clinical Instructor, Department of Internal Medicine, Medical Faculty Of Universitas Airlangga - Dr. Soetomo General Hospital, Jl. Prof. Dr. Moestopo 6-8, Surabaya, Indonesia, <sup>4</sup>Professor, Institute Of Tropical Disease, Universitasairlangga Campus C, Surabaya, Indonesia, <sup>5</sup>Professor, Department of Physiology And Medical Biochemistry, Medical Faculty Of Universitas Airlangga, Jl. Prof. Dr. Moestopo 47, Surabaya, Indonesia

## Abstract

**Background.** Hepatitis B infection can lead to advanced liver disease (ALD) which causes serious health problems. Several factors that are thought to play a role in advanced liver disease are P53, Transforming Growth Factor- $\beta$ 1 (TGF- $\beta$ 1), and Interleukin-10 (IL-10). Therefore, this study aimed to determine the association between P53, TGF- $\beta$ 1, and IL-10 serum levels with ALD.

**Methods.** We collected 68 sera from patients with HBV infection. P53, TGF- $\beta$ 1, and IL-10 serum levels were measured by ELISA. We also detected SNPs from P53 and TGF- $\beta$ 1 genes to determines their associations.

**Results and Conclusions:** In this study, we obtained 41.18% CH patients and 58.82% ALD patients. Male patients outnumbered female in all groups. There was a significant relationship between serum P53 and TGF- $\beta$ 1 levels with ALD ( $p = 0.03$  and  $0.01$ , respectively), but not serum IL-10 levels. However, there was no significant relationship between SNP gene P53 and serum P53 levels nor between SNP gene TGF- $\beta$ 1 and serum TGF- $\beta$ 1 levels ( $p = 0.73$  and  $0.23$ , respectively). Both P53 and TGF- $\beta$ 1 serum levels can act as biomarkers of prognosis and target therapy for ALD patients.

**Keywords:** P53, Transforming Growth Factor Beta-1, interleukin-10, advanced liver disease, Hepatitis B Virus infection

## Introduction

Hepatitis B Virus (HBV) infection is a global health problem, especially in developing countries<sup>1,2</sup>. Untreated early HBV infection can lead to persistent chronic infection with manifestations range from asymptomatic to cirrhosis, decompensated liver disease, and hepatocellular carcinoma (HCC) in up to 40% of patients<sup>3,4</sup>. Annually, 3% patients with chronic hepatitis B develop liver cirrhosis and 5% develop decompensated liver disease. The main therapy for end-stage liver disease is liver transplantation, however, this treatment modality cannot be accessed by the majority

of patients<sup>4</sup>. Liver cirrhosis and HCC are advanced liver disease (ALD) conditions with complex molecular pathogenesis, involving the interactions of various cells and cytokines<sup>5,6</sup>. Several factors that are thought to play a role in advanced liver disease are Transforming Growth Factor- $\beta$ 1 (TGF- $\beta$ 1), P53, and Interleukin-10 (IL-10).

TGF- $\beta$ 1 is a cytokine that can trigger apoptosis and control cell growth, but on the other hand, this cytokine can cause the transition of Hepatic Stellate Cell (HSC) to myofibroblasts which stimulate extracellular matrix synthesis and inhibit its degradation<sup>7,8</sup>. Chronic

inflammatory conditions cause hepatocytes to become resistant to the anti-tumor activity of TGF- $\beta$ 1 and a change in the signaling pathway converts TGF- $\beta$ 1 to be pro-fibrotic and pro-carcinogenic<sup>9,10</sup>. Changes in TGF- $\beta$ 1 levels play an important role in the pathogenesis of various diseases, including liver disease<sup>11</sup>. TGF- $\beta$ 1 is also suggested to be a more specific biomarker than Alpha Feto Protein (AFP) for HCC<sup>12</sup>.

P53 also plays an important role in the pathogenesis of ALD, as the gene can be defective and mutated in HBV infection<sup>13,14</sup>. This protein acts as a regulator which suppresses the cell cycle and induced apoptosis, thus preventing carcinogenesis, including liver cancer. As P53 protein has an important role in the progression of ALD, its expression can be used to monitor and identify ALD at an early stage.<sup>13,15,16</sup>.

Interleukin-10 (IL-10) is a cytokine produced by macrophages and dendritic cells to inhibit the activation of these two cells. Experimental data from animal models and clinical data from patients suggest that IL-10 is involved in the development of liver injury<sup>17</sup>. Even though IL-10 inhibits the production of various inflammatory cytokines (Th1 cytokines), including IL-1, TNF, and IL-12<sup>18</sup>, it suppresses immune system against HBV infection (rolisaxena). IL-10 inhibits T cell activity, either directly or indirectly. The direct effect of IL-10 inhibition on T cells is by inhibiting their proliferation, functional differentiation, and activity, while the indirect inhibition is by reducing the molecular expression of presenting cell antigens, thus interfering with T cell maturation<sup>19</sup>.

To date, data regarding the relationship between TGF- $\beta$ 1, P53, and IL-10 levels and the incidence of CLD in patients with chronic HBV infection, especially in the Indonesian population, are still limited. Several other studies also linked the Single Nucleotide Polymorphism (SNP)s of TGF- $\beta$ 1 and P53 genes with the expression, even though this yield conflicting results<sup>20-23</sup>. Therefore, this study aimed to analyze the relationship between serum TGF- $\beta$ 1, P53, and IL-10 levels and HBV-related advanced liver disease in the Indonesian population.

## Materials and Methods

### Sampling

This was a cross-sectional study that used 68 stored blood samples from chronically infected HBV patients. Samples were taken at Inpatient/Outpatient Clinics of Internal Medicine Department, Dr. Soetomo General Hospital, Surabaya, Indonesia in 2017 and stored in -80°C freezer at Hepatitis laboratory, Institute of Tropical Disease, Surabaya, Indonesia. Inclusion criteria were: Adult patients with chronic HBV infection (HBsAg was positive for more than 6 months), clinically stable, and never had Hepatitis B vaccination. Exclusion criteria were: HCV and HIV coinfection, diabetes mellitus/ autoimmune diseases. Patients were categorized into chronic hepatitis without ALD if there were no sign of liver cirrhosis or hepatocellular carcinoma through physical, laboratory, and radiologic examination, while they were categorized into ALD if liver cirrhosis or HCC was found.

This study had received approval from the Ethical Committee of Faculty of Medicine, Universitas Airlangga, Surabaya (Ethical clearance No. 186/EC/KEPK/FKUA/2020). Informed consent was explained and voluntarily signed by participants beforehand, in accordance with the declaration of Helsinki.

**Measurement of TGF- $\beta$ 1, P53, and IL-10 serum levels.** TGF- $\beta$ 1 serum level was measured by: Human TGF- $\beta$ 1 ELISA Kit with Catalog No: E-EL-H0110 (Elabscience Biotechnology Inc, USA) according to the manufacturer's instructions. P53 serum level was measured by: Human TP53 (Tumor Protein p53) ELISA Kit with Catalog No: E-EL-H0910 (Elabscience Biotechnology Inc, USA). IL-10 serum level was measured by: Human TGF- $\beta$ 1 ELISA Kit with Catalog No: E-EL-H0103 (Elabscience Biotechnology Inc, USA). Optical Density for TGF- $\beta$ 1, P53, and IL-10 were measured by Microplate Reader: iMark (Biorad) S/N 12908.

### Host DNA Extraction

Host DNA was extracted using QIAamp DNA Extraction kit (Qiagen, Germany) Cat.No.51104 according to the manufacturer's instructions.

### PCR-RFLP of TGF- $\beta$ 1 and p53 Genes

PCR was carried out as our previous work<sup>24</sup>. TGF- $\beta$ 1 gene SNP was checked at locus -509, while P53

gene SNP was checked at codon 72. PCR products of TGF-β1-509 SNP were incubated using DdeI, while p53 SNP used BstUI.

TGF-β1 -509 SNP showed 120 bp (T allele), while wild type (C allele) showed 74 and 46 bp fragments. p53 codon 72 SNP showed 231 and 165 bp fragments (G allele), while wild type showed 396 bp (C allele).

### Statistical Analysis

Associations between TGF-β1, P53, and IL-10 with ALD were analyzed for each parameter by Mann-Whitney test, while the associations between SNPs of TGF-β1 and P53 with their expression were analyzed by Kruskal Wallis test. A P-value < 0.05 following a

two-tailed analysis was considered to be statistically significant. All statistical analyses were performed using SPSS version 23.

## Results

### Characteristics of the participants

This study involved 68 participants with HBV infection, consisting of 41.18% CH patients and 58.82% ALD patients. Data summary regarding gender, mean ± SD age and age range of the participants can be seen in Table 1. The youngest patients with HBV infection in this study were 16 years old and the oldest was 72 years old, with a mean age of 47 years. There were more males (79.41%) than females (20.59%). The age range of male patients in this study was not much different from that of women, with the mean value being almost the same.

**Table 1. Characteristics of the participants**

Characteristics	CH (n=28)	ALD (n=40)	Total (n=68)	P-value
Gender				
Female	7	7	14	0.55
Male	21	33	54	
Age, years	44.36±16.31	49.23±9.52	47.22±12.87	0.16
P53 SNP Genotype				0.17
CC	9	8	17	
CG	15	22	37	
GG	4	10	14	0.28
Allele	33	38	71	
C	23	42	65	
G				
TGF-β1 SNP Genotype				0.44
CC	8	8	16	
CT	14	18	32	
TT	6	14	20	0.22
Allele				
C	30	34	64	
T	26	46	72	
P53 serum level (pg/mL)	114.99±164.73	244.75±374.62	191.32±311.07	0.03*
TGF-β1 serum level (ng/mL)	0.16±0.39	0.29±0.27	0.21±0.35	0.01*
IL-10 serum level (pg/mL)	9.38±9.14	14.34±34	12.3±26.69	0.76

\* statistically significant

In both CH and ALD, more patients were male than female, although there was no significant difference in gender distribution in the CH and ALD groups. From statistical analysis, there was no significant difference between gender and age with ALD. For both SNPs, heterozygote genotype were predominant in either CH or ALD. SNP analysis showed that there was no difference in the distribution of genotypes or SNP alleles of TGF- $\beta$ 1 and p53 between CH and ALD. Serum P53, TGF-b1, and IL-10 levels in ALD patients were higher than those of CH patients. There was a significant relationship between serum P53 and TGF-b1 levels with ALD, but not serum IL-10 levels. However, there was no significant relationship between SNP gene P53 and serum P53 levels nor between SNP gene TGF-b1 and serum TGF-b1 levels  $p = 0.73$  and  $0.23$ , respectively).

### Discussion

In this study, 68 samples of patients with HBV infection were obtained, consisting of CH and ALD. The number of male patients were more than women. This is in accordance with several previous studies which stated that there were more people with cirrhosis of the liver with HBV infection in men than in women<sup>25-27</sup>. This is due to the protective role of estrogen for the liver. Estrogen can inhibit stellate cell proliferation and fibrogenesis which plays an important role in the course of cirrhosis. In experimental animal models with liver cirrhosis, administration of estradiol causes a decrease in collagen types I and III and proliferation of stellate cells.<sup>28</sup> Besides being able to reduce viral RNA transcription and HBx expression, estrogen is also able to regulate HBV activity through direct inhibition of the binding of Hepatocyte Nuclear Factor (HNF)-4 $\alpha$  with HBV EnhI, thus estrogen signals can suppress the production of pro-inflammatory cytokines in the liver<sup>29,30</sup>. ALD patients in this study were dominated by the age range 45-50 years. It is said that the mean age of incidence of HCC associated with HBV infection is 50 years, except in high-risk populations which is 40 years<sup>31</sup>.

In this study, it was found that the levels of P53, TGF-b1, and IL-10 were higher in men than women. Levels of P53, TGF-b1, and IL-10 were also found to be higher in ALD than in CH. In statistical analysis, there

were significant differences in serum P53 and TGF-b1 levels between the ALD and CH ( $p < 0.05$ ). It was said that the expression of P53 increased in malignancy and pre-malignancy, even overexpression of P53 was not uncommon in malignant conditions, including in terms of it's in liver cancer. In liver cancer there may also be a mutation in the P53 gene that increases expression and cannot be distinguished from normal P53 expression<sup>32</sup>. TGF-b1 levels also indicate hepatic function impairment, which is increasing in number in patients with cirrhosis of the liver. In the liver, high levels of TGF-b1 play a role in the occurrence of hepatic fibrogenesis, regulation of cell growth, and tumor development.<sup>33</sup> In a study by Kohla et al (2017), it was also stated that TGF-b1 levels increased with disease severity and progression in hepatocellular carcinoma, and TGF-b1 level could be complementary to alpha fetoprotein (AFP) in the diagnosis of HCC.<sup>34</sup> In this study, there was no significant relationship between serum IL-10 levels and ALD. This is in line with the meta-analysis conducted by Shakiba et al (2018) which states that there are differences in IL-10 levels between HCC patients and normal people, but there is no difference between HCC and CH patients. In that study, it was also stated that the increase in IL-10 levels in ALD was due more to the cirrhosis process than tumor load<sup>35</sup>.

When associated with previous studies, there was no relationship between Single Nucleotide Polymorphism (SNP) TGF- $\beta$ 1 -509T/C and P53 Arg72Pro genes with TGF- $\beta$ 1 and P53 serum levels. In vitro, it is known that there are several external factors that can influence the expression of the five TGF- $\beta$  isoforms, including TGF- $\beta$ 1. In addition, the TGF- $\beta$ 1 signaling process is quite complex, involving the intracellular protein Smad which acts on various transcription factors in the nucleus. The expression of TGF- $\beta$ 1 is also influenced by a variety of different cytokines and chemical compounds. Other locus polymorphisms such as C466T and T869C can also affect TGF- $\beta$ 1 expression<sup>36</sup>. Likewise, P53 levels are not only affected by the Arg72Pro polymorphism, but also SNPs in other P53 gene regions, including non-coding regions that can affect gene splicing<sup>37</sup>.

This study has several limitations, the first, it did not compare between ALD and CH with healthy controls. The second, it could not explain the role of other cytokines and epigenetic pathways in relation to

the pathogenesis of ALD. Further research is needed to understand the mechanism pathways for increasing levels of P53 and TGF- $\beta$ 1 in ALD patients other than through gene expression and polymorphisms. Studies regarding the various cytokines and molecular signaling involved can help understand the pathogenesis and are used as therapeutic pathways in ALD patients with HBV infection.

### Conclusion

P53 and TGF- $\beta$ 1 levels were significantly elevated in ALD patients with HBV infection. Both parameters can act as biomarkers of prognosis and target therapy for ALD patients.

**Ethical approval:** Ethical Committee of Faculty of Medicine, Universitas Airlangga, Surabaya (Ethical clearance No. 186/EC/KEPK/FKUA/2020).

**Patient' Informed Consent:** Patient's informed consent has been obtained.

**Conflict of Interest:** Nil

### Funding

The authors would like to thank Universitas Airlangga for giving research fund through "Penelitian Unggulan Fakultas" year 2020.

### References

1. Xu H-Z, Liu Y-P, Guleng B, Ren J-L. Hepatitis B Virus-Related Hepatocellular Carcinoma: Pathogenic Mechanisms and Novel Therapeutic Interventions. *Gastrointest Tumors* [Internet]. 2014;1(3):135–45. Available from: <http://www.karger.com?doi=10.1159/000365307>
2. Zampino R, Boemio A, Sagnelli C, Alessio L, Adinolfi LE, Sagnelli E, et al. 2015 Advances in Hepatitis B virus hepatitis B virus burden in developing countries. *World J Gastroenterol*. 2015;21(42):11941–53.
3. Weisberg IS, Brown RS, Sigal SH. Hepatitis B and End-Stage Liver Disease. *Clin Liver Dis*. 2007;11(4):893–916.
4. Guan R, Lui HF. Treatment of Hepatitis B in Decompensated Liver Cirrhosis. *Int J Hepatol* [Internet]. 2011;2011:1–11. Available from: <http://www.hindawi.com/journals/ijh/2011/918017/>
5. Dewidar B, Meyer C, Dooley S, Meindl-Beinker AN. TGF- $\beta$  in Hepatic Stellate Cell Activation and Liver Fibrogenesis-Updated 2019. *Cells*. 2019;8(11):1–35.
6. Zhou WC, Zhang QB, Qiao L. Pathogenesis of liver cirrhosis. *World J Gastroenterol*. 2014;20(23):7312–24.
7. Bordiny M El, Hanafy SM, Abdo A. Impact of single nucleotide polymorphism of TGF- $\beta$ 1 gene (SNP-Codon10) on hepatocellular carcinoma risk in Egyptian patients following HCV infection. *Aust J Basic Appl Sci*. 2011;5(9):1814–21.
8. Mohy A, Fouad A. Role of transforming growth factor- $\beta$ 1 in serum and -509 C>T promoter gene polymorphism in development of liver cirrhosis in Egyptian patients. *Meta Gene* [Internet]. 2014;2:631–7. Available from: <http://dx.doi.org/10.1016/j.mgene.2014.08.002>
9. Falleti E, Fabris C, Toniutto P, Fontanini E, Cussigh A, Bitetto D, et al. TGF-Beta1 genotypes in cirrhosis: Relationship with the occurrence of liver cancer. *Cytokine*. 2008;44(2):256–61.
10. Ozaki I, Hamajima H, Matsuhashi S, Mizuta T. Regulation of TGF- $\beta$ 1-induced pro-apoptotic signaling by growth factor receptors and extracellular matrix receptor integrins in the liver. *Front Physiol*. 2011;2 OCT(October):1–8.
11. Matsuzaki K, Murata M, Yoshida K, Sekimoto G, Uemura Y, Sakaida N, et al. Chronic inflammation associated with hepatitis C virus infection perturbs hepatic transforming growth factor  $\beta$  signaling, promoting cirrhosis and hepatocellular carcinoma. *Hepatology*. 2007;46(1):48–57.
12. Yao D-F, Dong Z-Z, Yao M. Specific molecular markers in hepatocellular carcinoma. *Hepatobiliary Pancreat Dis Int* [Internet]. 2007;6(3):241–7. Available from: <papers3://publication/uuid/E679ED79-36DF-452B-A23D-F0B4D180D28A>
13. Wang Z, Gou W, Liu M, Sang W, Chu H, Zhang W. Expression of P53 and HSP70 in Chronic Hepatitis, Liver Cirrhosis, and Early and Advanced Hepatocellular Carcinoma Tissues and Their Diagnostic Value in Hepatocellular Carcinoma: An Immunohistochemical Study. *Med Sci Monit* [Internet]. 2015;21:3209–15. Available from: <http://www.medscimonit.com/abstract/index/idArt/895592>
14. Hussain SP, Schwank J, Staib F, Wang XW, Harris

- CC. TP53 mutations and hepatocellular carcinoma: Insights into the etiology and pathogenesis of liver cancer. *Oncogene*. 2007;26(15):2166–76.
15. Gouas DA, Villar S, Ortiz-Cuaran S, Legros P, Ferro G, Kirk GD, et al. TP53 R249S mutation, genetic variations in HBX and risk of hepatocellular carcinoma in The Gambia. *Carcinogenesis*. 2012;33(6):1219–24.
  16. Hu S, Zhao L, Yang J, Hu M. The association between polymorphism of P53 Codon72 Arg/Pro and hepatocellular carcinoma susceptibility: Evidence from a meta-analysis of 15 studies with 3,704 cases. *Tumor Biol* [Internet]. 2014;35(4):3647–56. Available from: <http://dx.doi.org/10.1016/j.mgene.2013.09.010>
  17. Knolle PA, Gerken G. Local control of the immune response in the liver. *VN-readcube.com. Immunol Rev* [Internet]. 2000;174:21–34. Available from: [c:\Users\Costr005\Desktop\5CPhD backups\5CDell Backup 130701\5 CDELL\5 CPHD\5 COther\5CReview and Literature-PDFs\5 CSystematic Review\5CPDFs for systematic review\5CMedline Search\5CNOT ACCEPTED\5 CExcluded but important\5CLocal control of the immune res](http://c:\Users\Costr005\Desktop\5CPhD backups\5CDell Backup 130701\5 CDELL\5 CPHD\5 COther\5CReview and Literature-PDFs\5 CSystematic Review\5CPDFs for systematic review\5CMedline Search\5CNOT ACCEPTED\5 CExcluded but important\5CLocal control of the immune res)
  18. Abbas AK, Lichtman AH, Pillai S, Baker DL, Baker A. *Cellular and Molecular Immunology*. 8th ed. Philadelphia: Elsevier Saunders; 2015.
  19. Özgüler M, Akbulut HH, Akbulut A. Evaluation of interleukin-10 levels in patients diagnosed with chronic hepatitis. *West Indian Med J*. 2015;64(2):71–5.
  20. Grochola LF, Zeron-Medina J, Meriaux S, Bond GL. Single-nucleotide polymorphisms in the p53 signaling pathway. *Cold Spring Harb Perspect Biol*. 2010;2(5):1–18.
  21. Bond GL, Levine AJ. A single nucleotide polymorphism in the p53 pathway interacts with gender, environmental stresses and tumor genetics to influence cancer in humans. *Oncogene*. 2007;26(9):1317–23.
  22. Khani M, Amani D, Taheripanah R, Sanadgol N, Feizollahzadeh S, Rahmani Z. Transforming growth factor beta-1 (TGF- $\beta$ 1) gene single nucleotide polymorphisms (SNPs) and susceptibility to pre-eclampsia in Iranian women: A case-control study. *Pregnancy Hypertens* [Internet]. 2015;5(4):267–72. Available from: <http://dx.doi.org/10.1016/j.preghy.2015.01.002>
  23. Xu S, Yang S, Sun G, Huang W, Zhang Y. Transforming Growth Factor-Beta Polymorphisms and Serum Level in the Development of Osteosarcoma. *DNA Cell Biol*. 2014;33(11):802–6.
  24. Wungu CDK, Amin M, Ruslan SEN, Purwono PB, Kholili U, Maimunah U, et al. Association between host TNF- $\alpha$ , TGF- $\beta$ 1, p53 polymorphisms, hbv x gene mutation, hbv viral load and the progression of hbv-associated chronic liver disease in Indonesian patients. *Biomed Reports*. 2019;11(4):145–53.
  25. Bobek V, Kolostova K, Pinterova D, Kacprzak G, Adamiak J, Kolodziej J, et al. A clinically relevant, syngeneic model of spontaneous, highly metastatic B16 mouse melanoma. *Anticancer Res*. 2010;30(12):4799–804.
  26. Lee MH, Kim DY, Kim JK, Chang HY, Kang SH, Ryu HJ, et al. Combination of preS deletions and A1762T/G1764A mutations in HBV subgenotype C2 increases the risk of developing HCC. *Intervirology*. 2012;55(4):296–302.
  27. Başıyigit S, Asiltürk Z, Sapmaz F, Kefeli A, Yeniova AÖ, Uzman M, et al. Hepatitis B virus is still the most common etiologic factor of liver cirrhosis: Results from a single center in Turkey. *Dicle Med J / Dicle Tip Derg* [Internet]. 2015;42(4):416–21. Available from: <http://dergipark.gov.tr/doi/10.5798/diclemedj.0921.2015.04.0601>
  28. Brady CW. Liver disease in menopause. *World J Gastroenterol*. 2015;21(25):7613–20.
  29. Montella M, D’Arena G, Crispo A, Capunzo M, Nocerino F, Grimaldi M, et al. Role of Sex Hormones in the Development and Progression of Hepatitis B Virus-Associated Hepatocellular Carcinoma. *Int J Endocrinol*. 2015;2015:1–9.
  30. Wang SH, Yeh SH, Lin WH, Yeh KH, Yuan Q, Xia NS, et al. Estrogen Receptor  $\alpha$  Represses Transcription of HBV Genes via Interaction With Hepatocyte Nuclear Factor 4 $\alpha$ . *Gastroenterology* [Internet]. 2012;142(4):989–998.e4. Available from: <http://dx.doi.org/10.1053/j.gastro.2011.12.045>
  31. Yan H, Yang Y, Zhang L, Tang G, Wang Y, Xue G, et al. Characterization of the Genotype and Integration Patterns of Hepatitis B Virus in Early- and Late-Onset Hepatocellular Carcinoma. *Hepatology*. 2015;61:1821–31.
  32. Lalisang TJM, Moenadjat Y, Siregar NC, Stephanie

- M. Overexpression of p53 in extra large (More than 10 cm) hepatocellular carcinoma. *Med J Indones.* 2018;27(2):3.
33. Flisiak R, Pytel-Krolczuk B, Prokopowicz D. Circulating transforming growth factor  $\beta$ 1 as an indicator of hepatic function impairment in liver cirrhosis. *Cytokine.* 2000;12(6):677–81.
34. Kohla MAS, Attia A, Darwesh N, Obada M, Taha H, Youssef MF. Association of serum levels of transforming growth factor b1 with disease severity in patients with hepatocellular carcinoma. *Hepatoma Res.* 2017;3(12):294.
35. Shakiba E, Sadeghi M, Shakiba M. A systematic review and meta-analysis of evaluation of serum interleukin 8 levels in hepatocellular carcinoma. *Clin Exp Hepatol.* 2019;5(2):123–8.
36. Panek M, Pietras T, Fabijan A, Ziolo J, Wieteska Ł, Małachowska B, et al. Identification and association of the single nucleotide polymorphisms, C-509T, C+466T and T+869C, of the TGF-1 $\beta$  gene in patients with asthma and their influence on the mRNA expression level of TGF- $\beta$ 1. *Int J Mol Med.* 2014;34(4):975–86.
37. Zhang G, Xu Q, Wang Z, Sun L, Lv Z, Liu J, et al. P53 protein expression affected by TP53 polymorphism is associated with the biological behavior and prognosis of low rectal cancer. *Oncol Lett.* 2019;18(6):6807–21.