

J. Indian Chem. Soc. Vol. 96 No. 12
December 2019

JOURNAL OF THE INDIAN CHEMICAL SOCIETY



Published by Indian Chemical Society
92, Acharya Prafulla Chandra Road
Kolkata-700 009, India



✉ INDI3478@DATAONE.IN

☎ CALL US:

(../)


[HOME](#) [ABSTRACTS](#) [TEMPLATES](#) [INFORMATION TO AUTHOR \(DOCS/INFORM_AUTH.PDF\)](#)

[SPECIAL ISSUE](#) [EDITORIAL BOARD](#) [LOGIN](#) [ICS HOME](#)

[All Volumes & Issues \(new_rjournal.php?journal_type=R\)](#)

VOLUME 96, ISSUE 12, DECEMBER 2019


In this issue (13 articles)

 *Original Paper*

Revisiting the synthesis and applications of graphene oxide

[\(abstract_details.php?journal=MjUw\)](#)


Sujata Kumari, Vandana Yadav, Pratibha Sharma and Sudip Majumder*

 *Original Paper*

Volumetric, ultrasonic and conductance behaviour of metformin hydrochloride (MH) in water and aqueous sorbitol at different temperatures

(abstract_details.php?journal=MjUx)


Shashi Kant Lomesh*, Inesh Kumar, Dinesh Kumar and Madhu Bala

 Original Paper

Graph theoretical study of stabilization of some IPR fullerenes by cage closure

(abstract_details.php?journal=MjUy)


Bankim Chandra Ghosh

 Original Paper

Comparative study on denitrification kinetics of nitrified effluent with and without organic carbon by activated sludge process

(abstract_details.php?journal=MjUz)


Roumi Bhattacharya* and Debabrata Mazumder

 Original Paper

Preparation and characterization of PEGylated capric acid liposomes for intravenous delivery system

(abstract_details.php?journal=MjU0)


V. R. Eh Suk *a, N. A. Basrowi a, K. Khalid b and M. Misran a

 Original Paper

Preparation of carboxylic acid functionalized organosilan on ferrite-silica nanoparticles and evaluation of their catalytic activity in synthesis of oxindoles

(abstract_details.php?journal=MjU1)

Hassan Hassani *a and Sorayya Ebrahimi b

 Original Paper

Isolation of cytotoxic sesquiterpenes from *Curcuma comosa* and characterization of their structures

(abstract_details.php?journal=MjU2)

Khun Nay Win Tun a,b, Nanik Siti Aminah *a, Alfinda Novi Kristanti a, Rico Ramadhan a, Yoshiaki Takaya c and Hnin Thanda Aung d



Original Paper

Food preservative chemistry: Effects and side effects

([abstract_details.php?journal=MjU3](http://indianchemicalsociety.com/journal/abstract_details.php?journal=MjU3))

Debashree Mandal



Original Paper

A simplified approach for modelling of an aerobic fixed bed hybrid bioreactor

([abstract_details.php?journal=MjU4](http://indianchemicalsociety.com/journal/abstract_details.php?journal=MjU4))

Sushovan Sarkar *a and Debabrata Mazumder b



Original Paper

Analytical method for the assay of Perindopril Erbumine in formulations by ion association complex formation using Tropaeolin OOO (TPOOO)

([abstract_details.php?journal=MjU5](http://indianchemicalsociety.com/journal/abstract_details.php?journal=MjU5))

Thuttgartunta Manikya Sastry *a and Karripeddi Ramakrishna b



Original Paper

Biodegradation of *p*-Chloro Meta Xylenol (PCMX) and modelling of degradation kinetic analysis using *Pandoraea sp.*

([abstract_details.php?journal=MjYw](http://indianchemicalsociety.com/journal/abstract_details.php?journal=MjYw))

Bhanupriya Brahma a and Priyabrata Sarkar *b



Original Paper

Epoxidation of alkenes using cost-effective green oxidant under eco-friendly reaction condition

([abstract_details.php?journal=MjYx](http://indianchemicalsociety.com/journal/abstract_details.php?journal=MjYx))

Uday Sankar Agarwalla



Original Paper

Effect of alkali treatment on single species wood pulp fiber properties for the application of nursing pad absorbent core

([abstract_details.php?journal=MjYy](http://indianchemicalsociety.com/journal/abstract_details.php?journal=MjYy))

Aswini M.

© Copyrights [Indian Chemical Society \(index.html\)](#) . All rights reserved. [Privacy policy](#) | [Terms of use](#)



Honorary Editors

Physical & Industrial Chemistry Section

Professor A C Bhasikuttan



Radiation & Photochemistry Division

BARC, Mumbai, Mumbai – 400 085

E-mail: bhasikuttan@gmail.com

http://www.hbni.ac.in/faculty/BARC/barcm_chem_bhasikuttan_ac.htm

Inorganic & Analytical Chemistry Section

Professor Sundargopal Ghosh



Department of Chemistry

Indian Institute of Technology Madras, Chennai – 600 036

E-mail: ghoshsundargopal@gmail.com

<http://chem.iitm.ac.in/profghosh/index.html>

Organic and Biochemistry Section

Professor Chhanda Mukhopadhyay



Head, Department of Chemistry

University of Calcutta

E-mail: cmukhop@yahoo.co.in

<https://www.caluniv.ac.in/academic/department/Chemistry/Chhanda-Mukhopadhyay.pdf>

International Executive Editor

Bimal Krishna Banik, PhD. C. Chem. F. R. S. C.

**Vice President of Research and Education Development Community Health
Systems of South Texas Edinburg, Texas 78539, USA**

E-mail: bimal.banik@chsst.org ; bimalbanik10@gmail.com

Phone Numbers: 956-627-4094 and 281-813-2104

International Editors

Professor Takashiro Akitsu

Department of Chemistry, Faculty of Science, Tokyo University of Science

1-3 Kagurazaka, Shinjuku-ku, Tokyo 162-8601, Japan

Tel. +81-3- 5228-8271, Fax. +81-3-5261-4631

E-mail: akitsu@rs.kagu.tus.ac.jp

<http://www.rs.kagu.tus.ac.jp/akitsu/index.htm>

Professor Ebbe Nordlander

Department of Chemical Physica

Lund University

Box 117, 221 00 Lund, Sweden

E-mail: ebbe.nordlander@chemphys.lu.se

<https://www.lunduniversity.lu.se/>

Dr. Anil Kumar FNA FASc FNASc

J C Bose National Fellow

Physical & Materials Chemistry Division

CSIR-National Chemical Laboratory (NCL)

Pune 411008, INDIA

Email: a.kumar@ncl.res.in

<http://academic.ncl.res.in/a.kumar>

Editor (Guest)

Dr. Shivendu Ranjan, B.Tech, Ph.D, PDF, MICS, FBSS (India), FLS (London)

Scientist DST-Centre for Policy Research Lucknow, Uttar Pradesh, India

And

Senior Research Associate (Adjunct) Faculty of Engineering & Built Environment

**University of Johannesburg, Johannesburg (Auckland Park Kingsway Campus)
South Africa**

Email: shivenduranjan@gmail.com

<https://sites.google.com/view/shivenduranjan>



SJR

Scimago Journal & Country Rank

Enter Journal Title, ISSN or Publisher Name

[Home](#)[Journal Rankings](#)[Country Rankings](#)[Viz Tools](#)[Help](#)[About Us](#)

Journal of the Indian Chemical Society

Country [India](#) - [SIR Ranking of India](#)**Subject Area and Category**[Chemistry](#)[Electrochemistry](#)[Inorganic Chemistry](#)[Organic Chemistry](#)[Physical and Theoretical Chemistry](#)[Pharmacology, Toxicology and Pharmaceutics](#)[Drug Discovery](#)**Publisher**[Scientific Publishers](#)**Publication type**[Journals](#)**ISSN**

00194522

Coverage

1973-1985, 1987-1991, 1996-ongoing

Scope

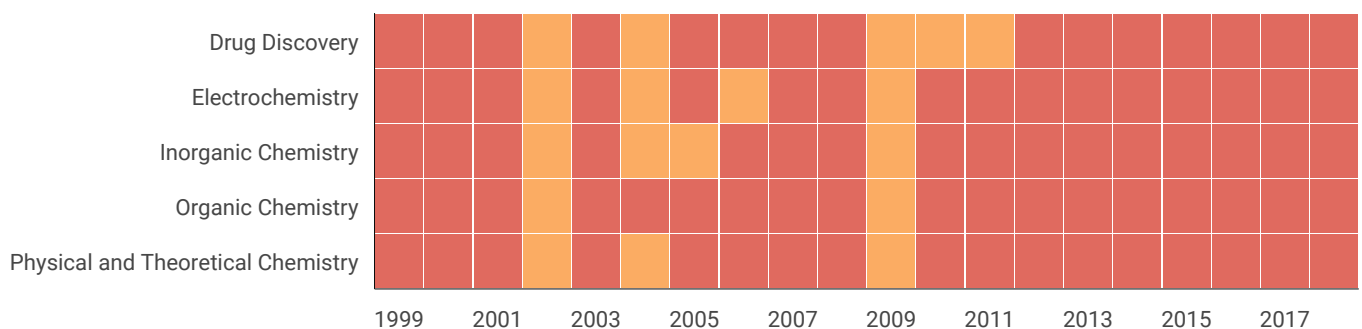
The Journal of the Indian Chemical Society publishes original, fundamental, theoretical, experimental research work of highest quality in all areas of chemistry, biochemistry, medicinal chemistry, electrochemistry, agrochemistry, chemical engineering and technology, food chemistry, environmental chemistry, etc.

[Homepage](#)[How to publish in this journal](#)[Contact](#)[Join the conversation about this journal](#)

31

H Index

Quartiles

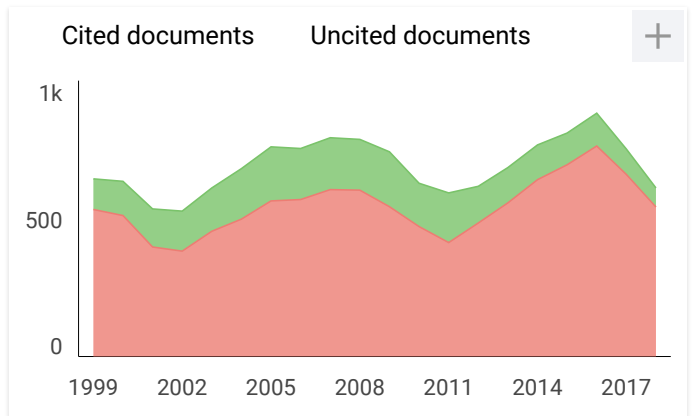
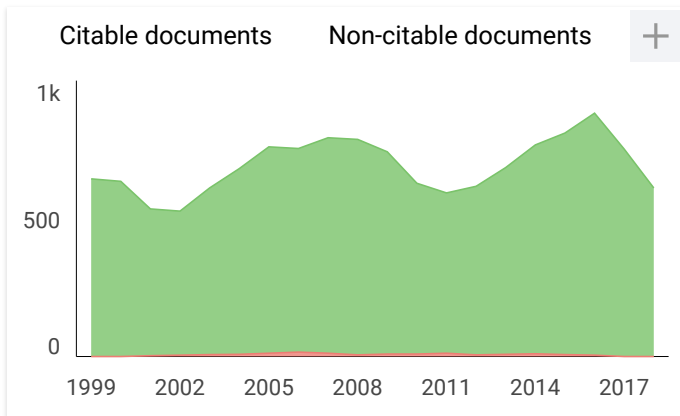
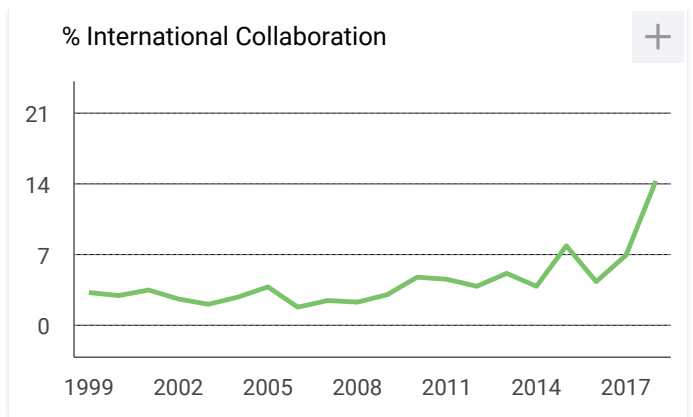
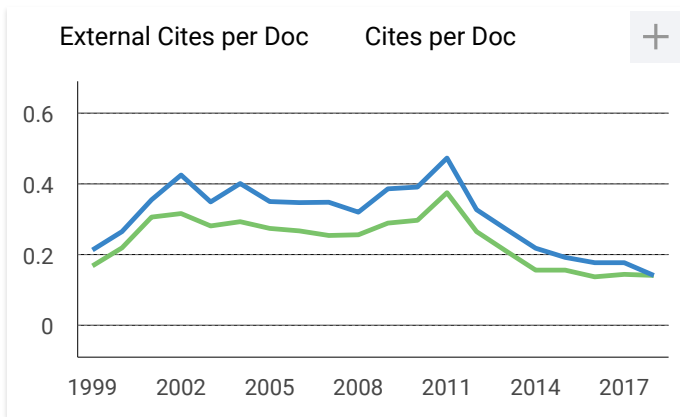
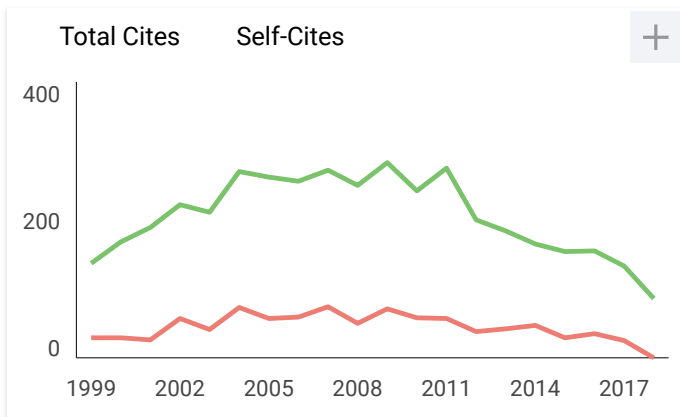
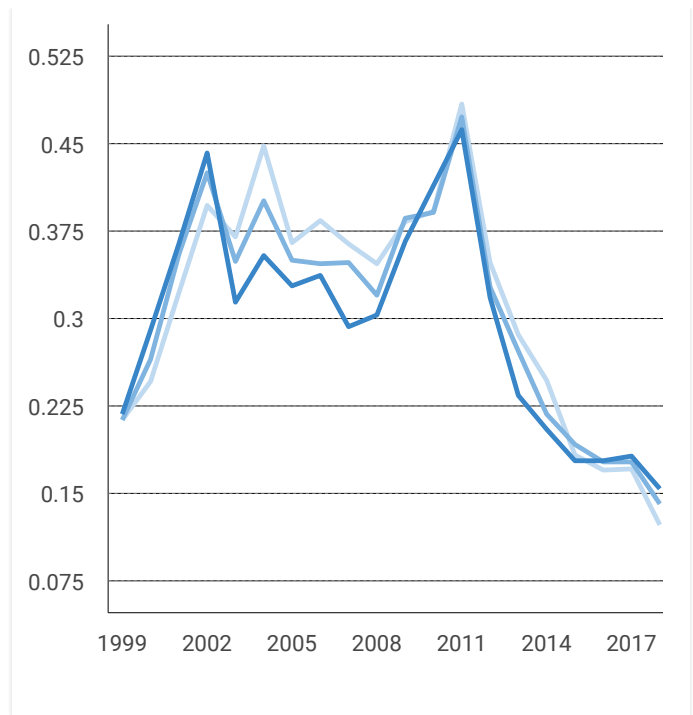
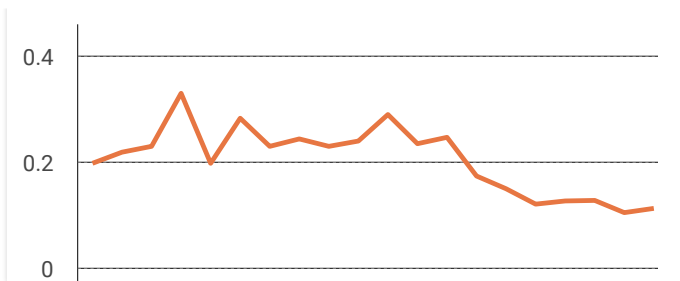


SJR



Citations per document





Journal of the Indian Chemical Society

← Show this widget in your own website

Q4 Drug Discovery
best quartile

SJR 2018 0.11

powered by scimagojr.com

Just copy the code below and paste within your html code:

```
<a href="https://www.scimaç
```




Isolation of cytotoxic sesquiterpenes from *Curcuma comosa* and characterization of their structures

Khun Nay Win Tun^{a,b}, Nanik Siti Aminah^{*a}, Alfinda Novi Kristanti^a, Rico Ramadhan^a, Yoshiaki Takaya^c and Hnin Thanda Aung^d

^aDepartment of Chemistry, Faculty of Science and Technology, Universitas Airlangga, Komplek Kampus C UNAIR, Jl. Mulyorejo, Surabaya, Indonesia

^bDepartment of Chemistry, Taunggyi University, Shan State (South), Myanmar

^cFaculty of Pharmacy, Meijo University, Tempaku, Nagoya, Japan

^dDepartment of Chemistry, Mandalay University, Mandalay, Myanmar

E-mail: nanik-s-a@fst.unair.ac.id

Manuscript received online 21 November 2019, revised and accepted 02 December 2019

Three guaiane-type sesquiterpenes named curcumenol (**1**), zedoarondiol (**2**), and (1*S*,4*S*,5*S*,10*R*)-isozedoarondiol (**3**) were isolated from the rhizomes of *Curcuma comosa*. Their structures were elucidated on the basis of extensive spectroscopic analysis. The cytotoxic activities of all isolated compounds were tested by MTT assay. Compounds **2** and **3** showed the most potent activities against T47D cell line with IC₅₀ values 12.13 and 10.93 µg/mL, respectively.

Keywords: *Curcuma comosa*, Zingiberaceae, guaiane-type sesquiterpenes, MTT assay.

Introduction

Curcuma comosa (Zingiberaceae), widely grown in tropical and subtropical area of Asia, including Thailand, Indonesia, Malaysia and Taunggyi¹⁻³ (Shan State of Myanmar). Taunggyi is the fifth largest city of Myanmar. Taunggyi has a humid subtropical climate. The climate usually comprises three seasons: hot summer, rainy monsoon, and cold winter.

In Taunggyi, the rhizome of *Curcuma comosa* is called **Sa-nwin-ga** and local people have used it as a traditional medicine for stomach ache, diabetes mellitus and hypertension. In Thailand, the rhizome of *C. comosa* is called **Waam chak mod luuk** and it has been used for the treatment of reproductive disorders in women, and for relief of unpleasant menopausal symptoms among postmenopausal women, and it is also widely used as an aromatic stomachic and anti-inflammatory agent. Several compounds have been isolated from the rhizomes of *C. comosa*. Three major groups of structures reported include sesquiterpenes, diarylheptanoids, and flavonoids glycosides. The structure of sesquiterpenes can be classified into five sub-groups: (i) Germacrane type sesquiterpene, (ii) Guaiane type sesquiterpene, (iii) Bisaborane

type sesquiterpene, (iv) Carabrane type sesquiterpene, and (v) Eudesmane type sesquiterpene. Pharmacological investigations on diarylheptanoids have displayed significant biological activities, including estrogenic, anti-bacteria, anti-inflammatory, and anti-osteoclastogenic properties³⁻¹⁰.

This research focused on chemical investigation of a methanolic extract of *C. comosa* resulting in isolation and structure elucidation of three guaiane-type sesquiterpenes. Moreover, cytotoxic activity of all the isolated compounds also evaluated.

Experimental

General:

All chemical solvents used were of analytical grade and were purchased in Surabaya, Indonesia. The solvents used for extraction and chromatography were distilled at their boiling points. Column chromatography (CC) was carried out on silica gel 60. TLC was carried out on silica gel 60GF₂₅₄ pre-coated plates (Merck). Melting points were determined using appropriate apparatus. The ¹H NMR and ¹³C NMR spectra were recorded at 600 and 151 MHz, in CDCl₃, methanol-

d_4 and the residual solvent peaks were used as internal standard. Chemical shifts are reported in parts per million (δ) and coupling constants in Hertz. NMR assignments were obtained from investigation of 1D and 2D experiment (^1H NMR, ^{13}C NMR, DQF-COSY, NOESY, HMBC, HSQC, and DEPT). JEOL JMS HX-110 mass spectrometer was used to obtain HRFAB-MS spectra. The infrared spectra (IR) were obtained on FT IR-8400S (Shimadzu) using KBr. A Buichi Rotary Evaporator with a high vacuum pump was used for evaporation of solvents under reduced pressure at 40°C.

Table 1. ^1H NMR data of compounds **1**, **2** and **3**

H	1 ^a	2 ^b	3 ^c
1	1.95 (m)	1.98 (m)	2.79 (m)
2	1.96 (m)	1.66 (m)	1.63 (m)
3	1.90 (m)	1.71 (m)	1.73 (m)
4	1.93 (m)	–	–
5	–	1.37 (d, 12.9 Hz)	2.02 (d, 12.9 Hz)
6	2.11 (d, 16.9 Hz), 2.66 (d, 16.9 Hz)	2.02 (m), 2.86 (d, 15.0 Hz)	2.52 (d, 13.9 Hz), 1.91 (d, 13.9 Hz)
7	–	–	–
8	–	–	–
9	5.77 (s)	2.51 (d, 12.7 Hz) 2.98 (d, 12.7 Hz)	2.30 (dd, 16.1 Hz, 1.2 Hz) 3.34 (d, 16.1 Hz)
10	–	–	–
11	–	–	–
12	1.60 (s)	1.90 (s)	1.97 (s)
13	1.82 (s)	1.86 (s)	1.88 (s)
14	1.03 (d, 6.4 Hz)	1.17 (s)	1.39 (s)
15	1.67 (s)	1.11 (s)	1.19 (s)

^aSpectra recorded at 600 MHz (CDCl_3), ^{b,c}Spectra recorded at 600 MHz (methanol- d_4).

Plant material:

The rhizomes of *Curcuma comosa* were collected in September 2016 from Taunggyi, Shan State, Myanmar.

Extraction and isolation:

The air-dried rhizomes of *Curcuma comosa* (1 kg) were successively extracted with methanol (3000 mL) at room temperature for three weeks. After concentration of the solvent under reduced pressure, the methanol extract (41.16 g) was partitioned with hexane/MeOH (100 mL×3, v/v). MeOH extract (15 g) was separated by vacuum liquid chromatography (VLC), eluting with hexane/EtOAc (100:0, 95:5, 80:20, 70:30, 0:100) step gradient as eluents to afford five fractions

(CT-1 to CT-5). Fraction CT-2 (438 mg) was purified over silica gel by column chromatography, eluting with solvent mixtures of hexane/EtOAc (100:0, 90:10, 80:20, 0:100) to yield compound **1** (250.6 mg). Fraction CT-4 (1 g) was subjected to column chromatography using hexane/EtOAc (90:10, 80:20, 70:30, 0:100) step gradient solvent mixtures as eluents to give compounds **2** (150.2 mg) and **3** (10 mg).

MTT assay:

Cytotoxicity of the isolated compounds on HeLa and T47D cells were carried out, using MTT assay *in vitro*. The isolated compounds were dissolved in DMSO (100 μL) to obtain various concentrations. Doxorubicin was used as a positive control. The media control solution consists of media culture and cell control solutions consist of culture and cell media. Sufficient amount of HeLa cells and T47D cells (213.4×10^4 cells/well and 167×10^4 cell/well) solution were prepared and 100 μL were placed in each well in a 96-well plates. The plates containing the cancer cells were treated with compound and incubated for 24 h. The cells were washed and treated by the 100 μL MTT per well. Plates were incubated at 37°C in a 5% CO_2 atmosphere for 4 h, and 0.1 mL of the extraction buffer (10% sodium dodecyl sulfate in 0.01% HCl) was added. After an overnight incubation at 37°C, the absorbance was measured at 595 nm using an ELISA reader and the results were compared with the control cultures without compound. To determine cell viability, percent viability was calculated by the formula

$$\% \text{ Viability} = \frac{A_t - A_{mc}}{A_{ng} - A_{mc}} \times 100$$

where, A_t = absorbance of treatment, A_{mc} = absorbance of media control and A_{nc} = absorbance of negative control.

Results and discussion

Chromatographic isolation work on the extracts of *Curcuma comosa* successfully yielded three guaiane-type sesquiterpenes, curcumenol (**1**), zedoarondiol (**2**), and (1S,4S,5R,10R)-isozedoarondiol (**3**). Their structures were elucidated by extensive spectroscopic techniques and by comparison with the data reported in the literature.

Compound (**1**) was obtained as colorless crystals. Its melting point was 98–100°C. The IR spectrum showed the absorption band for O-H stretching vibration at 3371 and 3321 cm^{-1} , C=C stretching vibration at 1695 and 1658 cm^{-1} , C-C-

O stretching vibration of alcohol group at 1274 cm^{-1} . The proton NMR spectrum of compound (**1**) revealed the presence of four methyl singlet signal peaks including a doublet peaks at δ_{H} 1.03 (6.4 Hz) and three singlet peaks at δ_{H} 1.60, 1.67 and 1.82. Three methylene signal peaks including two multiplet peaks at δ_{H} 1.97 and 1.90, and a doublet peak at 2.66/2.11 (16.9 Hz), and two methine multiplet signals at 1.91 and 1.95 were also observed. In addition, there were an olefinic singlet signal peak at δ_{H} 5.77 and a hydroxyl broad singlet signal peak at δ_{H} 6.06. ^{13}C NMR along with DEPT 135 and 90 experiments allowed the identification of four sp^3 methyl carbon, three sp^3 methylene carbons, two sp^3 methine carbons, one sp^2 methine carbon and a carbonyl carbon, respectively. Detailed structure of compound (**1**) was established by ^1H - ^1H COSY and ^1H - ^{13}C HMBC experiments (Figs. 1a and 1b). The two methyl signals showed HMBC correlation of H-12 (δ_{H} 1.60) with C-13/C-7/C-11, H-13 (δ_{H} 1.82) with C-12/C-7/C-11. An olefinic proton at δ_{H} 5.77 also showed long-range HMBC correlation to C-1 (δ_{C} 51.3), C-8 (δ_{C} 101.5) and C-15 (δ_{C} 20.9). Furthermore, the HMBC correlation showed one methyl doublet signal at δ_{H} 1.03 (d, 6.4 Hz) with C-1 (δ_{C} 51.3), C-9 (δ_{C} 125.6) and C-10 (δ_{C} 139.2), and another singlet methyl at δ_{H} 1.67 with C-1 (δ_{H} 51.3), C-9 (125.6) and C-10 (139.2). The configuration of compound (**1**) was considered the same with the previous data based on NOE correlations (Fig. 1c), as well as 1D, 2D NMR, and MS spectral data. Thus, based on above data, compound (**1**) was identified as curcumenol^{11–14}.

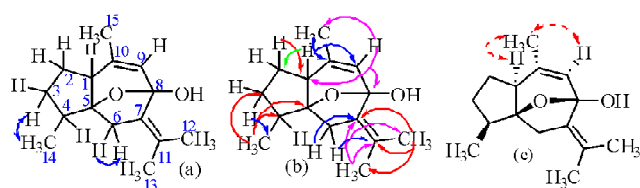


Fig. 1. DQF-COSY (a), HMBC (b), and NOESY (c) correlation in compound (**1**).

Compound (**2**) was obtained as yellow oil. The IR spectrum showed the presence of hydroxyl (3387 cm^{-1}), conjugated ketone (1708 and 1664 cm^{-1}), and olefinic (1694 cm^{-1}) groups. The proton NMR spectrum of compound (**2**) revealed the presence of four tertiary methyl signals at δ_{H} 1.11, 1.17, 1.86 and 1.90 (each 1H, s), four set of methylene protons at δ_{H} 1.66 (2H, m) 1.71 (2H, m), 2.02 (1H, m)/2.86

(1H, d, 15 Hz), 2.51 (1H, d, 12.7)/2.98 (1H, d, 12.7), two methine proton at δ_{H} 1.37 (1H, d, 12.9 Hz), 1.98 (1H, m). The ^{13}C NMR and DEPT 135 and 90 spectra revealed the presence of four methyl carbon signals (δ_{C} 18.7, 20.9, 21.1, 21.7), four methylene carbon signals (δ_{C} 21.5, 28.1, 38.7, 59.5), two oxygenated carbon signals (δ_{C} 71.8, 79.1), two methine carbons signals (δ_{C} 51.5, 55.7), two olefinic quaternary carbon signals (δ_{C} 135.1, 141.8) and a carbonyl signal (δ_{C} 204.3), respectively. Based on the above spectra data, compound (**2**) might be a guaiane-type sesquiterpenes. This assumption was further determined through a variety of 2D NMR spectroscopic techniques. In the ^1H - ^1H COSY correlations (Fig. 2a), the proton signal at δ_{H} 2.02 was coupled with the proton signal at δ_{H} 2.86; the proton signal at δ_{H} 2.51 was coupled with the proton signal at δ_{H} 2.98; the proton signal at δ_{H} 1.37 was coupled with the proton signal at δ_{H} 1.98; and the proton signal at δ_{H} 1.66 was coupled with the proton signal at δ_{H} 1.71, respectively. Furthermore, the ^1H - ^{13}C HMBC long range correlation (Fig. 2b) of H-12 with C-7/C-8/C-11, H-13 (δ_{H} 1.86) with C-7/C-8/C-11, H-14 with C-3/C-4/C-5, H-15 with C-1/C-9/C-10, H-5 with C-1/C-4/C-7/C-10, H-6a with C-1/C-4/C-5/C-7/C-8/C-11, H-9a with C-1/C-7/C-8/C-10, H-9b with C-1/C-8/C-10/C-15, respectively. The relative configuration of compound (**2**) was considered to be the same with previous data^{1,15,16}, based on the NOE correlations (Fig. 2c) between H-5 and H-15, H-1 and H-14. Based on these spectra data and comparison with the previous report, compound (**2**) was identified as zedoarondiol^{1,15,16}.

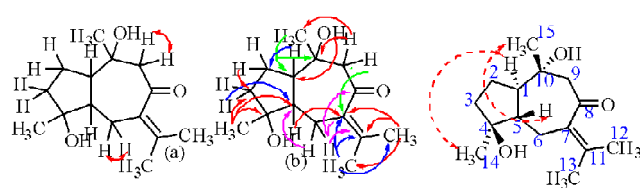


Fig. 2. DQF-COSY (a), HMBC (b), and NOESY (c) correlation in compound (**2**).

Compound (**3**) was also obtained as yellow oil. The IR spectrum showed absorption band due to conjugated ketone (1701 and 1665 cm^{-1}), a double bond (1612 cm^{-1}) and a hydroxyl group (3394 cm^{-1}). The ^1H NMR spectrum of the compound (**3**) exhibited characteristic signals for guaiane-type sesquiterpenes skeleton, including four methyl protons at δ_{H} 1.19, 1.39, 1.88, 1.97 (each 3H, s), two methine pro-

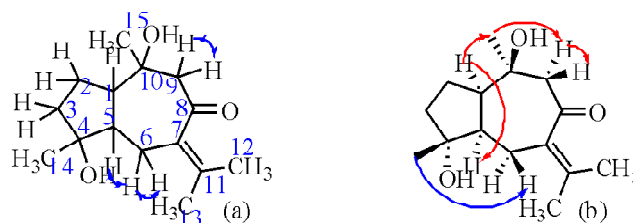
Table 2. ^{13}C NMR of compounds **1**, **2** and **3**

C	1 ^a	2 ^b	3 ^c
1	51.3	55.7	51.2
2	27.6	21.5	24.6
3	31.2	38.7	36.0
4	40.4	79.1	81.7
5	85.7	51.5	52.6
6	37.3	28.1	27.0
7	137.4	135.1	134.0
8	101.5	204.3	204.7
9	125.6	59.5	49.9
10	139.2	71.8	72.5
11	122.2	141.8	143.0
12	22.3	21.7	20.9
13	18.9	20.9	21.8
14	11.8	21.1	23.4
15	20.9	18.7	31.3

^aSpectra recorded at 151 MHz (CDCl_3), ^{b,c}Spectra recorded at 151 MHz (methanol- d_4).

tons at δ_{H} 2.02 (1H, d, 12.9 Hz) and 2.79 (1H, m), four methylene protons at δ_{H} 1.63 (2H, m), 1.73 (2H, m), 2.52 (1H, d, 13.9 Hz)/1.91 (1H, d, 13.9 Hz), 3.34 (1H, d, 16.1 Hz)/2.30 (1H, dd, 16.1, 1.2 Hz). The ^{13}C NMR and HSQC experiment revealed the presence of 15 carbon signals due to one carbonyl function (δ_{C} 204.7), two sp^2 quaternary carbons (δ_{C} 134.0, 143.0), two sp^3 quaternary carbonyl (δ_{C} 72.5, 81.7), two sp^3 methine carbon (δ_{C} 51.2, 52.6), four sp^3 methylene (δ_{C} 27.0, 24.6, 36.0, 49.9) and four sp^3 methyl carbons (δ_{C} 20.9, 21.8, 23.4, 31.3). HMBC analysis exhibited correlation of H-15 with C-5 and C-10; H-14 with C-1, C-3, C-4 and C-5; H-13 with C-7, C-8, C-11 and C-12; H-12, C-7, C-8, C-11, C-13; H-9a with C-1, C-8 and C-10; H-9b with C-1, C-7, C-8 and C-10; H-6a with C-1, C-4, C-7 and C-11; H-6b with C-5, C-7 and C-11; H-5 with C-2, C-3 and C-4; H-1 with C-2 and C-10. In the ^1H - ^1H COSY spectrum, the following cross peak correlations were observed (Fig. 3a): the proton signal at δ_{H} 3.34 was correlated with the proton signal at δ_{H} 2.30; the proton signal at δ_{H} 2.52 was correlated with the proton signal at δ_{H} 1.91; the proton signal at δ_{H} 2.02 was correlated with the proton signal at δ_{H} 1.63; the proton signal at δ_{H} 3.34 was correlated with the proton signal at δ_{H} 1.73, respectively. In nuclear overhauser effect spectroscopy (NOESY) spectrum, the signal at δ 2.79 (H-1) had NOE enhancements

with the signals at δ 1.19 (Me-15) and δ 2.02 (H-5). And NOE effects of Me-14 (δ 1.39) with H-1 (δ 2.79) and H-6 (δ 2.51) were also observed (Fig. 3b). With the aid of above data and previous report compound (**3**) was named as (1*S*,4*S*,5*S*,10*R*)-isozedoarondiol^{16,17}.

**Fig. 3.** DQF-COSY (a) and NOE (b) correlation in compound (**3**).

All isolated compounds (**1-3**) were examined for their cytotoxic activities against HeLa and T47D cell lines. Compound **2** and **3** showed the most potent activities against T47D cell line with IC_{50} values of 12.13 and 10.93 mg/mL.

Table 3. Cytotoxic activity of compounds (**1-3**) against HeLa and T47D

Compd.	IC_{50} ($\mu\text{g/mL}$)	
	HeLa	T47D
1	142.67	213.55
2	99.83	12.13
3	170.83	10.93
Doxorubicin	2.69	0.04

Biosynthesis pathway to compound (**1-3**) have been reported earlier¹⁸.

Conclusions

In summary, this paper described the isolation and structure elucidation of three quaiane-type sesquiterpenes, namely curcumenol (**1**), zedoarondiol (**2**), and (1*S*,4*S*,5*S*,10*R*)-isozedoarondiol (**3**). The cytotoxic activity of all isolated compounds against two cancer cell lines was also reported. Compounds **2** and **3** showed the most potent activities against T47D cell lines with IC_{50} values 12.13 and 10.93 mg/mL.

Acknowledgements

This work was supported by the Ailangga Development Scholarship (ADS) and "RISET MANDAT GRANT" of Universitas Airlangga, Surabaya, Indonesia.

Supporting information

Supporting information accompanies this paper are described.

References

1. R. Chockchaisiri, P. Pimkaew, P. Piyachaturawat, R. Chalermglin and A. Suksamrarn, *Rec. Nat. Prod.*, 2014, **8(1)**, 46.
2. A. Suksamrarn, M. Ponglikit-mongkol, K. Wongkrajang, A. Chindaduang, S. Kittidanairak, A. Jankam, B. Yingyongnarongkul, N. Kittipanumat, R. Chokchaisiri, P. Khetkam and P. Piyachaturawat, *Bioorg. Med. Chem.*, 2008, **16(14)**, 6891.
3. F. Xu, S. Nakamura, Y. Qu, H. Matsuda, Y. Pongpiriyadacha, L. Wu and M. Yoshikawa, *Chem. Pharm. Bull.*, 2008, **56(12)**, 1710.
4. N. Sornkaew, Y. Lin, F. Wang, G. Zhang, R. Chokchaisiri, A. Zhang, K. Wongkrajang, P. Suebsakwong, P. Piyachaturawat and A. Suksamrarn, *Nat. Prod. Commun.*, 2015, **10(1)**, 89.
5. Y. Qu, F. Xu, S. Nakamura, H. Matsuda, Y. Pongpiriyadacha, L. Wu and M. Yoshikawa, *J. Nat. Med.*, 2009, **63(1)**, 102.
6. T. M. Jurgnes, E. G. Frazier, J. M. Schaeffer, T. E. Jones, D. L. Zink and R. P. Borris, *J. Nat. Prod.*, 1994, **57(2)**, 230.
7. T. Matsumoto, S. Nakamura, S. Nakashima, M. Yoshikawa, K. Fujimoto, T. Ohta, A. Morita, R. Yasui, E. Kashiwazaki and H. Matsuda, *Bioorg. Med. Chem. Lett.*, 2013, **23(18)**, 5178.
8. V. Keeratinijakal and S. Kongkiatpaiboon, *Rev. Bras. Farmacogn.*, 2017, **27(3)**, 290.
9. C. Tipbunjong, Y. Kitiyanant, G. Chaturapanich, N. Sornkaew, A. Suksamrarn, N. Kitiyanant, K. A. Esser and C. Pholpramool, *Med. Chem. Res.*, 2016, **26(1)**, 274.
10. R. Chokchaisiri, P. Innok and A. Suksamrarn, *Phytochem. Lett.*, 2012, **5(2)**, 361.
11. O. A. A. Hamdi, L. J. Ye, M. N. A. Kamarudin, H. Hazni1, M. Paydar, C. Y. Looi, J. A. Shilpi, H. A. Kadir and K. Awang, *Rec. Nat. Prod.*, 2015, **9(3)**, 349.
12. P. Sugita, S. O. Firdaus, A. Ilmiawati and D. U. C. Rahayu, *J. Chem. Pharm. Res.*, 2018, **10(6)**, 68.
13. D. Wang, W. Huang, Q. Shi, C. Hong, Y. Cheng, Z. Ma and H. Qu, *Nat. Prod. Commun.*, 2008, **3(6)**, 861.
14. M. A. Sukari, T. S. Wah, S. M. Saad, N. Y. Rashid, M. Rahmani, N. H. Lajis and T. Y. Hin, *Nat. Prod. Res.*, 2010, **24(9)**, 838.
15. I. Takano, I. Yasuda, K. Takeya and H. Itokawa, *Phytochemistry*, 1995, **40(4)**, 1197.
16. M. Kuroyanagi, A. Ueno, K. Ujiie and S. Sato, *Chem. Pharm. Bull.*, 1987, **35(1)**, 53.
17. N. Jia, M. X. Chi, X. X. Lan, J. H. Wei and G. Dean, *Magn. Reson. Chem.*, 2008, **46(2)**, 178.
18. H. Hikino, C. Konno and T. Takemoto, *Chem. Pharm. Bull.*, 1972, **20(5)**, 987.

