

Prevalence of Helicobacter pylori genotypes and their association with upper gastrointestinal diseases: A cross sectional study in southern IranOriginal Submission
Muhammad Miftahussurur, M.D., Ph.D

Back

Edit Review

Print

Submit Review to Editorial Office

Recommendation: Reject**Transfer Authorization**

If this submission is transferred to another publication, do we have your consent to include your identifying information?

Response

No

If this submission is transferred to another publication, do we have your consent to include your original review?

No

Custom Review Question(s):

Publons Reviewer Recognition. Springer Nature can send verification of this review directly to Publons (a subsidiary of Clarivate Analytics). If you would like to take advantage of this service, please click on the "Yes" option below. Your name, email address, title of the reviewed manuscript, name of the journal, and date of your review submission (the "Review Data") will then be transmitted to Publons upon publication of the manuscript. If you have already registered at Publons, they will notify you of the receipt of this review and update your profile as per your settings and their policy. If you are not registered with Publons, you will receive an email from them asking you to register in order for them to be able to recognize your review on your new profile page. Publons may use the Review Data to generate derivative metadata for the benefit of Publons and you as a reviewer, carefully considering the sensitivity of such information. For example, Publons may verify your record as a reviewer by updating your profile published on its webservice if you have registered for such service or help editors to identify candidate reviewers. Please find the details of processing in Publons' privacy policy <https://publons.com/about/terms>

Response

No

Level of interest

Please indicate how interesting you found the manuscript:

An article of limited interest

Quality of written English

Please indicate the quality of language in the manuscript:

Needs some language corrections before being published

Declaration of competing interests

Please complete a declaration of competing interests, considering the following questions:

I declare that I have no competing interests

1. Have you in the past five years received reimbursements, fees, funding, or salary from an organisation that may in any way gain or lose financially from the publication of this manuscript, either now or in the future?
2. Do you hold any stocks or shares in an organisation that may in any way gain or lose financially from the publication of this manuscript, either now or in the future?
3. Do you hold or are you currently applying for any patents relating to the content of the manuscript?
4. Have you received reimbursements, fees, funding, or salary from an organization that holds or has applied for patents relating to the content of the manuscript?
5. Do you have any other financial competing interests?
6. Do you have any non-financial competing interests in relation to this paper?

If you can answer no to all of the above, write 'I declare that I have no competing interests' below. If your reply is yes to any, please give details below.

Reviewer Comments to Author

1. Although there is a correlation between cagA and hiatal hernia, and also the normal esophageal finding is correlated with glmM genotype, the author failed to explain the possible mechanisms of this relationship. Whereas association is not the same as causation, perhaps, this is just a coincidence factor. However, if this is not a coincidence, the relationship in between them should be explained. For example, the presence of vacAs1bm1 and vacAs1bm2 had a significant association with gastric erythema; according to this passage, the relationship in between them can still be understood. This result may be influenced by the use of endoscopic method, instead of histology. Hence, result could be misleading.
2. It is necessary to carry out a multivariate analysis to rule out any accidental relationships. Multivariate analysis could be done by determining and entering significant variables.
3. Discussion section need to described relevant reasons within the study, rather than explaining results or comparing with other studies. In addition, please state the study limitation.

Gut Pathogens

Prevalence of Helicobacter pylori genotypes and their association with upper gastrointestinal diseases: A cross sectional study in southern Iran

--Manuscript Draft--

Manuscript Number:	GUTP-D-20-00232
Full Title:	Prevalence of Helicobacter pylori genotypes and their association with upper gastrointestinal diseases: A cross sectional study in southern Iran
Article Type:	Research
Abstract:	<p>Background</p> <p>Investigating the prevalence of vacuolating cytotoxin (vacA), cytotoxin associated gene A (cagA), glm M genotypes, and subtypes of vacA of Helicobacter pylori (H. pylori) isolate in Jahrom, Southern Iran. DNA extracted from H. pylori samples retrieved from gastric biopsy isolated from 113 dyspeptic patients with positive rapid urease test (RUT). Genotyping was done by polymerase chain reaction (PCR) technique, using primers for vacA (s1a, s1b, s1c, s1, s2, m2, and m1), cagA, and glmM. Endoscopy was done for all the patients to screen upper gastrointestinal (GI) disorders.</p> <p>Results</p> <p>GlmM was detected in 100% of the cases. VacA subtypes s1am2, s2m2, s1a, s1b, and s1c were detected in 27.9%, 25.6%, 50%, 3.5% and 2.4% of the isolates, respectively, while cagA was detected in 60.5% of the isolates. VacA alleles m1, s1, and s2 were detected in 54%, 50%, and 44% of isolates respectively. Also, 60.5% of the isolates were cagA-vacA-positive. A significant correlation was observed between vacAs1bm1 and gastroesophageal reflux disease (GERD) and glmM and normal esophagus. The presence of vacAs1bm1 and vacAs1bm2 has a significant association with gastric erythema. The presence of cagA showed a significant association with normal esophagus and hiatal hernia.</p> <p>Conclusions</p> <p>In our research, the number of glmM and cagA positive isolates is higher among other genotypes and cagA is correlated with hiatal hernia, and normal esophageal finding is correlated with glmM genotype. There was no association between age or sex of the patients and bacterial genotype.</p>

[Click here to view linked References](#)

1 **Prevalence of Helicobacter pylori genotypes and their association with upper**
2 **gastrointestinal diseases: A cross sectional study in southern Iran**

3 Rahim Raufi ¹, Reza Shahriarirad ^{2,3}, SeyedehMaryam Pishva ³ and Nikta Taghipour ^{4*}

4 ¹ Department of Infection Disease, Jahrom University of Medical Sciences, Jahrom, Iran

5 ² Thoracic and Vascular Surgery Research Center, Shiraz University of Medical Sciences,
6 Shiraz, Iran

7 ³ Student Research Committee, Shiraz University of Medical Sciences, Shiraz, Iran

8 ⁴ Student Research Committee, Jahrom University of Medical Sciences, Jahrom, Iran

9
10 * Correspondence: Nikta Taghipour, Student Research Committee, Jahrom University of
11 Medical Sciences, Jahrom, Iran; Email: Niktataghipoor114@gmail.com .

12
13 **Abstract**

14 **Background:** Investigating the prevalence of vacuolating cytotoxin (vacA), cytotoxin
15 associated gene A (cagA), glm M genotypes, and subtypes of vacA of Helicobacter pylori (H.
16 pylori) isolate in Jahrom, Southern Iran. DNA extracted from H. pylori samples retrieved from
17 gastric biopsy isolated from 113 dyspeptic patients with positive rapid urease test (RUT).
18 Genotyping was done by polymerase chain reaction (PCR) technique, using primers for vacA
19 (s1a, s1b, s1c, s1, s2, m2, and m1), cagA, and glmM. Endoscopy was done for all the patients
20 to screen upper gastrointestinal (GI) disorders.


21 **Results:** GlmM was detected in 100% of the cases. VacA subtypes s1am2, s2m2, s1a, s1b,
22 and s1c were detected in 27.9%, 25.6%, 50%, 3.5% and 2.4% of the isolates, respectively,

23 while *cagA* was detected in 60.5% of the isolates. *VacA* alleles m1, s1, and s2 were detected
24 in 54%, 50%, and 44% of isolates respectively. Also, 60.5% of the isolates were *cagA-vacA*-
25 positive. A significant correlation was observed between *vacAs1bm1* and gastroesophageal
26 reflux disease (GERD) and *glmM* and normal esophagus. The presence of *vacAs1bm1* and
27 *vacAs1bm2* has a significant association with gastric erythema. The presence of *cagA* showed
28 a significant association with normal esophagus and hiatal hernia.

29 **Conclusions:** In our research, the number of *glmM* and *cagA* positive isolates is higher among
30 other genotypes and *cagA* is correlated with hiatal hernia, and normal esophageal finding is
31 correlated with *glmM* genotype. There was no association between age or sex of the patients
32 and bacterial genotype.

33 **Keywords:** Prevalence; *Helicobacter pylori*; genotype; Gastrointestinal diseases; Iran

35 Background

36  *Helicobacter pylori* (*H. pylori*) is a gram-negative bacillus and one of the most common
37 bacterial infections which affect nearly half of the world's population, which has naturally
38 colonized humans for at least 100,000 years, and probably throughout human evolution [1]. *H.*
39 *pylori* colonize the stomach in approximately 50% of the world's human population. In the
40 United States and other developed countries, the prevalence of *H. pylori* is approximately 30%
41 of the population [2]. Moreover, DNA-level analyses have indicated that *H. pylori* are one of
42 the most genetically diverse bacterial species [3].

43 The sequencing of the *H. pylori* genome in 1997 has led to considerable progress in the
44 understanding of the biology of this organism. *H. pylori* are considered an important etiological
45 agent in the development of gastritis, peptic ulcers, and gastric carcinoma [4, 5]. Colonization
46 with this organism is the main risk factor for peptic ulceration as well as for gastric

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47 adenocarcinoma and gastric mucosa-associated lymphoid tissue (MALT) lymphoma [6]. H.
48 pylori infection is virtually always associated with chronic active gastritis, but less than 15%
49 of patients develop peptic ulcer disease (PUD), gastric adenocarcinoma, or gastric lymphoma
50 [7]. Studies suggested that more than 90% of duodenal ulcers (DU) and 85% of gastric ulcers
51 (GU) are associated with H. pylori [7]. There is a strong connection between H. pylori and
52 gastric lymphoma. H. pylori antigen stimulates T-cells which promotes B-cell proliferation,
53 leading to low-grade gastric MALT lymphoma. Eradication of H. pylori can result in regression
54 or cure of this lymphoma [8].

55 The clinical outcome following infection with this pathogen has been related to environmental
56 conditions, host immunological factors, and microorganism virulence [9]. H. pylori genotypes
57 and their geographic distribution are linked to the severity of PUD [10, 11]. The VacA protein
58 induces vacuolation and apoptotic processes in epithelial cells, as well as immunosuppressive
59 actions in immunological cells [12].

60 The vacA, cagA, and glmM are the most commonly studied virulence factors of H. pylori, and
61 in this study, we investigate the prevalence of the genotypes and their association with GI
62 pathologies.

63 The clinical outcome of this bacterial infection seems to be influenced by the distribution of
64 the above-mentioned pathogenic factors in H. pylori strains [13]. Due to the lack of H. pylori
65 genotyping in southern Iran, in this study we investigate H. pylori genotyping in Jahrom,
66 southern Iran. Furthermore, we aim to establish the main virulence strain and its association
67 with clinical outcomes.

68

69 **Results**

70 A total of 113 patients with an average age of 39.15 (SD:16.18, range 13-89) were studied
71 which consisted of 44 (38.9%) male and 69 (61.1%) female patients. Furthermore, 15.9% of

72 the patients were smokers. Based on PCR evaluation for H. pylori, 86 samples out of 113
 73 (76.1%) patients were positive. Table 1 demonstrates the study population characteristics.

74

75 **Table 1** Characteristic of patients with persistent gastrointestinal problems in our study

Variable	Frequency n=113	Percentage
Gender	Male	44 38.9
	Female	69 61.1
Positive PCR for H.Pylori		86 76.1
Endoscopy finding	NUD	52 46
	Erosive gastropathy	47 41.6
	GU	3 2.7
	DU	11 9.7
Erythema in NUD	Antrum	45 39.8
	Antrum with extension to body or fundus	6 5.3
Edema in NUD	Antrum	4 5.3
	Antrum with extension to body or fundus	2 1.8
Nodularity	Antrum	5 4.4
	Antrum with extension to body or fundus	0 0
Esophageal finding	Esophagitis	9 8
	GERD	5 4.4
	Hiatal hernia	57 50.4
	Normal	42 37.2

PCR: polymerase chain reaction; H, pylori: Helicobacter Pylori; NUD: non-ulcer dyspepsia; GU: Gastric ulcer; DU: Duodenal ulcer; GERD: Gastroesophageal reflux disease;

76
1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

77 Vac A gene was present in all H. pylori samples and was accompanied by the Cag A gene in
78 60.5% of samples. Table 2 demonstrates the result of different genotypes that we found in our
79 samples.

80
81 **Table 2** Distribution of H. pylori genotypes in southern Iran n=86

Genotypes	Positive		Negative	
	Frequency	Percentage	Frequency	Percentage
Vac Am1	39	45.3	47	54.7
Vac Am2	47	54.7	39	45.3
Vac As1a	43	50	43	50
Vac As1b	3	3.5	83	96.5
Vac As1c	2	2.3	84	97.7
Vac As2	38	44.2	48	55.8
Cag a	52	60.5	34	39.5
Glm m	86	100	0	0

82
83 The vac A gene and its subtypes distribution are shown in table 3. The most common genotypes
84 were vac As1am2 with a prevalence of 28% then vac As2bm2 and vac As1am1.

85
86 **Table 3** Distribution of the vac A gene and its subtypes in southern Iran.

Genotypes	Positive		Negative	
	Frequency	Percentage	Frequency	Percentage
vacAs1am1	19	22.1%	67	77.9
vacAs1am2	24	27.9	62	72.1

vacAs1bm1	2	2.3	84	97.7
vacAs1bm2	1	1.2	85	98.8
vacAs1cm1	1	1.2	85	98.8
vacAs1cm2	1	1.2	85	98.8
vacAs2m1	16	18.6	70	81.4
vacAs2m2	22	25.6	64	74.4

87

88 The patients were divided into 4 groups based on their endoscopic findings: a) Non-
89 ulcer dyspepsia (NUD): no erosion, but erythema, edema, or nodularity was observed. The
90 location of these findings in the stomach was also studied. b) Erosive gastropathy c) Gastric
91 Ulcer (GU) d) Duodenal ulcer (DU).

92 Out of 52 patients with NUD, 51 of them had gastric erythema, 6 of them had gastric
93 edema and 5 patients had gastric nodularity. The erythema was located in the antrum for 39.8%
94 of them. Out of 6 patients with edema, 5.3% had antrum involvement.

95 Esophageal findings of patients are shown in table 1. Esophagitis in 8% of the cases,
96 Gastroesophageal reflux in 4.4% of the cases, and hiatal hernia in 50.4% of the cases.

97 Table 4 shows the relationship between the different H. pylori genotypes and age, gender,
98 smoking, gastric endoscopy, and esophageal findings.

99 **[INSERT TABLE 4]**

100 There was no significant relationship found between the age, gender, smoking of the patients,
101 and the genotype of H. pylori they were colonized with. However, the Vac As1cm1 genotype
102 had the highest mean age (70 ± 0) and the vacAs1am1 gene had the lowest mean age ($33.6 \pm$
103 10.8). Also, there is no relationship between gastric endoscopy finding and the genotype of H.
104 pylori.

105 Based on table 4, there is a significant correlation among Vac As1bm1 genotype and GERD
106 (p=0.002); Cag A genotype and hiatal hernia (p=0.02); Cag A genotype and normal esophagus;
107 Glm M genotype and normal esophagus (p=0.02). We found that genotype vac As1bm2 is
108 linked with gastric edema (P>0.001); however, there is only 1 person in this group.

109

110 **Discussion**

111 H. pylori genotypes, environmental and epidemiological factors can play a role in its
112 pathogenicity. The genetic diversity and epigenetic modifications of H. pylori give rise to the
113 different levels of pathogenicity. Several studies have shown that the incidence and/or severity
114 of gastroduodenal pathologies related to H. pylori may vary between geographic areas, such as
115 studies have shown genotype of H. pylori in Europe differs from South-East Asia but is the
116 same for the North of Iran and Uzbekistan [14, 15]. The predominant H. pylori strain circulating
117 among geographic locations differs with regard to the genomic structure [16]. The present
118 study reports common H. pylori genotypes in Jahrom, Iran, and their clinical relevance.

119 The vacA gene was present in all H. pylori strains and it is a useful marker in predicting disease
120 outcome [17]. The vacA strain's structure determines its in vitro cytotoxic activity, with m1
121 vacA type being more active than m2 type, s1a more active than s1b, and s2 vacA not producing
122 detectable activity [18]. We found an apparent correlation between vacAs1bm1 positive cases
123 and GERD (P=0.002). Genotype vacA s1/m2 is a dominant H.pylori genotype in Iran, but no
124 relationships were found between these genotypes and clinical outcomes. In this study, we
125 found the vac A gene of H. pylori has 8 different combinations with its subtypes, and their
126 prevalence are; vacAs1am1 (22.1%), vacAs1am2 (27.9%), vacAs1bm1 (2.3%), vacAs1bm2
127 (1.2%), vacAs1cm1 (1.2%), vacAs1cm2 (1.2%), vacAs2m1 (18.6%) and vacAs2m2 (25.6%).
128 The most common genotype is vac A s1am2 with a prevalence of approximately 28%;
129 however, S1m1 was the most common genotype in some research studies conducted on

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

130 Afghan, Iranian, Turkish, and Thai patients [19-22], however in our research prevalence of
131 s1m1(22.1%) is the third common genotype. A study from Shiraz, southern Iran reported that
132 vacA-positive strains were more frequently found in PUD patients than in NUD patients [23].
133 Another study in Tehran reported that the vacA s1 genotype was detected in 79% and 68% of
134 patients with PUD and NUD, respectively [24]. However, in our study, we didn't observe any
135 correlation between any vacA subtypes and PUD or NUD.

136 The cagA toxin can induce severe inflammation of gastric mucosa and is related to peptic ulcer
137 and gastric cancer [18]. Based on literature regarding H. pylori genotype in Iran and around
138 the world, the prevalence of cag A genotype varies in different areas. The prevalence of cag A
139 genotype is high based on studies conducted on Iranian and Iraqi patients [24]. However, in
140 research conducted on Malaysian and Jordanian patients, the prevalence of cag A is relatively
141 low [25, 26]. The cagA was present in 60-70% of isolates from the Western population [27].
142 In our research cagA was present in 60.5% of isolates. We observed a significant relationship
143 between cag A and hiatal hernia (P=0.02). Several European and North American studies have
144 shown that infection with cagA-positive H. pylori strains also increases the risk for atrophic
145 gastritis and gastric cancer [27]. However, several studies in Asian populations did not confirm
146 these relationships, indicating that there are important geographic differences, which is
147 consistent with our research finding [28].

148 The association of the cagA-positive, vacA s1 genotypes with peptic ulcer disease (PUD) and
149 gastric cancer was reported in Western countries, which was not consistent with our research
150 and a research in East Asian countries [16]. Patients infected with less virulent genotypes are
151 more likely to have mild gastritis throughout their entire life, whereas patients infected with
152 more virulent genotypes have a higher probability of developing peptic ulcer disease, atrophic
153 gastritis, and eventually gastric carcinoma [28].

154 Our results showed that the most common allele in our isolates is m2 (55.9%), and s1 (55.9%),
155 then s2 (44.2%). Although in other research on Iranian and Afghan patients [21] the most
156 common allele is s1.

157

158 **Conclusions**

159 In this study, we analysed the *H. pylori* gene and its association with clinical outcomes. Our
160 data suggested that *cagA* is associated with normal esophageal findings, which support the
161 hypothesis of virulent strains may provide some protection for the esophagus. Furthermore, no
162 significant relationship was observed between the *H. pylori* gene and patients' characteristics
163 (such as age, sex, smoking status). This study provides the first report of *H. pylori* gene
164 diversity in Jahrom, Iran, and serves as an epidemiological tool for future studies on this
165 bacterium to better understand the clinical outcome of this pathogen.

167 **Materials and methods**

168 **Patients**

169 We used a descriptive cross-sectional method in this study. Patients who visited Jahrom's
170 Honary clinic (a city in Fars province, south of Iran) with persistent gastrointestinal problems,
171 above 18 years of age, and did not respond to 6 months treatment with proton pump inhibitor
172 (PPI), and positive rapid urease test (RUT) were studied. Esophagogastroduodenoscopy (EGD)
173 and 2 biopsy samples were retrieved from their stomach antrum.

174 This study was approved by the ethics committee of Jahrom University of Medical Sciences.
175 All the participants were informed about the study and signed consent forms and reassured
176 their confidentiality.

177 The patients included must have dyspepsia and positive RUT at the time of EGD. Patients were
 178 excluded from this study if they had a history of antibiotic therapy in the last month or if they
 179 used a PPI or H2 blocker in the last week.

180

181 **DNA Extraction and PCR**

182 Two samples were retrieved by EGD from every patient gastric antrum and with microbiology
 183 techniques one sample was used for RUT and another was kept on a transport medium which
 184 contains; Agar 1.3 g/L and yeast extract 3%. If the urease test came positive, PCR was
 185 conducted for the second sample. We used fermentas Company kits for DNA extraction.

186 The primers that were used for genotyping and PCR conditions are shown in table 5. The
 187 following cycle conditions were used: for vacA: 35 cycles of 1 min at 94°C, 1 min at 53°C,
 188 and 1 min at 72°C; for cagA: 1 min at 94°C, 1 min at 56°C, and 1 min at 72°C; for glmM: 1
 189 min at 93°C, 1 min at 55°C, and 1 min at 72°C. GlmM PCR product has 294 bp, cagA has 298
 190 bp and vacA subtypes (s1/s2, s1a, s1b, s1c, m1/m2) has 259, 286, 190, 187, 213, 567 and 642
 191 bp respectively.

192 **Table 5** Primer sequence and polymerase chain reaction (PCR) conditions

Genes	Primer sequence (5' → 3')	PCR		References
		product (bp)	PCR conditions	
glmM	AAGCTTTTAGGGGTGTTAGGGGTTT	294	93 °C, 1 min; 55 °C, 1	[29]
	AAGCTTACTTTCTAACACTAACGC		min; 72 °C, 1 min (35 cycles)	
vacA				
s1/s2	ATGGAAATACAACAAACACAC	259/286		[18, 30]

1		CTGCTTGAATGCGCCAAAC		94 °C, 1 min; 52 °C, 1	
2				min; 72 °C, 1 min (35	
3				cycles)	
4					
5					
6					
7		GTCAGCATCACACCGCAAC		94 °C, 1 min; 52 °C, 1	
8				min; 72 °C, 1 min (35	
9	s1a	CTGCTTGAATGCGCCAAAC	190	min; 72 °C, 1 min (35	[18]
10				cycles)	
11					
12					
13					
14		AGCGCCATACCGCAAGAG		94 °C, 1 min; 52 °C, 1	
15				min; 72 °C, 1 min (35	
16	s1b	CTGCTTGAATGCGCCAAAC	187	min; 72 °C, 1 min (35	[18]
17				cycles)	
18					
19					
20					
21		CTCTCGCTTTAGTGGGGYT		94 °C, 1 min; 52 °C, 1	
22				min; 72 °C, 1 min (35	
23	s1c	CTGCTTGAATGCGCCAAAC	213	min; 72 °C, 1 min (35	[31]
24				cycles)	
25					
26					
27					
28		CAATCTGTCCAATCAAGCGAG		94 °C, 1 min; 52 °C, 1	
29				min; 72 °C, 1 min (35	
30	m1/m2	GCGTCAAATAATTCCAAGG	567/642	min; 72 °C, 1 min (35	[18]
31				cycles)	
32					
33					
34					
35		ATAATGCTAAATTAGACAACCTTGAGCGA		94 °C, 1 min; 60 °C, 1	
36				min; 72 °C, 1 min (45	
37	cagA	TTAGAATAATCAACAAACATCACGCCAT	298	min; 72 °C, 1 min (45	[32]
38				cycles)	
39					
40					
41					

42 193

46 194 **Data analysis**

49 195 Based on Prevalence of 69%, ci=95%, absolute error=80% and population(N) of 1,000,000, a
52 196 sample size of n=113 was calculated [33]. Data collected from this study were analyzed using
54 197 chi-square, fisher's exact test and spearman methods, and SPSS software ver 22.0.

57 198

59 199 **Abbreviations**

200 vacA: Vacuolating cytotoxin A; cagA: Cytotoxin associated gene A; H. pylori: Helicobacter
201 pylori; RUT: Rapid urease test; PCR: Polymerase chain reaction; GI: Gastrointestinal; GERD:
202 Gastroesophageal reflux disease; MALT: Mucosa-associated lymphoid tissue; PUD: Peptic
203 ulcer disease; DU: Duodenal ulcers; GU: Gastric ulcers; NUD: Non-ulcer dyspepsia; PPI:
204 Proton pump inhibitor; EGD: Esophagogastroduodenoscopy.

206 **Declarations**

207 **Ethical approval of the study**

208 Written inform consent was obtained from the patients in our study. The purpose of this
209 research was completely explained to the patient and was assured that their information will be
210 kept confidential by the researcher. This research was approved by the ethical committee of
211 Jahrom University of Medical Sciences issued with code No.: Jums.REC.1392.028.

213 **Consent for publication**

214 Consent was obtained from the patients regarding the publication of this study.

216 **Availability of data and materials**

217 SPSS data of the participant can be requested from the authors. Please write to the
218 corresponding author if you are interested in such data.

220 **Competing interests**

221 The authors declare that they have no competing interests.

223 **Funding**

224 No financial support was received for this study.

225

226 **Authors' contributions**

227 Study concept and design: Rahim Raufi, Nikta Taghipour. Acquisition of data: Rahim Raufi,

228 Seyedeh Maryam Pishva. Analysis and interpretation of data: Reza Shahriarirad, Nikta

229 Taghipour. Drafting of the manuscript: Seyedeh Maryam Pishva, Reza Shahriarirad. Critical

230 revision of the manuscript for important intellectual content: Rahim Raufi, Nikta Taghipour .

231 Statistical analysis: Reza Shahriarirad, Nikta Taghipour. Study supervision: Rahim Raufi. All

232 authors have approved the final version of the manuscript.

233

234 **Acknowledgments**

235 We would like to express our sincere gratitude to the Clinical Research Development Unit of

236 Motahari Educational and Research and Therapeutic Center affiliated to Jahrom University of

237 Medical Sciences for providing suitable environmental facilities and research requirements.

238

239 **Authors' information**

240 ¹ Department of Infection Disease, Jahrom University of Medical Sciences, Jahrom, Iran. ²

241 Thoracic and Vascular Surgery Research Center, Shiraz University of Medical Sciences,

242 Shiraz, Iran. ³ Student Research Committee, Shiraz University of Medical Sciences, Shiraz,

243 Iran. ⁴ Student Research Committee, Jahrom University of Medical Sciences, Jahrom, Iran.

244

245 **References**

246 1. Ansari S, Yamaoka Y. Helicobacter pylori Virulence Factors Exploiting Gastric

247 Colonization and its Pathogenicity. Toxins. 2019;11(11):677.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

248 2. Salih B. Helicobacter pylori infection in developing countries: The burden for how long?
249 Saudi J Gastroenterol. 2009;15(3):201.

250 3. Akopyanz N, Bukanov NO, Westblom TU, Kresovich S, Berg DE. DNA diversity among
251 clinical isolates of Helicobacter pylori detected by PCR-based RAPD fingerprinting. Nucleic
252 Acids Res. 1992;20(19):5137-42.

253 4. Atherton JC. The clinical relevance of strain types of Helicobacter pylori. Gut.
254 1997;40(6):701-3.

255 5. De Luca A, Iaquinto G. Helicobacter pylori and gastric diseases: a dangerous association.
256 Cancer Lett. 2004;213(1):1-10.

257 6. Burkitt MD, Duckworth CA, Williams JM, Pritchard DM. Helicobacter pylori-induced
258 gastric pathology: insights from in vivo and ex vivo models. Dis Model Mech. 2017;10(2):89-
259 104.

260 7. Kusters JG, van Vliet AHM, Kuipers EJ. Pathogenesis of Helicobacter pylori Infection. Clin.
261 Microbiol. Rev.. 2006;19(3):449-90.

262 8. Park JB. Helicobacter pylori infection in gastric mucosa-associated lymphoid tissue
263 lymphoma. World J. Gastroenterol. 2014;20(11):2751.

264 9. Suerbaum S, Michetti P. Helicobacter pylori Infection. N. Engl. J. Med. 2002;347(15):1175-
265 86.

266 10. Covacci A. Helicobacter pylori Virulence and Genetic Geography. Science.
267 1999;284(5418):1328-33.

268 11. van Doorn LJ, Figueiredo C, Rossau R, et al. Typing of Helicobacter pylori vacA Gene and
269 Detection of cagA Gene by PCR and Reverse Hybridization. J. Clin. Microbiol.
270 1998;36(5):1271-6.

271 12. Gebert B, Fisher W, Haas R. The *Helicobacter pylori* vacuolating cytotoxin: from cellular
1 vacuolation to immunosuppressive activities. *Rev. Physiol. Biochem. Pharmacol.* 2004. p.
2 272
3 205-20.
4 273
5
6
7 274 13. Yamaoka Y, Kato M, Asaka M. Geographic Differences in Gastric Cancer Incidence Can
8
9 be Explained by Differences between *Helicobacter pylori* Strains. *J. Intern. Med.*
10 275
11 2008;47(12):1077-83.
12 276
13
14 277 14. Fallahi G-H, Maleknejad S. *Helicobacter pylori* culture and antimicrobial resistance in Iran.
15
16 *Indian J Pediatr.* 2007;74(2):127-30.
17 278
18
19 279 15. Torres LE, Melián K, Moreno A, et al. Prevalence of *vacA*, *cagA* and *babA2* genes in
20
21 Cuban *Helicobacter pylori* isolates. *World J. Gastroenterol.* 2009;15(2):204.
22 280
23
24 281 16. Saribasak H, Salih BA, Yamaoka Y, Sander E. Analysis of *Helicobacter pylori* Genotypes
25
26 and Correlation with Clinical Outcome in Turkey. *J. Clin. Microbiol.* 2004;42(4):1648-51.
27 282
28
29 283 17. Hosseini E, Poursina F, de Wiele TV, Safaei HG, Adibi P. *Helicobacter pylori* in Iran: A
30
31 systematic review on the association of genotypes and gastroduodenal diseases. *J Res Med Sci.*
32 284
33 2012 Mar;17(3):280-92.
34 285
35
36 286 18. Qiao W. *cagA* and *vacA* genotype of *Helicobacter pylori* associated with gastric diseases in
37
38 Xi'an area. *World J. Gastroenterol.* 2003;9(8):1762.
39 287
40
41 288 19. Caner V. *H pylori iceA* alleles are disease-specific virulence factors. *World J. Gastroenterol.*
42
43 2007;13(17):2581.
44 289
45
46 290 20. Chomvarin C, Namwat W, Chaicumpar K, et al. Prevalence of *Helicobacter pylori vacA*,
47
48 *cagA*, *cagE*, *iceA* and *babA2* genotypes in Thai dyspeptic patients. *Int. J. Infect. Dis.*
49 291
50 2008;12(1):30-6.
51 292
52
53 293 21. Dabiri H, Bolfion M, Mirsalehian A, et al. Analysis of *Helicobacter pylori* Genotypes in
54
55 Afghani and Iranian Isolates. *Pol. J. Microbiol.* 2010;59(1):61-6.
56 294
57
58
59
60
61
62
63
64
65

- 295 22. Kargar M, Souod N, Doosti A, Ghorbani-Dalini S. Prevalence of *Helicobacter pylori*
1
2
3 296 vacuolating cytotoxin A gene as a predictive marker for different gastroduodenal diseases. Iran
4
5 297 J Clin Infect Dis. 2011;6(2):85-9.
6
7 298 23. Jafari F, Shokrzadeh L, Dabiri H, et al. *vacA* genotypes of *Helicobacter pylori* in relation
8
9 299 to *cagA* status and clinical outcomes in Iranian populations. Jpn. J. Infect. Dis. 2008;61(4):290.
10
11 300 24. Hussein NR, Mohammadi M, Talebkhan Y, et al. Differences in Virulence Markers
12
13 301 between *Helicobacter pylori* Strains from Iraq and Those from Iran: Potential Importance of
14
15 302 Regional Differences in *H. pylori*-Associated Disease. J. Clin. Microbiol. 2008;46(5):1774-9.
16
17 303 25. Amjad N. Clinical significance of *Helicobacter pylori cagA* and *cagE* genotype status. World
18
19 304 J. Gastroenterol. 2010;16(35):4443.
20
21 305 26. Nimri LF, Matalka I, Bani-Hani KE, Ibrahim M. *Helicobacter pylori* genotypes identified
22
23 306 in gastric biopsy specimens from Jordanian patients. BMC Gastroenterol. 2006;6(1).
24
25 307 27. Infection with *cagA*-positive *H. pylori* strains is associated with gastric carcinoma risk. Nat
26
27 308 Rev Clin Oncol 4, 678 (2007). <https://doi.org/10.1038/ncponc0967>
28
29 309 28. Nogueira C, Figueiredo C, Carneiro F, et al. *Helicobacter pylori* Genotypes May Determine
30
31 310 Gastric Histopathology. Am. J. Pathol.. 2001;158(2):647-54.
32
33 311 29. Lu J-J, Perng C-L, Shyu R-Y, et al. Comparison of Five PCR Methods for Detection of
34
35 312 *Helicobacter pylori* DNA in Gastric Tissues. J. Clin. Microbiol. 1999;37(3):772-4.
36
37 313 30. Atherton JC, Cao P, Peek RM, Tummuru MKR, Blaser MJ, Cover TL. Mosaicism in
38
39 314 Vacuolating Cytotoxin Alleles of *Helicobacter pylori*. J. Biol. Chem. 1995;270(30):17771-7.
40
41 315 31. Yamazaki S, Yamakawa A, Okuda T, et al. Distinct Diversity of *vacA*, *cagA*, and *cagE*
42
43 316 Genes of *Helicobacter pylori* Associated with Peptic Ulcer in Japan. J. Clin. Microbiol.
44
45 317 2005;43(8):3906-16.
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

318 32. Hamlet A, Thoreson AC, Nilsson O, Svennerholm AM, Olbe L. Duodenal Helicobacter
1 pylori infection differs in cagA genotype between asymptomatic subjects and patients with
2
3 319
4
5 320 duodenal ulcers. *Gastroenterology*. 1999;116(2):259-68.
6
7 321 33. Niknam R, Seddigh M, Fattahi MR, Dehghanian A, Mahmoudi L. Prevalence of
8
9 322 Helicobacter pylori in Patients With Dyspepsia. *Jundishapur J Microbiol*. 2014;7(9).
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Table 4 Relationship between different H.pylori genotypes and variables in our study

Variable	Genotype n=86																				
	S1am1 n=19	P	S1am2 n=24	P	S1bm1 n=2	P	S1bm2 n=1	P	S1cm1 n=1	P	S1cm2 n=1	P	S2m1 n=16	P	S2m2 n=22	P	Cag a n=52	P	Glm m n=86	P	
Mean Age (SD)	33.6 (10.8)	-	39.2 (14.3)	-	40.5 (27.6)	-	29 (0)	-	70 (0)	-	60 (0)	-	38.1 (17.2)	-	41.8 (19.3)	-	37.8 (15)	-	38.9 (16.1)	-	
Gender	<i>Male</i>	7 (36.8)		9 (37.5)		2 (100)		1 (100)		1 (100)		1 (100)		4 (25)		9 (40.9)		20 (35.5)		34 (39.5)	
	<i>Female</i>	12 (63.2)	1	15 (62.5)	1	0 (0)	0.149	0 (0)	0.389	0 (0)	0.389	0 (0)	0.389	12 (75)	0.275	13 (59.1)	1	32 (61.5)	1	52 (60.5)	1
Smoker	<i>Yes</i>	3 (15.8)	1	3 (12.5)	0.76	1 (50)	0.294	0 (0)	1	0 (0)	1	1 (100)	0.159	3 (18.8)	0.718	6 (4.5)	0.19	10 (29.2)	0.4	12 (14)	0.4
	<i>No</i>	16 (84.2)		21 (87.5)		1 (50)		1 (100)		1 (100)		0 (0)		13 (81.2)		21 (95.5)		42 (80.8)		74 (86)	
Endoscopy	<i>NUD</i>	7 (36.8)	0.454	13 (54.2)	0.489	1 (50)	1	1 (100)	0.46	1 (100)	0.46	0 (0)	1	8 (50)	0.791	11 (50)	0.812	24 (46.2)	1	42 (48.8)	0.4
	<i>Erosive gastropathy</i>	10 (52.6)	0.316	6 (25)	0.101	1 (50)	1	0 (0)	1	0 (0)	1	0 (0)	1	7 (43.8)	1	9 (40.9)	1	19 (36.56)	0.3	33 (38.4)	0.3
	<i>GU</i>	0 (0)	1	2 (8.3)	0.114	0 (0)	1	0 (0)	1	0 (0)	1	0 (0)	1	0 (0)	1	1 (4.5)	0.481	1 (1.9)	1	3 (3.5)	1
	<i>DU</i>	2 (10.5)	1	3 (12.5)	0.698	0 (0)	1	0 (0)	1	0 (0)	1	1 (100)	0.097	1 (6.2)	1	1 (4.5)	0.689	8 (15.4)	0.1	8 (9.3)	0.7
Esophageal Findings	<i>Esophagitis</i>	0 (0)	0.353	3 (12.5)	0.398	0 (0)	1	0 (0)	1	0 (0)	1	0 (0)	1	2 (12.5)	0.613	1 (4.5)	1	3 (5.8)	0.5	5 (5.8)	0.4
	<i>GERD</i>	1 (5.3)	1	1 (4.2)	1	2 (100)	0.002	0 (0)	1	0 (0)	1	0 (0)	1	0 (0)	1	0 (0)	0.581	3 (5.8)	0.6	4 (4.7)	1
	<i>Hiatal Hernia</i>	9 (47.4)	0.806	10 (41.7)	0.366	1 (50)	1	0 (0)	0.496	0 (0)	0.496	1 (100)	1	7 (43.8)	0.6	11 (50)	1	20 (38.4)	0.02	39 (45.3)	0.07
	<i>Normal</i>	9 (47.4)	0.439	10 (41.7)	0.813	0 (0)	0.524	1 (100)	0.381	1 (100)	0.381	0 (0)	1	7 (43.8)	0.782	10 (45.5)	0.468	26 (50)	0.02	38 (44.2)	0.02

PCR: polymerase chain reaction; H.pylori: Helicobacter Pylori; NUD: Non-ulcer dyspepsia; GU: Gastric ulcer; DU: Duodenal ulcer; GERD: Gastroesophageal reflux disease;



Click here to access/download
Supplementary Material
Cover_Letter_h.pilory- gut path.docx

