

Bioorganic & Medicinal Chemistry Letters

des man les entres aleste alles alles des

The Tetrahadron Journey for Nessanch at the Letterfloor of Deserbary and Hedge



EducationOtest Origit A. BEROOM

C²ScienceDirect

BIOORGANIC & MEDICINAL CHEMISTRY LETTERS

Editor-in-Chief: Professor D. L. BOGER

Department of Chemistry, BCC 483A, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, California 92037, USA.

Facsimile: (1) 619-699-6700

European Regional Editor: Professor Stephen Neidle, Department of Pharmaceutical & Biological Chemistry, The School of Pharmacy, University of London, 29/39 Brunswick Square, London WC1N 1AX, UK. Facsimile: 020 7753 5970

Japanese Regional Editor: Professor M. Shibasaki, Graduate School of Pharmaceutical Sciences, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033, Japan. Facsimile: (81) 3 5684 5206

EXECUTIVE BOARD OF EDITORS FOR TETRAHEDRON PUBLICATIONS

Chairman: Professor C. H. Wong

Editor Emeritus: Professor H. H. Wasserman

Professor D. L. Boger, Department of Chemistry, The Scripps Research Institute, La Jolla, CA 92037, USA

Professor S. G. Davies, Dyson Perrins Laboratory, Department of Chemistry, University of Oxford, Oxford OX1 3QY, UK

Professor B. Ganem, Department of Chemistry & Chemical Biology, Baker Laboratory, Cornell University, Ithaca, NY 14853-1301, USA

Professor L. Ghosez, l'Institut Européen de Chimie et de Biologie (IECB), 33607 Pessac Cedex, France

Professor Lin Guo-Qiang, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, 354 Fenglin Road, Shanghai 200032, China

Professor Y. Hashimoto, Institute of Molecular & Cellular Biosciences, The University of Tokyo, III-Yayoi, Bunkyo-ku, Tokyo 113-0032, Japan

Professor T. Hayashi, Department of Chemistry, Faculty of Science, Kyoto University, Kyoto 606, Japan

Professor Barbara Imperiali, Department of Chemistry, Massachusetts Institute of Technology, Cambridge, MA 01239, USA

Professor S. F. Martin, Department of Chemistry & Biochemistry, University of Texas, Austin, TX 78712, USA

Professor W. B. Motherwell, Department of Chemistry, University College, 20 Gordon Street, London WC1H 0AJ, UK

Professor Stephen Neidle, The School of Pharmacy, Department of Pharmaceutical & Biological Chemistry, University of London, London WC1N 1AX, UK Professor M Shibasaki Graduate School of Pharmaceutical Sciences

The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033, Japan

Professor R. J. K. Taylor, Department of Chemistry, University of York, Heslington, York YO10 5DD, UK (Associate Editors, Dr P. A. O'Brien and Dr D. K. Smith)

Professor K. Tomioka, Graduate School of Pharmaceutical Sciences, Department of Synthetic Medicinal Chemistry, Kyoto University, Kyoto 606-8501, Japan

Professor E. J. Thomas, Department of Chemistry, University of Manchester, Brunswick Street, Manchester M13 9PL, UK (Associate Editor, Professor J. A. Joule)

Professor H. Waldman, Max-Planck-Inst. für Molekular Physiology, Department of Chemistry, Otto-Hahn-Strasse 11, 44227 Dortmund, Germany

Professor H. H. Wasserman, Department of Chemistry, Yale University, PO Box 208107, New Haven, CT 06520-8107, USA

Professor R. M. Williams, Department of Chemistry, Colorado State University, Fort Collins, CO 80523

Professor C.-H. Wong, Department of Chemistry, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, CA 92037, USA

Professor J. Wood, Department of Chemistry, Colorado State University, Fort Collins, CO 80523-1872

Professor Y. Yamamoto, Department of Chemistry, Graduate School of Science, Tohoku University, Sendai 980-8578, Japan (Associate Editor, Professor M. Hirama)

NC

Professor S. Z. Zard, Laboratoire de Synthèse Organique, Ecole Polytechnique, F-91128 Palaiseau Cedex, France (Associate Editor, Dr B. Sire)

BOARD OF CONSULTING EDITORS

P. S. Anderson, Wilmington, DE	A. R. Fersht, Cambridge	J. Kelly, La Jolla, CA	P. G. Schultz, La Jolla, CA
KH. Altmann, Zürich	D. M. Floyd, Princeton, NJ	P. Krogsgaard-Larsen, Copenhagen	P. Seeberger, Zürich
P. G. Baraldi, Ferrara	G. I. Georg, Minneapolis, MN	R. A. Lerner, La Jolla, CA	O. Seitz, Berlin
C. F. Barbas, III, La Jolla, CA	A. Giannis, Leipzig	H. Liu, Austin, TX	K. Shokat, San Francisco, CA
J. K. Barton, Pasadena, CA	B. Giese, Basel	A. McKillop, Northumberland	R. Silverman, Evanston, IL
C. R. Bertozzi, Berkeley, CA	P. Gmeiner, Erlangen	R. Metternich, Berlin	J. Stubbe, Cambridge, MA
R. C. Breslow, New York, NY	H. B. Gray, Pasadena, CA	S. Mignani, Vitry-sur-Seine	C. T. Supuran, Firenze
T. C. Bruice, Santa Barbara, CA	G. L. Grunewald, Lawrence, KS	L. A. Mitscher, Lawrence, KS	G. L. Verdine, Cambridge, MA
A. R. Chamberlin, Irvine, CA	P. Herrling, Basel	K. C. Nicolaou, La Jolla, CA	S. Walker, Cambridge, MA
E. J. Corey, Cambridge, MA	D. Hilvert, Zürich	H. L. Pearce, Indianapolis, IN	C. T. Walsh, Boston, MA
B. Cravatt, La Jolla, CA	L. C. Hsieh-Wilson, Pasadena, CA	C. D. Poulter, Salt Lake City, UT	P. A. Wender, Stanford, CA
S. J. Danishefsky, New York, NY	K. Janda, La Jolla, CA	J. Rebek, Jr., La Jolla, CA	G. Whitesides, Cambridge, MA
P. B. Dervan, Pasadena, CA	W. L. Jorgensen, New Haven, CT	B. Samuelsson, Stockholm	R. V. Wolfenden, Chapel Hill, 1
A. Eschenmoser, Zürich	A. R. Katritzky, Gainesville, FL	J. Saunders, San Diego, CA	
JM. Fang, Taipei	J. A. Katzenellenbogen, Urbana, IL	S. L. Schreiber, Cambridge, MA	

PUBLISHED TWICE MONTHLY

Orders, claims, and journal enquiries: Please contact the Regional Sales Office nearest you:

Orlando: Elsevier, Customer Service Department, 6277 Sea Harbor Drive, Orlando, FL 32887-4800, USA; phone: (877) 839 7126 [toll free within the USA]; (+1) (407) 563 6022 [outside the USA]; fax: (+1) (407) 363 1354; e-mail: JournalCustomerService-usa@elsevier.com

Amsterdam: Elsevier, Customer Service Department, PO Box 211, 1000 AE Amsterdam, The Netherlands; phone: (+31) (20) 4853757; fax: (+31) (20) 4853432; e-mail: JournalsCustomerServiceEMEA@elsevier.com

Tokyo: Elsevier, Customer Service Department, 4F Higashi-Azabu, 1-Chome Bldg., 1-9-15 Higashi-Azabu, Minato-ku, Tokyo 106-0044, Japan; phone: (+81) (3) 5561 5037; fax: (+81) (3) 5561 5047; e-mail: JournalsCustomerServiceJapan@elsevier.com

Singapore: Elsevier, Customer Service Department, 3 Killiney Road, #08-01 Winsland House I, Singapore 239519; phone: (+65) 63490222; fax: (+65) 67331510; e-mail: JournalsCustomerServiceAPAC@elsevier.com



Bioorganic & Medicinal Chemistry Letters Vol. 18, No. 13, 2008

Contents

ARTICLES

Studies on the structure-activity relationship of bicifadine analogs as monoamine transporter inhibitorspp 3682-3686Mingzhu Zhang, Florence Jovic, Troy Vickers, Brian Dyck, Junko Tamiya, Jonathan Grey,
Joe A. Tran, Beth A. Fleck, Rebecca Pick, Alan C. Foster, Chen Chen*pp 3682-3686



Insight from molecular modeling into different conformation and SAR of natural steroids and unnatural 7-oxa-steroids

Fu-An Kang^{*}, Xin Chen, Nareshkumar Jain, George Allan, Pamela Tannenbaum, Scott Lundeen, Zhihua Sui



Molecular modeling, in vivo activity, pharmacokinetic and metabolic properties of unnatural 7-oxa-steroids are reported.

Protein subtype-targeting through ligand epimerization: Talose-selectivity of galectin-4 and galectin-8 Christopher T. Öberg, Helen Blanchard, Hakon Leffler, Ulf J. Nilsson^{*} pp 3691-3694

pp 3687-3690



Galectin-4 C-terminal domain and galectin-8 N-terminal domain were found to prefer the p-talopyranose configuration to the natural ligand p-galactopyranose configuration. Methyl β -p-talopyranosides derivatized at O2 and O3 were synthesized and discovered to be selective submillimolar inhibitors of galectin-4C and galectin-8N.



Chrobisiamone A, a new bischromone from *Cassia siamea* and a biomimetic transformation of 5-acetonyl-7-hydroxy-2-methylchromone into cassiarin A

Shiori Oshimi, Yuichiro Tomizawa, Yusuke Hirasawa, Toshio Honda, Wiwied Ekasari, Aty Widyawaruyanti, Marcellino Rudyanto, Gunawan Indrayanto, Noor Cholies Zaini, Hiroshi Morita^{*}



A new bischromone, chrobisiamone A (1) with an antiplasmodial activity has been isolated from the leaves of *Cassia siamea* and the structure was elucidated by 2D NMR analysis. Cyclization of 5-acetonyl-7-hydroxy-2-methylchromone (2) in the presence of ammonium acetate resulted in generation of cassiarin A (3) with an unprecedented tricyclic skeleton, supporting a biogenetic pathway proposed for **3**.

2-Aminopropane-1,2,3-tricarboxylic acid: Synthesis and co-crystallization with the class A β-lactamase BS3 of *Bacillus licheniformis*

Joséphine Beck, Eric Sauvage, Paulette Charlier, Jacqueline Marchand-Brynaert*



Synthesis and biochemical evaluation against β-lactamases of amino analog of citric acid are presented. The structure of the complex aminocitrate-BS3 has been analyzed by X-ray diffraction and compared to ones with citrate and isocitrate.

Pyrrolo[2,1-**c**][1,4]benzodiazepine-β-glucuronide prodrugs with a potential for selective therapy of solid tumors by PMT and ADEPT strategies

Ahmed Kamal*, Venkatesh Tekumalla, P. Raju, V. G. M. Naidu, Prakash V. Diwan, Ramakrishna Sistla

RO

MeC



15a R = A15b R = B

=0_{ОН}

Two new diterpenes, sucutiniranes A (1) and B (2), have been isolated from *Bowdichia nitida*. Sucutinirane A (1) and $\beta\alpha$ -acetoxyvouacapane (3) showed a moderate cytotoxicity and $\beta\alpha$, $\beta\beta$ -diacetoxyvouacapane (4) showed in vitro antiplasmodial activity against parasite *Plasmodium falciparum* 3D7.





pp 3761-3763

pp 3764-3768

()⁺

pp 3769-3773

Dopamine/serotonin receptor ligands. Part 17: A cross-target SAR approach: Affinities of azecine-styled ligands for 5-HT_{2A} versus D₁ and D₂ receptors

Christoph Enzensperger, Tilo Görnemann, Heinz H. Pertz, Jochen Lehmann*



pK, for D1: 6.79 to 9.45; for D2: 6.41 to 8.45; pA2 for 5-HT2A : 7.21 to 9.97

A cross-target SAR was conducted with 13 azecine-styled compounds on D₁, D₂ and 5-HT_{2A} receptors. Surprisingly, molecular modifications aect the affinity for the D₁ receptor in the same manner as the 5-HT_{2A} receptor. The protein-ligand interactions were discussed with respect to the dierent binding pockets.

Synthesis of 5"-branched derivatives of cyclic ADP-carbocyclic-ribose, a potent Ca²⁺-mobilizing agent: The first antagonists modified at the N1-ribose moiety

Natsumi Sakaguchi, Takashi Kudoh, Takayoshi Tsuzuki, Takashi Murayama, Takashi Sakurai, Akira Matsuda, Mitsuhiro Arisawa, Satoshi Shuto*



OTHER CONTENTS

Summary of instructions to authors

*Corresponding author (i)+ Supplementary data available via ScienceDirect

COVER

Overlay of high resolution co-crystal structures of R-22-ADP (cyan) and 1-ADP (green) bound in an allosteric binding site of the mitotic kinesin KSP. [Roecker, A. J.; Coleman, P. J.; Mercer, S. P.; Schreier, J. D.; Buser, C. A.; Walsh, E. S.; Hamilton, K.; Lobell, R. B.; Tao, W.; Diehl, R. E.; South, V. J.; Davide, J. P.; Kohl, N. E.; Yan, Y.; Kuo, L. C.; Li, C.; Fernandez-Metzler, C.; Mahan, E. A.; Prueksaritanont, T.; Hartman, G. D. Bioorg. Med. Chem. Lett. 2007, 17, 5677.]

Indexed/Abstracted in: Beilstein, Biochemistry & Biophysics Citation Index, CANCERLIT, Chemical Abstracts, Chemistry Citation Index, Current Awareness in Biological Sciences/BIOBASE, Current Contents: Life Sciences, EMBASE/Excerpta Medica, MEDLINE, PASCAL, Research Alert, Science Citation Index, SciSearch, TOXFILE



pp 3814-3818

3681

рI

Contents lists available at ScienceDirect



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Sucutiniranes A and B, new cassane-type diterpenes from Bowdichia nitida

Yosuke Matsuno^a, Jun Deguchi^a, Yusuke Hirasawa^a, Kunio Ohyama^b, Hiroo Toyoda^b, Chieko Hirobe^c, Wiwied Ekasari^d, Aty Widyawaruyanti^d, Noor Cholies Zaini^d, Hiroshi Morita^{a,*}

^a Faculty of Pharmaceutical Sciences, Hoshi University, Ebara 2-4-41, Shinagawa-ku, Tokyo 142-8501, Japan

^b School of Pharmacy, Tokyo University of Pharmacy & Life Science, 1432-1 Horinouchi, Hachioji, Tokyo 192-0392, Japan

^c Seisen University, Higashi Gotanda 3-16-21, Shinagawa-ku, Tokyo 141, Japan

^d Airlangga University, Jalan Dharmawangsa Dalam, Surabaaya 60286, Indonesia

ARTICLE INFO

Article history: Received 21 March 2008 Revised 19 April 2008 Accepted 9 May 2008 Available online 16 May 2008

Keywords: Diterpene Sucutinirane A Sucutinirane B Bowdichia nitida Cytotoxicity Antiplasmodial activity

ABSTRACT

Two new cassane-type diterpenes, sucutiniranes A (1) and B (2), have been isolated from the seeds of *Bowdichia nitida* together with 6α -acetoxyvouacapane (3) and 6α , 7β -diacetoxyvouacapane (4), and the structures of 1 and 2 were elucidated by using 2D NMR data and chemical correlations. Sucutinirane A (1) and 3 showed a moderate cytotoxicity against human colon carcinoma COLO201 cells, and 6α , 7β -diacetoxyvouacapane (4) showed in vitro antiplasmodial activity against parasite *Plasmodium falciparum* 3D7. © 2008 Elsevier Ltd. All rights reserved.

Bowdichia nitida Spruce ex Benth., common name 'sucupira', is distributed in the Brazilian Amazon, and the seeds of this plant are used for rheumatic, antipyretic, and gouty agents.¹ So far, alkaloids, triterpenes, isoflavonoids, benzofuranes, and benzopyranes have been isolated from the genus *Bowdichia*.^{2–4}

Our efforts on identifying new natural products from the seeds of *B. nitida* resulted in the isolation of two new cassane-type diterpenes, sucutiniranes A (1) and B (2). This Letter describes the structure elucidation of 1 and 2 on the basis of spectroscopic data and chemical correlations as well as cytotoxicity against human colon carcinoma COLO201 cells and antiplasmodial activity.

Structures of sucutiniranes A (1) and B (2). The seeds of Bowdichia nitida were extracted with MeOH, and the extract was partitioned between EtOAc and 3% tartaric acid. EtOAc-soluble materials were subjected to a silica gel column (hexane/EtOAc and CHCl₃/MeOH) and an ODS column (MeOH/H₂O) followed by HPLC (MeOH/H₂O) to afford sucutiniranes A (1, 0.0002% yield) and B (2, 0.0006%) together with 6α -acetoxyvouacapane (3, 0.02%)⁵ and 6α ,7 β -diacetoxyvouacapane (4, 0.0008%).⁶

Sucutinirane A {1, $[\alpha]_D^{22}$ -24 (*c*, 1.0, CHCl₃)} was revealed to have the molecular formula C₂₂H₃₂O₅, by HRESITOFMS [*m*/*z* 399.2142 (M+Na)⁺, Δ -0.5 mmu]. IR absorptions implied the presence of hydroxyl (3480 cm⁻¹) and carbonyl (1740 cm⁻¹) groups. The ¹H and ¹³C NMR data (Table 1) suggested the presence of two carbonyl carbons, one sp² methine, one sp² quaternary carbon, five sp³ methylenes, five sp³ methines, three sp³ quaternary carbons, and five methyl groups. The presence of the α , β -unsaturated γ -lactone moiety was substantiated by the signals of one sp² methine ($\delta_{\rm C}$ 114.2), one sp² quaternary carbon ($\delta_{\rm C}$ 175.0), one sp³ quaternary carbonyl carbon ($\delta_{\rm C}$ 107.6), and one carbonyl carbon ($\delta_{\rm C}$ 173.4).

Partial structures **a** (C-1 to C-3) and **b** (C-5 to C-9, C-11, C-14, and C-17) were deduced from a detailed analysis of 2D NMR data of 1 (Fig. 1). The HMBC cross-peaks of H₃-19 to C-3, C-4, C-5, and C-18 indicated the connection among C-3, C-5, C-18, and C-19 through C-4. HMBC correlations for H₃-20 to C-1, C-5, C-9, and C-10 indicated connection among C-1, C-5, C-9, and C-20 through C-10. On the other hand, HMBC correlations for H-14 to C-12 and C-15, H₃-17 to C-13, and H-15 to C-12 and C-16 supported the location of the methyl group at C-14, and the α , β -unsaturated γ lactone moiety at C-12 and C-13. Furthermore, the presence of an acetoxy group at C-6 was elucidated by the HMBC correlation for H-6 and H₃-22 to C-21. Thus, the gross structure of sucutinirane A was assigned to be **1** with a cassane-type skeleton⁷ with the methyl group at C-14 and the α . β -unsaturated γ -lactone moiety at C-12 and C-13. The existence of cassane butenolides is rare as compared to that of cassane furanoditerpenes.

To assign the relative stereochemistry at the hemiketal C-12 position, **1** was acetylated with acetic anhydride in pyridine at

^{*} Corresponding author. Tel./fax: +81 354985778.

E-mail address: moritah@hoshi.ac.jp (H. Morita).



 Table 1
 1 H [δ _H (J, Hz)] and 13 C [δ _C] NMR Data of sucutiniranes A (1) and B (2)

	1 ^a		2 ^b	
	¹ H	¹³ C	¹ H	¹³ C
1a	1.10 (1H, m)	40.7	1.02 (1H, m)	39.7
1b	1.74 (1H, m)		1.71 (1H, m)	
2a	1.48 (1H, m)	19.5	1.47 (1H, m)	18.4
2b	1.59 (1H, m)		1.52 (1H, m)	
3a	1.29 (1H, m)	44.6	1.24 (1H, m)	43.4
3b	1.38 (1H, m)		1.39 (1H, m)	
4		34.1		33.1
5	1.31 (1H, d, 11.0)	58.4	1.22 (1H, m)	57.2
6	5.12 (1H, ddd, 11.0, 11.0,	73.5	5.07 (1H, ddd, 11.1, 11.1,	72.0
	4.1)		4.3)	
7a	1.49 (1H, m)	38.0	1.41 (1H, m)	36.7
7b	1.84 (1H, m)		1.88 (1H, m)	
8	1.84 (1H, m)	40.9	1.82 (1H, m)	38.7
9	1.54 (1H, m)	45.6	1.25 (1H, m)	44.4
10		39.5		38.7
11a	1.25 (1H, m)	38.9	0.99 (1H, m)	33.7
11b	2.38 (1H, dd, 12.9, 3.3)		2.50 (1H, ddd, 11.8, 6.4, 3.0)	
12		107.6	4.84 (1H, dd, 11.4, 6.4)	79.1
13		175.0		176.1
14	2.96 (1H, m)	37.3	2.95 (1H, m)	35.7
15	5.74 (1H, s)	114.2	5.68 (1H, s)	110.9
16		173.4		173.4
17	1.17 (3H, d, 7.3)	12.9	1.08 (3H, d, 7.3)	13.9
18	1.10 (3H, s)	37.2	1.06 (3H, s)	36.5
19	0.91 (3H, s)	23.0	0.89 (3H, s)	22.5
20	0.91 (3H, s)	15.8	0.88 (3H, s)	15.3
21		172.2		170.3
22	2.04 (3H, s)	21.9	2.06 (3H, s)	21.9

^a In CD₃OD.

^b In CDCl₃.



Figure 1. Selected 2D NMR correlations for sucutinirane A (1).

room temperature to afford the monoacetylated product **1a**. The relative stereostructure of **1a** as shown in computer-generated 3D drawing (Fig. 2) was deduced from cross-peaks observed in the NOESY spectrum and ³*J* coupling constants. The NOESY correlation of H₃-17/H₃-24 indicated to be α -orientation for CH₃-17 and CH₃-24. Antiperiplanar conformation between H-5 and H-6 was preferred because of the coupling constant, ³*J*_{H5/H6} = 11.0 Hz. The β -configuration of H-6, CH₃-19, and CH₃-20 was



supported by the NOESY cross-peaks among H-6, H₃-19, and H₃-20, while the α -configuration of both H-5 and H-9 was supported by the NOESY cross-peak between H-5 and H-9. Furthermore, oxidation of the furan ring of 6α -acetoxyvouacapane (**3**) with *m*CPBA gave sucutinirane A (**1**) as shown in Scheme 1. Thus, the structure of sucutinirane A including relative stereo-chemistry was assigned as shown in Figure 2.

Sucutinizate B {**2**, $[\alpha]_D^{22} - 33$ (*c*, 0.2, CHCl₃)} was revealed to have the molecular formula C₂₂H₃₂O₄, by HRESITOFMS [*m/z* 361.2389 (M+H)⁺, Δ +2.1 mmu]. IR absorptions implied the presence of carbonyl (1735 cm⁻¹) group. The ¹H and ¹³C NMR data (Table 1), and 2D NMR correlations (Fig. 3) suggested that **2** had the same cassane-type skeleton as that of **1**, except for the presence of an oxymethine at C-12 (δ_H 4.84, δ_C 79.1). The relative stereochemistry of sucutinirane B (**2**) was deduced by NOESY spectrum (Fig. 4) and ³*J* coupling constants. The configuration of β-oriented H-6, H-8, CH₃-19, and CH₃-20 was supported by the NOESY correlations of H-6/H-8, H-6/H₃-19, and H-8/H₃-20. The coupling constant, ³*J*_{H5/} H₆ = 11.1 Hz indicated antiperiplanar conformation between H-5 and H-6. The α-configuration of H-5, H-9, H-12, and CH₃-17 was supported by the NOESY cross-peaks of H-5/H-9 and H-12/H-9 and H₃-17. Thus, the structure of **2** was assigned as 12-deoxy-sucutinirane A.

The absolute stereochemistry of sucutiniranes A (1) and B (2) was deduced by applying CD curves for γ -lactone chromophore.⁸ The sign of the CD curve in MeOH [1: λ_{max} 222 nm ($\Delta \varepsilon$ –0.8) and 244 nm ($\Delta \varepsilon$ –0.6), 2: λ_{max} 218 nm ($\Delta \varepsilon$ –1.1)] was negative, indicating that the chirality at C-12 of 1 and 2 was as shown in Figures 3 and 4.

To confirm the proposed structure for **2**, treatment of 6α -acetoxyvouacapane (**3**) with *m*CPBA in the presence of 1 drop 12 N HCl in CHCl₃ afforded sucutinirane B (**2**) together with two byproducts, compounds **5** and **6**, which was elucidated by 2D NMR correlations as shown in Figure 5. Stereochemistry of



Figure 2. Selected NOESY correlations and relative stereochemistry of compound la.



Scheme 1. Oxidation of 6α-acetoxyvouacapane (**3**) by *mCPBA*.



Figure 3. Selected 2D NMR correlations for sucutinirane B (2).



Figure 4. Selected NOESY correlations and relative stereochemistry of sucutinirane B (2).

a hydroxyl at C-11 and spiro carbon at C-13 for **5** was assigned by NOESY data of H-8/H-11 and H-15/H₃-17. Compound **5**, which was derived from oxidative intermediate at C-11 followed by epoxidation at Δ^{12} and Pinacol-type rearrangement, possesses a spirojoined β , γ -unsaturated γ -lactone, and cyclopentane bicycles with a hydroxyl group at C-11. On the other



Figure 5. Selected 2D NMR correlations for 5 and 6.

hand, compound **6**, which was produced from oxidative intermediate at C-11 followed by epoxidation at Δ^{15} and cleavage between C-11 and C-12 bond accompanied with cleavage of the epoxide, contains an aldehyde moiety at C-11 and an α , β -unsaturated γ -lactone moiety at C-14 (Scheme 1). These structures of **5** and **6** were also supported by HMBC (Fig. 5) and NOESY correlations.

Sucutinirane A (1) and 6α -acetoxyvouacapane (3) showed a moderate cytotoxicity against human colon carcinoma COLO201 cells with IC₅₀ 37.3 and 86.6 µg/mL, respectively, while sucutinirane B (2), 6α , 7β -diacetoxyvouacapane (4), and compounds 5 and 6 were inactive (IC₅₀ > 100 µg/mL).

Each compound was also tested for its ability to inhibit *Plasmodium falciparum* growth.⁹ 6α , 7β -diacetoxyvouacapane (**4**) showed promising in vitro antiplasmodial activity against parasite *P. falciparum* 3D7 (IC₅₀ 0.39 µg/mL) and a good selectivity index with regard to the cytotoxicity on COLO201 cells (IC₅₀ > 100 µg/mL), whereas other compounds were inactive at a concentration of 1 µg/mL (Chloroqine: IC₅₀ 0.006 µg/mL).

Acknowledgments

This work was supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan, and grants from the Research Foundation for Pharmaceutical Sciences and The Open Research Center Project. We also acknowledge the financial support provided by Faculty of Pharmacy, Airlangga University, Indonesia.

References and notes

- 1. Hashimoto, G. Illustrated Cyclopedia of Brazilian Medicinal Plants 1996, 642.
- 2. (a) Torrenegra, G. R.; Escarria, R. S.; Bauereiss, P.; Achenbach, H. Planta Medica **1985**, 3, 276; (b) Barbosa-Filho, J. M.; Guedes, A. J. R.; Carlos, C. V.; Leitao, E. V.; Sobral, M.; Braz-Filho, R. *Journal of Asian Natural Products Research* **2004**, *1*, 11. Melo, F. N.; Navarro, V. R.; Marcelo, S.; Emidio, V. L.; Barbosa-Filho, J. M.; Braz-
- 3. Filho, R. Natural Products Letters 2001, 4, 261.
- Brown, M. P.; Thomson, R. H.; Hausen, B. M.; Simatupang, M. H. Justus Liebigs Annalen der Chemie **1974**, *8*, 1295.
 Mendes, F. N. P.; Silveira, E. R. Phytochemistry **1993**, 35, 1499.
 Mahajan, J. R.; Monterio, M. B. J. Chem. Soc. **1973**, 5, 520.
 Pudhom, K.; Sommit, D.; Suwankitti, N.; Petsom, A. J. Nat. Prod. **2007**, 70, 15120

- 1542.
- Stöcklin, W.; Waddell, T. G.; Geissman, T. A. *Tetrahedron* **1970**, *26*, 2397.
 Trager, W.; Jensen, J. B. *Science* **1976**, *193*, 673.

