

# Acute Interval and Continuous Moderate-Intensity Exercise Enhanced Circadian Thermogenic Activity through Browning-related Genes in Obese Adolescent Female

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**Abstract** Thermogenesis is associated with oxidation activity in muscle and fat tissue, the target of non-pharmacological therapy in preventing the increase in obesity. This research was designed to reveal the circadian profile of thermogenic gene expression after the acute interval and continuous moderate-intensity exercise. The subjects were 22 randomly selected obese adolescent females who met the predetermined inclusion criteria. The study subjects were then divided into three groups: control group (CG), acute interval moderate-intensity exercise group (AIMIE), and acute continuous moderate-intensity exercise group (ACMIE). Acute interval and continuous exercise were performed by running on a treadmill for 40-45 minutes, while moderate-intensity was defined as 60%-70% of the maximum heart rate (HR<sub>max</sub>). The blood samples were collected initially (pre-exercise), followed by 10 minutes, 6 hours, and 24 hours post-acute interval and continuous moderate-intensity exercise treatment. Measurement of peroxisome proliferator-activated receptor-gamma coactivator 1 alpha (PGC-1 $\alpha$ ) and fibronectin type III domain 5 (FNDC-5) expressions in protein level were confirmed by enzyme-linked immunosorbent assay (ELISA) method. Data were analyzed using one way-ANOVA and two way-ANOVA with a significant level of 5%. The findings suggest a substantial increase in the expression of PGC-1 $\alpha$  and FNDC-5 after exercise compared to before the workout. A significant difference in PGC-1 $\alpha$  and FNDC-5 expressions between the control group compared to AIMIE and ACMIE ( $p \leq 0.05$ ) has been observed. However, there is no significant difference in PGC-1 $\alpha$  and FNDC-5 expressions after exercise between AIMIE and ACMIE ( $p \geq 0.05$ ). In conclusion, acute interval and continuous moderate-intensity exercise increase the expression of thermogenesis-related genes. Hence, acute interval and continuous moderate-intensity exercise might be potential non-pharmacological therapy to prevent, reduce, and control the increasing prevalence of obesity.

**Keywords:** Thermogenic, exercise, PGC-1 $\alpha$ , FNDC-5, obesity

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## Introduction

Obesity has been classified as a global disease [1] that severely impacts human health [2]. Obesity is a disease that has caused a high risk of complications [3], disability [4] and premature death [5], and

morbidity and mortality in the global community [6]. This is because obesity can lead to an increased risk of cardiovascular disease [7], type 2 diabetes [8], mitochondrial dysfunction [9], metabolic disorders [10], and several types of cancer [6]. Previous studies recommend that obesity prevention and control can be carried out using approaches, such as eating behavior transformation [11], reducing calorie intake diet, giving nutritional supplements consumption [6], lifestyle modification [12], changing environmental conditions [1], and also a pharmacological approach by providing drugs [13]. However, some of the methods recommended by previous studies are still considered ineffective [1] because they increase risk factors for complications and cause dependence [6], and require significant costs [2]. This approach is considered effective in the short term [8] because it can only reduce lean body weight, lower basal metabolism, and increase the risk of nutritional deficiencies [11]. Therefore, a non-pharmacological approach based on exercise is required. Exercise carries physiological effects that significantly affect the body [13], one of which it increases the expression of the thermogenesis gene [14] and activates the adipose network, decreasing lipid accumulation [15]. Therefore, the preparation of an exercise program as a non-pharmacological therapy for obese individuals through increasing activity of thermogenic genes is interesting to be developed further [16].

Exercise increases muscle contraction, a contributor to thermogenesis induced by the activation of the uncoupling protein-1 (UCP-1) thermogenic gene [17]. Exercise increases the need for adenosine triphosphate (ATP), which will change the AMP/ATP ratio [18]. The change in the AMP/ATP ratio stimulates an increase in peroxisome proliferator-activated receptor-gamma coactivator 1 alpha (PGC-1 $\alpha$ ) [19] and fibronectin type III domain-containing protein 5 (FNDC-5), transformed into irisin molecules and is often found in circulation [20]. Increased irisin activates UCP-1 [21], causing white adipose tissue browning activity (WAT) [22]. Meanwhile, the browning in WAT triggers thermogenesis activity [9] via extracellular-signal-regulated kinase (ERKs) and p38 mitogen-activated protein kinase (p38-MAPK) pathways [23] and activation of lipolysis [24]. The increase in lipolysis causes the breakdown of triglycerides as an increased energy source [25], increases energy expenditure via oxidative phosphorylation [14], regulates glucose and lipid homeostasis [20], which affects the metabolic health [11]. Thus, thermogenesis is an attractive potential non-pharmacological therapy target for obesity prevention and management [24].

Exercise provides opportunities as a form of non-pharmacological therapeutic approach capable of involving integrative and adaptive responses at the tissue, organ, cellular, molecular, and system levels simultaneously [26]. Furthermore, exercise intensity has an essential role in response and adaptation [27], which affects the expression of thermogenic genes [28]. The results of previous *in vivo* animal models concluded that high-intensity exercise increased transformation to thermogenic genes faster than moderate-intensity exercise [29]. Inversely, other studies have concluded that moderate-intensity exercise is more beneficial than a high-intensity exercise in increasing response and adaptation to the browning gene and cause no systemic injury to muscles [15]. High-intensity exercise increases the browning gene adaptation faster, but it carries a higher risk of inflammation and systemic injury [17]. In addition, high-intensity exercise can also increase stressors and disrupt homeostasis [30] and triggers tissue damage [31]. Hence, moderate-intensity exercise is a safe exercise for obesity prevention and its complications [11]. Besides, it also follows the level of physical fitness of the general public, especially subjects with obesity [32].

Increased thermogenesis in exercise is an important marker that links metabolic regulation with targeted therapy to prevent and treat metabolic syndrome and obesity [33]. After exercise, therapeutic gene expression has been observed as a healing target in preventing and treating overweightness, globally. In addition, exercise is predicted to be an effective and strategic method for improving metabolic health [34] and improve metabolic profiles [35]. Although previous studies claim that exercise can enhance the performance of thermogenic gene expression related to metabolism [15, 29], but the effect of acute interval training and continuous moderate-intensity exercise on changing the thermogenesis profile of circadian genes in obese individuals has not been investigated, further. Therefore, this study seeks to uncover changes in the circadian profile of thermogenic gene expression after the acute interval and continuous moderate-intensity exercise. The acute interval and continuous moderate-intensity exercise are synthesized to enhance the circadian profile of thermogenic gene expression in obese individuals.

## Materials and Methods

### Experimental Design and Exercise Treatment

Based on the ethical standards of the Declaration of Helsinki 1975, this research procedure has received approval from the Health Research Ethics Commission (IRB) Faculty of Medicine, University of Brawijaya Malang (number 26/EC/KEPK-S1/02/2020). In this study, a truly experimental approach with a Basic Time Series Design using adolescent females was employed. The inclusion criteria for the subjects were

obese adolescent females with body mass index of 27-35 kg/m<sup>2</sup>, age of 18-22 years, percentage of body fat > 30%, normal blood pressure, resting heart rate (60-80 bpm), fasting blood glucose < 100 mg/dL, normal hemoglobin, excellent category of VO<sub>2</sub>max and classified as healthy based on a doctor's examination. All subjects had received information directly and in writing about the objectives, procedures, rights, and obligations in this study. Twenty-two prospective subjects met the inclusion criteria and stated their willingness to become research subjects. Then, the research subjects were divided randomly into three groups: control group (CG), acute interval moderate-intensity exercise group (AIMIE), and acute continuous moderate-intensity exercise group (ACMIE). The subjects were fasted for 8 hours starting at 10.00 p.m. before the exercise treatment on the next day.

The exercise intervention was given to the research subjects at 6.30–10.30 a.m. [36], which begins with a medical examination and blood sampling before exercise (pretest) PGC-1 $\alpha$ , FNDC-5, and UCP-1 examinations. After taking blood samples (before exercise), the subject was given a drink containing 5% glucose levels [37], then the subjects took a rest for 30 minutes [38]. After 30 minutes of resting, the subject was given an exercise intervention. The exercise was carried out in the Fitness Center of the Health Ministry of Malang with room temperature between 22–25 °C [39] with a humidity level of 50-70% [40]. Exercise interventions were completed using a treadmill (Richter Treadmill Semi-Commercial Evolution (4.0 HP DC); Richter Fitness, Taipei, Taiwan) with inclination levels of 0% [38]. It was carried out by running on a treadmill at intervals for group 2 (AIMIE) and continuously for group 3 (ACMIE) with moderate intensity. Before the core workout, a 5-minutes warm-up and 5 minutes cool-down after the core workout with low-intensity were completed. Meanwhile, the AIMIE group exercised for 45 minutes, consisting of running 5 minutes, followed by a rest period of 2.5 minutes (ratio of work to rest 1:1/2), repeated five times. The rest was carried out using active rest by running low-intensity [41], while ACMIE did a continuous exercise of moderate-intensity for 40 minutes continuously (without rest). The moderate intensity and light intensity were equal to 60%–70% and 50%–60% of HRmax, respectively [42]. Intensity measurement was carried out using a measurement formulation of HRmax: HRmax – age in years (220 – age in years) [39]. The heart rate was monitored during exercise using a polar heart rate monitor (Polar H 10 Heart Rate Sensor, USA. Inc).

### Baseline Data Collection

Anthropometric measurements of height used was a stadiometer (SECA, Chino, CA) while measuring body weight, body mass index, percentage of body fat, Fat Mass, Free Fat Mass, Muscle Mass, and Basal Metabolic Rate, were carried out using TANITA (Body Composition Analyzer DC3607601 (2) - 1604 FA, TANITA Corporation of America, Inc). Furthermore, pressure measurements used an automated device (OMRON Model HEM-7130 L, Omron Co., JAPAN) and for heart rate measurement using Polar (Polar H10 Heart Rate Sensor, USA, Inc). Measuring VO<sub>2</sub>max was completed using the Astrand-method with an ergo cycle device (Monark 828 E, Version 1010 Art. No: 7950-296, Vansbro, Sweden). Blood samples were collected four times for expression examination of PGC-1 $\alpha$ , FNDC5, and UCP-1, before exercise, 10 minutes after exercise, 6 hours after exercise, and 24 after exercise. After the second blood sample was taken (10 minutes after the exercise), the subjects were given fruit prayer consumption, food, drink and brought back to the guest house for rest and preparation for the third and fourth blood sampling. The subjects were fasted again for 2 hours before blood sampling for the third and fourth blood sampling. Meanwhile, the control group did not do any exercise but get the same procedure for blood sampling as the intervention group (AIMIE and ACMIE).

### Serological Assay

Blood samples were taken from 5 ml cubital veins [43]. The blood that had been drawn was put into a 6 ml vacutainer, then centrifuged for 15 minutes at 3000 rpm speed. Serum was separated and stored at -80 °C for level analysis of PGC-1 $\alpha$ , FNDC5, and UCP-1 in the Physiology Laboratory Faculty of Medicine Universitas Brawijaya Malang. Measurement of PGC-1 $\alpha$  levels was completed using an ELISA kit (Catalog No. E-EL-H1359; Elabscience, Inc., China, 2020) with a sensitivity level of PGC-1 $\alpha$  in the kit 0.10 ng/mL. Measurement of FNDC5 levels used an ELISA kit (Catalog No. E-EL-H2254; Elabscience, Inc., China, 2020) with a sensitivity level of FNDC-5 in the kit 0.1 ng mL. Meanwhile, the UCP levels -1 was measured by the ELISA method (Catalog No. E-EL-H1661; Elabscience, Inc., China, 2020) with UCP-1 sensitivity level in the 0.10 ng/mL kit. The sandwich and competitive ELISA method was used as a measurement method. Measurement of Fasting Blood Glucose used ACCU-CHEK (ACCU-CHEK® Performance, Mannheim, Germany) with mg/dL concentration units. Measurement of Hb using Easy Touch (Easy Touch GCHb, Taiwan) was carried out with a g/dL concentration unit.

### Data Analysis

The normality test was completed using the Shapiro-Wilk test, followed by the homogeneity test using the Levene test. Different tests using one-way-ANOVA, two-way-ANOVA and followed by Tukey's Honestly Significant Difference (HSD) post hoc test and Pearson product-moment correlation test were used to determine the relationship between PGC-1 $\alpha$  and anthropometrics and biomarkers of

thermogenesis. All data are presented as mean ± Standard Deviation (SD). All statistical analyzes used a significant level ( $p \leq 0.05$ ).

## Results and Discussion

The one Way-ANOVA Test results suggest no significant difference in the characteristics between the control group, the acute interval moderate-intensity exercise group, and the acute continuous moderate-intensity exercise group ( $p \geq 0.05$ ) (see **Table 1**, **Figure 1**, and **Figure 2**). Furthermore, no significant difference in the expression of the thermogenic genes PGC-1 $\alpha$ , FNDC-5, and UCP-1 baseline (before exercise) between CG, AIMIE, and ACMIE ( $p \geq 0.05$ ) (**Table 2**) has been observed. Importantly, the UCP-1 thermogenic gene expression at 10 minutes, 6 hours, and 24 hours after exercise is concluded based on the significant difference between CG, AIMIE, and ACMIE ( $p \leq 0.05$ ). Further analysis of UCP-1 thermogenic gene expression at 10 minutes following the exercise based on Tukey's HSD post hoc test results showed a significant difference between AIMIE and CG ( $p \leq 0.001$ ), along with ACMIE and CG ( $p \leq 0.001$ ). Simultaneously, there is no significant difference between AIMIE and ACMIE ( $p \geq 0.05$ ). The UCP-1 gene expression tested 6 hours after exercise of the control group is significantly different from the AIMIE and ACMIE groups ( $p \leq 0.001$ ). Meanwhile, no significant difference between AIMIE and ACMIE ( $p \geq 0.05$ ) has been observed. Similar results were also found in the measurement of UCP-1 expression at 24 hours post-exercise (**Figure 3**).

**Table 1.** Characteristics of the Research Subjects

Variable	Group			One Way-ANOVA <i>p-values</i>
	CG (n=7)	AIMIE (n=7)	ACMIE (n=8)	
Age (years)	19.71±1.49	19.57±1.51	20.00±1.52	0.854
Body Weight (kg)	72.99±5.71	73.58±8.34	74.47±7.45	0.923
Body Height (m)	1.57±0.04	1.58±0.06	1.58±0.05	0.963
BMI (kg/m <sup>2</sup> )	29.21±1.03	29.11±1.55	29.74±1.62	0.666
SBP (mmHg)	114.71±2.87	114.43±2.82	114.62±2.45	0.980
DBP (mmHg)	77.43±3.78	76.29±3.49	77.88±3.72	0.699
RHR (bpm)	73.86±8.69	70.57±8.10	74.50±4.24	0.544
VO <sub>2max</sub> (mL/kg/min)	28.61±3.81	26.23±1.14	27.31±1.22	0.193
FBG (mg/dL)	92.43±5.09	92.42±3.31	87.87±5.46	0.124
Hb (g/dL)	15.74±1.82	15.39±0.88	14.25±1.03	0.090
PBF (%)	45.71±3.83	43.91±2.24	44.02±4.05	0.563
FM (kg)	34.53±6.41	32.46±4.52	33.00±5.71	0.775
FFM (kg)	40.63±4.10	41.31±4.61	41.52±2.99	0.901
MM (kg)	38.24±3.76	38.86±4.23	39.05±2.73	0.905
BMR (kcal)	1361.00±142.23	1366.43±151.35	1380.12±97.29	0.958
Waist (cm)	93.29±13.88	85.57±8.79	88.37±7.99	0.392
Hip (cm)	108.14±4.29	108.00±6.73	108.50±5.42	0.984
WHR	0.82±0.03	0.79±0.05	0.81±0.04	0.305

**Description:** BMI: Body Mass Index; SBP: Systolic Blood Pressure; DBP: Diastolic Blood Pressure; RHR: Resting Heart Rate; FBG: Fasting Blood Glucose; PBF: Percentage Body Fat; FM: Fat Mass; FFM: Free Fat Mass; MM: Muscle Mass; BMR: Basal Metabolic Rate; WHR: Waist to Hip Ratio; VO<sub>2max</sub>: Volume Oxygen Maximum; CG: Control Group; AIMIE: acute interval moderate-intensity exercise; ACMIE: acute continuous moderate-intensity exercise. All data are presented with Mean±SD.

The results of serological testing for other thermogenic gene expression PGC-1 $\alpha$  at 6 hours and 24 hours after exercise showed no significant difference between CG, AIMIE, and ACMIE ( $p \geq 0.05$ ) (**Table 2**). However, the expression of the thermogenic PGC-1 $\alpha$  gene at 10 minutes after exercise showed significantly different results among the three groups ( $p \leq 0.05$ ). Besides, the results of the further analysis with Tukey's HSD post hoc test showed a significant difference between ACMIE and CG ( $p \leq 0.05$ ), no significant difference between AIMIE and CG ( $p \leq 0.05$ ), along with no substantial difference between AIMIE and ACMIE ( $p \geq 0.05$ ) (**Figure 4**). Interestingly, the thermogenic FNDC-5 gene expression analysis at 10 minutes, 6 hours, and 24 hours after exercise also presents a significant distinction between CG, AIMIE, and ACMIE of ( $p \leq 0.05$ ) (**Table 2**). The results of Tukey's HSD post hoc test of thermogenic gene expression FNDC5 at 10 minutes after exercise demonstrated a significant

difference between AIMIE and CG of ( $p \leq 0.05$ ), ACMIE, and CG of ( $p \leq 0.05$ ). However, no substantial difference between ACMIE and AIMIE ( $p \geq 0.05$ ) has been observed from that test results. Moreover, the expression of the thermogenic gene FNDC5 at 6 hours after exercise also presents a meaningful difference between AIMIE and CG ( $p \leq 0.05$ ), as well as between ACMIE and CG ( $p \leq 0.05$ ). However, there is no significant difference between ACMIE and AIMIE ( $p \leq 0.05$ ). Expression testing at 24 hours post-exercise showed a significant difference between ACMIE of and CG of ( $p \leq 0.05$ ), but no significant difference between AIMIE and CG ( $p \leq 0.05$ ) and ACMIE and AIMIE ( $p \geq 0.05$ ) (Figure 5). In the relationship analysis with the Pearson Product Moment correlation test, a significant relationship between PGC-1 $\alpha$  and body weight ( $r = -0.564$ ,  $p \leq 0.001$ ), BMI ( $r = -0.705$ ,  $p \geq 0.05$ ), PBF ( $r = -0.506$ ,  $p \geq 0.05$ ), and FM ( $r = -0.617$ ,  $p \leq 0.001$ ). PGC-1 $\alpha$  expression was also closely related to irisin ( $r = 0.759$ ,  $p \leq 0.001$ ), UCP-1 ( $r = 0.718$ ,  $p \leq 0.001$ ), betatrophin ( $r = -0.659$ ,  $p \leq 0.001$ ), MDA ( $r = -0.796$ ,  $p \leq 0.001$ ) and cortisol ( $r = -0.538$ ,  $p \leq 0.001$ ) have been identified (Table 3 and Figure 6).

**Table 2.** Analysis Results of PGC-1 $\alpha$ , FNDC-5, and UCP-1 Levels

Variable	Duration	Group			One Way-ANOVA <i>p-values</i>
		CG (n=7)	AIMIE (n=7)	ACMIE (n=8)	
PGC-1 $\alpha$ (ng/mL)	Baseline	2.31 $\pm$ 1.55	2.23 $\pm$ 1.99	2.25 $\pm$ 0.56	0.994
	10 min post-exercise	2.06 $\pm$ 1.26	3.57 $\pm$ 0.89*	3.95 $\pm$ 1.68*	$p \leq 0.05$
	6 hr post-exercise	2.32 $\pm$ 1.49	3.28 $\pm$ 1.76	3.02 $\pm$ 0.79	0.420
	24 hr post-exercise	2.27 $\pm$ 1.01	2.94 $\pm$ 1.79	2.62 $\pm$ 0.93	0.629
FNDC-5 (ng/mL)	Baseline	10.84 $\pm$ 3.13	10.67 $\pm$ 3.99	10.66 $\pm$ 1.86	0.992
	10 min post-exercise	10.48 $\pm$ 3.25	13.67 $\pm$ 0.95*	14.21 $\pm$ 0.77*	$p \leq 0.05$
	6 hr post-exercise	10.58 $\pm$ 3.41	13.55 $\pm$ 1.06*	13.88 $\pm$ 0.43*	$p \leq 0.05$
	24 hr post-exercise	10.38 $\pm$ 3.71	13.26 $\pm$ 0.69	13.63 $\pm$ 0.57*	$p \leq 0.05$
UCP-1 (ng/mL)	Baseline	3.67 $\pm$ 0.42	3.60 $\pm$ 1.28	3.91 $\pm$ 1.39	0.862
	10 min post-exercise	3.71 $\pm$ 0.63	5.57 $\pm$ 0.40*	6.39 $\pm$ 0.96*	$p \leq 0.001$
	6 hr post-exercise	3.69 $\pm$ 0.77	5.21 $\pm$ 0.51*	5.51 $\pm$ 0.38*	$p \leq 0.001$
	24 hr post-exercise	3.70 $\pm$ 0.33	4.95 $\pm$ 0.52*	5.12 $\pm$ 0.59*	$p \leq 0.001$

\*Significant vs. control group (CG) ( $p \leq 0.05$ ). All data are presented with Mean $\pm$ SD.

In general, the results suggest a significant difference in the expression of thermogenic genes in individual circadian cycles within obese subjects between both acute interval moderate-intensity exercise and acute continuous moderate-intensity exercise compared to the control group. The difference in the expression of the activity of thermogenic genes is strongly presumed to be an effect of the intervention because the characteristics of the subjects are associated with changes in the circadian cycle of thermogenic gene activity, such as UCP-1 and PGC-1 $\alpha$  [44]. Substantially, obese subjects are susceptible to specific exercises compared to normal weight subjects [7]. Obese individuals can carry out moderate-intensity exercise because they tend to have low muscle mass [45], high fat mass, and easy fatigue [30], while high-intensity exercise should be avoided [16]. Physiologically and psychologically, moderate-intensity exercise is very suitable, comfortable, and enjoyable for obese subjects [7], so based on our hypothesis, moderate-intensity exercise will significantly affect changes in the circadian cycle of thermogenic genes in the subjects of this study.

Based on the analysis results in the moderate-intensity interval and continuous exercise, the changes in the circadian cycle of the UCP-1 thermogenic gene at 10 minutes, 6 hours, and 24 hours post-intervention are different with the control group. It is in line with the results of previous studies that the increased expression of the UCP-1 thermogenic gene in obese non-diabetic adults after giving 70%-80% intensity exercise [46]. In their study, Tanaka *et al.* [15] also concluded an increase in UCP-1 expression in adult men with average body mass index after moderate-intensity walking exercise [15]. Research using experimental animals with weight exercise also concluded that exercise increases the UCP-1 thermogenic gene [47], activity of thermogenic genes such as PGC-1 $\alpha$ , UCP-1, as well as acid transport genes after moderate-intensity continuous exercise and high-intensity interval exercise [29]. The increase in the UCP-1 thermogenic gene is not only observed in regular exercise but also in acute exercise [20]. This increase is the effect of increased PGC-1 $\alpha$  expression caused by muscle contraction during exercise [48], which in turn increases FNDC5-induced irisin release [49]. Meanwhile, the increased irisin release enhances the activation of the UCP-1 gene [20], causes an increase in browning activity which accelerates the thermogenetic activity [9]. In addition, moderate-intensity acute exercise is a myokine stimulant [18], which will carry changes in the circadian cycle of thermogenic genes [50].



Interestingly, in our study with UCP-1 examination at 6 hours and 24 hours after exercise, the circadian cycle of the UCP-1 gene is lower compared to UCP-1 levels at 10 minutes after exercise. However, the decrease in UCP-1 levels at 6- and 24-hours post-workout remains higher than that of placebo. In line with our findings, Kim *et al.* stated that UCP-1 levels decrease at 12 hours post [51]. It is hypothesized that the increase of UCP-1 posts 10-minute moderate-intensity exercise strongly correlates to the irisin level. Also, previous study has mentioned that irisin expression grows highest 1 hour after exercise and decreases significantly 6 hours after exercise [52], while the increasing expression of the thermogenic UCP-1 gene activates irisin [20]. As the result, decreased irisin expression affects the UCP-1 thermogenic gene activity [53]. Interestingly, our results suggest that the UCP-1 circadian cycle changes have the same pattern as the PGC-1 $\alpha$  and FNDC-5 gene changes cycles (Figure 4-5), so that elevation of UCP-1 can be a potential for obesity exercise therapy [20].

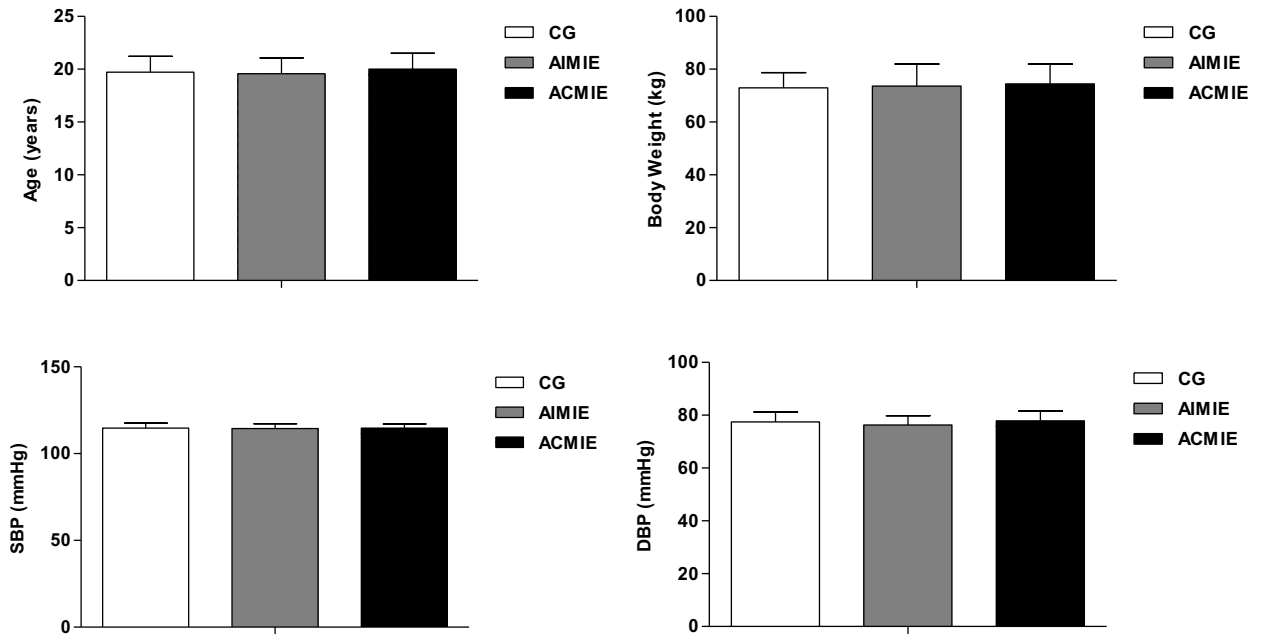
**Table 3.** Linear Correlation Analysis between PGC-1 $\alpha$  and Subjects' Characteristics

Parameter	PGC-1 $\alpha$	
	<i>r</i>	<i>p-values</i>
Age (years)	0.362	0.090
Weight (kg)	-0.564*	$p \leq 0.001$
Height (m)	-0.261	0.228
BMI (kg/m <sup>2</sup> )	-0.705*	$p \leq 0.001$
SBP (mmHg)	-0.385	0.069
DBP (mmHg)	-0.180	0.412
RHR (bpm)	-0.354	0.097
VO <sub>2</sub> max (mL/kg/min)	0.226	0.301
FBG (mg/dL)	-0.439*	0.036
PBF (%)	-0.506*	0.014
FM (kg)	-0.617*	$p \leq 0.001$
FFM (kg)	-0.379	0.075
MM (kg)	-0.449*	0.032
BMR (kcal)	-0.557	0.128
Waist (cm)	-0.362	0.090
Hip (cm)	-0.342	0.111
WHR	-0.209	0.338
Irisin (ng/mL)	0.750*	$p \leq 0.001$
UCP-1 (ng/mL)	0.718*	$p \leq 0.001$
IGF-1 (ng/mL)	0.743*	$p \leq 0.001$
Betatrophin (pg/mL)	-0.659*	$p \leq 0.001$
MDA (ng/mL)	-0.796*	$p \leq 0.001$
Cortisol (ng/mL)	-0.538*	$p \leq 0.001$

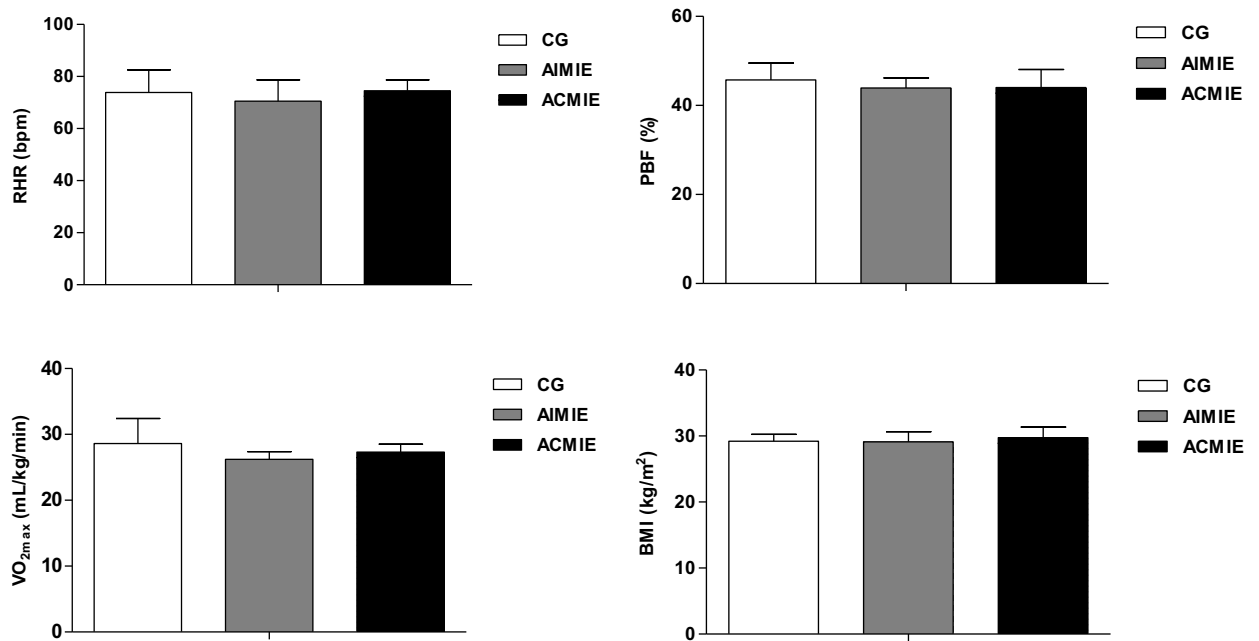
\* Significant with  $p \leq 0.05$  by Pearson's product-moment correlation test.

Based on the analysis results of moderate-intensity exercise, both interval and continuous exercise increase the circadian cycle changes of the thermogenic genes PGC-1 $\alpha$  and FNDC-5 at 10 minutes, 6 hours, and 24 hours after exercise, compared to the control group. The increase in PGC-1 $\alpha$  after exercise stimulates a growth in FNDC-5 protein, which cleavages to form irisin [54], activating the UCP-1 thermogenic gene to enhance thermogenesis [14]. Previous studies also reported higher PGC-1 $\alpha$  in experimental mice after swimming treatment than in the control group [55]. The same results have been reported on the mouse model after 30 minutes of treadmill exercise [53]. Lee *et al.* [56], in their research, also concluded that moderate-intensity workout (60%-70% VO<sub>2</sub>max) increases PGC-1 $\alpha$  expression [56]. The results of other studies also claim that 45 minutes of exercise with 70% VO<sub>2</sub>max intensity increases PGC-1 $\alpha$  gene expression in individual circadian cycles. That peak PGC-1 $\alpha$  gene expression is estimated to occur 2 hours after exercise [57]. Several other studies also reported that administering moderate-intensity, high-intensity, and exercise at the anaerobic threshold of peak PGC-1 $\alpha$  gene expression occurred 3 hours after exercise [58]. The peak is observed between 2-3 hours [59], while in experimental animals, it is estimated 3 hours after exercise [48]. Furthermore, the decreasing trend of PGC-1 $\alpha$  in the mouse model occurred 6 hours after 30 minutes of treadmill exercise [53]. Changes in the circadian cycle of the PGC-1 $\alpha$  thermogenic gene are closely related to irisin [60] in triggering browning [28]. Meanwhile, moderate-intensity exercise has a better response to the circadian cycle change of thermogenic gene

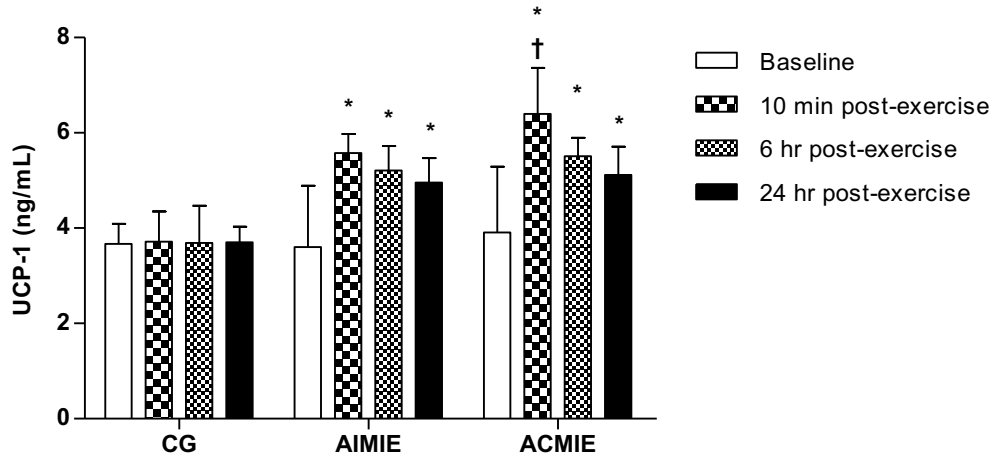
expression than high intensity or anaerobic threshold [58]. The increase in the PGC-1 $\alpha$  thermogenic gene in moderate-intensity exercise is extremely substantial as it modulates various metabolic processes, increases mitochondrial biogenesis, increases browning and thermogenesis so that it becomes a potential therapeutic target [61]. Hence, moderate-intensity exercise is expected to attract public interest to adopt a healthy and active lifestyle for non-pharmacological therapy, especially in obese subjects.



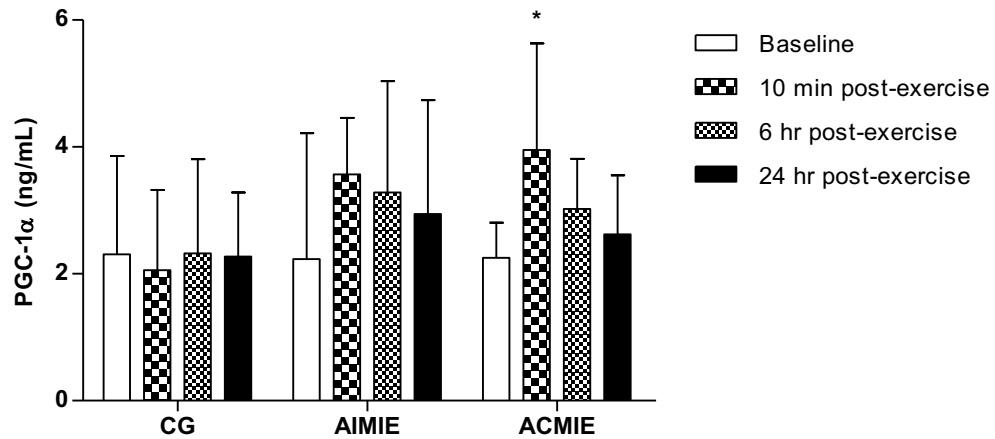
**Figure 1.** Bodyweight, age, blood pressures (systole and diastole) between the research subject groups



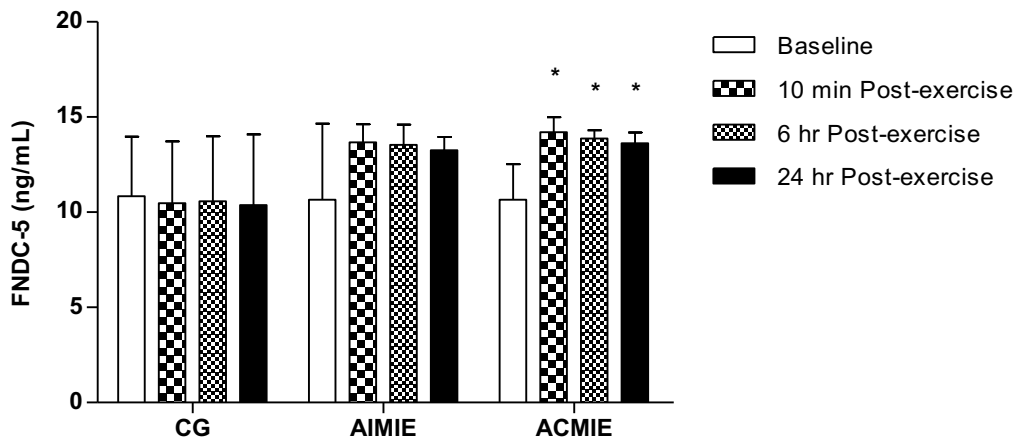
**Figure 2.** RHR, FBF, VO<sub>2max</sub>, and BMI of Each Research Subject Group



**Figure 3.** Pretest and Posttest UCP-1 Expression on Each Group  
 \*Significant vs. baseline ( $p \leq 0.05$ ). †Significant vs 24 hr post-exercise ( $p \leq 0.05$ ).

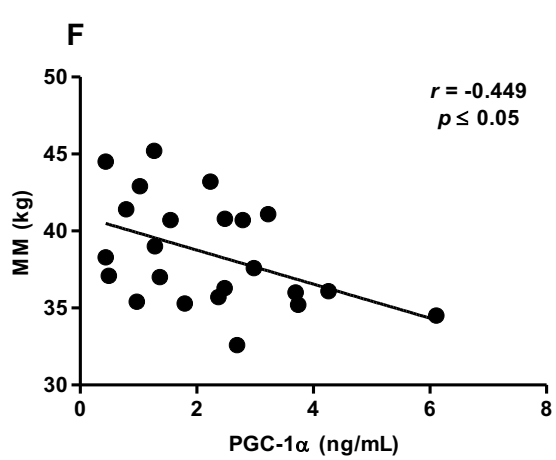
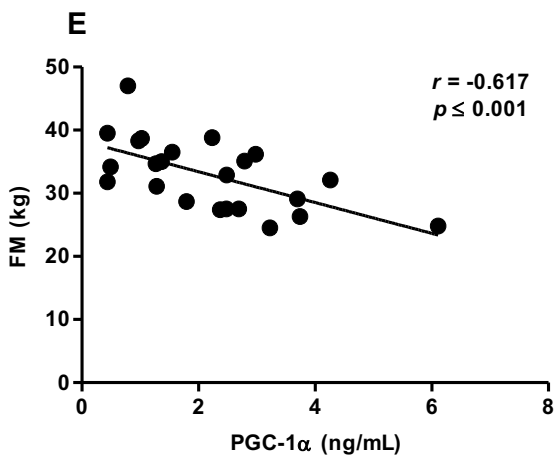
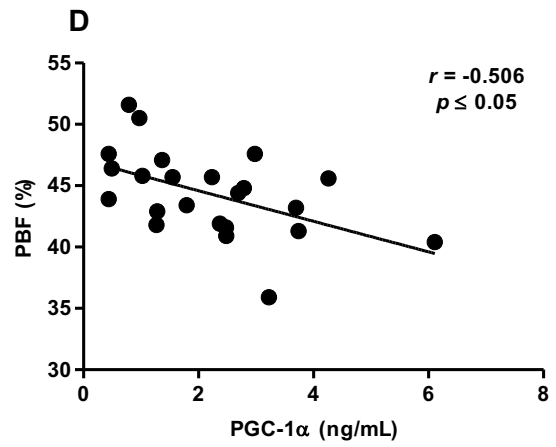
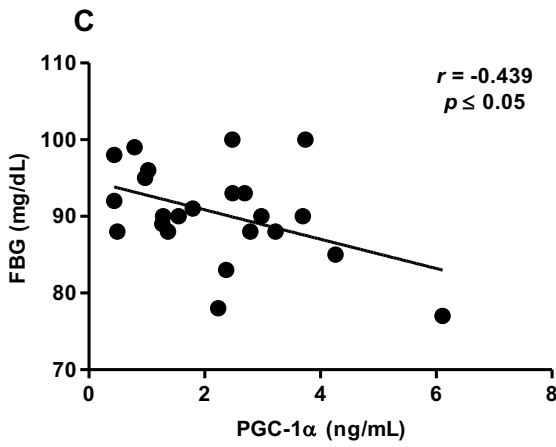
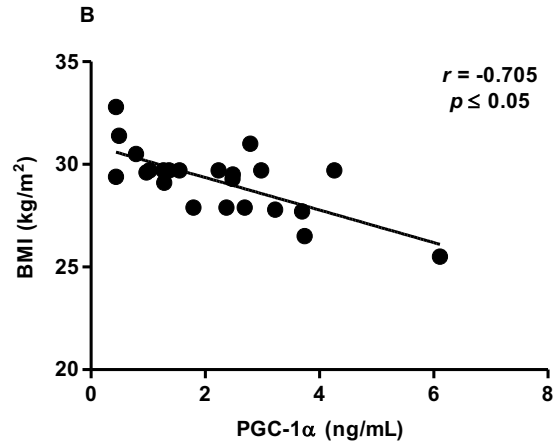
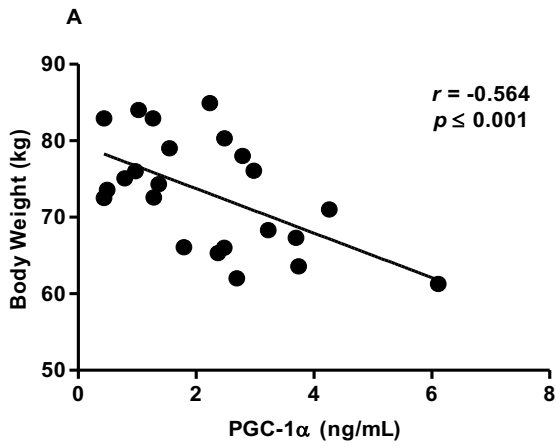


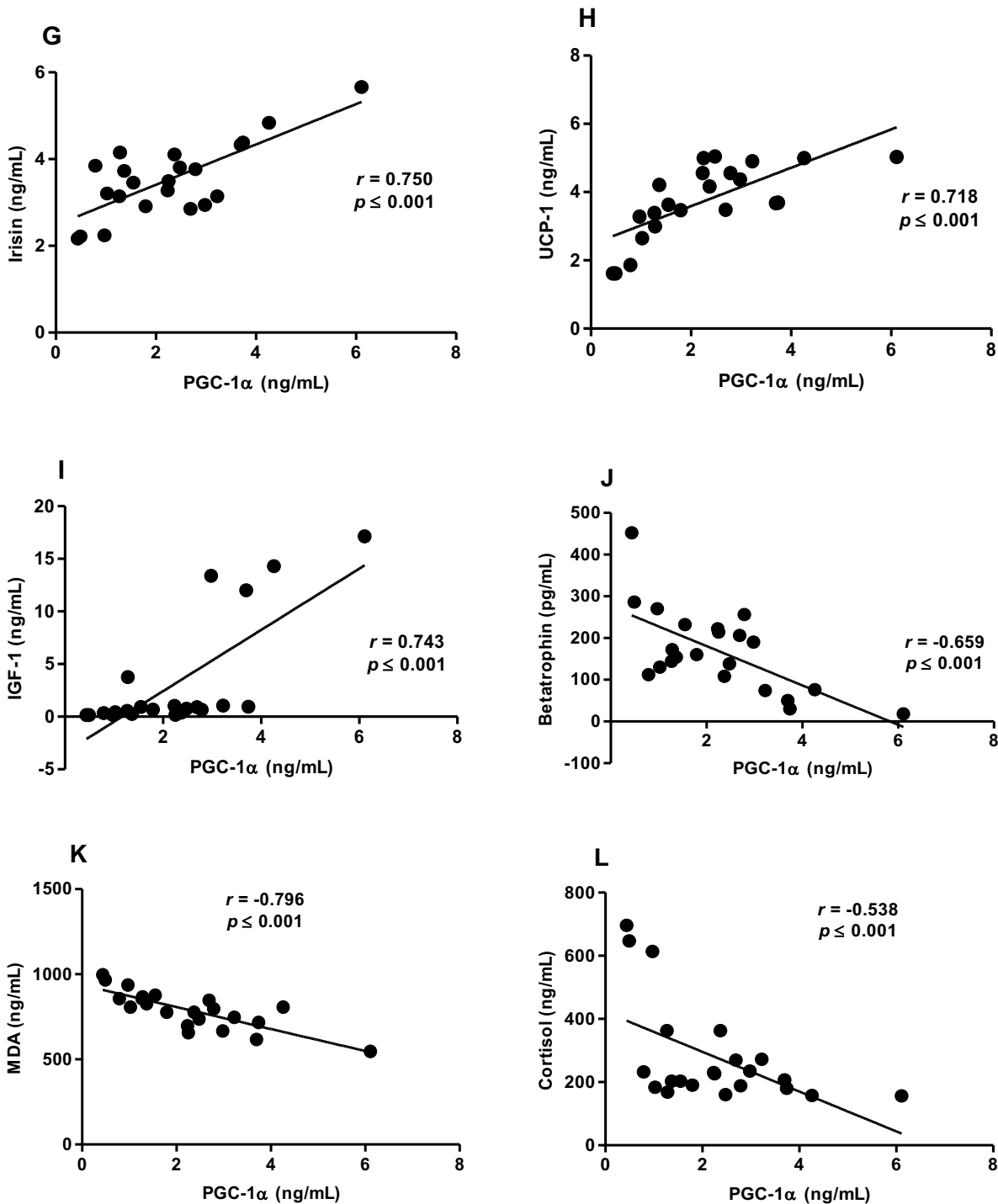
**Figure 4.** Pretest and Posttest PGC-1α Expression on Each Group  
 \*Significant vs. baseline ( $p \leq 0.05$ ).



**Figure 5.** Pretest and Posttest FNDC5 Expression on Each Group  
 \*Significant vs. baseline ( $p \leq 0.05$ ).







**Figure 6.** The relation between PGC-1α, anthropometric measurements, metabolism, and thermogenic genes, along with physiological and oxidative stress. The significant linear correlation between parameters was visualized in the plot model ( $p \leq 0.05$ ). Picture **A**. Correlation between PGC-α and bodyweight; **B**. Correlation between PGC-1α and BMI; **C**. Correlation between PGC-1α with FBG; **D**. Correlation between PGC-1α and PBF; **E**. Correlation between PGC-1α and FM; **F**. Correlation between PGC-1α and MM; **G**. Correlation between PGC-1α and irisin; **H**. Correlation between PGC-1α and UCP-1; **I**. Correlation between PGC-1α and IGF-1; **J**. Correlation between PGC-1α and Betatrophin; **K**. Correlation between PGC-1α and MDA; and **L**. Correlation between PGC-1α and Cortisol.

Changes in the circadian cycle of thermogenic genes in this study present a decreasing trend at 6 hours to 24 hours after exercise compared to 10 minutes after exercise, but it remains higher than in the control group (Table 2 and Figure 3-5). Our results found coordinated circadian cycle changes in the thermogenic genes PGC-1 $\alpha$ , FNDC-5, and UCP-1 at 10 minutes, 6 hours, and 24 hours after the interval and continuous exercise in obese female subjects. Previous studies also identify coordinated changes in expression of irisin, FNDC-5, and PGC-1 $\alpha$  after 1 hour of exercise in mice in which the expression of these genes turns similar to the control group 24 hours after exercise [58]. However, in our study, 24 hours after exercise, the thermogenic gene expression of PGC-1 $\alpha$ , FNDC-5, and UCP-1 remain higher than the control group. Previous studies have also reported changes in the circadian cycle of the thermogenic gene PGC-1 $\alpha$  6 hours after exercise [59]. Besides, the results of this study also find a positive correlation between PGC-1 $\alpha$  and FNDC-5, irisin and UCP-1 (Table 3 and Figure 6). It signifies that thermogenic genes are interrelated and influence each other, with PGC-1 $\alpha$  as the master regulator of biogenesis [62]. It is hypothesized that 24 hours after exercise, the thermogenic gene is expected to remain active. This condition is favorable and attractive for the browning process in maintaining homeostasis in response to decrease AMP/ATP ratio [63] and increase biogenesis in mitochondria [64]. A decrease in the AMP/ATP ratio leads to active AMPK, which is observed to occur 3 hours after exercise [59]. AMPK activation triggers activation of the PGC-1 $\alpha$  gene receptor [61], stimulates an increase in FNDC-5, which then cleaves into circulating irisin to activate UCP-1 [65]. Consequently, PGC-1 $\alpha$ -induced UCP-1 activation increases browning [46]. This process enhances energy generation through oxidative phosphorylation and energy expenditure [61]. It also improves the browning capacity, which can oxidize glucose and lipids as an energy source [4]. Thus, it becomes an attractive target for controlling obesity [66].

Furthermore, our results indicated that circadian cycle changes in thermogenic gene expression are detected 6 hours and 24 hours after exercise. It is intriguing to see that the thermogenic gene cycle is non-permanent and transient (Figure 3-5). Previous studies have also reported changes in the thermogenic gene cycle PGC-1 $\alpha$ , FNDC-5, and UCP-1 decreased after acute exercise [53] and chronicles [49]. The activity of thermogenic genes is essential as a regulator of energy metabolism [67] to enhance mitochondrial biogenesis [61] and increase oxidative phosphorylation [33]. This is presumed to be able to prevent the development of obesity [11]. Therefore, previous studies recommend that exercise should be done repeatedly and continuously [68], with the right intensity [32]. This will improve the PGC-1 $\alpha$ , FNDC-5, and UCP-1 thermogenic genes [22]. The increasing thermogenic gene on exercise balances glucose through affected gene expressions, such as glucose transporter type 4 (GLUT4) and peroxisome proliferator-activated receptor- $\alpha$ , which play a role in regulating glycogenase (PCK1) and gluconeogenesis (PYG) [23]. The results of this study also identify a negative correlation of the PGC-1 $\alpha$  thermogenic gene with betatrophin (Table 3 and Figure 6). Similarly, previous studies have also reported an increase in PGC-1 $\alpha$  expression followed by a decrease in betatrophin [69]. Decreased expression of betatrophin affects the active lipoprotein lipase (LPL) enzyme [25] and increases the use of triglycerides as an energy source [23]. Consequently, it increases oxidative phosphorylation in the mitochondria [70]. Therefore, the PGC-1 $\alpha$ , FNDC5, and UCP-1 thermogenic genes activities must be maintained to increase browning activity with increased thermogenesis [1]. The increase in thermogenesis affects the increasing energy expenditure through oxidation so that it decreases the fat deposits [22]. However, exercise must be done with the right intensity and according to ability [11] considering that it also carries the potential to increase the physiological stress hormone cortisol [71].

Importantly, the correlation analysis results in this study indicate a negative relationship between thermogenic gene expression and physiological and oxidative stress (Table 3 and Figure 6). Physiological stress reduces ATP, increases the ATP/AMP ratio, and activates AMPK, thereby inhibiting PGC-1 $\alpha$  expression [72]. It also causes metabolic disorders and lowers immunity [73], and is a risk factor for increased disease and a cause for exercise maladaptations [74]. On the other hand, oxidation stress is a cause of tissue damage, especially in the mitochondrial membrane [75]. This signifies that thermogenic genes also regulate antioxidant gene expression in the mitochondria to prevent oxidative injury and mitochondrial dysfunction [33]. However, in obese subjects, thermogenic genes such as the PGC-1 $\alpha$  gene and UCP-1 are lower than in normal-weight subjects [76]. This low thermogenic gene may also be related to the low IGF-1 as a regulator of muscle mass and muscle hypertrophy [77]. The greater the muscle mass increases the PGC-1 $\alpha$  expression and the biogenesis ability [78]. Obese subjects also had low UCP-1 thermogenic gene activity [79], along with minimum influenced mitochondrial function and browning [34]. Obese subjects are observed to have lower antioxidant activity compared to normal-weight subjects, so that they have higher chance to get oxidative stress [33]. Obese subjects also have a relatively high level of physiological stress [80], while exercise is classified as a potential stressor for the body [30]. Therefore, exercise for these people must be properly regulated, with an appropriate and measured intensity [81]. In addition to improving physical fitness [32], immunity health [30], and increase endorphins [82], exercise also plays a role in improving the performance of thermogenic genes, which

prevents the increase in malondialdehyde (MDA) [83]. Thus, the exercise is essential as a non-pharmacological strategy and method [84]. However, the exercise intensity must be measured and following the body's ability, so that exercise can be enjoyable and cause no stress [72]. A measurable exercise is predicted to have an optimal effect on skeletal muscle performance and energy metabolism [28]. Exercise has an appeal for society for lifestyle modifications that contribute to preventing obesity, metabolic syndrome and metabolic disorders, and increasing the physiological capacity of the body [11]. Acute moderate-intensity interval and continuous exercise in this study contribute to new theoretical development as a non-pharmacological and physiological approach in preventing obesity and reducing its risk factors. Exercise with moderate-intensity intervals continuously changes the circadian cycle of thermogenesis genes, positively impacting the improvement of metabolic health in obese subjects. Therefore, moderate-intensity exercise is the proper workout that follows the subject's ability, so it will be a strategic and attractive approach for vulnerable or untrained groups in preventing obesity and reducing its risk factors. The use of interval and continuous exercise with moderate-intensity in obese female subjects is a new approach expected to become a model for active lifestyle modification in preventing obesity in the future.

Even though our results demonstrate a significant effect on thermogenic genes' activity in the circadian cycle of obese individuals, this research still has some limitations. First, the research did not measure the circadian cycle of thermogenic genes that changes every hour, so the thermogenic gene's circadian peak was not investigated. Second, this research only employed obese subjects. It did not compare the changes in the circadian cycle of thermogenic genes with healthy subjects or normal-weight subjects after acute exercise. However, the circadian cycle pattern of thermogenic genes changes in obese individuals and its relationship with stress parameters can provide primary data. Besides, it also gives an initial picture that changes in the circadian cycle of thermogenic genes have an extensive effect on metabolism and can be used as a strategy for non-pharmacological therapy in preventing and treating, obesity, metabolic syndrome, and other metabolic disorders.

## Conclusions

Acute and continuous exercise with moderate intensity can increase the expression of thermogenic genes in the circadian cycle of obese subjects. Hence, acute exercise with moderate-intensity may be a model for developing potential non-pharmacological therapies to prevent, treat and control the increase in obesity and other metabolic disorders.

## Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

## Conflicts of interest

The authors declare that they have no competing interests.

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## References

- [1] M. F. Hussain, A. Roesler, and L. Kazak, "Regulation of adipocyte thermogenesis: mechanisms controlling obesity," *The FEBS journal*, vol. 287, no. 16, pp. 3370–3385, 2020. <https://doi.org/10.1111/febs.15331>.
- [2] N. K. Tayagi, A. Solanky, S. N. Jamali, M. Azharuddin, K. Ali, and I. Ahmad, "Aerobic Training, in Combination with Listening Music, Changes Post-Exercise Cardiac Autonomic Function in Collegiate Overweight and Obese Individuals," *Asian Journal of Sports Medicine*, vol. 11, no. 1, pp. e97122, 2020. <https://doi.org/10.5812/asjasm.97122>.
- [3] J. Kim, and Y. Jee, "EMS-effect of Exercises with Music on Fatness and Biomarkers of Obese Elderly Women," *Medicina (Kaunas)*, vol. 56, no. 4, pp. 158, 2020. <https://doi.org/10.3390/medicina56040158>.
- [4] E. A. Castro, E. V. Carraça, R. Cupeiro, B. López-Plaza, P. J. Teixeira, D. González-Lamuño, and A. B. Peinado, "The Effects of the Type of Exercise and Physical Activity on Eating Behavior and Body Composition in Overweight and Obese Subjects. *Nutrients*, vol. 12, no. 2, pp. 557, 2020. <https://doi.org/10.3390/nu12020557>.
- [5] L. G. Leal, M. A. Lopes, and M. L. Batista, Exercise-Induced Myokines and Muscle-Adipose Tissue Crosstalk: A Review of Current Knowledge and the Implications for Health and Metabolic Diseases. *Frontiers in Physiology*, vol. 9, 2018. <https://doi.org/10.3389/fphys.2018.01307>.
- [6] Wylie-Rosett, J., and S. S. Jhangiani, "Obesity and Disease in an Interconnected World: A Systems Approach to Turn Huge Challenges into Amazing Opportunities," *Sharjah: Bentham Science Publishers*, pp. 3–29, 2015. <https://doi.org/10.2174/9781681080369115010006>.
- [7] I. L. P. Bonfante, M. P. T. Chacon-Mikahil, D. T. Brunelli, A. F. Gáspari, R. G. Duft, A. G. Oliveira, T. G. Araujo, M. J. A. Saad, and C. R. Cavaglieri, "Obese with higher FNDC5/Irisin levels have a better metabolic profile, lower lipopolysaccharide levels and type 2 diabetes risk," *Archives of Endocrinology and Metabolism*, vol. 61, no. 6, pp. 524–533, 2017. <https://doi.org/10.1590/2359-3997000000305>.
- [8] G. Sanchez-Delgado, B. Martinez-Tellez, J. Olza, C. M. Aguilera, Á. Gil, and J. R. Ruiz, "Role of Exercise in the Activation of Brown Adipose Tissue," *Annals of Nutrition and Metabolism*, vol. 67, no. 1, pp. 21–32, 2015. <https://doi.org/10.1159/000437173>.
- [9] Y. Onder, and C. B. Green, "Rhythms of metabolism in adipose tissue and mitochondria. *Neurobiology of Sleep and Circadian Rhythms*, vol. 4, pp. 57–63, 2018. <https://doi.org/10.1016/j.nbscr.2018.01.001>.
- [10] A. W. C. Man, H. Li, and N. Xia, "Impact of Lifestyles (Diet and Exercise) on Vascular Health: Oxidative Stress and Endothelial Function," *Oxidative Medicine and Cellular Longevity*, vol. 2020, Article ID 1496462, pp. 1–22, 2020. <https://doi.org/10.1155/2020/1496462>.
- [11] A. Petridou, A. Siopi, and V. Mougios, "Exercise in the management of obesity," *Metabolism*, vol. 92, pp. 163–169, 2019. <https://doi.org/10.1016/j.metabol.2018.10.009>.
- [12] J. J. Ruiz-Ramie, J. L. Barber, and M. A. Sarzynski, "Effects of exercise on HDL functionality," *Current Opinion in Lipidology*, vol. 30, no. 1, pp. 16–23, 2019. <https://doi.org/10.1097/MOL.0000000000000568>.
- [13] H. Bekele, A. Asefa, B. Getachew, A. M. Belete, "Barriers and Strategies to Lifestyle and Dietary Pattern Interventions for Prevention and Management of TYPE-2 Diabetes in Africa, Systematic Review," *Journal of Diabetes Research*, vol. 2020, Article ID 7948712, pp. 1–14, 2020. <https://doi.org/10.1155/2020/7948712>.
- [14] P. C. Dinas, I. M. Lahart, J. A. Timmons, P-A. Svensson, Y. Koutedakis, A. D. Flouris, and G. S. Metsios, "Effects of physical activity on the link between PGC-1 $\alpha$  and FNDC5 in muscle, circulating Irisin and UCP1 of white adipocytes in humans: A systematic review," *F1000Research*, vol. 6, 286, 2017. <https://doi.org/10.12688/f1000research.11107.1>.
- [15] R. Tanaka, S. Fuse, M. Kuroiwa, S. Amagasa, T. Endo, A. Ando, R. Kime, Y. Kurosawa, T. Hamaoka, "Vigorous-Intensity Physical Activities Are Associated with High Brown Adipose Tissue Density in Humans," *International Journal of Environmental Research and Public Health*, vol. 17, no. 8, pp. 2796, 2020. <https://doi.org/10.3390/ijerph17082796>.
- [16] Tanaka, H., Ueno, S., Aoyagi, R., Hatamoto, Y., Jackowska, M., Shiose, K., and Higaki, Y, "Easily performed interval exercise induces to increase in skeletal muscle PGC-1 $\alpha$  gene expression." *Integrative Molecular Medicine*, vol. 4., no. 4, pp. 1–4, 2017. <https://doi.org/10.15761/IMM.1000293>.
- [17] J. G. Knudsen, M. Murholm, A. L. Carey, R. S. Biensø, A. L. Basse, T. L. Allen, J. Hidalgo, B. A. Kingwell, M. A. Febbraio, J. B. Hansen, and H. Pilegaard, "Role of IL-6 in Exercise Training- and Cold-Induced UCP1 Expression in Subcutaneous White Adipose Tissue," *PLoS ONE*, vol. 9, no. e84910, 2014. <https://doi.org/10.1371/journal.pone.0084910>.
- [18] C. Laurens, A. Bergouignan, and C. Moro, "Exercise-Released Myokines in the Control of Energy Metabolism," *Frontiers in Physiology*, vol. 11, no. 91, 2020. <https://doi.org/10.3389/fphys.2020.00091>.
- [19] L. Elizondo-Montemayor, A. M. Gonzalez-Gil, O. Tamez-Rivera, C. Toledo-Salinas, M. Peschard-Franco, N. A. Rodríguez-Gutiérrez, C. Silva-Platas, and G. Garcia-Rivas, "Association between Irisin, hs-CRP, and Metabolic Status in Children and Adolescents with Type 2 Diabetes Mellitus," *Mediators of Inflammation*, vol. 2019, Article ID 6737318, pp. 1–13, 2019. <https://doi.org/10.1155/2019/6737318>.
- [20] Ikeda, K., and T. Yamada, "UCP1 Dependent and Independent Thermogenesis in Brown and Beige Adipocytes," *Frontiers in Endocrinology*, vol. 11, no. 498, 2020. <https://doi.org/10.3389/fendo.2020.00498>.
- [21] L. Catalano-Iniesta, V. Sánchez Robledo, M. C. Iglesias-Osma, A. G. Albiñana, S. Carrero, E. J. Blanco, M. Carretero-Hernández, J. Carretero, and M. J. García-Barrado, "Evidences for Expression and Location of ANGPTL8 in Human Adipose Tissue," *Journal of Clinical Medicine*, vol. 9, no. 2, pp. 512, 2020. <https://doi.org/10.3390/jcm9020512>.
- [22] N. Perakakis, G. A. Triantafyllou, J. M. Fernández-Real, J. Y. Huh, K. H. Park, J. Seufert, C. S. Mantzoros, "Physiology and role of irisin in glucose homeostasis," *Nature Reviews Endocrinology*, vol. 13, no. 6, pp. 324–337, 2017. <https://doi.org/10.1038/nrendo.2016.221>.
- [23] M. Enteshary, F. Esfarjani, and J. Reisi, "Comparison of the Effects of Two Different Intensities of Combined Training on Irisin, Betatrophin, and Insulin Levels in Women with Type 2 Diabetes," *Asian Journal of Sports Medicine*, vol. 10, no. 2, pp. e68943, 2019. <https://doi.org/10.5812/asjasm.68943>.



- [24] F. Rabiee, L. Lachinani, S. Ghaedi, M. H. Nasr-Esfahani, T. L. Megraw, and K. Ghaedi, "New insights into the cellular activities of Fndc5/Irisin and its signaling pathways," *Cell & Bioscience*, vol. 10, no. 51, 2020. <https://doi.org/10.1186/s13578-020-00413-3>.
- [25] M. Abu-Farha, D. Sriraman, P. Cherian, I. AlKhairi, N. Elkum, K. Behbehani, and J. Abubaker, "Circulating ANGPTL8/Betatrophin Is Increased in Obesity and Reduced after Exercise Training," *PLoS One*, vol. 11, no. e0147367, 2016. <https://doi.org/10.1371/journal.pone.0147367>.
- [26] G. Z. Schaun, C. L. Alberton, D. O. Ribeiro, and S. S. Pinto, "Acute effects of high-intensity interval training and moderate-intensity continuous training sessions on cardiorespiratory parameters in healthy young men," *European Journal of Applied Physiology*, vol. 117, no. 11, pp. 1437–1444, 2017. <https://doi.org/10.1007/s00421-017-3636-7>.
- [27] C. Granata, R. S. F. Oliveira, J. P. Little, K. Renner, and D. J. Bishop, "Sprint-interval but not continuous exercise increases PGC-1 $\alpha$  protein content and p53 phosphorylation in nuclear fractions of human skeletal muscle," *Scientific Reports*, vol. 7, no. 44227, 2017. <https://doi.org/10.1038/srep44227>.
- [28] D. V. Popov, "Adaptation of Skeletal Muscles to Contractile Activity of Varying Duration and Intensity: The Role of PGC-1 $\alpha$ ," *Biochemistry (Moscow)*, vol. 83, no. 6, pp. 613–628, 2018. <https://doi.org/10.1134/S0006297918060019>.
- [29] M. Khalafi, H. Mohebbi, M. E. Symonds, P. Karimi, A. Akbari, E. Tabari, M. Faridnia, and K. Moghaddami, "The Impact of Moderate-Intensity Continuous or High-Intensity Interval Training on Adipogenesis and Browning of Subcutaneous Adipose Tissue in Obese Male Rats," *Nutrients*, vol. 12, no. 4, pp. 925, 2020. <https://doi.org/10.3390/nu12040925>.
- [30] C. A. M. Gonçalves, P. M. S. Dantas, I. K. dos Santos, M. Dantas, D. C. P. da Silva, B. G. A. T. Cabral, R. O. Guerra, and G. B. C. Júnior, "Effect of Acute and Chronic Aerobic Exercise on Immunological Markers: A Systematic Review," *Frontiers in Physiology*, vol. 10, no. 1602, 2020. <https://doi.org/10.3389/fphys.2019.01602>.
- [31] Sugiharto, D. Merawati, R. G. Kinanti, H. Susanto, A. Taufiq, and Sunaryono, "The Attenuation of Physical-Physiological Stresses through Musical-High Intensity Exercise Co-Treatment in Non-Athlete Individual," *Journal of Physics: Conference Series*, vol. 1093, no. 012026, 2018. <https://doi.org/10.1088/1742-6596/1093/1/012026>.
- [32] B. M. Antunes, F. E. Rossi, L. M. Oyama, J. C. Rosa-Neto, and F. S. Lira, "Exercise intensity and physical fitness modulate lipoproteins profile during acute aerobic exercise session," *Scientific Reports*, vol. 10, Article number 4160, 2020. <https://doi.org/10.1038/s41598-020-61039-6>.
- [33] S. Rius-Pérez, I. Torres-Cuevas, I. Millán, Á. L. Ortega, and S. Pérez, "PGC-1 $\alpha$ , Inflammation, and Oxidative Stress: An Integrative View in Metabolism," *Oxidative Medicine and Cellular Longevity*, vol. 2020, Article ID 1452696, pp. 1–20, 2020. <https://doi.org/10.1155/2020/1452696>.
- [34] C. Larson, M. Opichka, M. L. McGlynn, C. W. Collins, and D. Slivka, "Exercise- and Cold-Induced Human PGC-1 $\alpha$  mRNA Isoform Specific Responses," *International Journal of Environmental Research and Public Health*, vol. 17, no. 16, pp. 5740, 2020. <https://doi.org/10.3390/ijerph17165740>.
- [35] I. G. Fatouros, "Is irisin the new player in exercise-induced adaptations or not? A 2017 update," *Clinical Chemistry and Laboratory Medicine (CCLM)*, vol. 56, no. 4, pp. 525-548, 2018. <https://doi.org/10.1515/cclm-2017-0674>.
- [36] H. Nygaard, G. Slettaløkken, G. Vegge, I. Hollan, J. E. Whist, T. Strand, B. R. Rønnestad, and S. Ellefsen, "Irisin in Blood Increases Transiently after Single Sessions of Intense Endurance Exercise and Heavy Strength Training," *PLOS ONE*, vol. 10, no. e0121367, 2015. <https://doi.org/10.1371/journal.pone.0121367>.
- [37] L. Parker, C. S. Shaw, L. Banting, I. Levinger, K. M. Hill, A. J. McAinch, and N. K. Stepto, "Acute Low-Volume High-Intensity Interval Exercise and Continuous Moderate-Intensity Exercise Elicit a Similar Improvement in 24-h Glycemic Control in Overweight and Obese Adults," *Frontiers in Physiology*, vol. 7, no. 661, 2017. <https://doi.org/10.3389/fphys.2016.00661>.
- [38] Y. Tsuchiya, S. Mizuno, and K. Goto, "Irisin response to downhill running exercise in humans," *J Exerc Nutrition Biochem*, vol. 22, no. 2, pp. 12–17, 2018. <https://doi.org/10.20463/jenb.2018.0011>.
- [39] V. O. A. Santos, R. A. V. Browne, D. C. Souza, V. A. F. Matos, G. A. D. Macêdo, L. F. Farias-Junior, J. C. Farias-Júnior, E. C. Costa, and A. P. T. Fayh, "Effects of High-Intensity Interval and Moderate-Intensity Continuous Exercise on Physical Activity and Sedentary Behavior Levels in Inactive Obese Males: A Crossover Trial," *J Sports Sci Med*, vol. 18, no. 3, pp. 390–398, 2019.
- [40] S. Qiu, E. Bosnyák, G. Treff, J. M. Steinacker, A. M. Niefß, K. Krüger, F. C. Mooren, M. Zügel, and U. Schumann, "Acute exercise-induced irisin release in healthy adults: Associations with training status and exercise mode," *European Journal of Sport Science*, vol. 18, no. 9, pp. 1226–1233, 2018. <https://doi.org/10.1080/17461391.2018.1478452>.
- [41] G. A. Tew, D. Leighton, R. Carpenter, S. Anderson, L. Langmead, J. Ramage, J. Faulkner, E. Coleman, C. Fairhurst, M. Seed, and L. Bottoms, "High-intensity interval training and moderate-intensity continuous training in adults with Crohn's disease: a pilot randomised controlled trial," *BMC Gastroenterology*, vol. 19, no. 1, pp. 19, 2019. <https://doi.org/10.1186/s12876-019-0936-x>.
- [42] K. A. Dias, C. B. Ingul, A. E. Tjonna, S. E. Keating, S. R. Gomersall, T. Follstad, M. S. Hosseini, S. M. Hollekim-Strand, T. B. Ro, M. Haram, E. M. Huuse, P. S. W. Davies, P. A. Cain, G. M. Leong, and J. S. Coombes, "Effect of High-Intensity Interval Training on Fitness, Fat Mass and Cardiometabolic Biomarkers in Children with Obesity: A Randomised Controlled Trial," *Sports Medicine*, vol. 48, no. 3, pp. 733-746, 2017. <https://doi.org/10.1007/s40279-017-0777-0>.
- [43] J. Zhao, Z. Su, C. Qu, and Y. Dong, "Effects of 12 Weeks Resistance Training on Serum Irisin in Older Male Adults," *Frontiers in Physiology*, vol. 8, no. 171, 2017. <https://doi.org/10.3389/fphys.2017.00171>.
- [44] C-F. Cheng, H-C. Ku, and H. Lin, "PGC-1 $\alpha$  as a Pivotal Factor in Lipid and Metabolic Regulation," *International Journal of Molecular Sciences*, vol. 19, no. 11, pp. 3447, 2018. <https://doi.org/10.3390/ijms19113447>.
- [45] F. Giolo De Carvalho, and L. Sparks, "Targeting White Adipose Tissue with Exercise or Bariatric Surgery as Therapeutic Strategies in Obesity," *Biology*, vol. 8, no. 1, pp. 16, 2019. <https://doi.org/10.3390/biology8010016>.



- [46] B. Otero-Díaz, M. Rodríguez-Flores, V. Sánchez-Muñoz, F. Monraz-Preciado, S. Ordoñez- Ortega, V. Becerril-Elias, G. Baay-Guzmán, R. Obando-Monge, E. García-García, B. Palacios-González, M. T. Villarreal-Molina, M. Sierra-Salazar, and B. Antuna-Puente, "Exercise Induces White Adipose Tissue Browning Across the Weight Spectrum in Humans," *Front. Physiol.*, vol. 9, no. 1781, 2018. <https://doi.org/10.3389/fphys.2018.01781>.
- [47] J. Reisi, K. Ghaedi, H. Rajabi, and S. M. Marandi, "Can Resistance Exercise Alter Irisin Levels and Expression Profiles of FNDC5 and UCP1 in Rats?," *Asian J Sports Med*, vol. 7, no. 4, 2016. <https://doi.org/10.5812/asjms.35205>.
- [48] N. Brandt, M. M. Dethlefsen, J. Bangsbo, and H. Pilegaard, "PGC-1 $\alpha$  and exercise intensity dependent adaptations in mouse skeletal muscle," *PLOS ONE*, vol. 12, no. e0185993, 2017. <https://doi.org/10.1371/journal.pone.0185993>.
- [49] S. Ringholm, J. G. Knudsen, L. Leick, A. Lundgaard, M. Munk Nielsen, and H. Pilegaard, "PGC-1 $\alpha$  Is Required for Exercise and Exercise Training-Induced UCP1 Up-Regulation in Mouse White Adipose Tissue," *PLOS ONE*, vol. 8, no. e64123, 2013. <https://doi.org/10.1371/journal.pone.0064123>.
- [50] N. Chen, Q. Li, J. Liu, and S. Jia, "Irisin, an exercise-induced myokine as a metabolic regulator: an updated narrative review: Irisin as a Metabolic Regulator," *Diabetes/Metabolism Research and Reviews*, vol. 32, no. 1, pp. 51–59, 2016. <https://doi.org/10.1002/dmrr.2660>.
- [51] N. Kim, J. Kim, C. Yoo, K. Lim, T. Akimoto, and J. Park, "Effect of acute mid-intensity treadmill exercise on the androgen hormone level and uncoupling protein-1 expression in brown fat tissue of mouse," *Journal of Exercise Nutrition & Biochemistry*, vol. 22, no. 1, pp. 15–21, 2018. <https://doi.org/10.20463/jenb.2018.0003>.
- [52] J. Y. Huh, V. Mougios, A. Kabasakalis, I. Fatourus, A. Siopi, I. I. Douroudos, A. Filippaios, G. Panagiotou, K. H. Park, and C. S. Mantzoros, "Exercise-induced irisin secretion is independent of age or fitness level and increased irisin may directly modulate muscle metabolism through AMPK activation," *Journal of Clinical Endocrinology and Metabolism*, vol. 99, no. 11, pp. E2154–E2161, 2014. <https://doi.org/10.1210/jc.2014-1437>.
- [53] M. Pang, J. Yang, J. Rao, H. Wang, J. Zhang, S. Wang, X. Chen, and X. Dong, "Time-Dependent Changes in Increased Levels of Plasma Irisin and Muscle PGC-1 $\alpha$  and FNDC5 after Exercise in Mice," *The Tohoku Journal of Experimental Medicine*, vol. 244, no. 2, pp. 93–103, 2018. <https://doi.org/10.1620/tjem.244.93>.
- [54] C. M. Dieli-Conwright, J. L. Kiwata, C. T. Tuzon, T. M. Spektor, F. R. Sattler, J.C. Rice, and E. T. Schroeder, "Acute Response of PGC-1 $\alpha$  and IGF-1 Isoforms to Maximal Eccentric Exercise in Skeletal Muscle of Postmenopausal Women," *Journal of Strength and Conditioning Research*, vol. 30, no. 4, pp. 1161–1170, 2016. <https://doi.org/10.1519/JSC.0000000000001171>.
- [55] E. Badawy, N. A. El-laithy, S. M. Morsy, M. N. Ashour, T. R. Elias, M. M. Masoud, O. Aly, Role of swimming on muscle PGC-1 $\alpha$ , FNDC5 mRNA, and assessment of serum omentin, adiponin, and irisin in high carbohydrate high fat (HCHF) diet induced obesity in rats. *Egyptian Journal of Medical Human Genetics*, vol. 21, Article number 37, 2020. <https://doi.org/10.1186/s43042-020-00080-6>.
- [56] S. Lee, L. Garcia, A. D. Leon, R. Z. Bustos, B. Campos, J. A. Krentzel, L. Mandarino, and D. K. Coletta, "167-OR: Exercise Training Alters Peroxisome Proliferator-Activated Receptor Gamma Coactivator 1 Alpha (PGC-1 Alpha) DNA Methylation in Human Skeletal Muscle," *Diabetes*, vol. 69, no. 1, 2020. <https://doi.org/10.2337/db20-167-OR>.
- [57] F. Norheim, T. M. Langley, M. Hjorth, T. Holen, A. Kiehl, H. K. Stadheim, H. L. Gulseth, K. I. Birkeland, J. Jensen, and C. A. Drevon, "The effects of acute and chronic exercise on PGC-1 $\alpha$ , irisin and browning of subcutaneous adipose tissue in humans," *FEBS Journal*, vol. 281, no. 3, pp. 739–749, 2014. <https://doi.org/10.1111/febs.12619>.
- [58] Y. Fukushima, S. Kurose, H. Shinno, H. Cao Thi Thu, A. Tamanoi, H. Tsutsumi, T. Hasegawa, T. Nakajima, and Y. Kimura, "Relationships between serum irisin levels and metabolic parameters in Japanese patients with obesity: Irisin levels and metabolic parameters," *Obesity Science & Practice*, vol. 2, no. 2, pp. 203–209, 2016. <https://doi.org/10.1002/osp4.43>.
- [59] C. Skovgaard, N. Brandt, H. Pilegaard, and J. Bangsbo, Combined speed endurance and endurance exercise amplify the exercise-induced PGC-1 $\alpha$  and PDK4 mRNA response in trained human muscle. *Physiological Reports*, vol. 4, no. e12864, 2016. <https://doi.org/10.14814/phy2.12864>.
- [60] M. Cao, M. Quan, and J. Zhuang, "Effect of High-Intensity Interval Training versus Moderate-Intensity Continuous Training on Cardiorespiratory Fitness in Children and Adolescents: A Meta-Analysis," *International Journal of Environmental Research and Public Health*, vol. 16, no. 9, pp. 1533, 2019. <https://doi.org/10.3390/ijerph16091533>.
- [61] S. Yang, E. Loro, S. Wada, B. Kim, W.-J. Tseng, K. Li, T.S. Khurana, and Z. Arany, "Functional effects of muscle PGC-1 $\alpha$  in aged animals," *Skeletal Muscle*, vol. 10, Article number 14, 2020. <https://doi.org/10.1186/s13395-020-00231-8>.
- [62] J. F. Halling, and H. Pilegaard, "PGC-1 $\alpha$ -mediated regulation of mitochondrial function and physiological implications," *Applied Physiology, Nutrition, and Metabolism*, vol. 45, no. 9, pp. 927–936, 2020. <https://doi.org/10.1139/apnm-2020-0005>.
- [63] C. W. Taylor, S. A. Ingham, J. E. A. Hunt, N. R. W. Martin, J. S. M. Pringle, R. A. Ferguson, "Exercise duration-matched interval and continuous sprint cycling induce similar increases in AMPK phosphorylation, PGC-1 $\alpha$  and VEGF mRNA expression in trained individuals," *European Journal of Applied Physiology*, vol. 116, no. 8, pp. 1445–1454, 2016. <https://doi.org/10.1007/s00421-016-3402-2>.
- [64] G. L. McKie, and D. C. Wright, "Biochemical adaptations in white adipose tissue following aerobic exercise: from mitochondrial biogenesis to browning," *Biochemical Journal*, vol. 477, no. 6, pp. 1061–1081, 2020. <https://doi.org/10.1042/BCJ20190466>.
- [65] J.-S. Park, J. O. Holloszy, K. Kim, and J.-H. Koh, "Exercise Training-Induced PPAR $\beta$  Increases PGC-1 $\alpha$  Protein Stability and Improves Insulin-Induced Glucose Uptake in Rodent Muscles," *Nutrients*, vol. 12, no. 3, pp. 652, 2020. <https://doi.org/10.3390/nu12030652>.
- [66] C. Scheele, and S. Nielsen, Metabolic regulation and the anti-obesity perspectives of human brown fat. *Redox Biology*, vol. 12, pp. 770–775, 2017. <https://doi.org/10.1016/j.redox.2017.04.011>.

- [67] H. Islam, D. A. Hood, and B. J. Gurd, "Looking beyond PGC-1 $\alpha$ : emerging regulators of exercise-induced skeletal muscle mitochondrial biogenesis and their activation by dietary compounds," *Applied Physiology, Nutrition, and Metabolism*, vol. 45, no. 1, pp. 11–23, 2020. <https://doi.org/10.1139/apnm-2019-0069>.
- [68] J. A. Hawley, M. Hargreaves, M. J. Joyner, and J. R. Zierath, "Integrative Biology of Exercise," *Cell*, vol. 159, no. 4, pp. 738–749, 2014. <https://doi.org/10.1016/j.cell.2014.10.029>.
- [69] X. Xie, T. Gao, M. Yang, P. Chen, H. Jin, L. Yang, and X. Yu, "Associations of betatrophin levels with irisin in Chinese women with normal glucose tolerance," *Diabetology & metabolic syndrome*, vol. 7, Article number 26, 2015. <https://doi.org/10.1186/s13098-015-0019-2>.
- [70] E. Murawska-Cialowicz, P. Wolanski, J. Zuwała-Jagiello, Y. Feito, M. Petr, J. Kokstajn, P. Stastny, and D. Goliński, "Effect of HIIT with Tabata Protocol on Serum Irisin, Physical Performance, and Body Composition in Men," *International Journal of Environmental Research and Public Health*, vol. 17, no. 10, pp. 3589. <https://doi.org/10.3390/ijerph17103589>.
- [71] M. Gerber, C. Imboden, J. Beck, S. Brand, F. Colledge, A. Eckert, E. Holsboer-Trachslar, U. Pühse, and M. Hatzinger, "Effects of Aerobic Exercise on Cortisol Stress Reactivity in Response to the Trier Social Stress Test in Inpatients with Major Depressive Disorders: A Randomized Controlled Trial," *Journal of clinical medicine*, vol. 9, no. 5, pp. 1419, 2020. <https://doi.org/10.3390/jcm9051419>.
- [72] J. Luo, C. Tang, X. Chen, Z. Ren, H. Qu, R. Chen, and Z. Tong, "Impacts of Aerobic Exercise on Depression-Like Behaviors in Chronic Unpredictable Mild Stress Mice and Related Factors in the AMPK/PGC-1 $\alpha$  Pathway," *International Journal of Environmental Research and Public Health*, vol. 17, no. 6, pp. 2042, 2020. <https://doi.org/10.3390/ijerph17062042>.
- [73] M. Khammassi, N. Ouerghi, M. Said, M. Feki, Y. Khammassi, B. Pereira, D. Thivel, and A. Bouassida, "Continuous Moderate-Intensity but Not High-Intensity Interval Training Improves Immune Function Biomarkers in Healthy Young Men," *Journal of Strength and Conditioning Research*, vol. 34, no. 1, pp. 249–256, 2020. <https://doi.org/10.1519/JSC.0000000000002737>.
- [74] M. A. Stults-Kolehmainen, and R. Sinha, "The Effects of Stress on Physical Activity and Exercise," *Sports Med*. Vol. 44, no. 1, pp. 81–121, 2014. <https://doi.org/10.1007/s40279-013-0090-5>.
- [75] J. Williamson, and G. Davison, "Targeted Antioxidants in Exercise-Induced Mitochondrial Oxidative Stress: Emphasis on DNA Damage," *Antioxidants*, vol. 9, no. 11, pp. 1142, 2020. <https://doi.org/10.3390/antiox9111142>.
- [76] F. de Souza-Teixeira, J. Alonso-Molero, C. Ayán, L. Vilorio-Marques, A. J. Molina, C. González-Donquiles, V. Dávila-Batista, T. Fernández-Villa, J. A. de Paz, and V. Martín, PGC-1 $\alpha$  as a Biomarker of Physical Activity-Protective Effect on Colorectal Cancer. *Cancer prevention research (Philadelphia, Pa.)*, vol. 11, no. 9, pp. 523–534, 2018. <https://doi.org/10.1158/1940-6207.CAPR-17-0329>.
- [77] J. L. Ruas, J. P. White, R. R. Rao, S. Kleiner, K. T. Brannan, B. C. Harrison, N. P. Greene, J. Wu, J. L. Estall, B. A. Irving, I. R. Lanza, K. A. Rasbach, M. Okutsu, K. S. Nair, Z. Yan, L. A. Leinwand, and B. M. Spiegelman, "A PGC-1 $\alpha$  Isoform Induced by Resistance Training Regulates Skeletal Muscle Hypertrophy," *Cell*, vol. 151, no. 6, pp. 1319–1331, 2012. <https://doi.org/10.1016/j.cell.2012.10.050>.
- [78] K. Mannerkorpi, K. Landin-Wilhelmsen, A. Larsson, Å. Cider, O. Arodell, and J. L. Bjersing, "Acute effects of exercise on the serum insulin-like growth factor system in women with fibromyalgia," *BMC Musculoskeletal Disord*, vol. 18, no. 1, pp. 37, 2017. <https://doi.org/10.1186/s12891-017-1402-y>.
- [79] M. Wiecek, J. Szymura, M. Maciejczyk, M. Kantorowicz, and Z. Szygula, "Acute Anaerobic Exercise Affects the Secretion of Asprosin, Irisin, and Other Cytokines – A Comparison Between Sexes," *Frontiers in Physiology*, vol. 9, no. 1782, 2018. <https://doi.org/10.3389/fphys.2018.01782>.
- [80] E. S. van der Valk, M., Savas, and E. F. C. van Rossum, "Stress and Obesity: Are There More Susceptible Individuals?," *Current obesity reports*, vol. 7, no. 2, pp. 193–203, 2018. <https://doi.org/10.1007/s13679-018-0306-y>.
- [81] D. C. Nieman, and L. M. Wentz, "The compelling link between physical activity and the body's defense system," *Journal of Sport and Health Science*, vol. 8, no. 3, pp. 201–217, 2019. <https://doi.org/10.1016/j.jshs.2018.09.009>.
- [82] Sugiharto, "Physiological Effects of Music during Exercise Secretion of Hormones Cortisol and Endorphins," *Folia Medica Indonesia*, vol. 45, no. 72, pp.121–129, 2009.
- [83] C. Simioni, G. Zauli, A. M. Martelli, M. Vitale, G. Sacchetti, A. Gonelli, and L. M. Neri, Oxidative stress: role of exercise and antioxidant nutraceuticals in adulthood and aging," *Oncotarget*, vol. 9, no. 24, pp. 17181–17198, 2018. <https://doi.org/10.18632/oncotarget.24729>.
- [84] P. Vidal, and K. I. Stanford, "Exercise-Induced Adaptations to Adipose Tissue Thermogenesis. *Frontiers in Endocrinology*, vol. 11, no. 270, 2020. <https://doi.org/10.3389/fendo.2020.00270>.

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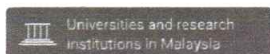
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