

The
Japan
Institute
of
Heterocyclic
Chemistry
Publication

HETEROCYCLES

An International Journal for Reviews and Communications in Heterocyclic Chemistry
Web Edition ISSN: 1881-0942

Published online by The Japan Institute of Heterocyclic Chemistry

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Published by	The Japan Institute of Heterocyclic Chemistry 503 1-1-7 Motoakasaka, Minato-ku, Tokyo 107-0051, Japan Tel: +81 3 3404 5019 Fax: +81 3 3497 9370 E-mail: editorial@heterocycles.com

Source details

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Scopus coverage years: from 1983 to Present

Publisher: Elsevier

ISSN: 0385-5414

Subject area: Pharmacology, Toxicology and Pharmaceuticals: Pharmacology Chemistry: Analytical Chemistry Chemistry: Organic Chemistry

Source type: Journal

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CiteScore 2019
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COUNTRY

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SUBJECT AREA AND CATEGORY

Chemistry
 └ Analytical Chemistry
 └ Organic Chemistry

Pharmacology, Toxicology and Pharmaceuticals
 └ Pharmacology

PUBLISHER

Japan Institute of Heterocyclic Chemistry

H-INDEX

63

PUBLICATION TYPE

Journals

ISSN

03855414

COVERAGE

1983-2020

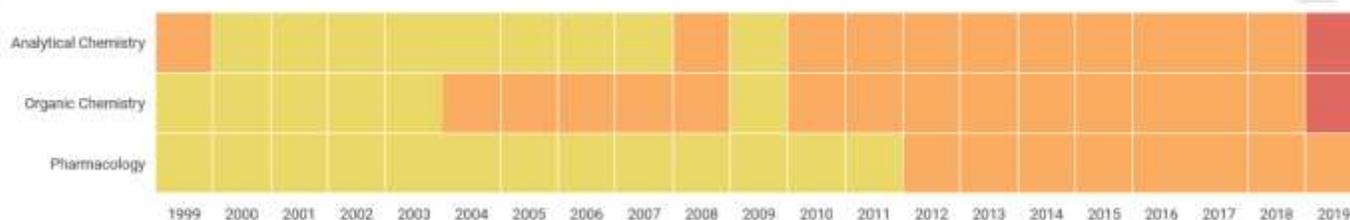
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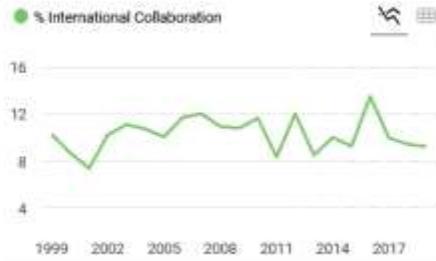
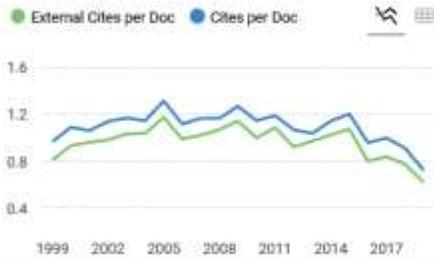
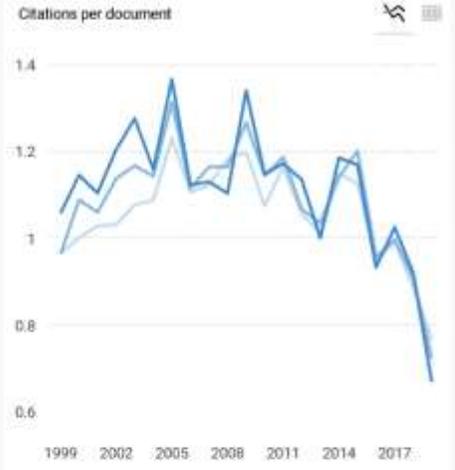
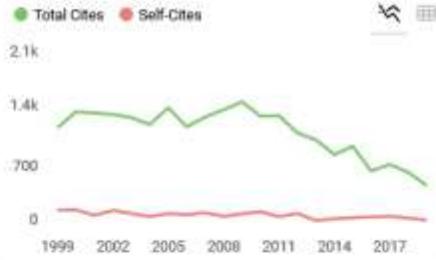
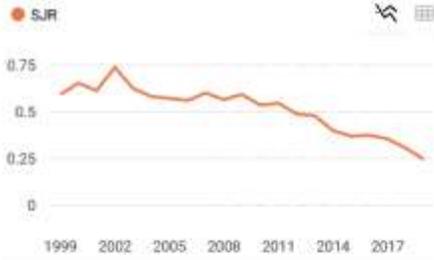
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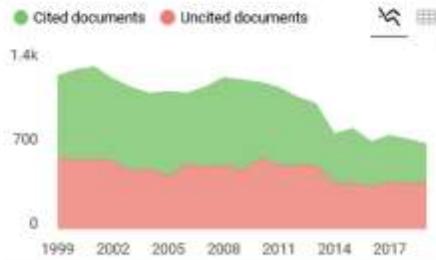
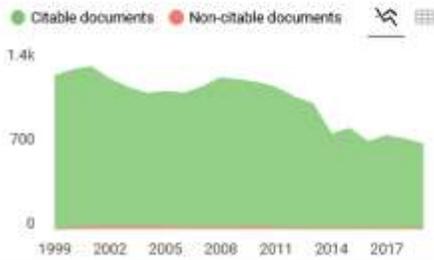
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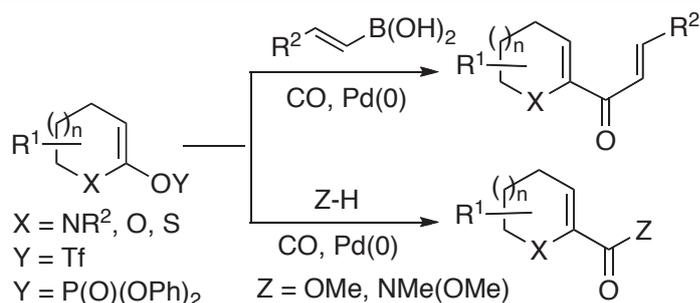
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■ REVIEWS

- 697 **Carbonylative Palladium-Catalyzed Reactions of Lactam-, Lactone-, and Thiolactone-Derived Vinyl Triflates and Phosphates for the Synthesis of *N*-, *O*-, and *S*-Heterocycles**

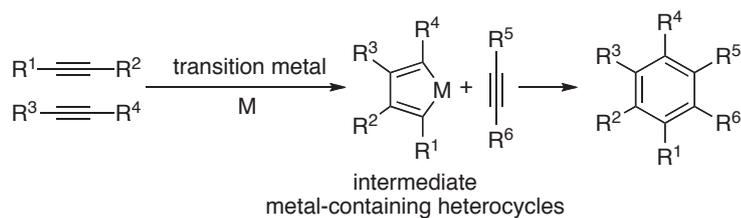
Ernesto G. Occhiato,* Dina Scarpi, and Cristina Prandi*



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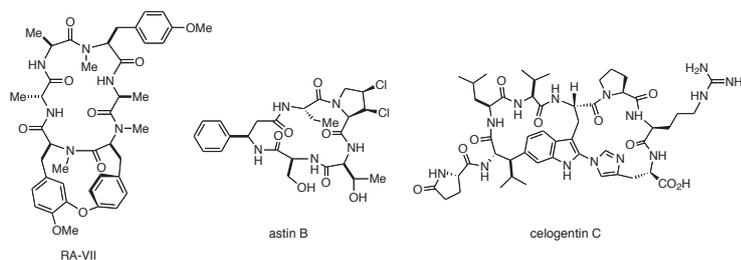
Lishan Zhou, Shi Li, Ken-ichiro Kanno, and Tamotsu Takahashi*



Aromatic Compound Intermolecular Coupling Reaction Alkyne Metallacyclopentadiene Selectivity

- 739 **Bioactive Cyclic Peptides from Higher Plants**

Hiroshi Morita* and Koichi Takeya

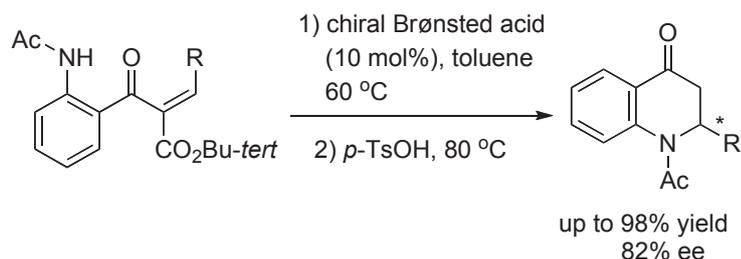


Cyclic Peptide Cyclopeptide Higher Plant

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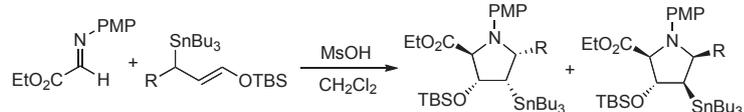
Zhen Feng, Qing-Long Xu, Li-Xin Dai, and Shu-Li You*



Asymmetric Catalysis *N*-Triflylphosphoramidate Enantioselectivity Michael Addition Organocatalysis

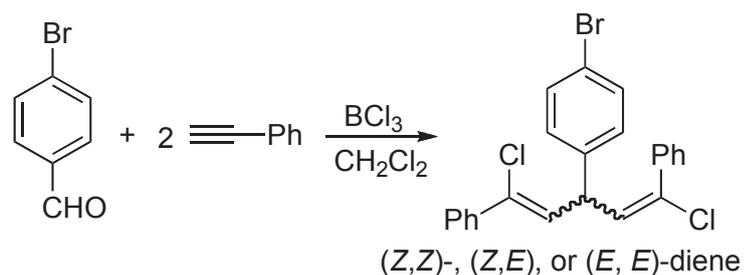
773 Facile Synthesis and Ring-Opening of 4-(Tributylstannyl)-pyrrolidine-2-carboxylates

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 Pyrrolidine-2-carboxylate Cyclization Allylstannane α -Iminoacetate Methanesulfonic Acid

779 Boron Trichloride Mediated Alkyne-Aldehyde Coupling Reactions

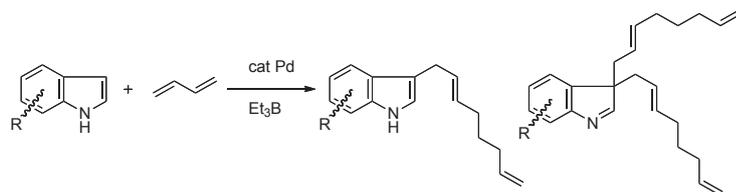
Min-Liang Yao, Michael P. Quinn, and George W. Kabalka*



Boron Halide Diene Aldehyde Reaction Synthesis

787 Allylic Alkylation of Indoles with Butadiene Promoted by Palladium Catalyst and Triethylborane

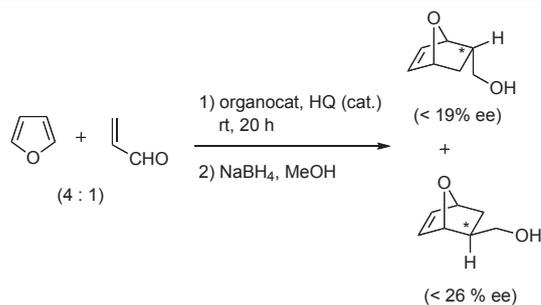
Masanari Kimura,* Katsumi Tohyama, Yumi Yamaguchi, and Tomohiko Kohno



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799 Organocatalytic Asymmetric Diels-Alder Reaction of Furan under High Pressure

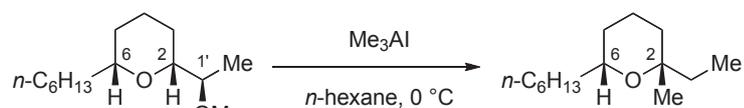
Akiko Mimoto, Keiji Nakano, Yoshiyasu Ichikawa, and Hiyoshizo Kotsuki*



Organocatalyst Asymmetric Diels-Alder Reaction Furan Acrolein High Pressure Reaction

805 Methyl Insertion Reactions of Tetrahydropyrans Having a C1'-Mesyloxy Group on the C2-Side Chain with Trimethylaluminum

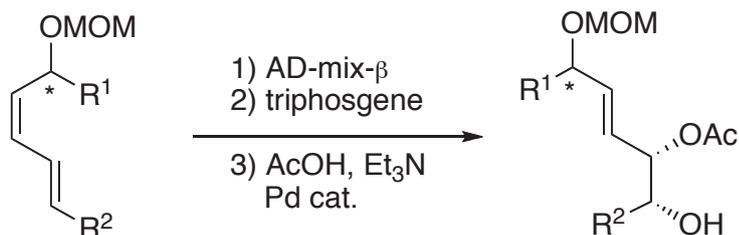
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Trimethylaluminum Tetrahydropyran Methyl Insertion Antiperiplanar 1,2-Hydride Shift

811 Synthesis of the 1,2-Anti Type of 3*E*-Alkene-1,2,5-triol Derivatives

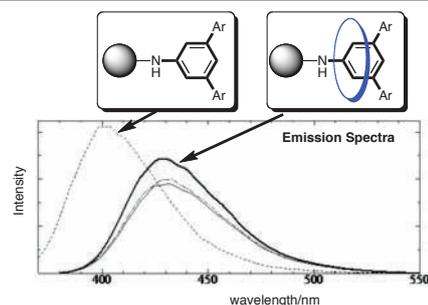
Yuichi Kobayashi,* Akira Takeuchi, and Hatsuhiko Hattori



Borate Boronate Ester Nickel Palladium Catalyzed Reaction Trioxilin A3

819 Preparation and Photochemical Properties of [2]Rotaxanes Containing an Aniline Moiety Encapsulated by Crown Ethers

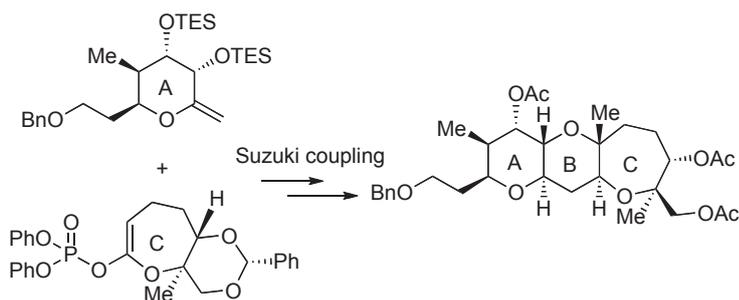
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Rotaxane Aniline Crown Ether UV-VIS and Emission Spectra

825 Synthesis of the ABC Ring Fragment of Brevisin, a New Dinoflagellate Polycyclic Ether

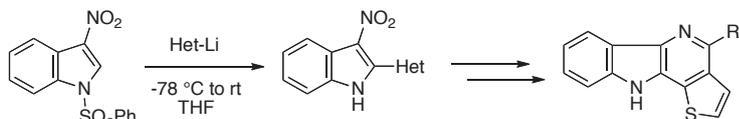
Naohito Ohtani, Ryosuke Tsutsumi, Takefumi Kuranaga, Tomohiro Shirai, Jeffrey L. C. Wright, Daniel G. Baden, Masayuki Satake,* and Kazuo Tachibana*



Suzuki-Miyaura Cross Coupling Reaction Ketene Acetal Phosphate Polycyclic Ether

831 Nucleophilic Addition of Hetarylithium Compounds to 3-Nitro-1-(phenylsulfonyl)indole: Synthesis of Tetracyclic Thieno[3,2-*c*]-5-carbolines

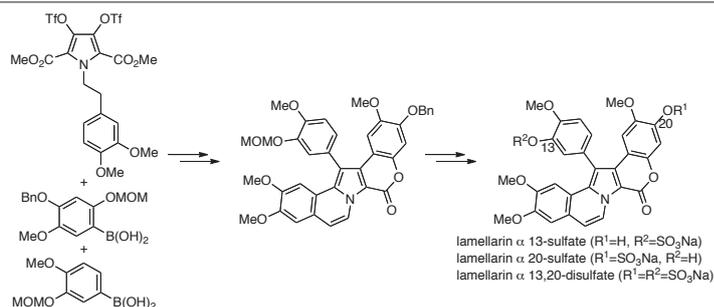
Philip E. Alford, Tara L. S. Kishbaugh, and Gordon W. Gribble*



3-Nitroindole Michael Addition Arylation Electron-Deficient Indole

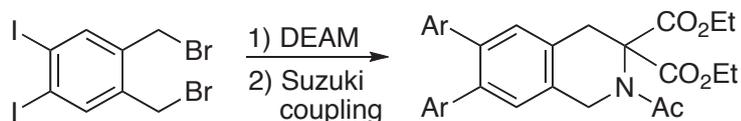
841 Divergent Synthesis of Lamellarin α 13-Sulfate, 20-Sulfate, and 13,20-Disulfate

Tsutomu Fukuda, Takeshi Ohta, Sho Saeki, and Masatomo Iwao*


 Lamellarin α 13-Sulfate Lamellarin α 20-Sulfate Lamellarin α 13,20-Disulfate

847 Diversity-Oriented Approach to 1,2,3,4-Tetrahydroisoquinoline-3-carboxylic Acid (Tic) Derivatives Using Diethyl Acetamidomalonate as a Glycine Equivalent: Further Expansion by Suzuki–Miyaura Cross-Coupling Reaction

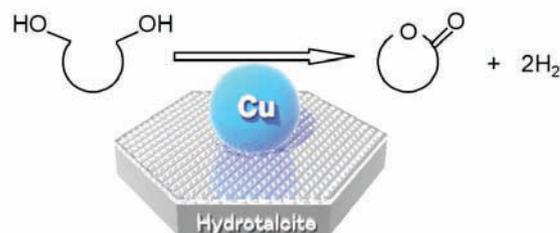
Sambasivarao Kotha,* Shilpi Misra, Nimita Gopal Krishna, Nagaraju Devunuri, Henning Hopf, and Abhilash Keecherikunnel



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855 Oxidant-Free Lactonization of Diols Using a Hydrotalcite-Supported Copper Catalyst

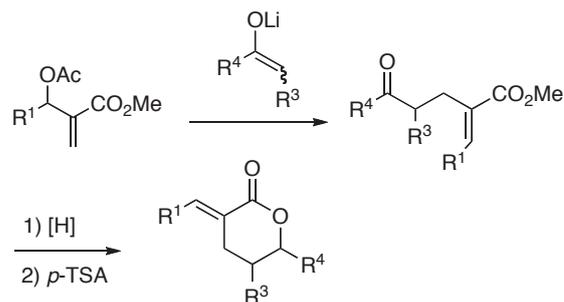
Yusuke Mikami, Kaori Ebata, Takato Mitsudome, Tomoo Mizugaki, Koichiro Jitsukawa, and Kiyotomi Kaneda*



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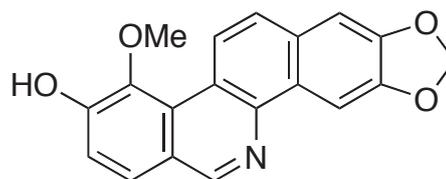
P. Veeraraghavan Ramachandran* and Annyt Bhattacharyya



Alkylidenevalerolactone Conjugate Addition Cyclization Keto Ester

873 Synthesis of Zanthoxyline and Its Related Compounds: Revision of the Reported Structure

Hitoshi Abe,* Naoko Kobayashi, Yasuo Takeuchi, and Takashi Harayama*

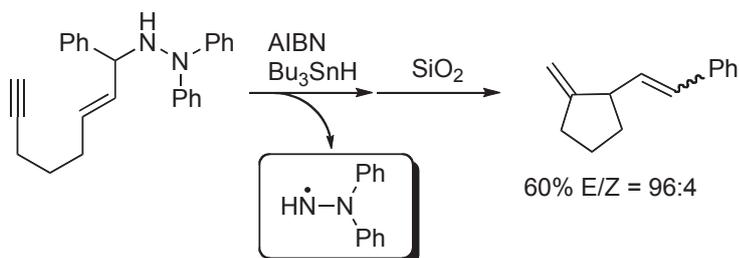


zanthoxyline (reported structure)

Benzo[*d*]phenanthridine Palladium Catalyzed Reaction Coupling Reaction

879 Novel Radical Cyclization Method Accompanied by Elimination of Hydrazyl Radical

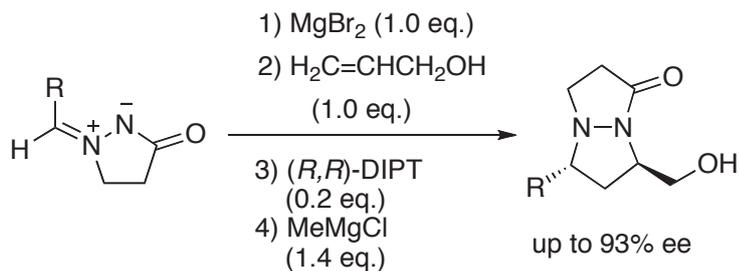
Shoji Kobayashi, Hidefumi Hirao, Tatsuro Kawauchi, and Ilhyong Ryu*



Hydrazine Hydrazyl Radical Vinyl Radical 5-Exo Cyclization 1,4-Diene

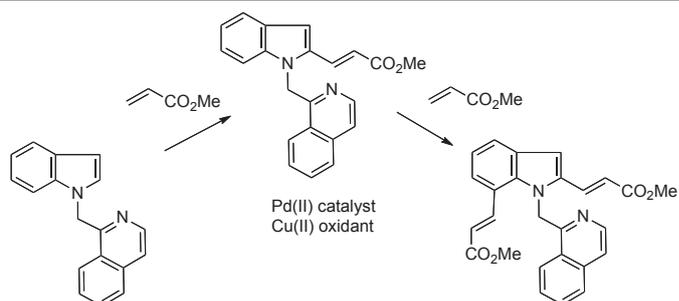
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Katsuyoshi Tanaka, Tomomitsu Kato, Yutaka Ukaji,* and Katsuhiko Inomata*


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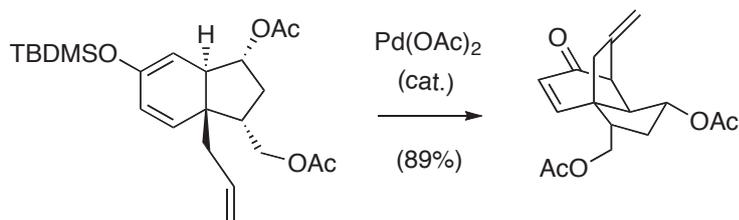
Guilia Fanton, Nicola M. Coles, Andrew R. Cowley, Jonathan P. Flemming, and John M. Brown*



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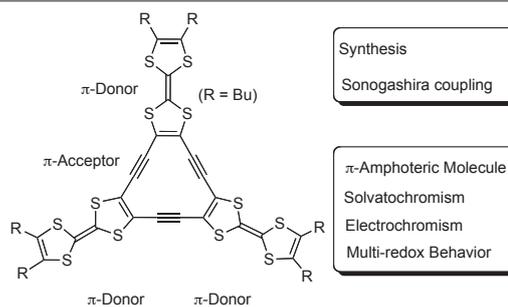
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Akihiro Ishihata, Megumi Saeki, Masaru Watanabe, Masataka Ihara, and Masahiro Toyota*


 Quadrone *Aspergillus terreus* Cycloalkenylation Palladium Acetate Intramolecular Michael Addition

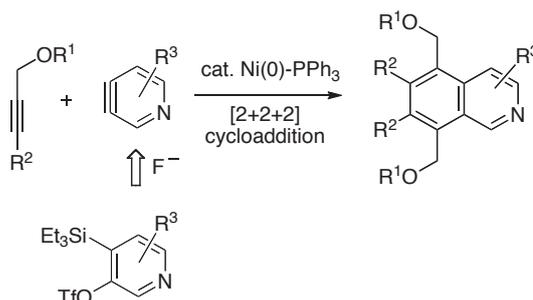
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Kenji Hara, Masashi Hasegawa, Yoshiyuki Kuwatani, Hideo Enozawa, and Masahiko Iyoda*


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917 Synthesis of Substituted Isoquinolines via Nickel-Catalyzed [2+2+2] Cycloaddition of Alkynes and 3,4-Pyridynes

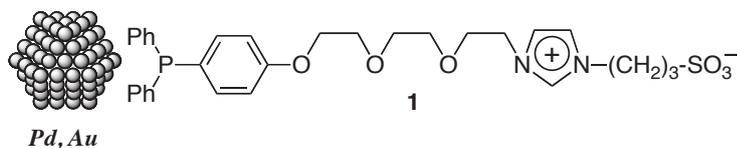
Toshihiko Iwayama and Yoshihiro Sato*



[2+2+2] Cycloaddition Nickel 3,4-Pyridyne 1,3-Diyne Isoquinoline

925 **Water-Soluble Palladium and Gold Nanoparticles Functionalized by a New Phosphine with Zwitterionic Liquid Based on Imidazolium Sulfonate Linked Ethylene Glycol Moiety**

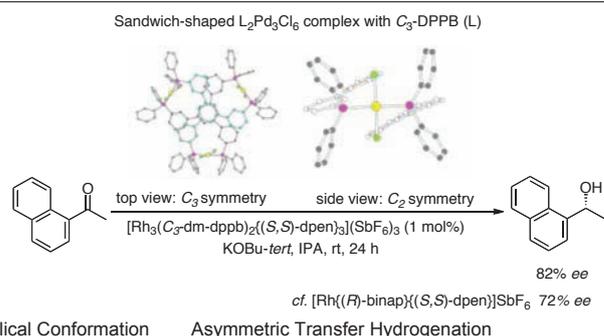
Taichi Akiyama, Chiharu Iбата, and Hisashi Fujihara*



Imidazolium Ion Palladium Nanoparticle Ionic Liquid Suzuki Coupling Reaction

933 **Helical Chirality Control of *Tropos* Sandwich-Shaped L_2M_3 Complexes with C_3 -Symmetric Tris(diphenylphosphinophenyl)benzene Ligand**

Kazuki Wakabayashi and Koichi Mikami*

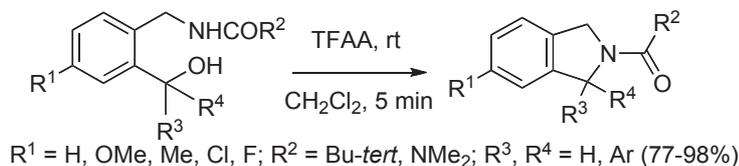


Tropos L_2M_3 Complex C_3 -Triphosphine Ligand Chirality Control Helical Conformation Asymmetric Transfer Hydrogenation

■ PAPERS

941 **A Simple and Convenient High Yielding Synthesis of Substituted Isoindolines**

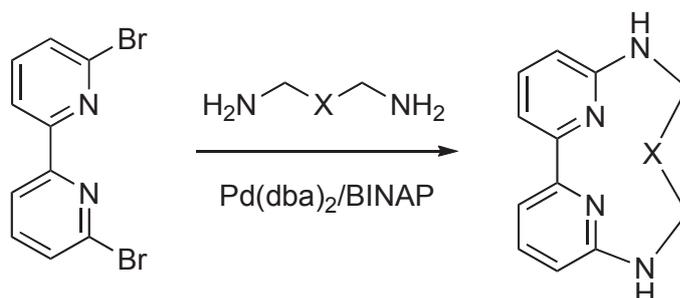
Keith Smith,* Gamal A. El-Hiti, Amany S. Hegazy, and Ahmed Fekri



Cyclization Dehydration Heterocycle Substituted Benzyl-*N,N*-dimethylurea Synthesis

957 **Synthesis of Polyazamacrocycles Comprising 6,6'-Diamino-2,2'-bipyridine Moieties *via* Pd-Catalyzed Amination**

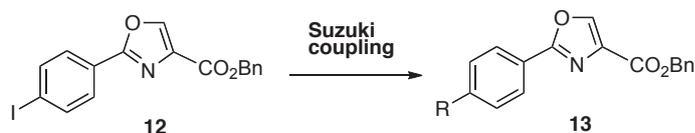
Alexei D. Averin,* Alexei N. Uglov, Alexei K. Buryak, Alla G. Bessmertnykh, Roger Guillard, and Irina P. Beletskaya*



2,2'-Bipyridine Amination Polyamine Pd Catalysis Macrocycle

977 Utilization of the Suzuki Coupling to Enhance the Antituberculosis Activity of Aryloxazoles

Garrett C. Moraski, Scott G. Franzblau, and Marvin J. Miller*

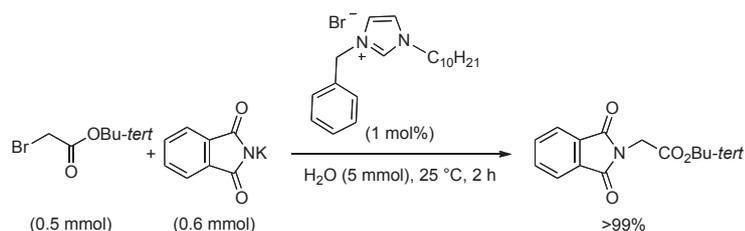


R = 2-chlorophenyl (**14**), 3-chlorophenyl (**15**), 4-chlorophenyl (**16**), 2-methoxyphenyl (**17**), 3-methoxyphenyl (**18**), 4-methoxyphenyl (**19**), 4-trifluoromethoxyphenyl (**20**), 4-cyanophenyl (**21**), 3-benzyloxyphenyl (**22**)

Antituberculosis Agent Suzuki Coupling Reaction Aryloxazole Palladium Catalyzed Cross Coupling Reaction

989 Design of Reaction Media for Nucleophilic Substitution Reactions by Using a Catalytic Amount of an Amphiphilic Imidazolium Salt in Water

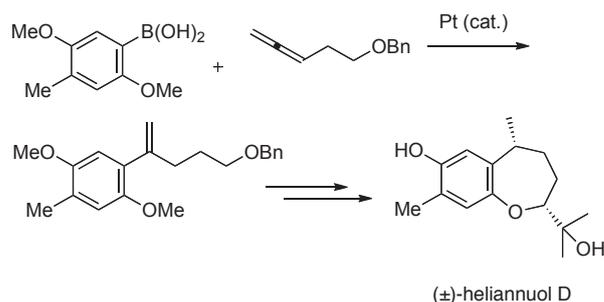
Keisuke Asano and Seiji Matsubara*



Imidazolium Salt Amphiphilicity Water Self-Assembly Hydrophobic Effect

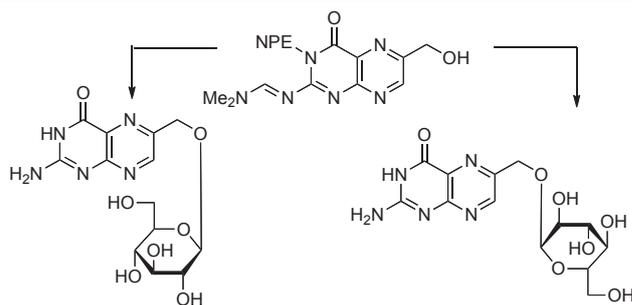
1003 Synthesis of (±)-Heliannuol D Based on Platinum Catalyzed Regioselective Addition of Arylboronic Acids to Allenes

Mayu Osaka, Makoto Kanematsu, Masahiro Yoshida, and Kozo Shishido*

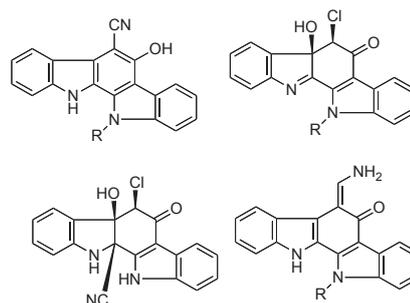

 (±)-Heliannuol D (±)-10-*epi*-Heliannuol D Total Synthesis Platinum Catalyzed Addition Allene

1013 Synthesis of 6-Hydroxymethylpterin α - and β -D-Glucosides

Tadashi Hanaya,* Hiroki Baba, Kazumasa Ejiri, and Hiroshi Yamamoto

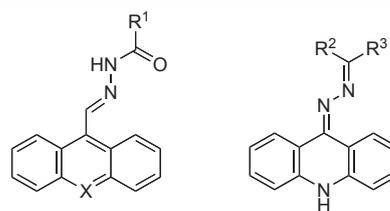

1027 Synthetic Study Directed toward Derivatives of Biologically Active Indolo[2,3-*a*]carbazole

Masako Sato, Yoshiaki Suzuki, Fumio Yamada, and Masanori Somei*



1047 Novel Carbohydrazone and Hydrazone Biomarkers Based on 9-Substituted Acridine and Anthracene Fluorogens

Zdenka Bedlovičová, Ján Imrich,* Pavol Kristian, Ivan Danihel, Stanislav Böhm, Danica Sabolová, Mária Kožurková, Helena Paulíková, and Karel D. Klika

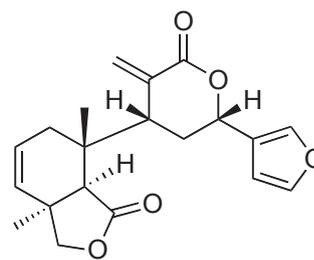


$X = N, CH$
 $R^1 = Me, C_6H_2, 3-Py, 4-Py, 9\text{-oxoacridin-4-yl}$
 $R^2 = H, Me$
 $R^3 = Me, Ph, 4-BrC_6H_4, 4-MeOC_6H_4, 2,4,6\text{-triMeC}_6H_2, 2,4,6\text{-triMeOC}_6H_2$

Acridine Carbohydrazone Hydrazone Fluorescence DFT

1067 Synthetic Study on Clutiolide Based on a Remote Chelation Controlled Ireland-Claisen Rearrangement

Jun Ishihara,* Okihisa Tokuda, Kazunori Shiraishi, Yukihiro Nishino, Keisuke Takahashi, and Susumi Hatakeyama*

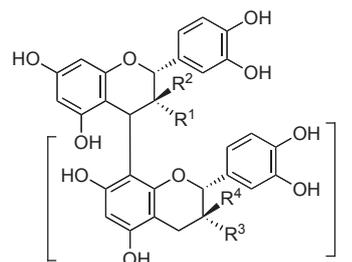


clutiolide

Clutiolide Diterpene Ireland-Claisen Rearrangement Chelation Control Diels-Alder Reaction

1081 Structure-Activity Relationships of Synthesized Procyanidin Oligomers: DPPH Radical Scavenging Activity and Maillard Reaction Inhibitory Activity

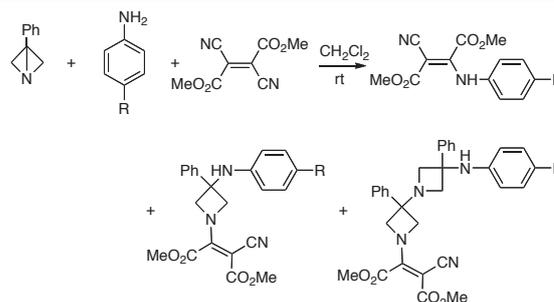
Akiko Saito and Noriyuki Nakajima*


 $R^1 = R^2 = R^3 = R^4: H, OH \text{ or } O\text{-Galloyl}, n = 0 - 4$

Condensed Tannin Antioxidant Tea Catechin Polyphenol Artificial Procyanidin Oligomer

1091 Three-Component Reactions with 3-Phenyl-1-azabicyclo-[1.1.0]butane, Dimethyl Dicyanofumarate, and Primary Aromatic Amines

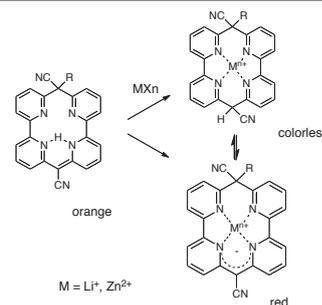
Grzegorz Mlostoń* and Heinz Heimgartner*



1-Azabicyclo[1.1.0]butane 2,3-Dicyanofumarate Zwitterionic Intermediate Three-Component Reaction Addition Reaction

1103 Unusual Reactions of the Highly Strained Macrocycles with Lithium Salts: Anion Control for the Reaction Rates and Elucidation of the Properties of Their Lithium Complexes

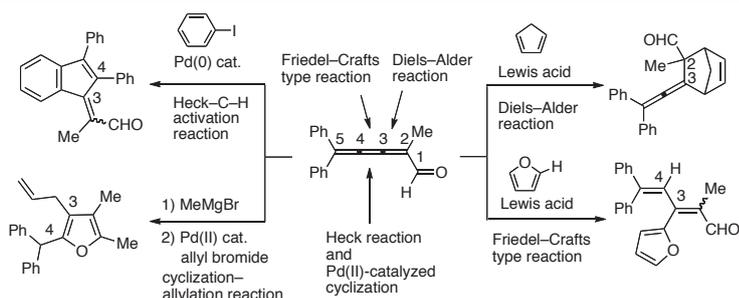
Junko Morita, Shinji Tsuchiya, Nao Yoshida, Nirei Nakayama, and Shojiro Ogawa*


 $M = Li^+, Zn^{2+}$

Tetraazamacrocycle Lithium Complex Unsymmetrical Structure Strained Molecule Anion Control

1125 Reaction Behavior of Cumulene: Diels–Alder, Friedel–Crafts, and Pd-Catalyzed Domino Reactions

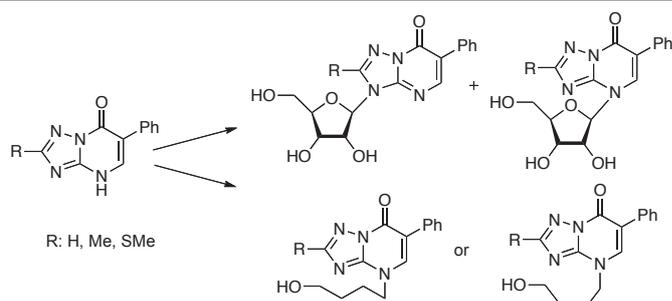
Tomohiro Asakawa, Mie Inuma, Yuko Wakasugi, Mayumi Kuno, Takumi Furuta,* Satoshi Fujii, Kiyoshi Tanaka, and Toshiyuki Kan*



Cumulene Diels–Alder Reaction Friedel–Crafts Reaction Heck Reaction Domino Reaction

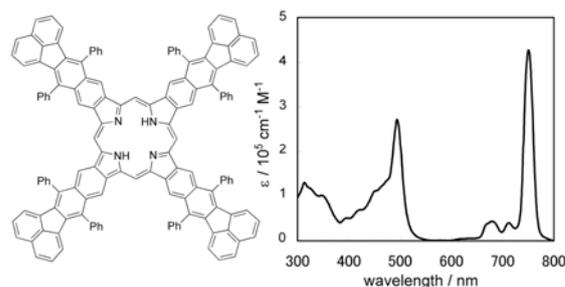
1149 Non-Natural Nucleosides Based on 1,2,4-Triazolo-[1,5-*a*]pyrimidin-7-ones

Oleg N. Chupakhin,* Tatiana S. Shestakova, Sergey L. Deev, Oleg S. Eltsov, and Vladimir L. Rusinov


 Non-Natural Nucleoside Nucleoside Analog 1,2,4-Triazolo[1,5-*a*]pyrimidin-7-one Glycosylation Alkyl Fragment

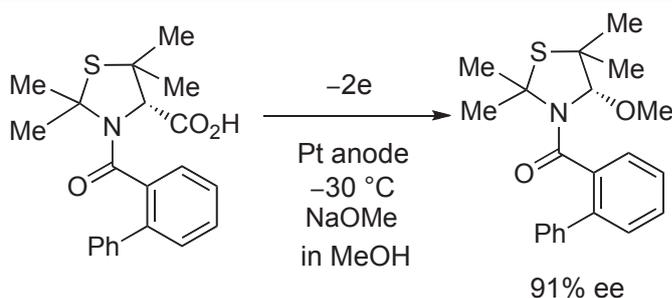
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Jun Nakamura, Tetsuo Okujima,* Yuya Tomimori, Naoki Komobuchi, Hiroko Yamada, Hidemitsu Uno, and Noboru Ono


 [2,3]Fluoranthobenzoporphyrin Retro Diels–Alder Reaction π -Expanded Porphyrin Strong Absorption in Near-IR Region

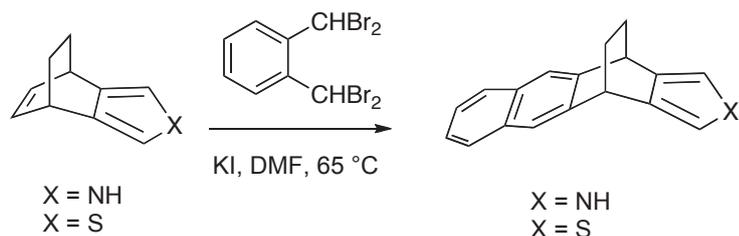
1177 Memory of Chirality in the Electrochemical Oxidation of Thiazolidine-4-carboxylic Acid Derivatives

George Ng'anNg'a Wanyoike, Yoshihiro Matsumura, Masami Kuriyama, and Osamu Onomura*


 Electrochemical Oxidation Memory of Chirality Carbon–Carbon Bond Cleavage Chiral *N,O*-Acetal Thiazolidine-4-carboxylic Acid

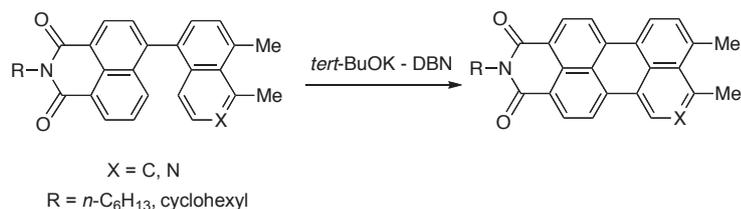
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Hiroki Uoyama, Cai Chenxin, Hiroyuki Tahara, Yusuke Shimizu, Hideki Hagiwara, Yasuaki Hanasaki, Hiroko Yamada, Tetsuo Okujima, and Hidemitsu Uno*


 Anthra[2,3-*c*]pyrrole Anthra[2,3-*c*]thiophene Diels–Alder Reaction X-Ray Structure σ -Quinodimethane

1197 Synthesis and Properties of Dicarboximide Derivatives of Perylene and Azaperylene

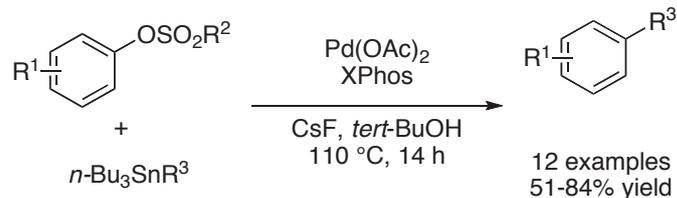
Yukinori Nagao,* Tatsuro Yoshida, Koji Arimitsu, and Kozo Kozawa



Coupling Reaction Ring Closure Reaction Absorption Spectrum Fluorescence Spectrum

1215 Stille Cross-Coupling Reactions of Aryl Mesylates and Tosylates Using a Biarylphosphine Based Catalyst System

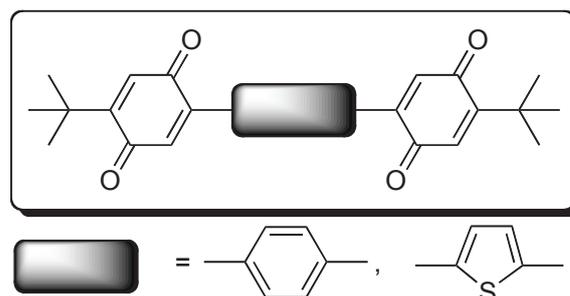
John R. Naber, Brett P. Fors, Xiaoxing Wu, Jonathon T. Gunn, and Stephen L. Buchwald*



Aryl Mesylate Palladium Catalysis C-C Bond Formation Stille Reaction Aryl Tosylate

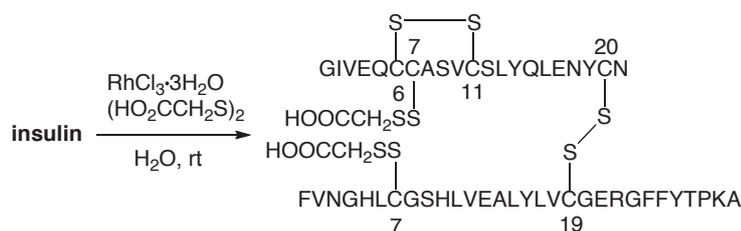
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Naoto Hayashi,* Teru Sakakibara, Takahiro Ohnuma, Junro Yoshino, and Hiroyuki Higuchi


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1239 RhCl₃-Catalyzed Disulfide Exchange Reaction of Insulin and Dithiodiglycolic Acid

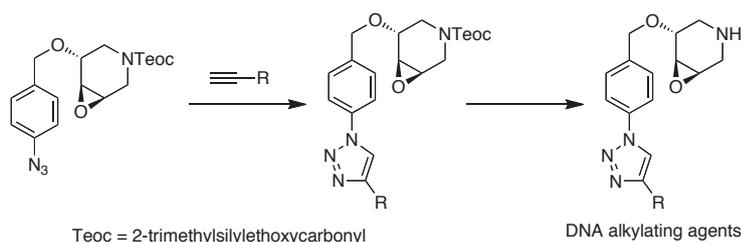
Mieko Arisawa, Manabu Kuwajima, Atsushi Suwa, and Masahiko Yamaguchi*



Rhodium Chloride Disulfide Exchange Reaction Catalysis A7/B7 Disulfide Insulin

1249 Synthesis and Evaluation of Novel 3,4-Epoxy piperidines as Efficient DNA Alkylating Agents

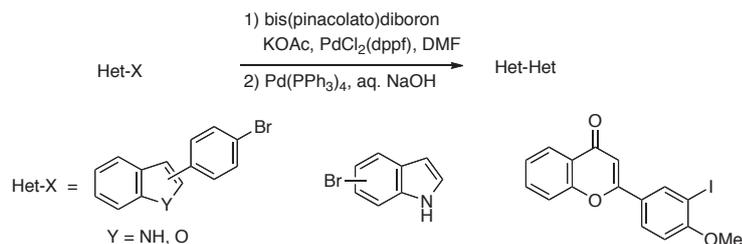
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DNA Alkylating Agent Epoxypiperidine Huisgen Reaction Anticancer Azinomycin

1267 Synthesis of Some New Biheterocycles by a One-Pot Suzuki-Miyaura Coupling Reaction

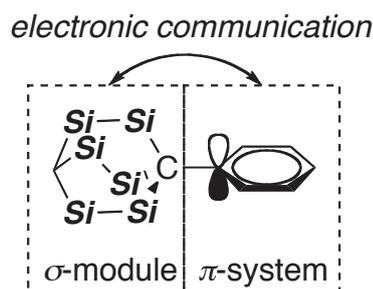
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1275 Synthesis and Photophysical Properties of 2,3,5,6,7,8-Hexasilabicyclo[2.2.2]octan-1-yl-substituted Arenes

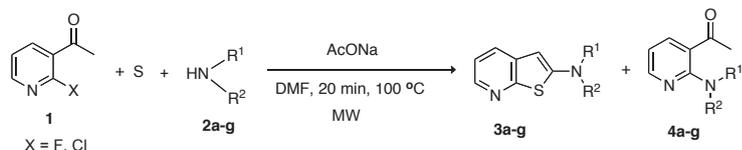
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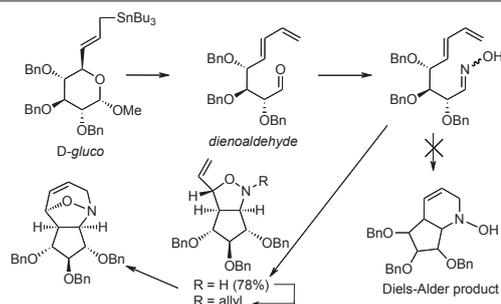
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Haribabu Ankati and Edward R. Biehl*


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1303 Application of Sugar Allyltin Derivatives for the Preparation of Heterocyclic Compounds

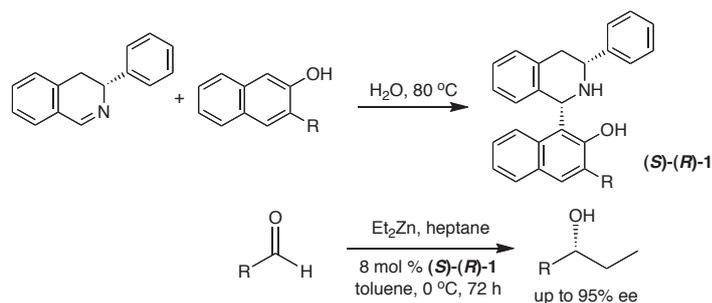
Marta Magdycz, Piotr Cmoch, and Sławomir Jarosz*



Oxime-Olefin Cyclization Nitronne Sugar Allyltin Ring Closing Metathesis

1319 Synthesis of Chiral 1,3-Disubstituted Tetrahydroisoquinolines and Their Use in the Asymmetric Addition of Diethylzinc to Aldehydes

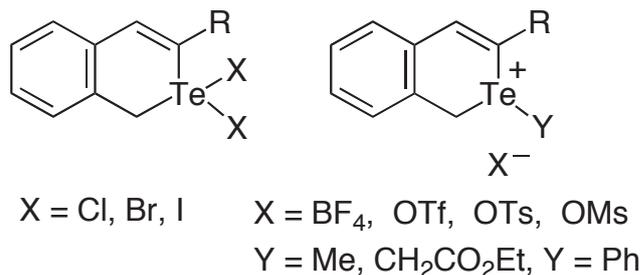
Patricia D. MacLeod, Amy M. Reckling, and Chao-Jun Li*



Chiral Tetrahydroisoquinoline Asymmetric Addition Diethylzinc Chiral Ligand Aza Friedel-Crafts Reaction

1339 2-Substituted Isotellurochromenium Salt Derivatives: Preparations, Structures, Spectroscopic Properties

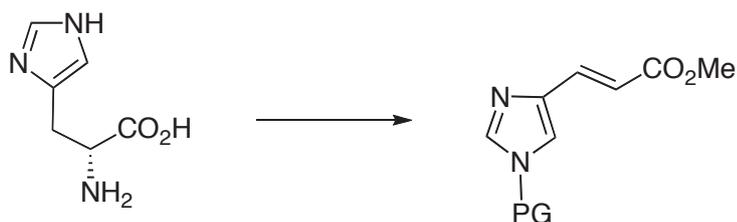
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Isotellurochromene Telluride Tellurium Salt Tellurane

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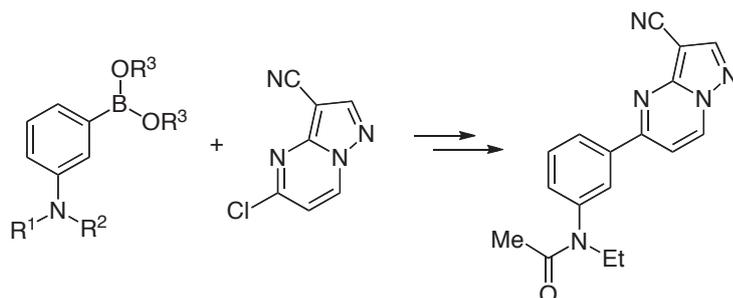
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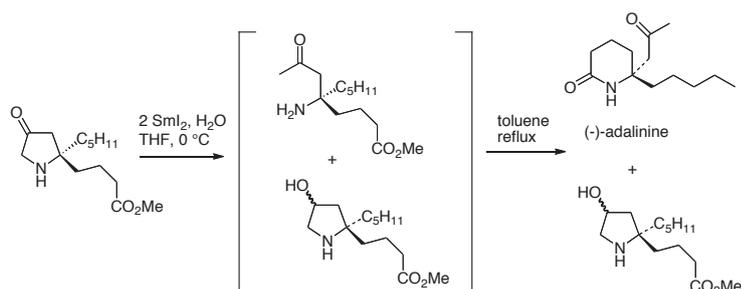
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Stanislav Rádľ,* Michaela Blahovcová, Marcela Tkadlecová, and Jaroslav Havlíček


 Zaleplon Regioisomer Synthesis Spectral Property 5-Arylpyrazolo[1,5-*a*]pyrimidine-3-carbonitrile Suzuki-Miyaura Cross Coupling Reaction

1381 Application of Samarium Diodide-Promoted Reductive Carbon-Nitrogen Bond Cleavage Reaction to 3-Oxopyrrolidine Derivatives: Alternative Synthesis of a Coccinellid Alkaloid, (-)-Adalinine

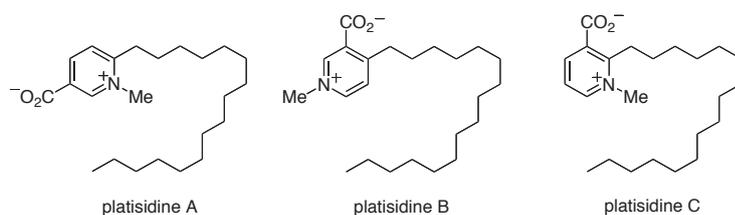
Toshio Honda* and Chihiro Hisa



Samarium Diodide Adalinine Carbon-Nitrogen Bond Cleavage Reaction 4-Hydroxyproline Chiral Synthesis

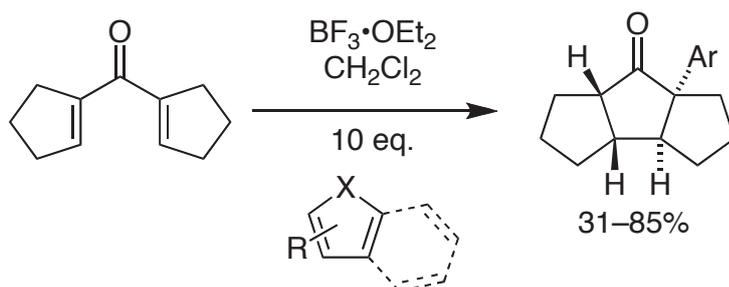
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Takaaki Kubota, Yuichiro Ishiguro, Sunao Yamamoto, Jane Fromont, and Jun'ichi Kobayashi*


 Sponge *Plakortis* species Pyridinium Alkaloid Platisidines A-C Acetylcholinesterase Inhibitor

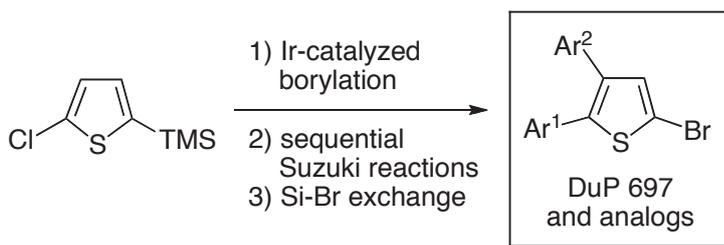
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Nazarov Reaction Domino Process Heteroaromatic Electrophilic Aromatic Substitution Triquinane

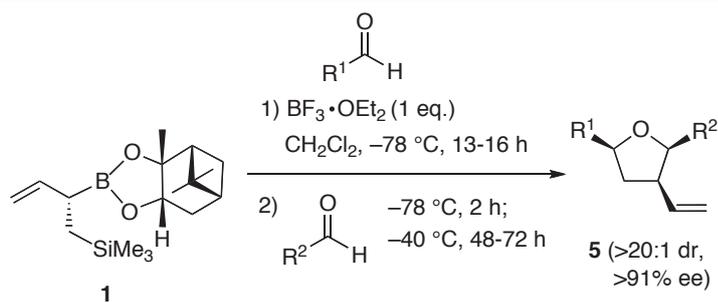
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Thiophene C-H Activation Boronic Ester Suzuki Coupling Reaction COX-2

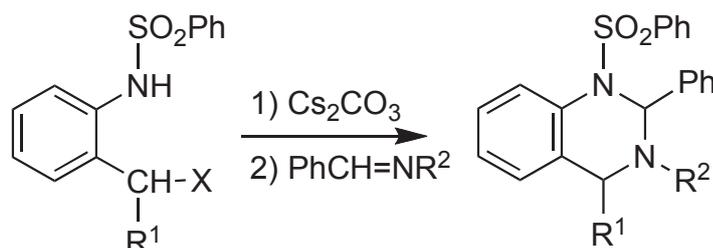
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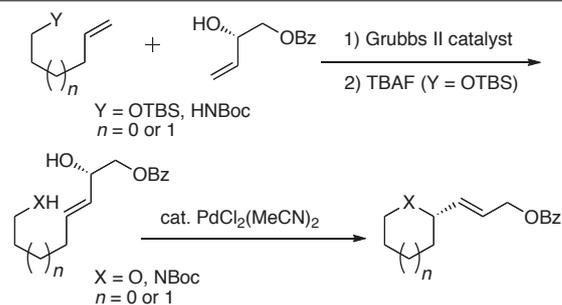
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 Giuseppe Cremonesi, Piero Dalla Croce,* Maddalena Gallanti,
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 Tetrahydroquinazoline *o*-Azaxylylene Aza Diels-Alder Reaction Imine

1463 A Short Access to Chiral Non-Racemic Oxa- and Azaheterocycles by Cross-Metathesis and Pd-Catalyzed Cyclization Sequence

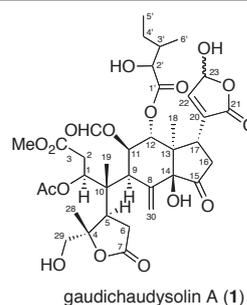
Jun'ichi Uenishi* and Yogesh S. Vikhe



Cyclization Palladium Catalyzed Reaction Cross Metathesis 1,3-Chirality Transfer Oxa- and Aza-Heterocycles

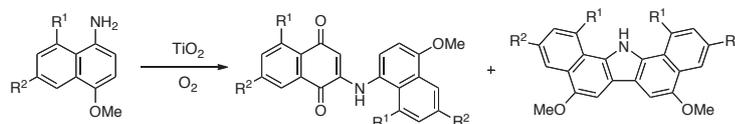
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Yuta Nagakura, Reiko Yamanaka, Yusuke Hirasawa, Takahiro Hosoya, Abdul Rahman, Idha Kusumawati, Noor Cholies Zaini, and Hiroshi Morita*


 Gaudichaudysolin A Limonoid *Dysoxylum gaudichaudianum* Meliaceae

1479 Oxidative Dimerization of 4-Methoxynaphthylamines in the Presence of Semiconductors

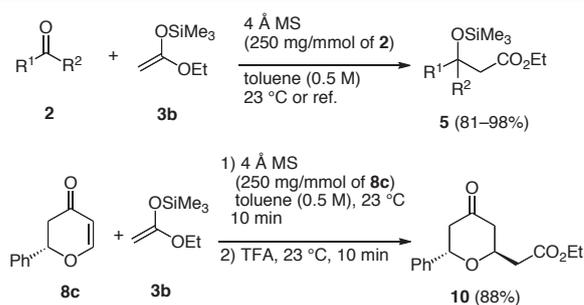
Tetsuya Takeya,* Yosuke Takahashi, Iwao Okamoto, and Osamu Tamura*



Molecular Oxygen Semiconductor 4-Methoxynaphthylamine Oxidative Dimerization Titanium Oxide

1489 The Mukaiyama Aldol and Mukaiyama–Michael Reactions Promoted by Commercially Available Molecular Sieves

Masahiro Anada, Takuya Washio, Yudai Watanabe, and Shunichi Hashimoto*



Mukaiyama Aldol Reaction Mukaiyama–Michael Reaction Molecular Sieves Silylketene Acetal Silyl Enol Ether

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HETEROCYCLES, Vol. 80, No. 2, 2010, pp. 1471 - 1477. © The Japan Institute of Heterocyclic Chemistry
 Received, 4th August, 2009, Accepted, 18th September, 2009, Published online, 24th September, 2009
 DOI: 10.3987/COM-09-S(S)106

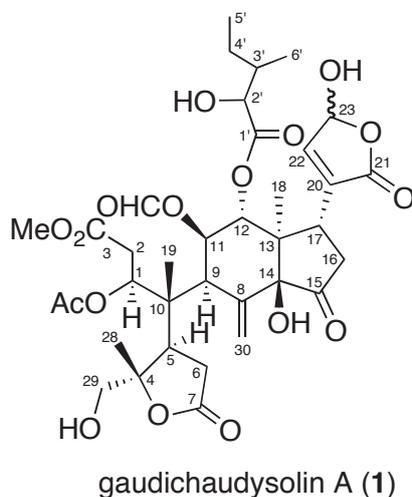
GAUDICHAUDYSOLIN A, A NEW LIMONOID FROM THE BARK OF *DYSOXYLUM GAUDICHAUDIANUM*

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Abstract – A new limonoid, gaudichaudysolin A (**1**) was isolated from the bark
 of *Dysoxylum gaudichaudianum* (Meliaceae) and the structure was elucidated by
 spectroscopic analysis.

Limonoids, highly oxidative unique secondary metabolites obtained from Meliaceae are produced by a unique biosynthetic route through tetranortriterpenoid nucleus,^{1,2} and are known to show various biological activities such as insecticidal, insect antifeedant, antibacterial, antifungal, antimalarial, anticancer, and antiviral activities.³ Recently, we have isolated new limonoids, ceramicines A – D⁴ with an unique tetranortriterpenoid skeleton from *Chisocheton ceramicus* and walsogyne A⁵ with a ring C–*seco* limonoid from *Walsura chrysogyne*. They showed an antiplasmodial and cytotoxic activities.^{4,5}



[†]Dedicated to Professor Emeritus Akira Suzuki, Hokkaido University, on the occasion of his 80th birthday.

In continuation of our research on limonoids containing in the plants belonging to Meliaceae family, we have isolated a new limonoid, gaudichaudysolin A (**1**) from the bark of *Dysoxylum gaudichaudianum*. Herein we report the structure elucidation of gaudichaudysolin A (**1**) by spectroscopic methods.

The bark of *D. gaudichaudianum* was extracted with MeOH, and the MeOH extract was in turn partitioned between EtOAc and H₂O. EtOAc-soluble materials were subjected to a silica gel column (hexane/EtOAc, 1:0→1:1; CHCl₃/MeOH, 1:0→0:1) and the fractions eluted by hexane/EtOAc (1:1) were subjected to a silica gel column (Toluene/EtOAc, 1:0→5:5; CHCl₃/MeOH, 1:0→5:5; CHCl₃/MeOH/H₂O, 5:5:1) followed by C₁₈ HPLC (40% CH₃CN/0.1%TFA) to afford gaudichaudysolin A (**1**, 0.00002 %).

Gaudichaudysolin A {**1**, [α]_D²³ -126 (c 0.2, MeOH)} was obtained as a colorless solid and was revealed to have the molecular formula C₃₆H₄₈O₁₇, by HRESITOFMS [*m/z* 775.2780 (M+Na)⁺, Δ -0.9 mmu]. IR absorptions implied the presence of hydroxyl (3425 cm⁻¹) and carbonyl (1755 and 1680 cm⁻¹) groups. UV spectrum (230 nm) indicated the presence of an unsaturated carbonyl group. ¹H and ¹³C NMR data (Table 1) revealed thirty six carbon resonances due to seven carbonyls, two sp² quaternary carbons, three sp³ quaternary carbons, one sp² methine, ten sp³ methines, one sp² methylene, five sp³ methylenes, and seven methyl groups. Among them, eight sp³ carbons (δ_C 67.3, 70.0, 72.0, 75.2, 76.3, 81.4, 93.1, and 98.8) and seven sp² carbons (δ_C 163.5, 172.1, 172.9, 173.2, 173.3, 177.8, and 208.0) were ascribed to those bearing an oxygen atom.

Six partial structures **a** (C-1 to C-2), **b** (C-5 to C-6), **c** (C-9 and C-11 to C-12), **d** (C-16 to C-17), **e** (C-22 to C-23), and **f** (C-2' to C-6') were deduced from ¹H-¹H COSY analysis of **1** in CD₃OD (Figure 1). Unit **A** composed of the partial structures **a** and **b** was assigned as shown in Figure 1 with a γ -lactone ring by using HMBC and NOESY correlations as follows. HMBC correlations of H₃-28 (δ_H 1.58) to C-4 (δ_C 93.1), C-5 (δ_C 43.8), and C-29 (δ_C 67.3), H₂-6 (δ_H 2.80) to C-4 and C-7 (δ_C 177.8), and H₂-29 (δ_H 3.72 and 3.78) to C-4 revealed the presence of a γ -lactone ring⁶ with a hydroxymethyl and a methyl groups at C-4. Acetoxy group at C-1 and a methyl carboxylate at C-2 were assigned by the HMBC correlations as shown in Figure 1. Connection between the partial structures **a** and **b** through C-10 (δ_C 49.6), was deduced by an HMBC correlation of H₂-6 to C-10 and the NOESY correlations as shown in Figure 2. Unit **B** composed of the partial structures **c** and **d** was assigned as an octahydroinden-1-one ring system with a methyl, an exo-methylene, a hydroxyl, and two ester functions as follows. These functions can be connected by the HMBC correlations of H₃-18 (δ_H 1.02) to C-12 (δ_C 75.2), C-13 (δ_C 51.5), C-14 (δ_C 81.4), and C-17 (δ_C 36.8), H-9 (δ_H 3.29) to C-8 (δ_C 142.0) and C-14, and H₂-30 (δ_H 5.75 and 5.84) to C-9 and C-14. This ring system and functions were also supported by the comparison of the ¹H and ¹³C NMR chemical shifts [C-14 (δ_C 81.4), C-15 (δ_C 208.0), and C-16 (δ_C 40.8)] with those [C-14 (δ_C 79.5), C-15 (δ_C 209.3), and C-16 (δ_C 42.0)] of rohituka 14⁷ isolated from the seeds of *Aphanamixis polystacha*. The presence of a formate at C-11 was indicated by an HMBC correlation of an aldehyde proton to C-11 (δ_C

72.0). Connection between the units **A** and **B** was indicated by HMBC correlations of H-9 to C-5 and C-10. Unit **C** composed of the partial structure **e**, which was attached at C-17 in the unit **B**, was assigned as an α -substituted γ -lactone ring system with a hydroxyl at γ position by HMBC correlations of H-17 (δ_{H} 3.86) and H-22 (δ_{H} 7.19) to C-20 (δ_{C} 136.5), H-23 (δ_{H} 6.10) to C-21 (δ_{C} 172.9), and IR absorption at 1755 cm^{-1} . Unit **D** composed of the partial structure **f** was assigned as a 2-hydroxy-3-methylpentanoic acid by the ^1H - ^1H COSY correlation in Figure 1 and the comparison of NMR data in turrapubesin D,⁸ which might be attached at the hydroxy group at C-12 (δ_{C} 75.2). Thus, the gross structure of **1** was assigned as A,B-*seco*-tetranorlimonoid skeletal system with a γ -butanolide at C-17.

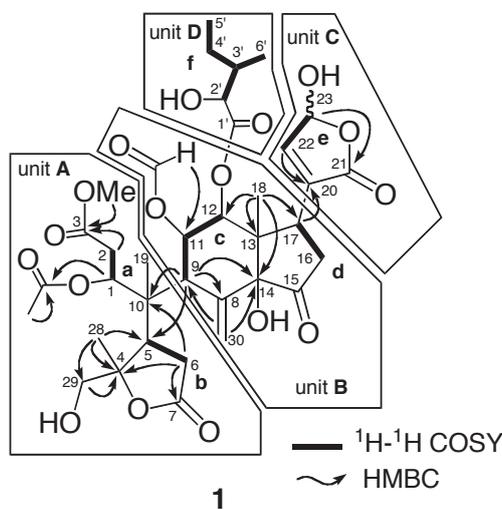


Figure 1. Selected 2D NMR correlations for **1**

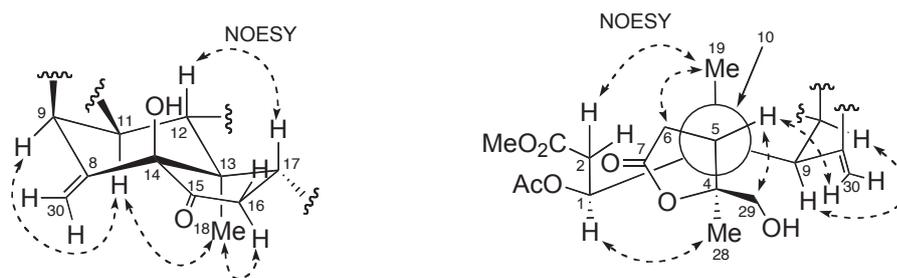


Figure 2. Selected NOESY correlations in unit **B** and rotation model of C-5/C-10 bond in unit **A** for **1**

The relative stereochemistry with selected NOESY correlations in unit **B** and rotation model of C-5/C-10 bond in unit **A** for **1** were elucidated by NOESY correlations as shown in Figure 2. Configurations of C-9, C-11, C-12, C-13, and C-17 in the unit **B** were elucidated by NOESY correlations of H₃-18/H-11 and H-16a, H-12/H-17, and H-9/H-11. As you can see the rotation model of C-5/C-10 bond, NOESY correlations of H₃-19/H-2 and H₂-6, H₃-28/H-1, and H-5/H₂-29 and H-30 indicated connectivity of C-5/C-10 bond and the relative stereochemistry in the unit **A** as shown in Figure 2. Thus, the relative configuration of **1** was assigned to be shown in computer-generated 3D drawing in Figure 3 except for

C-2' and C-3' in the 2-hydroxy-3-methylpentanoic acid.

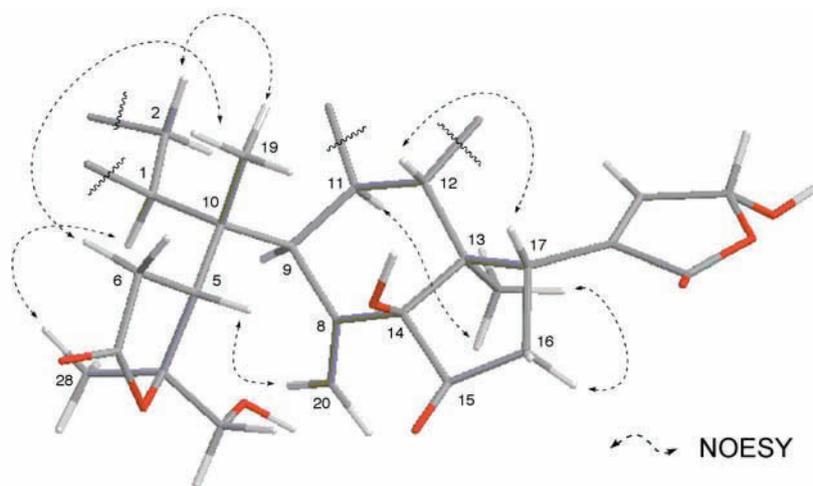


Figure 3. Selected NOESY correlations and relative configurations for **1**

From the bark of *Dysoxylum gaudichaudianum*, a series of dysoxylins A – D with a tetranortriterpenoid nucleus have already been isolated.⁹ Biogenetically, gaudichaudysolin A (**1**) may be derived by a unique oxidative route through this tetranortriterpenoid nucleus. Gaudichaudysolin A (**1**) was evaluated *in vitro* for cytotoxicity against five human cancer cell lines, HL60 (human blood premyelocytic leukemia), RPMI8226 (multiple myeloma), NCI-H226 (non-small cell lung carcinoma), HCT116 (human colon cancer), and MCF7 (human breast adenocarcinoma) cells, using MTT assay, but showed no inhibitory activity against five tested cell lines ($IC_{50} > 50 \mu\text{M}$).

EXPERIMENTAL

General Experimental Procedures. ^1H and 2D NMR spectra were recorded on a Bruker AV600 spectrometer and chemical shifts were reported using residual CD_3OD (δ_{H} 3.31 and δ_{C} 49.0) as internal standards. HSQC experiments were optimized for $^1J_{\text{CH}}=145$ Hz and HMBC experiments for $^nJ_{\text{CH}}=8$ Hz. Mass spectra were recorded on a Micromass LCT spectrometer.

Plant Material. The bark of *D. gaudichaudianum* was collected at Alas Purwo, Indonesia in 2007. A voucher specimen is deposited at the Purwodadi Botanical Garden, Indonesia.

Extraction and Isolation. The bark of *D. gaudichaudianum* (1370 g) was extracted with MeOH, and the MeOH extract was partitioned between EtOAc and H_2O . Water-soluble materials were extracted with BuOH. EtOAc-soluble materials were subjected to a silica gel column (hexane/EtOAc, 1:0→1:1; $\text{CHCl}_3/\text{MeOH}$, 1:0→0:1) and the fractions eluted by hexane/EtOAc (1:1) were subjected to a silica gel column (toluene/EtOAc, 1:0→5:5; $\text{CHCl}_3/\text{MeOH}$, 1:0→5:5, $\text{CHCl}_3/\text{MeOH}/\text{H}_2\text{O}$ 5:5:1) followed by C_{18} HPLC (40% $\text{CH}_3\text{CN}/0.1\%$ TFA) to afford gaudichaudysolin A (**1**, 1.3 mg, 0.00002 %).

Table 1. ^1H and ^{13}C NMR Data [δ_{H} (J , Hz) and δ_{C}] of Gaudichaudysolin A (**1**) in CD_3OD at 300K

Position	δ_{H}	δ_{C}
1	5.70 (1H, m)	70.0
2a	2.70(1H, dd,15.0,11.4)	36.0
2b	3.05 (1H, d, 14.4)	
3		173.2
4		93.1
5	3.16 (1H, m)	43.8
6	2.80 (2H, d, 10.2)	35.7
7		177.8
8		142.0
9	3.29 (1H, m)	55.4
10		49.6
11	5.38 (1H, t, 10.2)	72.0
12	5.97 (1H, d, 10.8)	75.2
13		51.5
14		81.4
15		208.0
16a	2.51 (1H, m)	40.8
16b	2.86 (1H, m)	
17	3.86 (1H, m)	36.8
18	1.02 (3H, s)	13.0
19	1.44 (3H, s)	20.0
20		136.5
21		172.9
22	7.19 (1H, br s)	148.6
23	6.10 (1H, br s)	98.8
28	1.58 (3H, s)	20.0
29a	3.72 (1H, d, 13.2)	67.3
29b	3.78 (1H, d, 13.2)	
30a	5.52 (1H, br s)	123.3
30b	5.84 (1H, br s)	
1-OAc	2.04 (3H, s)	21.1
		172.1
3-OMe	3.65 (3H, s)	52.4
11-OCHO	8.15	163.5
1'		173.3
2'	3.86 (1H, m)	76.3
3'	1.62 (1H, m)	39.3
4'a	1.28 (1H, m)	24.4
4'b	1.20 (1H, m)	
5'	0.86 (3H, s)	11.9
6'	0.93 (3H, s)	15.9

Gaudichaudysolin A (1): a colorless amorphous solid; $[\alpha]_D^{23}$ -126 (*c* 0.2, MeOH); IR (KBr) ν_{\max} 3425, 1755, 1680, 1630, 1585, 1440, 1390, 1200, 1135, 1075, 840, and 801 cm^{-1} ; UV (MeOH) λ_{\max} 230 (ϵ 9000); ^1H and ^{13}C NMR data (Table 1); ESIMS m/z 775 ($\text{M}+\text{Na}$) $^+$; HRESITOFMS m/z 775.2780 [$\text{M}+\text{Na}$] $^+$, calcd for $\text{C}_{36}\text{H}_{48}\text{O}_{17}$, 775.2789].

Cytotoxic Activity. Each cell line [HL60 (human blood premyelocytic leukemia), RPMI8226 (multiple myeloma), NCI-H226 (non-small cell lung carcinoma), HCT116 (human colon cancer), and MCF7 (human breast adenocarcinoma) cells] was seeded onto 96-well microtiter plates at 1×10^4 cells per well for HL60 and RPMI8226 and 5×10^3 cells per well for NCI-H226, HCT116, and MCF7, respectively. Cells were preincubated for 24 h at 37°C in humidified atmosphere of 5% CO_2 . Different concentrations of each compound (10 μL) were added to the cultures, and then the cells were incubated at 37°C for 48 h. On the third day, 15 μL MTT solution (5 mg/mL) was added into each well of the cultured medium. After further 2 h of incubation, 100 μL of 10% SDS-0.01N HCl solution was added to each well and the formazan crystals in each well were dissolved by stirring with a pipette. The optical density measurements were made using a micropipette reader (Benchmark Plus microplate spectrometer, BIO-RAD) equipped with a two wavelengths system (550 and 700 nm). In each experiment, three replicate of wells were prepared for each sample. The ratio of the living cells was determined based on the difference of the absorbance between those of samples and controls. These differences are expressed in percentage and cytotoxic activity was indicated as an IC_{50} value.

ACKNOWLEDGMENTS

This work was supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan, and grants from the Research Foundation for pharmaceutical Sciences and The Open Research Center Project.

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