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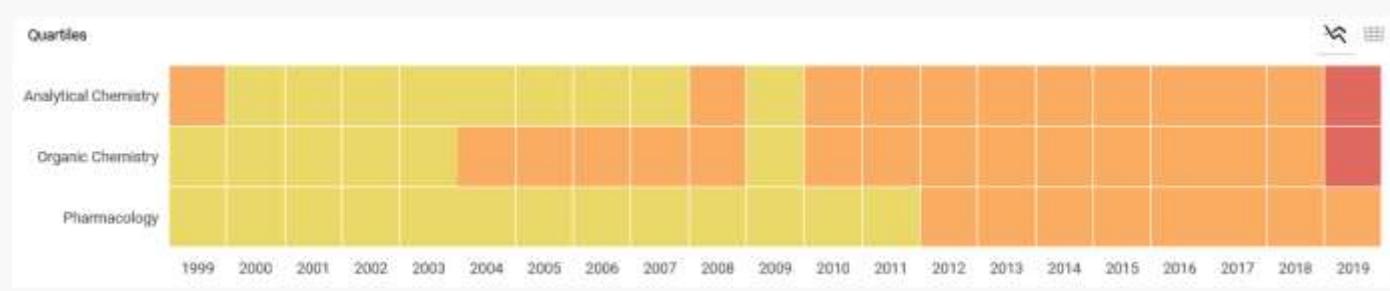
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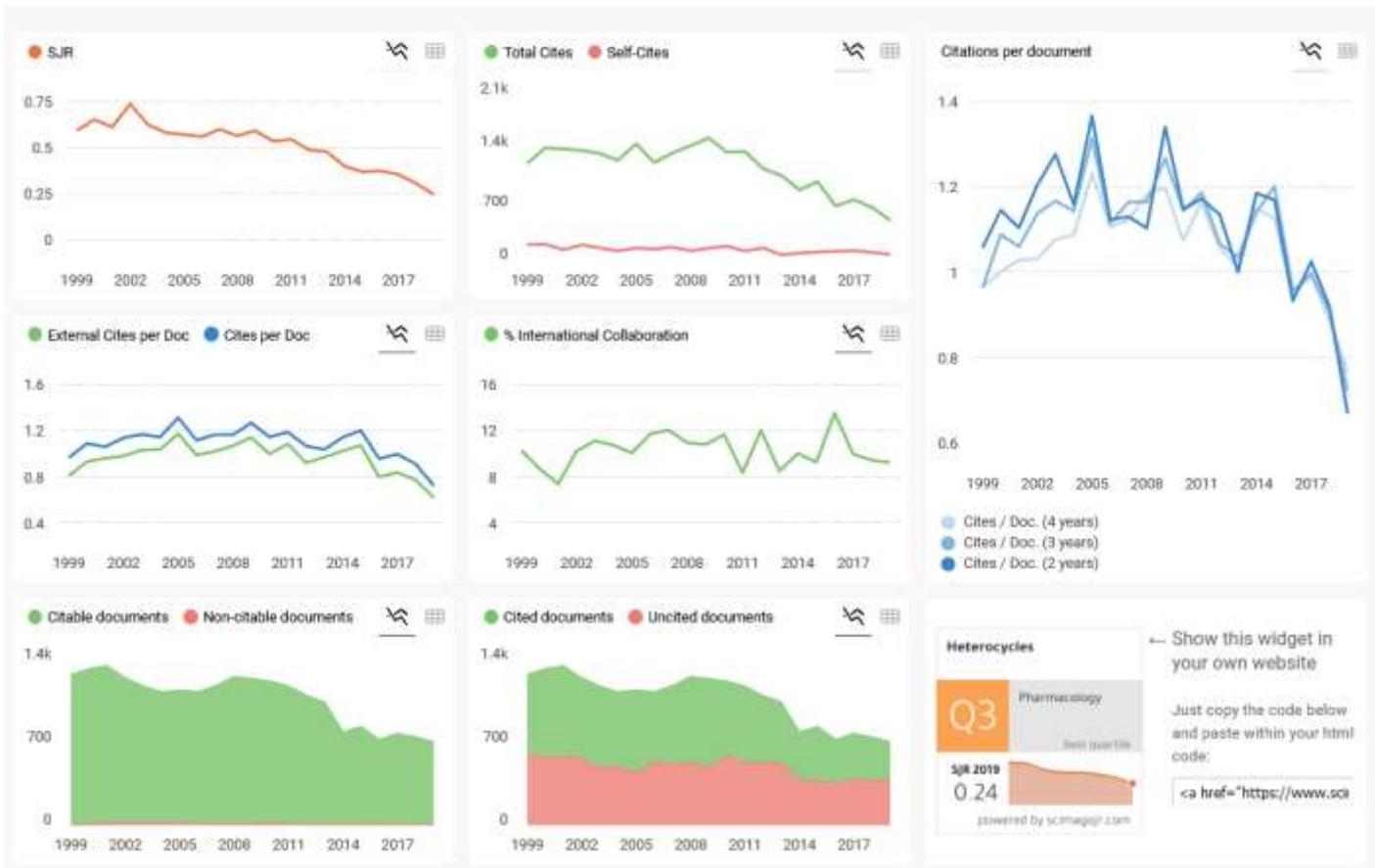
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SCOPE

Since its inception in 1973 HETEROCYCLES has provided a platform for the rapid exchange of research in the areas of organic, pharmaceutical, analytical, and medicinal chemistry of heterocyclic compounds in addition to communications, papers, reviews, a special section of the journal presents newly-discovered natural products whose structure has recently been established. Another section is devoted to the total synthesis of previously documented natural products with heterocyclic ring systems. Due to the fact that the journal is able to publish articles within two months of receipt of the manuscripts, researchers in this field can obtain up-to-date information on heterocyclic research by reading Heterocycles regularly. Audience: Organic and Physical Organic Chemists, Biochemists, Pharmacologists and Scientists studying heterocyclic compounds





■ CELEBRATION OF PROFESSOR ISAO KUWAJIMA

- 1 Preface to Heterocycles Issue
Honoring the 77th Birthday of Professor Isao Kuwajima
Eiichi Nakamura*

-
- 3 Preface to Heterocycles Issue
Honoring the 77th Birthday of Professor Isao Kuwajima
Keiji Tanino*

■ CURRICULUM VITAE

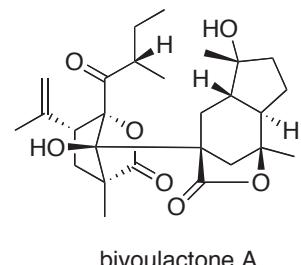
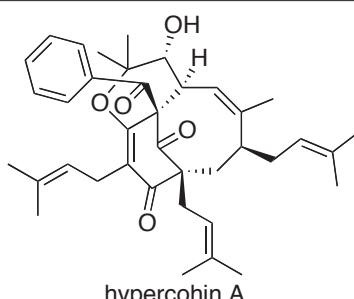
- 6 Curriculum Vitae
Isao Kuwajima*

■ PUBLICATIONS**7 List of Publication**

Isao Kuwajima*

**■ REVIEWS****23 Prenylated Acylphloroglucinols and Meroterpenoids from *Hypericum* Plants**

Naonobu Tanaka and Jun'ichi Kobayashi*

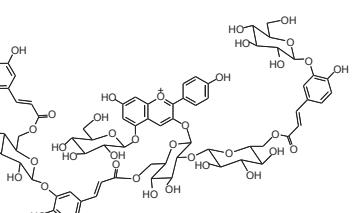
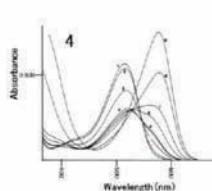


Prenylated Phloroglucinol

Meroterpenoid

Hypericum Plant**41 Light Absorption Spectral Patterns of Intact Garden Flowers in Relation to the Flower Colors and Anthocyanin Pigments**

Norio Saito, Fumi Tatsuzawa,* and Toshio Honda



Electronic spectra of ipomoea red anthocyanin 1

Anthocyanin Pigment

Intra- and Intermolecular Co-pigmentation

Self Association

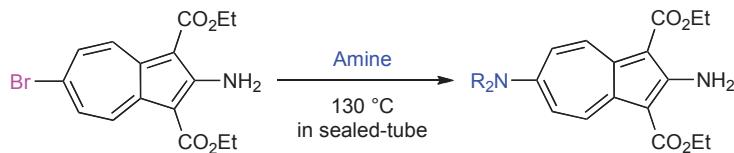
Light Absorption Spectral Pattern

Flower Color

■ COMMUNICATIONS

85 Synthesis of 2,6-Diaminoazulenes by the S_NAr Reaction with Cyclic Amines

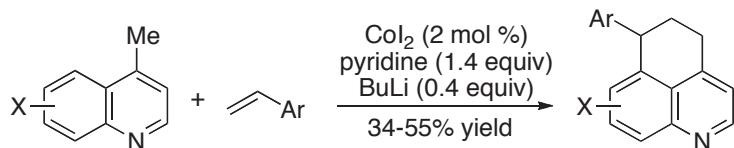
Taku Shoji,* Yuki Fujiwara, Akifumi Maruyama, Mitsuhsia Maruyama, Shunji Ito, Masafumi Yasunami (the late), Ryuji Yokoyama, and Noboru Morita



Azulene Amination Aromatic Nucleophilic Substitution

89 Cobalt-Catalyzed C5-Selective C-H Functionalization of 4-Me-Quinolines with Styrenes: An Approach to 5,6-Dihydro-4*H*-benzo[*d,e*]quinolines

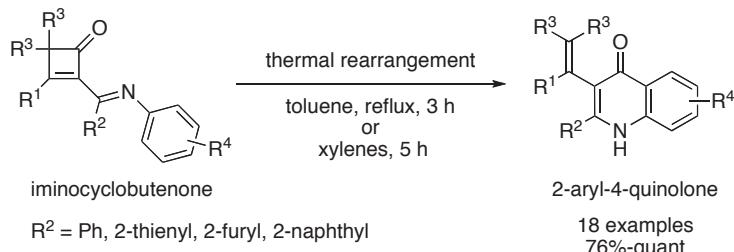
Shohei Yamamoto, Shigeki Matsunaga,* and Motomu Kanai*



C-H Functionalization Cobalt Catalyst Quinoline Transition Metal Catalysis Synthetic Method

97 2-Aryl-4-quinolone Synthesis Using the Thermal Rearrangement of Iminocyclobutenones

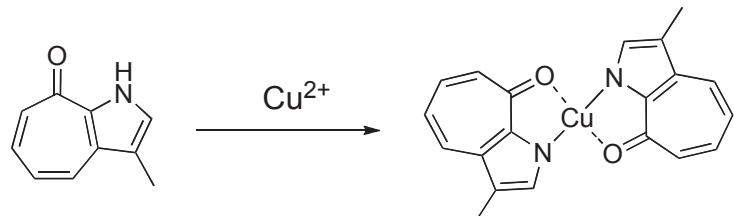
Iwao Hachiya, Keiichi Yokoyama, Akinori Ito, and Makoto Shimizu*



4-Quinolone Iminocyclobutene Thermal Rearrangement Alkynyl Imine Conjugate Addition Reaction

104 Crystal Structures of 3-Methylpyrrolo[2,3-*b*]tropone and Its Copper(II) Complex

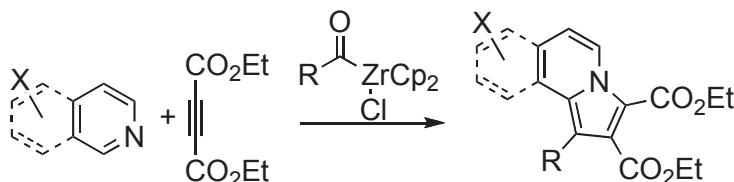
Kanji Kubo,* Taisuke Matsumoto, Keiko Ideta, and Akira Mori



Troponoid Pyrrolotropone Cu(II) Complex Crystal Structure X-Ray Analysis

108 Three-Component Synthesis of Indolizines from Azaaromatic-Acetylenedicarboxylate Zwitterions with Acylzirconocene Chloride Complexes

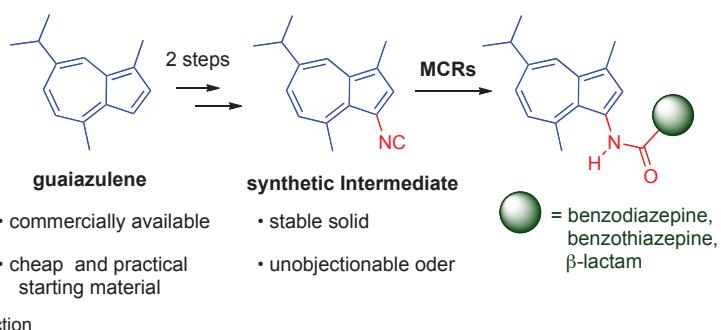
Akio Saito,* Naoki Yamashita, Kohei Sudo, and Yuji Hanzawa



Acylzirconocene Chloride Azaaromatic Compound Nucleophilic Addition Three-Component Synthesis Zwitterion

113 Facile Synthesis of Guaiazulene-Heterocycle Hybrids via Ugi Multicomponent Reactions

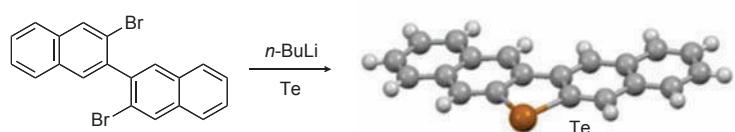
Naoko Takenaga,* Koh Fukazawa, Miki Maruko, and Koichi Sato*



Guaiazulene Isocyanide Multicomponent Reaction Ugi Reaction

121 Synthesis and Structural Characterization of a Novel Organotellurium Compound: Dinaphtho[2,3-*b*;2',3'-*d*]-tellurophene

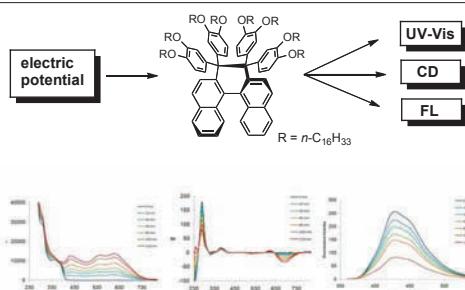
Mio Matsumura, Atsuya Muranaka, Naoki Kakusawa, Jyoji Kurita, Daisuke Hashizume, Masanobu Uchiyama,* and Shuji Yasuike*



Dinaphthotellurophene Tellurium Planar Structure Group 16 Element HOMO-LUMO Gap

126 Three-Way Output Molecular Response System Based on Tetrakis(3,4-dialkoxyphenyl)-3,4-dihydro[5]helicenes: Perturbation of Properties by Long Alkyl Chains

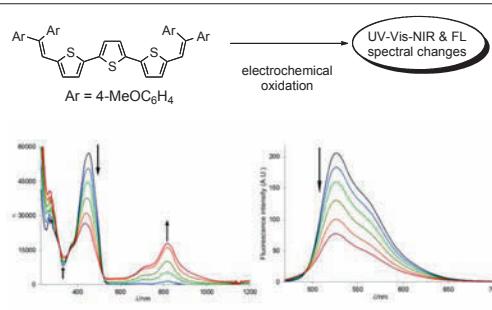
Yusuke Ishigaki, Satoshi Yoshida, Hidetoshi Kawai, Ryo Katono, Kenshu Fujiwara, Takanori Fukushima, and Takanori Suzuki*



Redox System Electrochromism Long Alkyl Chain Circular Dichroism Dihydroxepin

136 Bis(diarylethenyl)thiophene, -bithiophene, and -terthiophene: A New Series of Violene-Cyanine Hybrid-Type Electron Donors

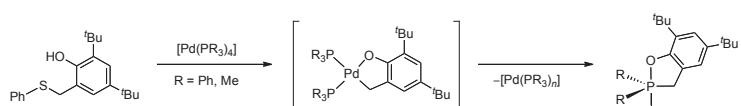
Yusuke Ishigaki, Hidetoshi Kawai, Ryo Katono, Kenshu Fujiwara, and Takanori Suzuki*



Electron Donor Redox System Electrochromism NIR Fluorescence

144 Palladium(0)-Mediated C-S and O-H Bonds Cleavage Reaction in 2-Hydroxybenzyl Phenyl Sulfide: Formations of Oxyphosphorane and 1,2-Oxapalladacycle

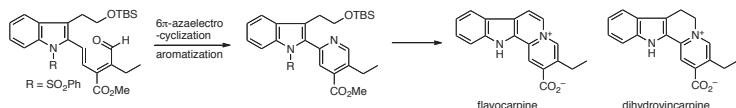
Norio Nakata, Noriyuki Furukawa, Hiroki Kobayashi, Izuru Suzuki, and Akihiko Ishii*



Oxyphosphorane Oxapalladacycle

150 Total Syntheses of Zwitterionic Indole Alkaloids, Flavocarpine and Dihydrovincarpine, by Extended Method for Substituted Pyridine Synthesis through Azaelectrocyclization

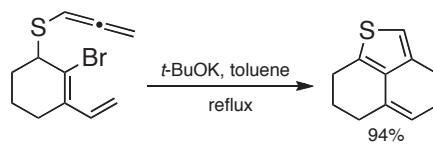
Yoshikatsu Hirose, Hiroshi Tsuchikawa,
Toyouharu Kobayashi,* and Shigeo Katsumura*



Total Synthesis Zwitterionic Indole Alkaloid Substituted Pyridine Azaelectrocyclization

157 Tandem [4+2] Cycloaddition/Aromatization Sequence of Allenyl 2-Bromo-3-vinylcyclohex-2-enyl Thioether to Naphtho[1,8-*b*c]thiophene

Noriyuki Hatae,* Aiichirou Kaji, Chiaki Okada, and Eiko Toyota

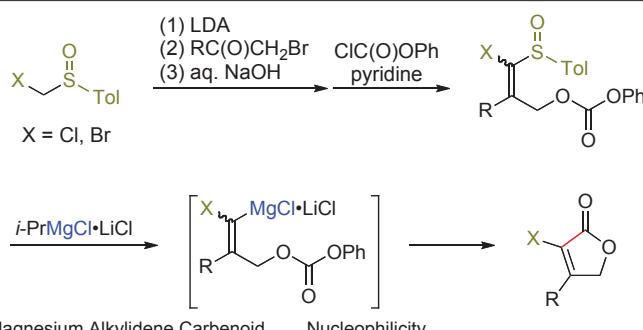


	ΔH (kcal/mol)	
	ΔH_{TS}	ΔH_{react}
X = H, Y = O	18.38	-44.17
X = Me, Y = O	24.85	-43.82
X = Br, Y = O	24.64	-49.26
X = Br, Y = S	31.86	-46.17

Naphtho[1,8-*b*c]thiophene Tandem [4+2] Cycloaddition/Aromatization Density Functional Study

163 Synthesis of α -Halobutenolides Using the Nucleophilicity of Magnesium Alkylidene Carbenoids

Tsutomu Kimura,* Kazuki Fukuda, Gaku Kashiwamura, and Tsuyoshi Satoh*

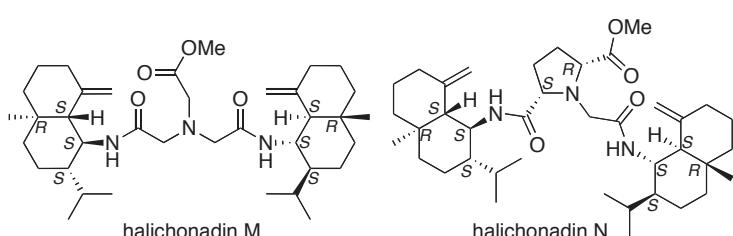


α -Halobutenolide 3-Halofuran-2(5*H*)-one Cyclization Reaction Magnesium Alkylidene Carbenoid Nucleophilicity

■ PAPERS

173 Halichonadins M-Q, Sesquiterpenes from an Okinawan Marine Sponge *Halichondria* sp.

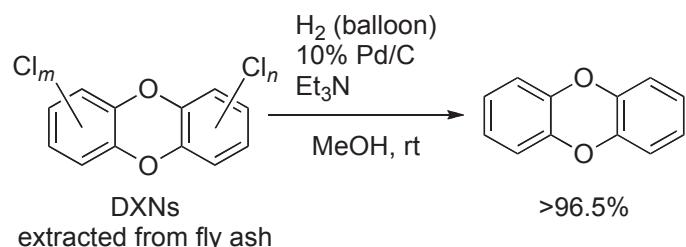
Naonobu Tanaka, Shohei Suto, Miki Asai, Taishi Kusama, Azusa Takahashi-Nakaguchi, Tohru Gono, Jane Fromont, and Jun'ichi Kobayashi*



Marine Sponge *Halichondria* sp. Sesquiterpene Halichonadins M-Q

186 Pd/C-Catalyzed Hydrodechlorination of Dioxins from Fly Ash under Ambient Pressure and Temperature

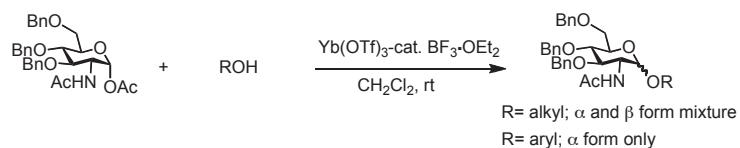
Yasunari Monguchi,* Akiko Ido, Miki Niikawa, Nobuharu Nagatsu, Ryosuke Mizukoshi, Hisamitsu Nagase, and Hironao Sajiki*



Hydrodechlorination Reaction Palladium on Carbon Dioxin Fly Ash Degradation

198 Formation of 2-Acetamido-2-deoxy-D-glucopyranosidic Linkages *via* Glycosidation Using a Combination of Two Lewis Acids

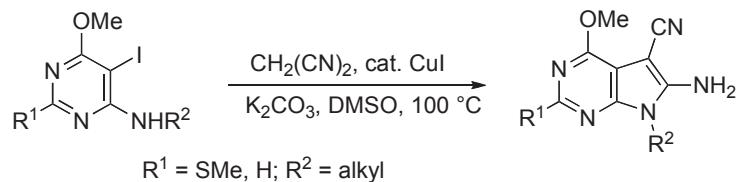
Yoshiki Oda, Masanobu Midorikawa, and Takashi Yamanoi*



Glycosylation Aryl Glycoside Arbutin 4-Hydroxyphenyl 2-Acetamido-2-deoxy- α -D-glucopyranoside Ytterbium Triflate

216 Synthesis of 7-Alkyl-6-amino-7*H*-pyrrolo[2,3-*d*]-pyrimidine-6-carbonitriles by the Copper-Catalyzed Reaction of 4-(Alkylamino)-5-iodopyrimidines with Propanedinitrile

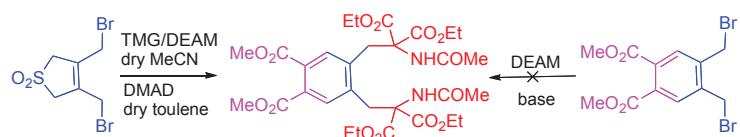
Kazuhiro Kobayashi,* Kazuya Nakazawa, Shohei Yuba, Hidetaka Hiyoshi, and Kazuto Umezawa



7*H*-Pyrrolo[2,3-*d*]pyrimidine Amino Nitrile Copper(I) Iodide Coupling Reaction Malononitrile

226 Diversity Oriented Approach to Phenylalanine Derivatives *via* the Diels-Alder Reaction Involving Sulfolene Intermediates

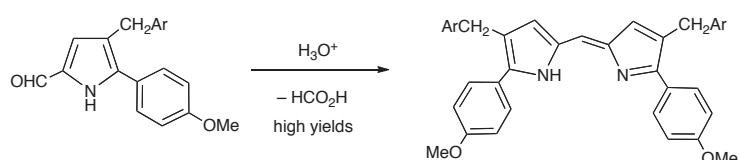
Sambasivarao Kotha* and Vijayalakshmi Bandi



α-Amino Acid Diels-Alder Reaction Sulfolene Diethyl Acetamidomalonate Unusual Amino Acid

238 Unusually Efficient Deformylative Synthesis of 1,2,8,9-Tetrasubstituted Dipyrromethenes from 4,5-Disubstituted Pyrrole-2-carbaldehydes

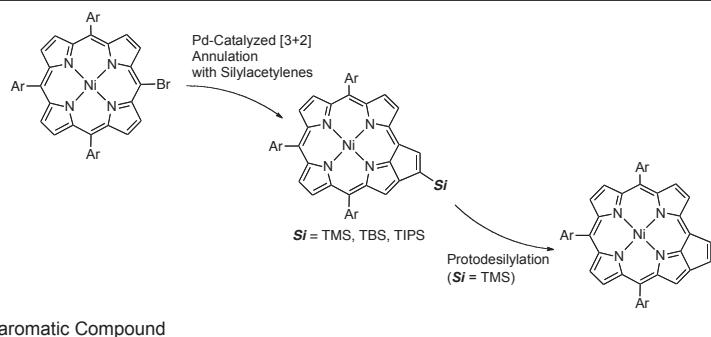
Mitsunori Oda,* Yurie Fujiwara, Yoshimitsu Kumai, Akira Ohta, and Ryuta Miyatake



Dipyrromethene Pyrrole Proton Sensor X-Ray Crystallographic Analysis BODIPY

252 Palladium-Catalyzed [3+2] Annulation of *meso*-Bromoporphyrin with Silylacetylenes and Desilylation of 8^a-Silyl-7,8-dehydropurpurin

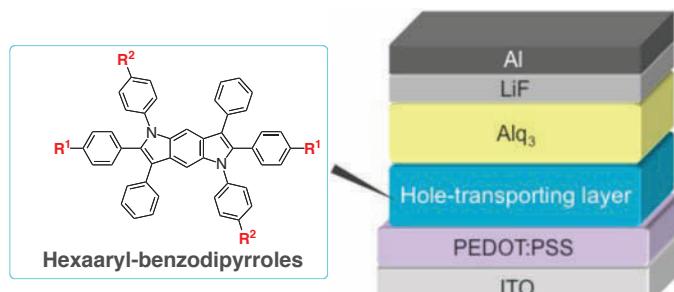
Norihito Fukui, Seiji Arai, Hiroshi Shinokubo, Hideki Yorimitsu,* and Atsuhiro Osuka*



Porphyrin Dehydropurpurin Annulation Pd Catalyst Antiaromatic Compound

261 Hexaaryl-benzodipyrroles: Properties and Application as Amorphous Carrier-Transporting Materials

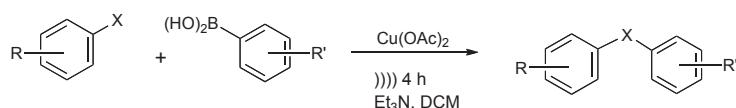
Hayato Tsuji,* Yuki Yokoi, Shunsuke Furukawa, and Eiichi Nakamura*



Pyrrole Carrier Mobility Semiconducting Material Light-Emitting Diode

271 Copper Mediated Formation of Carbon-Heteroatom Bonds Using Organoboron Reagents and Ultrasound

Bryan J. Musolino and George W. Kabalka*

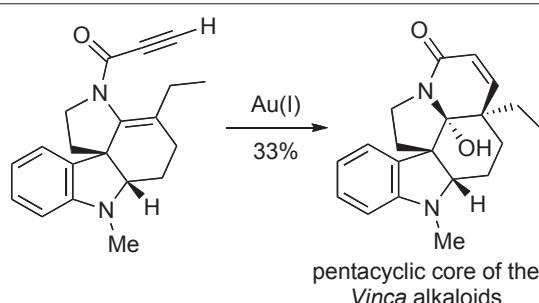


where X = OH, SH, NH₂ and R/R' = various functional groups

Heteroatom Organoboron Coupling Reaction Ultrasound Copper

298 ABC → ABCE/D Based Approaches to the Pentacyclic Ring System of the *Vinca* Alkaloids Using Intramolecular Hetero-[2+2]cycloaddition and Gold(I)-Catalyzed 6-*endo-dig* Cyclisation Protocols

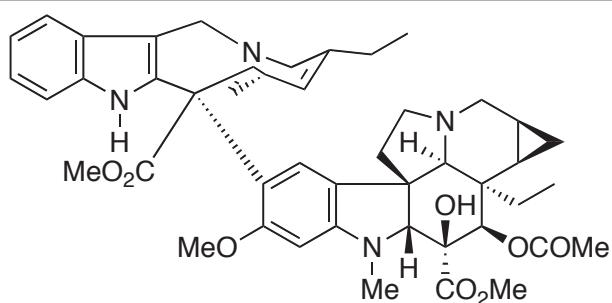
Lorenzo V. White, Martin G. Banwell,* and Anthony C. Willis



Vinca Alkaloid 6-*endo-dig* Cyclisation Au(I)-Catalysis Fischer Indole Synthesis

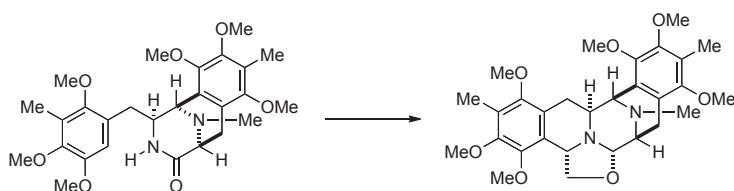
316 Bisindole Alkaloids Condensed with a Cyclopropane Ring, Part 2. Cyclopropano-vinorelbine and Its Derivatives

Péter Keglevich, László Hazai, Zsófia Dubrovay, Zsuzsanna Sánta, Miklós Dékány, Csaba Szántay, Jr., György Kalaus, and Csaba Szántay*



327 Synthetic Studies on Saframycin Antibiotics: An Improved Synthesis of Tricyclic Lactam Intermediate and Construction of the Core Ring System of Saframycin A

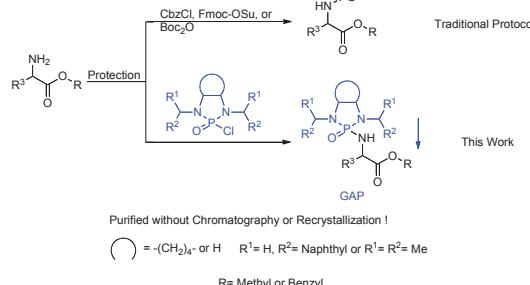
Shinya Kimura, Shintaro Kawai, Masayuki Azuma, Yoshifumi Umebara, Yu-ichi Koizumi, Masashi Yokoya, and Naoki Saito*



Aminonitrile Isoquinoline Cytotoxicity Saframycin A

344 Group-Assisted Purification (GAP) for Protection of Amino Acids Using *N*-Phosphonyl Functional Groups

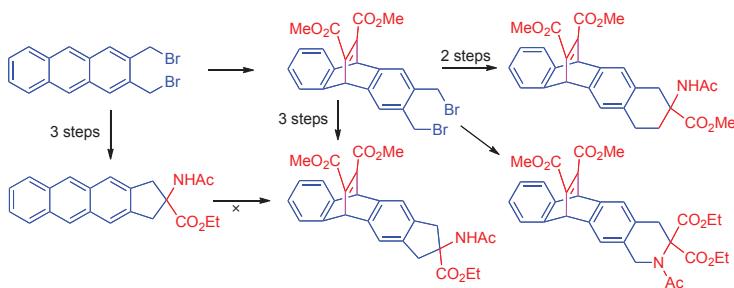
Guanghui An, Cole Seifert, Hao Sun, Yi Pan, and Guigen Li*



Group-Assisted Purification Phosphonyl Halide *N*-Phosphonyl Amino Acid

357 Design and Synthesis of Conformationally Constrained Bicyclo[2.2.2]octane-based Unusual α -Amino Acid Derivatives via the Diels–Alder Reaction

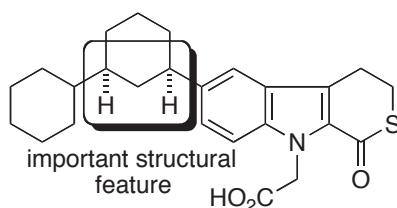
Sambasivarao Kotha* and Milind Meshram



Bicyclic α -Amino Acid Derivative Anthracene Derivative α -Quinodimethane Diels–Alder Reaction Glycine Equivalent

372 Synthesis and Evaluations of GLP-1 Secretion and Anti-Diabetic Effect in KKAY Mice of New Tricyclic Compounds

Daisuke Minehira, Daisuke Takeda, Shota Miyawaki, Atsushi Kato, Isao Adachi,* Akira Miyazaki, Ryuta Miyatake, Masahito Umezaki, Kyoko Miura, Yoshiro Kitahara, Kenji Sugimoto, Yuji Matsuya, and Naoki Toyooka*

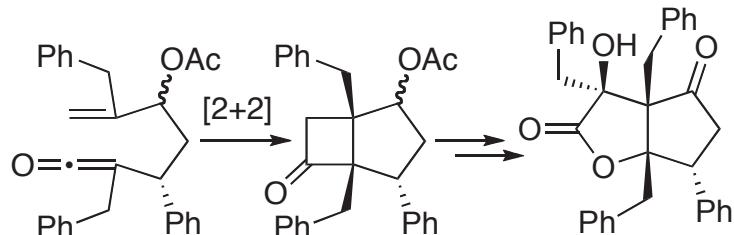


strong GLP-1 secretion and anti-diabetic properties

GLP-1 Secretion Tricyclic Compound Friedel-Crafts Reaction

405 Stereoselective Approach toward Ophiodilactones Based on an Intramolecular [2 + 2] Cycloaddition Reaction

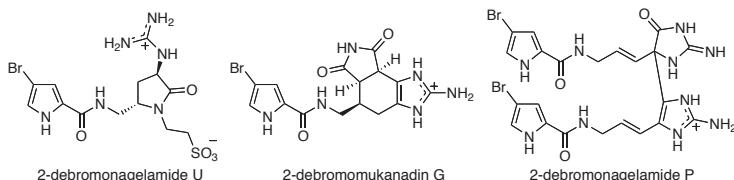
Takaaki Matsubara, Jun Ishihara, and Susumi Hatakeyama*



Baeyer–Villiger Oxidation [2 + 2] Cycloaddition Ketene Lactone Natural Product Synthesis

425 2-Debromonagelamide U, 2-Debromomukanadin G, and 2-Debromonagelamide P from Marine Sponge *Ageas* sp.

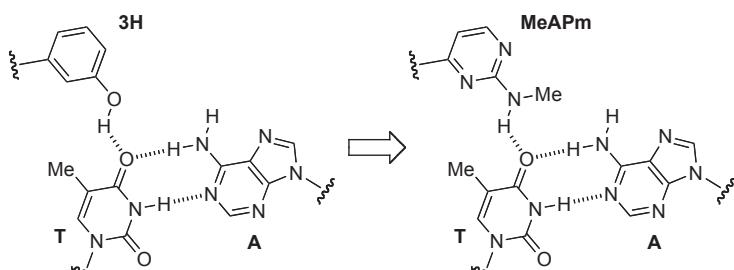
Kenta Nakamura, Taishi Kusama, Naonobu Tanaka, Kanae Sakai, Tohru Gonoi, Jane Fromont, and Jun'ichi Kobayashi*



Bromopyrrole Alkaloid *Ageas* sp. 2-Debromonagelamide U 2-Debromomukanadin G 2-Debromonagelamide P

432 Base Pair Recognition Ability of 2-(Methylamino)-pyrimidin-4-yl Nucleobase in Parallel Triplex DNA

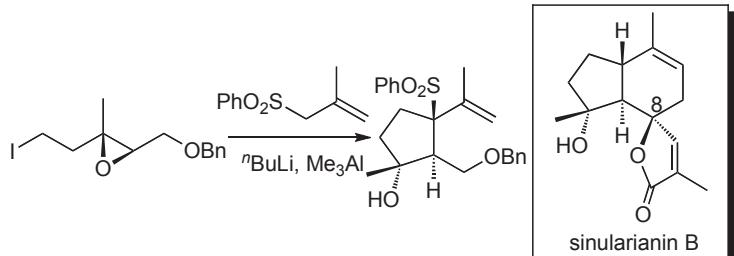
Yoshiyuki Hari,* Satoshi Kashima, Yuya Matsuda, Akihiro Sakata, Ryutaro Takamine, Shin Ijitsu, and Satoshi Obika*



Modified Nucleobase Triplex DNA Triplex-Forming Oligonucleotide

442 Total Synthesis of Marine Sesquiterpenoid Sinularianin B and 8-*epi*-Sinularianin B

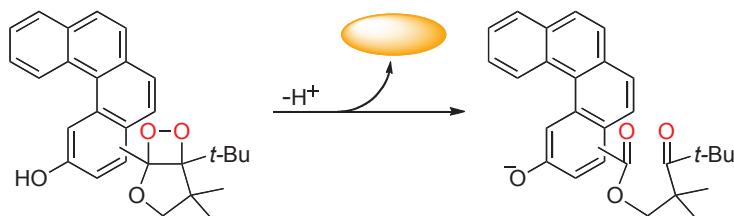
Koichiro Ota and Hiroaki Miyaoka*



Sinularianin B Total Synthesis Spirolactone One-Pot Synthesis Natural Product

462 Synthesis of Bicyclic Dioxetanes Bearing a Hydroxyphenanthrene or Hydroxy[4]helicene Moiety and Their Base-Induced Chemiluminescent Decomposition

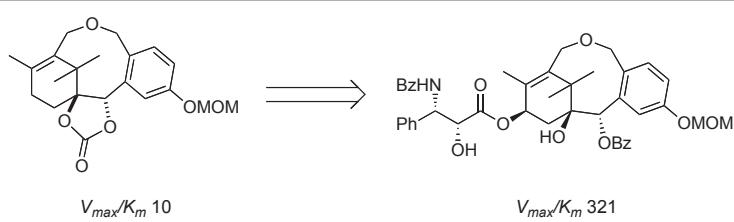
Yohei Koyama, Nobuko Watanabe, Hisako K. Ijuin, and Masakatsu Matsumoto*



Dioxetane Chemiluminescence Helicene

482 Synthesis and Biological Evaluation of C-Aromataxane Derivatives as P-Glycoprotein-Mediated Multi Drug Resistance Reversal Agents

Takayuki Doi,* Naoko Yamaguchi, Kosuke Ohsawa, Kazuoki Nakai, Masahito Yoshida, Kazuhiro Satake, Yuji Mitani, Hiroshi Nakagawa, Takashi Takahashi, and Toshihisa Ishikawa



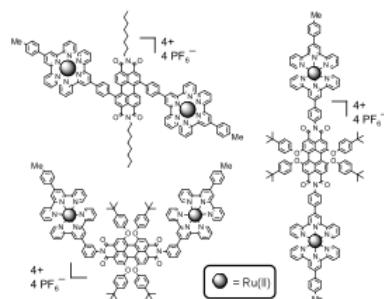
98% recovery of cytotoxicity of paclitaxel in the presence of 5.0 μ M **5a** (IC_{50} 30 nM against MDR KB-G2 cells)

9-Membered Cyclic Ether MDR Reversal Agent Paclitaxel P-Glycoprotein Cytotoxicity

502 Perylene-Based, *Bis*(terpyridine)-Ru(II) Complexes: Synthesis, Electrochemical and Photovoltaic Properties

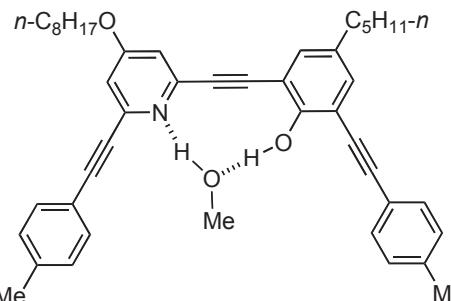
Hany El-Batal, Juan Manríquez Rocha, Perla F. Méndez, Luis A. Godínez, Kai Guo, Xiaopeng Li, Xiaocun Lu, Chrys Wesdemiotis, Charles N. Moorefield, and George R. Newkome*

Terpyridine Perylene *Bis*(terpyridine) Complex Ruthenium



515 A New Class of Structurally Simple and Highly Emissive Fluorophores with a Pyridine–Acetylene–Phenol Conjugate

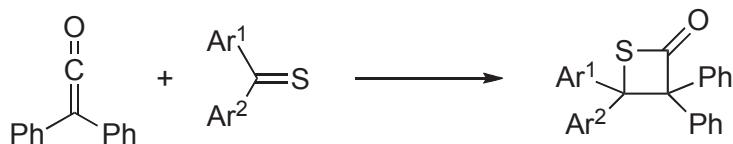
Yuki Ohishi, Hajime Abe,* and Masahiko Inouye*



Pyridine Phenol Hydrogen Bonding Optical Property Responsiveness to Acid and Base

529 Thermal [2+2]-Cycloadditions of Diphenylketene with Aryl- and Hetaryl-Substituted Thioketones

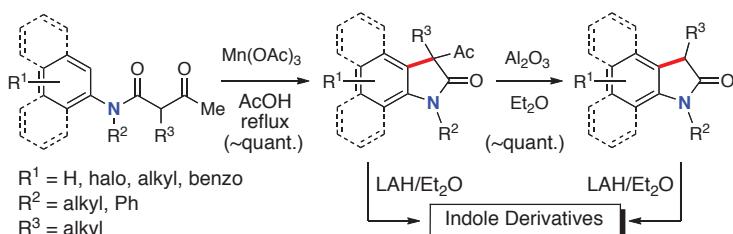
Grzegorz Młostorń,* Katarzyna Urbaniak, Anna Szychowska, Anthony Linden, and Heinz Heimgartner*



Cycloaddition Reaction Thioketone Diphenylketene Thietan-2-one X-Ray Crystallography

540 Mn(III)-Based Oxidative Cyclization of *N*-Aryl-3-oxobutanamides. Facile Synthesis and Transformation of Substituted Oxindoles

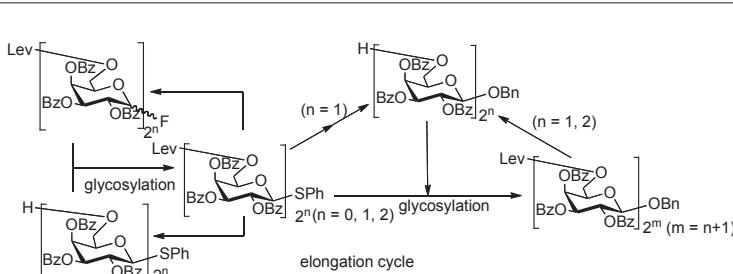
Nobutaka Kikue, Tetsuya Takahashi, and Hiroshi Nishino*



N-Aryl-3-oxobutanamide 3-Acetylindolin-2-one 1-H-Indole Oxidation Cyclization

563 Synthesis of Model Compounds Related to Linear β -D-(1 \rightarrow 6)-Galactosyl Side-Chains of Polysaccharides from *Astragalus mongholicus* Bunge

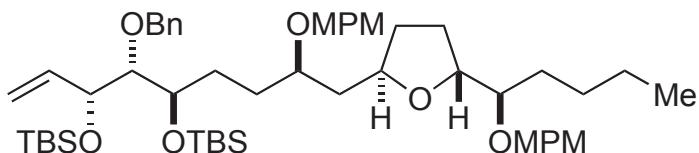
Noriyasu Hada, Ryo Shimura, Kyoko Hakamata, Hiroaki Kiyohara, Haruki Yamada, Tadahiro Takeda, and Fumiaki Kiuchi*



β (1 \rightarrow 6)-Oligogalactan *Astragalus mongholicus*

**579 Studies toward the Total Synthesis of Amphidinolide N:
Stereocontrolled Synthesis of the C13–C29 Segment**

Makoto Sasaki,* Yuki Kawashima, and Haruhiko Fuwa

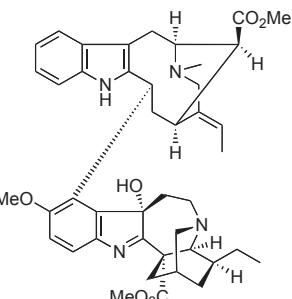


Amphidinolide N Marine Dinoflagellate Macrolide Total Synthesis

■ SHORT PAPERS

601 A New Indole Alkaloid from *Voacanga grandifolia*

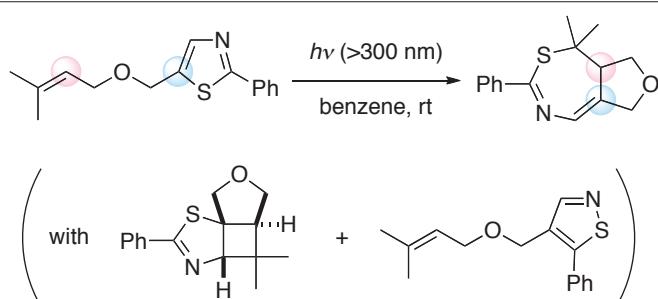
Azusa Haseo, Alfarius Eko Nugroho, Yusuke Hirasawa, Toshio Kaneda, Osamu Shirota, Abdul Rahman, Idha Kusumawati, Noor Cholies Zaini, and Hiroshi Morita*



Bisindole Alkaloid Voacalagine F *Voacanga grandifolia* Structure Elucidation

607 Novel Intramolecular Cyclization-Skeletal Reorganization of 2-Arylthiazoles under Photoirradiation

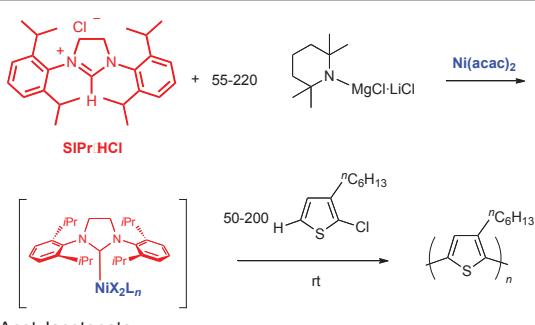
Noriyoshi Arai,* Moe Mizota, and Takeshi Ohkuma*



Photoreaction Cyclization Skeletal Reorganization Thiazole Thiazepine

617 Studies on the Effect of N -Heterocyclic Carbene as a Ligand for Nickel(II)-Catalyzed Polymerization of Thiophenes

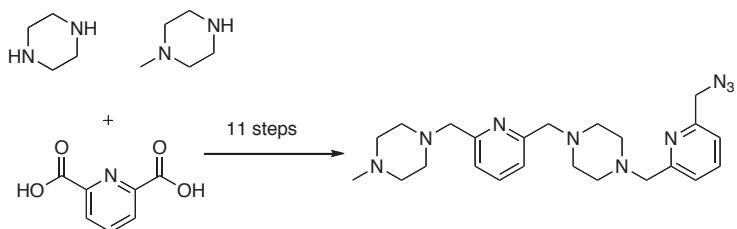
Atsunori Mori,* Makoto Fujio, and Shunsuke Tamba



N -Heterocyclic Carbene Nickel Catalyst Thiophene Polythiophene Acetylacetone

625 Synthesis of Unprotected CH₂-Skipped Piperazine-**Pyridine Alternating Cycles with Azide End-group**

Andi Kipper, Indrek Kalvet, Kaido Tämm, Lauri Sikk,
Peeter Burk, Kuldar Kõiv, and Uno Mäeorg*

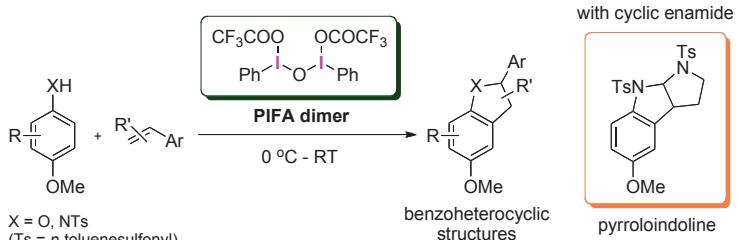
CH₂-Skipped Heterocycle

Pyridine-Piperidine Alternating Cycle

Organic Synthesis

631 Phenol and Aniline Oxidative Coupling with Alkenes by Using Hypervalent Iodine Dimer for the Rapid Access to Dihydrobenzofurans and Indolines

Toshifumi Dohi, Yosuke Toyoda, Tomofumi Nakae,
Daichi Koseki, Hiroko Kubo, Tohru Kamitanaka, and
Yasuyuki Kita*



Hypervalent Iodine

C-C Coupling Reaction

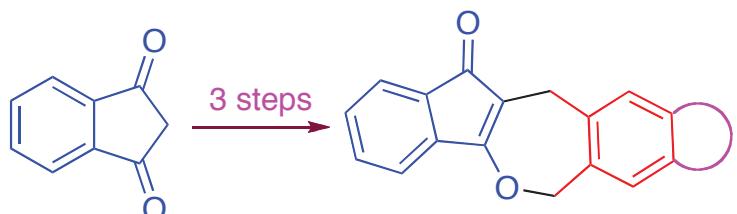
Oxidation

Cyclization

Benzoheterocycle

**645 Diversity Oriented Approach to Oxepine Derivatives:
Further Expansion *via* Diels–Alder Reaction**

Sambasivarao Kotha* and Rashid Ali



Oxepine

Diels–Alder Reaction

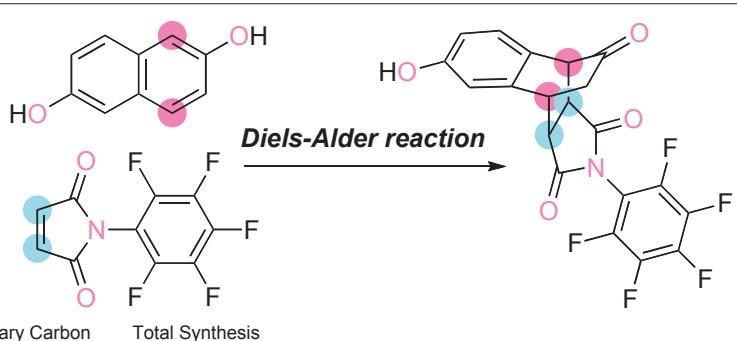
Rongalite

Sultine

Alkylation Reaction

659 N-(2,3,4,5,6-Pentafluorophenyl)maleimide as a Powerful Dienophile in Dearomatizing Diels–Alder Reactions

Koichi Hagiwara, Masafumi Iwatsu, Daisuke Urabe,
and Masayuki Inoue*



Diels–Alder Reaction

Dearomatization

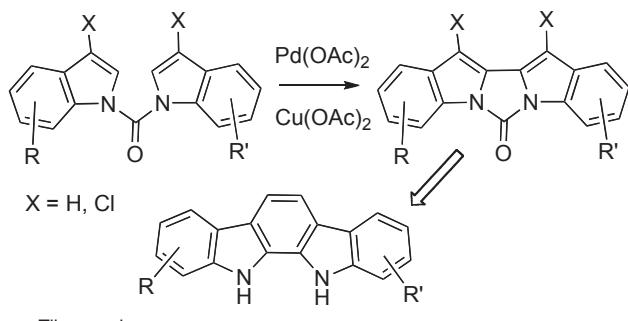
Carbocycle

Quaternary Carbon

Total Synthesis

673 Pd-Catalyzed Intramolecular Oxidative Coupling Reaction of 1,1'-Carbonyldiindoles

Takumi Abe and Minoru Ishikura*



2,2'-Biindolyl

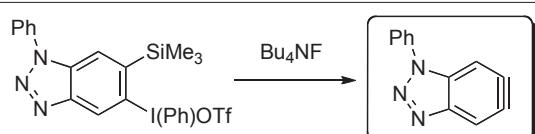
Pd-Catalyzed Coupling Reaction

Indolo[2,3-*b*]carbazole

Tjipanazole

681 Generation and Reactions of Heteroaromatic Arynes Using Hypervalent Iodine Compounds

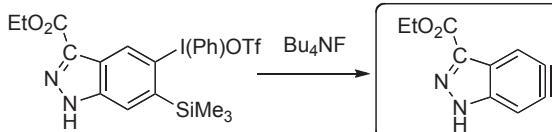
Keisuke Gondo, Jozo Oyamada, and Tsugio Kitamura*



Heteroaromatic Aryne

Hypervalent Iodine

Didehydrobenzotriazole

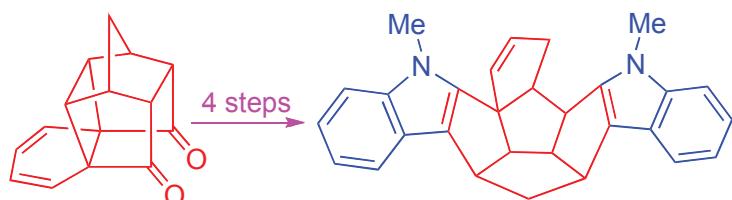


Didehydroindazole

Polycyclic Heteroaromatic Compound

690 Design of Aza-Polyquinanes *via* Fischer Indole Cyclization under Green Conditions

Sambasivarao Kotha* and Ajay Kumar Chinnam



Fischer Indole Synthesis

Polyquinane

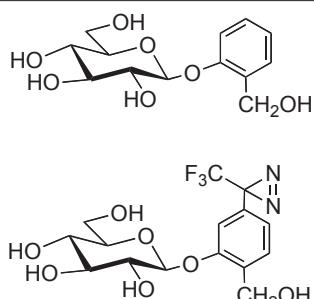
Low Melting Mixture

Caged Compound

Diels-Alder Reaction

698 Synthesis of Photoreactive Diazirinyl Salicin Derivative to Elucidate Functional Analysis of the Bitter Taste Receptor

Munenori Sakurai, Takuma Yoshida, Lei Wang, Yuta Murai, Katsuyoshi Masuda, Yasuko Sakihamama, Yasuyuki Hashidoko, Yasumaru Hatanaka, and Makoto Hashimoto*



Diazirine

Photoaffinity Label

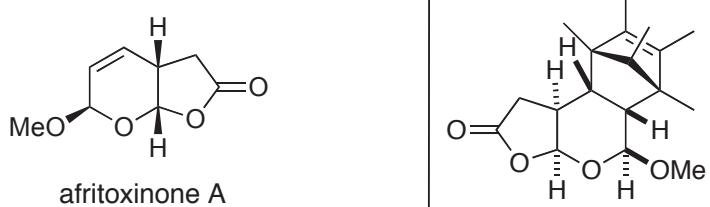
Salicin

Bitter Taste

Glucosidation

706 Synthetic Study of Afritoxinone A: Stereoselective Construction of Furopyanone Moiety

Hideki Abe, Toshihiro Yoshie, Takumi Wagatsuma, Toyoharu Kobayashi, and Hisanaka Ito*



Afritoxinone A

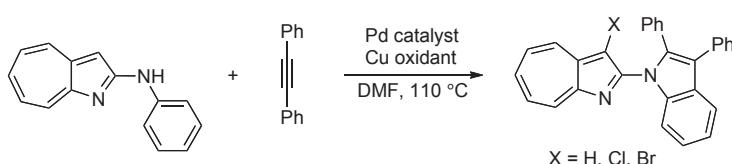
Dihydrofuropyanone

Stereoselective 1,4-Addition Reaction

Acetalization

715 The Cycloaddition of 2-Phenylamino-1-azaazulene with Diphenylacetylene Using Palladium Catalytic Systems

Hiroyuki Fujii,* Shigeki Oka, Ippei Nakamura, Yu Kawai, Reiko Ikeda, Takeo Konakahara, and Noritaka Abe



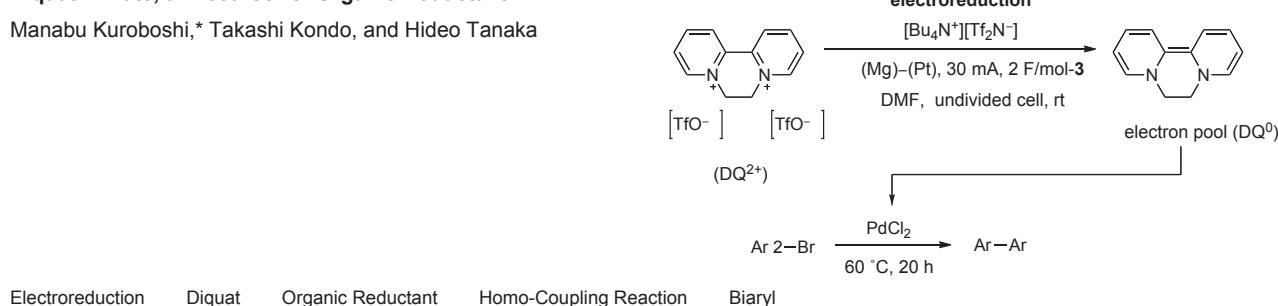
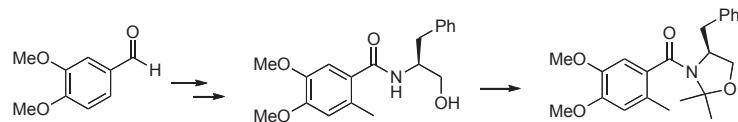
1-Azaazulene

Cycloaddition Reaction

Indole

723 Diquat Triflate, a Precursor of Organic Reductant

Manabu Kuroboshi,* Takashi Kondo, and Hideo Tanaka

**730 Synthesis and Crystal Structure of (4*S*)-4-Benzyl-3-(4,5-dimethoxy-2-methylbenzoyl)-2,2-dimethyl-1,3-oxazolidine**Maria Chrzanowska,* Zofia Meissner,
Joanna M. Chrzanowska, and Andrzej K. Gzella

Chiral Auxiliary Chiral Oxazolidine 8-Oxoberbine Oxidation of Aromatic Aldehyde X-Ray Analysis

Contributors To This Issue

- 515 Abe, Hajime
 706 Abe, Hideki
 715 Abe, Noritaka
 673 Abe, Takumi
 372 Adachi, Isao
 645 Ali, Rashid
 344 An, Guanghui
 607 Arai, Noriyoshi
 252 Arai, Seiji
 173 Asai, Miki
 327 Azuma, Masayuki
 226 Bandi, Vijayalakshmi
 298 Banwell, Martin G.
 625 Burk, Peeter
 690 Chinnam, Ajay Kumar
 730 Chrzanowska, Joanna M.
 730 Chrzanowska, Maria
 316 Dékány, Miklós
 631 Dohi, Toshifumi
 482 Doi, Takayuki
 316 Dubrovay, Zsófia
 502 El-Batal, Hany
 173, 425 Fromont, Jane
 715 Fujii, Hiroyuki
 617 Fujio, Makoto
 126, 136 Fujiwara, Kenshu
 85 Fujiwara, Yuki
 238 Fujiwara, Yurie
 113 Fukazawa, Koh
 163 Fukuda, Kazuki
 252 Fukui, Norihito
 126 Fukushima, Takanori
 144 Furukawa, Noriyuki
 261 Furukawa, Shunsuke
 579 Fuwa, Haruhiko
 502 Godínez, Luis A.
 681 Gondo, Keisuke
 173, 425 Gono, Tohru
 502 Guo, Kai
 730 Gzella, Andrzej K.
 97 Hachiya, Iwao
 563 Hada, Noriyasu
 659 Hagiwara, Koichi
 563 Hakamata, Kyoko
 108 Hanzawa, Yuji
 432 Hari, Yoshiyuki
 601 Haseo, Azusa
 698 Hashidoko, Yasuyuki
 698 Hashimoto, Makoto
 121 Hashizume, Daisuke
 157 Hatae, Noriyuki
 405 Hatakeyama, Susumi
 698 Hatanaka, Yasumaru
 316 Hazai, László
 529 Heimgartner, Heinz
 601 Hirasawa, Yusuke
 150 Hirose, Yoshikatsu
 216 Hiyoshi, Hidetaka
 41 Honda, Toshio
 104 Ideta, Keiko
 186 Ido, Akiko
 432 Ijitsu, Shin
 462 Ijuin, Hisako K.
 715 Ikeda, Reiko
 659 Inoue, Masayuki
 515 Inouye, Masahiko
 126, 136 Ishigaki, Yusuke
 405 Ishihara, Jun
 144 Ishii, Akihiko
 482 Ishikawa, Toshihisa
 673 Ishikura, Minoru
 97 Ito, Akinori
 706 Ito, Hisanaka
 85 Ito, Shunji
 659 Iwatsu, Masaumi
 271 Kabalka, George W.
 157 Kaji, Aiichirou
 121 Kakusawa, Naoki
 316 Kalaus, György
 625 Kalvet, Indrek
 631 Kamitanaka, Tohru
 89 Kanai, Motomu
 601 Kaneda, Toshio
 432 Kashima, Satoshi
 163 Kashiwamura, Gaku
 372 Kato, Atsushi
 126, 136 Katoono, Ryo
 150 Katsumura, Shigeo
 126, 136 Kawai, Hidetoshi
 327 Kawai, Shintaro
 715 Kawai, Yu
 579 Kawashima, Yuki
 316 Keglevich, Péter
 540 Kikue, Nobutaka
 327 Kimura, Shinya
 163 Kimura, Tsutomu
 625 Kipper, Andi
 631 Kita, Yasuyuki
 372 Kitahara, Yoshiro
 681 Kitamura, Tsugio
 563 Kiuchi, Fumiuki
 563 Kiyohara, Hiroaki
 144 Kobayashi, Hiroki
 23, 173, 425 Kobayashi, Jun'ichi
 216 Kobayashi, Kazuhiro
 150, 706 Kobayashi, Toyoharu
 625 Kőiv, Kuldar
 327 Koizumi, Yu-ichi

- 715 Konakahara, Takeo
 723 Kondo, Takashi
 631 Koseki, Daichi
 226, 357, 645, 690 Kotha, Sambasivarao
 462 Koyama, Yohei
 631 Kubo, Hiroko
 104 Kubo, Kanji
 238 Kumai, Yoshimitsu
 121 Kurita, Jyoji
 723 Kuroboshi, Manabu
 173, 425 Kusama, Taishi
 601 Kusumawati, Idha
 6, 7 Kuwajima, Isao
 344 Li, Guigen
 502 Li, Xiaopeng
 529 Linden, Anthony
 502 Lu, Xiaocun
 625 Mæorg, Uno
 113 Maruko, Miki
 85 Maruyama, Akifumi
 85 Maruyama, Mitsuhsisa
 698 Masuda, Katsuyoshi
 405 Matsubara, Takaaki
 432 Matsuda, Yuya
 462 Matsumoto, Masakatsu
 104 Matsumoto, Taisuke
 121 Matsumura, Mio
 89 Matsunaga, Shigeki
 372 Matsuya, Yuji
 730 Meissner, Zofia
 502 Méndez, Perla F.
 357 Meshram, Miliind
 198 Midorikawa, Masanobu
 372 Minehira, Daisuke
 482 Mitani, Yuji
 372 Miura, Kyoko
 442 Miyaoka, Hiroaki
 238, 372 Miyatake, Ryuta
 372 Miyawaki, Shota
 372 Miyazaki, Akira
 607 Mizota, Moe
 186 Mizukoshi, Ryosuke
 529 Młostów, Grzegorz
 186 Monguchi, Yasunari
 502 Moorefield, Charles N.
 104 Mori, Akira
 617 Mori, Atsunori
 601 Morita, Hiroshi
 85 Morita, Noboru
 698 Murai, Yuta
 121 Muranaka, Atsuya
 271 Musolino, Bryan J.
 186 Nagase, Hisamitsu
 186 Nagatsu, Nobuharu
 631 Nakae, Tomofumi
 482 Nakagawa, Hiroshi
 482 Nakai, Kazuoki
 1, 261 Nakamura, Eiichi
 715 Nakamura, Ippei
 425 Nakamura, Kenta
 144 Nakata, Norio
 216 Nakazawa, Kazuya
 502 Newkome, George R.
 186 Niikawa, Miki
 540 Nishino, Hiroshi
 601 Nugroho, Alfarius Eko
 432 Obika, Satoshi
 238 Oda, Mitsunori
 198 Oda, Yoshiki
 515 Ohishi, Yuki
 607 Ohkuma, Takeshi
 482 Ohsawa, Kosuke
 238 Ohta, Akira
 715 Oka, Shigeki
 157 Okada, Chiaki
 252 Osuka, Atsuhiro
 442 Ota, Koichiro
 681 Oyamada, Juzo
 344 Pan, Yi
 601 Rahman, Abdul
 502 Rocha, Juan Manríquez
 108 Saito, Akio
 327 Saito, Naoki
 41 Saito, Norio
 186 Sajiki, Hironao
 425 Sakai, Kanae
 432 Sakata, Akihiro
 698 Sakihama, Yasuko
 698 Sakurai, Munenori
 316 Sánta, Zsuzsanna
 579 Sasaki, Makoto
 482 Satake, Kazuhiro
 113 Sato, Koichi
 163 Satoh, Tsuyoshi
 344 Seifert, Cole
 97 Shimizu, Makoto
 563 Shimura, Ryo
 252 Shinokubo, Hiroshi
 601 Shirota, Osamu
 85 Shoji, Taku
 625 Sikk, Lauri
 108 Sudo, Kohei
 372 Sugimoto, Kenji
 344 Sun, Hao
 173 Suto, Shohei
 144 Suzuki, Izuru
 126, 136 Suzuki, Takanori
 316 Szántay, Csaba
 316 Szántay, Jr., Csaba
 529 Szychowska, Anna
 482 Takahashi, Takashi
 540 Takahashi, Tetsuya
 173 Takahashi-Nakaguchi, Azusa
 432 Takamine, Ryutaro
 372 Takeda, Daisuke
 563 Takeda, Tadahiro
 113 Takenaga, Naoko
 617 Tamba, Shunsuke
 625 Tämm, Kaido
 723 Tanaka, Hideo
 23, 173, 425 Tanaka, Naonobu
 3 Tanino, Keiji
 41 Tatsuzawa, Fumi
 631 Toyoda, Yosuke
 372 Toyooka, Naoki
 157 Toyota, Eiko
 150 Tsuchikawa, Hiroshi
 261 Tsuji, Hayato
 121 Uchiyama, Masanobu
 327 Umehara, Yoshifumi
 372 Umezaki, Masahito
 216 Umezu, Kazuto
 659 Urabe, Daisuke
 529 Urbaniak, Katarzyna
 706 Wagatsuma, Takumi
 698 Wang, Lei
 462 Watanabe, Nobuko
 502 Wesdemiotis, Chrys
 298 White, Lorenzo V.
 298 Willis, Anthony C.
 563 Yamada, Haruki
 482 Yamaguchi, Naoko
 89 Yamamoto, Shohei
 198 Yamanoi, Takashi
 108 Yamashita, Naoki
 121 Yasuike, Shuji
 85 Yasunami, Masafumi
 261 Yokoi, Yuki
 327 Yokoya, Masashi
 97 Yokoyama, Keiichi
 85 Yokoyama, Ryuji
 252 Yorimitsu, Hideki
 482 Yoshida, Masahito
 126 Yoshida, Satoshige
 698 Yoshida, Takuma
 706 Yoshie, Toshihiro
 216 Yuba, Shohei
 601 Zaini, Noor Cholies

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A NEW INDOLE ALKALOID FROM *VOACANGA GRANDIFOLIA*

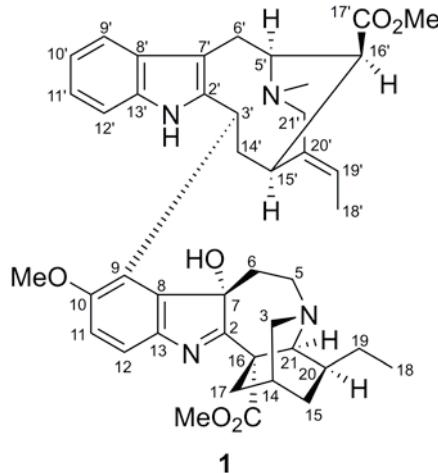
Azusa Haseo,^a Alfarius Eko Nugroho,^a Yusuke Hirasawa,^a Toshio Kaneda,^a Osamu Shirota,^b Abdul Rahman,^c Idha Kusumawati,^c Noor Cholies Zaini,^c and Hiroshi Morita^{a,*}

^aFaculty of Pharmaceutical Sciences, Hoshi University, Ebara 2-4-41 Shinagawa-ku, Tokyo 142-8501, Japan, ^bFaculty of Pharmaceutical Sciences at Kagawa Campus, Tokushima Bunri University, 1314-1 Shido, Sanuki City, Kagawa 769-2193, Japan, ^cFaculty of Pharmacy, Airlangga University, Jalan Dharmawangsa Dalam, Surabaya 60286, Indonesia

Abstract – A new bisindole alkaloid, voacalgin F (**1**), has been isolated from the bark of Indonesian *Voacanga grandifolia* (Miq.) Rolfe. Its structure was elucidated on the basis of 1D and 2D-NMR data analysis.

Voacanga is a small genus of the Apocynaceae family consisting of 12 species. Species of this genus are distributed mainly in the tropical Africa and Malaysia, and have been reported to contain vobasine, eburnane, iboga, and aspidosperma type of monoterpenoid indole alkaloids.¹ Various activities have been reported for monoterpenoid indole alkaloids, such as cytotoxicity,² anti-melanogenesis,³ anti-plasmodial,⁴ and vasorelaxant activities.⁵ In the search for new bioactive compounds from tropical plants,^{3,5-8} alkaloid constituents of *V. grandifolia* bark were investigated and a new bisindole alkaloid voacalgin F (**1**) was isolated together with voacamidine,⁹⁻¹² voacangine,⁹ voacanginehydroxyindolenine,¹³ and pagicerine.¹⁴ The isolation and structure elucidation of **1** are reported herein.

[†]Dedicated to the celebration of the 77th birthday of Prof. Dr. Isao Kuwajima, Professor emeritus of Tokyo Institute of Technology



Voacalgin F (**1**) was obtained as yellow amorphous solid and the molecular formula was determined as $C_{43}H_{52}N_4O_6$ from the HRESIMS data (m/z 721.3978 [$M+H]^+$, calcd for $C_{43}H_{53}N_4O_6$, 721.3965). The IR absorptions (3430 and 1720 cm^{-1}) implied the presence of hydroxyl and carbonyl functionalities. Analysis of the ^{13}C -NMR data (Table 1) showed that the chemical shift of 21 carbon signals is highly similar to the vobasine unit of voacamidine, suggesting the presence of a vobasine unit in **1**. The chemical shift of the other carbon signals is highly similar to that of voacangine hydroxyindolenine, except for downfield shift of C-9. These data suggested the structure of **1** as a new vobasine-iboga type of bisindole alkaloids as shown in Figure 1.

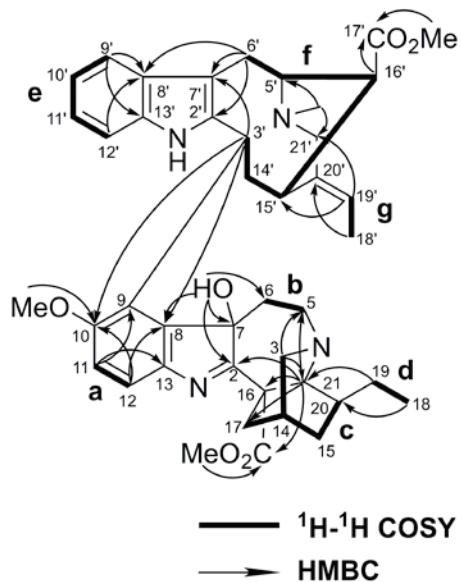


Figure 1. Selected 2D-NMR Correlations of **1**

The planar structure of **1** was further confirmed by 2D NMR analysis (^1H - ^1H COSY, HSQC and HMBC, Figure 1). Analysis of ^1H - ^1H COSY and HSQC data revealed the presence of 7 partial structure (**a-g**).

HMBC correlations of 7-OH to C-2, C-6, C-7 and C-8, H-11 to C-9 and C-13, and H-12 to C-8 and C-10 confirmed the presence of a 7-hydroxyindolenine moiety and the connection of C-6 and C-7. HMBC cross-peaks of H₃-18 to C-20 suggested the connectivity of C-19 and C-20, and the HMBC correlations of H₂-3 to C-5 and C-21, H₂-19 to C-21, H-21 to C-2, C-5, C-16, C-17 and a carbonyl (δ_C 174.4), and a methyl (δ_H 3.73) to δ_C 174.4 completed the structure of the iboga unit. HMBC cross-peaks of H-3' to C-7', H₂-6' to C-2', C-7', and C-8', H-9' to C-8' and C-13', and H-12' to C-8' suggested the presence of an indole unit and the connectivity of partial structure **f** to the indole unit. HMBC cross-peaks of H₃-18' to C-20', and H-19 to C15' and C-21' revealed the connectivity of partial structures **f**, **g**, and C-21' through C-20'. HMBC correlations of a methyl (δ_H 2.61) to C-5' and C-21' established the connections between C-5' and C-21' through a nitrogen atom, and HMBC cross-peaks of H-16' to C-17', and another methyl (δ_H 2.49) to C-17' suggested the presence of a methoxycarbonyl moiety at C-16'. Finally, the two units were confirmed to be connected by C-9 to C-3' bond by the HMBC correlations of H-3' to C-8 and C-10.

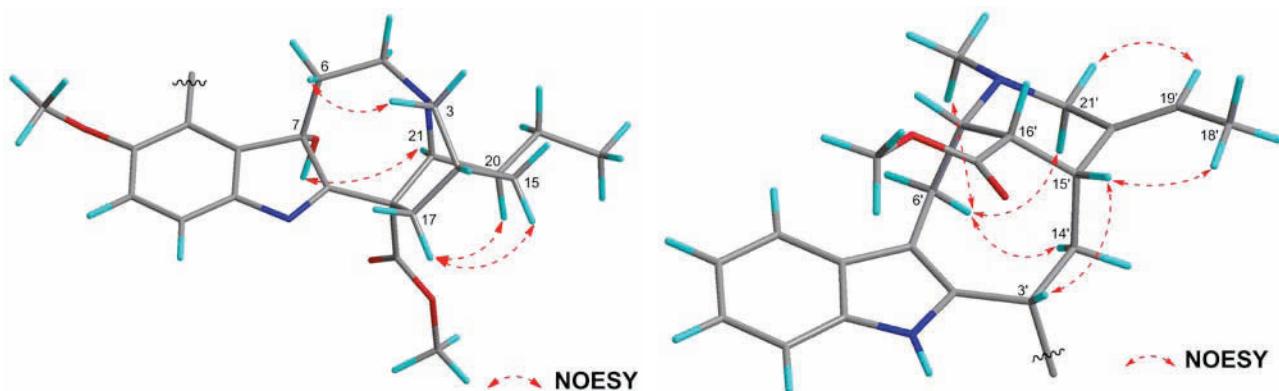


Figure 2. Selected NOESY Correlations of Each Indole Unit in **1**

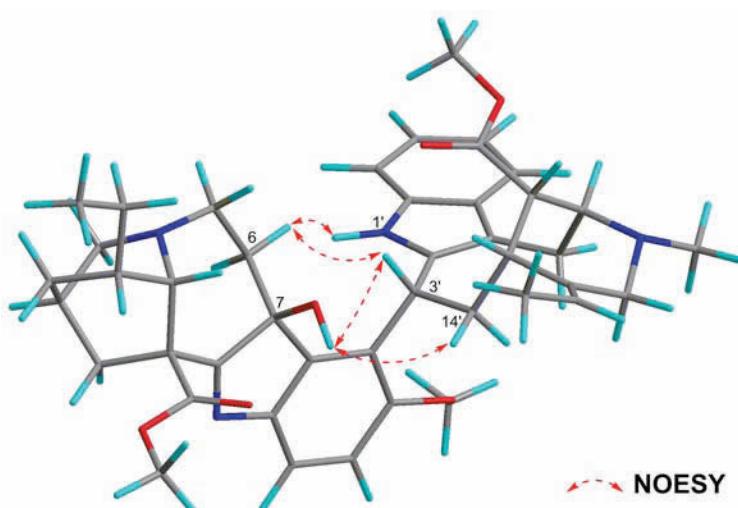


Figure 3. Selected NOESY Correlations Between Two Indole Unit of **1**

The relative configuration of **1** was assigned using the ^1H - ^1H coupling constant values, ^1H NMR chemical shift and NOESY correlations. The orientation of 7-OH and H-21 was assigned as α from the NOESY correlation 7-OH/H-21. The relative configuration C-14, C-16, C-20, C-21, C-3' and C-5' was assigned to be the same as in voacamine based on the NOESY correlations shown in Figure 2. The orientation of the methoxycarbonyl at C-16' was deduced from the highly shielded ^1H NMR chemical shift of the methoxy group (δ_{H} 2.49), and the configuration of the C-19'-C-20' was determined to be *E* from the NOESY correlation of H-19'/H₂-21'. Finally the relative configuration of the total molecule was deduced from the NOESY correlations of H-6a/NH and H-3', 7-OH/H-3' and H-14'a (Figure 3).

Table 1. ^1H (700 MHz) & ^{13}C (175 MHz) NMR Data of **1** in CDCl_3

	δ_{H} (J, Hz)	δ_{C}	δ_{H} (J, Hz)	δ_{C}
2		187.0	2'	136.7
3	2.85 (2H, m)	48.7	3'	5.28 (1H, d, 11.9)
5a	3.15 (1H, m)	49.2	5'	4.06 (1H, m)
5b	3.79 (1H, m)		6'a	3.20 (1H, m)
6a	2.24 (1H, td, 13.2, 3.6)	33.9	6'b	3.50 (1H, m)
6b	2.52 (1H, br. d, 13.2)		7'	110.0
7		89.9	8'	130.1
8		141.2	9'	7.53 (1H, d, 7.4)
9		130.1	10'	7.06 (1H, t, 7.4)
10		158.0	11'	7.05 (1H, t, 7.4)
11	6.68 (1H, d, 8.3)	112.6	12'	7.09 (1H, d, 7.4)
12	7.27 (1H, d, 8.3)	119.8	13'	135.3
13		145.3	14'a	1.83 (1H, m)
14	1.98 (1H, br. s)	27.0	14'b	3.13 (1H, td, 13.6, 3.8)
15a	1.15 (1H, d, 12.0)	32.1	15'	3.80 (1H, m)
15b	1.80 (1H, d, 12.0)		16'	2.75 (1H, br. s)
16		59.1	17'	171.6
17a	2.64 (1H, m)	34.0	18'	1.64 (3H, d, 6.8)
17b	2.70 (1H, d, 13.9)		19'	5.32 (1H, q, 6.8)
18	0.89 (3H, t, 7.0)	11.6	20'	138.1
19	1.45 (2H, m)	26.4	21'a	2.92 (1H, m)
20	1.41 (1H, m)	37.7	21'b	3.78 (1H, m)
21	3.87 (1H, br. s)	58.8	17'-OMe	2.49 (s)
CO_2Me		174.4	N-Me	2.61 (br. s)
CO_2Me	3.73 (s)	53.3	NH	7.15 (s)
10-OMe	3.39 (s)	56.5		
7-OH	4.15 (s)			

EXPERIMENTAL

General Experimental Procedures. Optical rotations were measured on a JASCO DIP-1000 automatic digital polarimeter. UV spectra were obtained on an Ultrospec 2100 pro spectrophotometer and IR

spectra were recorded on a JASCO FT/IR-4100 spectrophotometer. High-resolution ESI MS were obtained on a LTQ Orbitrap XL (Thermo Scientific). ¹H and 2D NMR spectra were recorded on a Bruker AV700 spectrometer and chemical shifts were referenced to the residual solvent peaks (δ_{H} 7.26 and δ_{C} 77.0 for chloroform-*d*). Standard pulse sequences were employed for the 2D NMR experiments.

Plant Material. The barks of *V. grandifolia* were collected at Purwodadi Botanical Garden, Indonesia in 2008. The botanical identification was made by Ms. Sri Wuryanti, Purwodadi Botanical Garden. A voucher specimen has been deposited in the herbarium at Purwodadi Botanical Garden, Pasuruan, Indonesia.

Extraction and Isolation. The dried and powdered bark of *V. grandifolia* (300 g) was extracted successively with MeOH. Part of the extract (17.0 g of 28.4 g) was dissolved in 3% aqueous tartaric acid (pH 2) and then partitioned with EtOAc. The aqueous layer was treated with saturated Na₂CO₃ (aq.) to pH 9 and was partitioned successively by CHCl₃ and *n*-BuOH. Part of the CHCl₃ soluble materials (5.0 g of 5.10 g) was subjected to an LH-20 column (CHCl₃/MeOH 1:1) to obtain 12 fractions.

Fraction 7 was fractionated by amino silica gel column chromatography (*n*-hexane/EtOAc, 1:0~1:1, CHCl₃/MeOH, 0:1~1:0) to obtain voacamine (100.8 mg, 0.032%). In addition, fraction eluted by CHCl₃/MeOH (80:1) was further separated by ODS HPLC (Inertsil ODS-3, 5 μm , 10 x 250 mm; 35% MeCN in 0.1% aqueous HCO₂H; flow rate 2 mL/min; UV detection at 254 nm) to obtain **1** (*t*_r 30 min., 2.7 mg, 0.001%).

Fraction 11 was separated by repeated amino silica gel column chromatography (*n*-hexane/EtOAc, 1:0~1:1, CHCl₃/MeOH, 0:1~1:0) and silica gel column chromatography (CHCl₃/MeOH, 0:1~1:0) to give voacangine (23.8 mg, 0.0043%), voacanginehydroxyindolenine (12.8 mg, 0.0079%), and pagicerine (3.6 mg, 0.0012%).

Voacalagine F (1): yellow amorphous solid; $[\alpha]_D^{22}$ -132 (*c* 1.0, MeOH); IR (KBr) ν_{max} 3430, 2940 and 1720 cm⁻¹; UV (MeOH) λ_{max} (ϵ) 225 (23400) and 290 (9000) nm; ¹H and ¹³C NMR data (Table 1); ESIMS *m/z* 721 (M+H)⁺; HRESIMS *m/z* 721.3978 (M+H⁺; calcd for C₄₃H₅₃N₄O₆, 721.3965).

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