



Dental Journal: Majalah Kedokteran Gigi

Accredited number: 158/E/KPT/2021

Certificate of Reviewing

Award presented to

Nanik Zubaidah

in acknowledgment of review contributed to journal under the title of manuscript:

Visualization of bubbles generation of electrical driven EndoActivator tips during solutions activation in a root canal model and a modified extracted tooth: a pilot study

Faculty of Dental Medicine Universitas Airlangga Mayjen Prof. Dr. Moestopo 47 60132 Surabaya INDONESIA T +62 31 5039478 F +62 31 5039478 E dental_journal@fkg.unair.ac.id W https://e-journal.unair.ac.id/MKG W https://fkg.unair.ac.id/en

The Editors of Dental Journal: Majalah Kedokteran Gigi E-ISSN: 2442-9740; P-ISSN: 1978-3728

ASSESSMENT FORM OF ORIGINAL ARTICLE DENTAL JOURNAL (MAJALAH KEDOKTERAN GIGI)

Manuscript title: Visualization of bubbles generation of electrical driven EndoActivator tips during solutions activation in a root canal model and a modified extracted tooth: a pilot study

Receive date:

Send date: Please comments written in English

REVIEW	YES*	NO*
1. Has this text ever been published on other media?		V
<u>Comment</u> :		
2. Is the title appropriate, concise, clear, and describes the contribution of scientific	V	V
development? (maximum 10 words, covering the variables studied)		
<u>Comment</u> :		
The title is more than 10 words, but it is appropriate.		
· · · · · · · · · · · · · · · · · · ·		
3. Research report:		I
a) Introduction includes the background clearly?	V	
<u>Comment</u> :		
b) The purpose clearly?	V	
<u>Comment</u> :		

	REVIEW	YES*	NO*
c)	Methods and research design in accordance with the purpose of the study?	V	
	<u>Comment</u> :		
	This is a descriptive basic experiment, no samples and no statistical analysis,		
	but this kind of experiment could also be found in other reputable journals		
	such as: J Endod and Lasers Med Sci.		
d)	The research procedure is described precisely and in detail, thus ensuring		V
	internal/external validity?		
	Comment:		
	A schematic figure on how the process of observation along with photos of		
	the utilities used during the experiment should be added.		
	These figures can be combined for optimalization, Figure 1 and 2 can be		
	merged.		
e)	The results can answer the research question?	V	
	Comment:		
	Aside from the supplemented videos (YouTube), a detailed photos of the		
	observation regarding the bubbles must be added to better understand the		
	bubble formation, it can be taken from the steps and please provide		
	annotations and arrows in the photos to clarify the proposed results, including		
	the photos taken both from the glass/acrylic model and extracted tooth for		
	better viewing.		
f)	- The discussion does not repeat the results?	V	
	- Aligned with the scope of the study and compared with similar research		
	results?		
	- Explain the meaning of research results in answering the problem?		
	<u>Comment</u> :		
g)	References are aligned with the research material and use the literature of the		V
	last 10 years?		
	Comment:		
	Most of the references are above 10 years (except reference number 11, 14,		
	17 no page provided, and 19), please replace with current references (less		
	than 10 years).		

	REVIEW	YES*	NO*	
h)	- The conclusion matches the title and the problem?	V		
	- The research results contibute to the development of dentistry?			
	- Perform synthesis base on similar research results that precede?			
	<u>Comment</u> :			
i)	References needs to be added/substracted**?			
	<u>Comment</u> :			
	Yes, please replace the references above 10 years with the current ones (less	than 10 years	6).	
4. Is	there a section that needs to be added/summarized**?			
<u>Co</u>	omment:			
	Yes, please add detailed pictures or photograph of the procedures on this research	arch.		
	In the discussion, regarding the preparation of natural tooth with balanced force	e technique using		
	Protaper hand file, what is your reasoning of using this technique and file? Fror	n the photo o	f the	
	natural tooth, the root canal anatomy is somewhat not straight, why do you cho	-		
	Please add your reasons in the discussion.		-	

Recommendation for Editors

- [......] 1. The script can be published without changes.
- [... V...] 2. The manuscript can be published with corrections according to the direction of the Reviewer (suggestions for improvement please write directly to the script). <u>Comment</u>: This is an interesting study on EndoActivator. There are some corrections and additions to be made for improvement as listed at the end of the manuscript. Please conduct plagiarism check (Turnitin / iThenticate).
 [......] 3. The manuscript could not be published. <u>Reason</u>:

Date: 13 January 2022 Reviewer, NZ

Visualization of bubbles generation of electrical driven EndoActivator tips during solutions activation in a root canal model and a modified extracted tooth: a pilot study

ABSTRACT

Background: EndoActivator, a sonically-driven canal irrigation system (Dentsply Tulsa Dental Specialties, Tulsa, OK), has been developed for activating root canal irrigants, and has recently been released onto the market.

Purpose: To obtain an initial understanding of bubbles generation of electrical EndoActivator tips during activation of the irrigant in a transparent root canal model and a modified extracted tooth.

Methods: A modified extracted tooth and a straight glass model were filled with a solution containing 17% EDTA or 3% NaOCl. A medium activator tip 22 mm polymer noncutting #25, 0.04 file driven by an electrical sonic hand-piece at 190 Hz (highest level) induced pressure waves that produced macro- and micro-bubbles. The physical mechanisms involved were visualized using a Miro 320S high-speed imaging system (Phantom, Wayne, NJ, USA) with high temporal and spatial resolutions. The imaging system acquired images at 25,000 frames per second with 320Å~240 pixels per image, and attached a 60 mm f/2.8 macro lens (Nikon, Tokyo, Japan).

Results: The end of the tip did not generate bubbles formation. Disruption of surface tension at the air–solution system in the glass canal model by an electrical sonic driven EndoActivator tip generated bubbles in the solution. However, it did not occur at the system of solution–air interfaces in the glass canal and modified extracted tooth.

Conclusion: The physical mechanism of the solution activated by an electrical sonic driven EndoActivator tip in generating bubbles formation is because the surface tension at the air–solution system disruption. No bubbles formation occurred in the solution in the restricted space either is the solution-air system or modified extracted tooth.

Clinical Relevance: Better understanding of the physical mechanisms that relate specifically to the activation behavior of EndoActivator tips in solutions is key to improving the cleaning mechanism that applies during root canal treatment.

Keywords: bubbles formation; EndoActivator; endodontics; sonic

Introduction

Non-automated agitation of the irrigant can be produced using a file¹ or a gutta-percha cone² inserted in the shaped and cleaned canal subsequently moved periodically and rapidly by the operator. Electrical ultrasonic and sonic devices are examples of automated agitation.³ These devices use a fine non-cutting polymer or metal tip that is vibrated within the canal space, with different frequencies according to manufacturer's instruction. Sonically activated instruments are driven with a frequency range of 1,000-6,000 Hz and will generate a single node near the point at which the file is attached, as well as an antinode at the tip of the file.⁴⁻⁶ In ultrasonic solutions agitation, a tip oscillates at frequencies of 25 to 30 kHz in a pattern of motion consisting of nodes and anti-nodes along its length.⁷ EndoActivator, a sonically-driven canal irrigation system (Dentsply Tulsa Dental Specialties, Tulsa, OK), has been developed for activating root canal irrigants, and has recently been released onto the market. It comprises a portable hand-piece with three modes (low, medium, and high) and 3 types of disposable flexible non-cutting polymer tips of different sizes. Its design allows for the safe activation of various irrigating solutions and could produce vigorous fluid dynamic in the canal space.⁸ Electrical devices driven sonically has been found to be effective for cleaning root canals in many studies.^{3,9,10} However, for the last decade there has been some conflicting results in the literature on the efficacy of the system and its superiority over other systems. Some researchers have found that sonically activated irrigation is less effective than ultrasonically activated irrigation, while others have suggested that the two methods produce comparable results.^{11,12} To our best knowledge, there have been no previous studies of the physical mechanism through which the electrical sonic driven EndoActivator tip generates bubbles during activation of irrigation solution. In the present study, the mechanism was investigated by acquiring real-time data using a modified extracted tooth and a transparent glass model of root canals in order to visualize the oscillation amplitude of an EndoActivator tip in the solution. Hence, more knowledge of mechanism underlying the activation behavior of the EndoActivator tips in the solutions is crucial to improve the outcomes of the root canal treatment.

MATERIALS AND METHODS

Preparation of a modified extracted tooth

A single-rooted human mandibular first premolar with straight canal and mature apex, and a tooth length of 25 mm was selected. The tooth was not decoronated, and the intact crown (10 mm) acted as a reservoir for the irrigation solution. The preparation of the root canal was completed to the working length (WL) using a Pro Taper hand file (Dentsply Maillefer) up to a size of 25/0.06 as the master apical file (MAF) with the balanced force technique. The distal aspect of the root of the extracted tooth was carefully cut (from cervical to apex) using a fissure diamond bur (Dentsply, Tulsa, OK, USA) until the canal space was exposed (Figure 1a and b), then the surface was ground successively with 240-, P400, and 600-grit sandpaper to give a smooth surface. A glass cover was then carefully glued to the surface without entering the canal, and this was covered with composite (Figure 1c and d). The apex was then sealed with composite and the outer surface of the tooth was covered with nail varnish, ensuring that there were no visible cracks, to allow conditions within the root canal in vivo to be simulated. The tooth was filled with the solution until it reached 5 mm below of the cervical level. The tip of the sonic instrument was inserted 2 mm shorter than the WL. A single modified extracted tooth was used for uniformity in the width and size of the root canal.

Figure 1. (A) A root sample with a longitudinal cut in the distal aspect of the root canal. (B) Exposed root canal space. (C) Exposed tooth with a glass cover (0.15 mm thick). (D) The glass cover glued to the tooth and shaped to fit the shape of the root.

Root Canal Model.

The conditions within a straight root canal were simulated and visualized using a glass model with artificial canal and pulp cavity (which acted as a reservoir). The model was a glass root canal model (Kimia Farma, Bandung, West Java, Indonesia), with canal inner diameter 0.4 mm at the apex, crown height 8 mm, crown diameter 6 mm, a taper of 0.06 and an overall length of 25 mm. The canal was filled with a solution containing 17% EDTA or 3% NaOC1. The apex of the model was sealed with composite to allow the conditions within a root canal to be simulated. Two approaches were used, one

using the air–solution interface (in the coronal portion) and the other using the solution– air interface (in the apical portion). The solution–air interface system represents air entrapment in the apical region (Figure 2). The models were filled with the solution until the level reached 5 mm below the coronal end (air-solution interface system), while in the solution-air interface system, the model was filled with solution starting 5 mm from the apex (air entrapment model). The tip of the sonic instrument was inserted into the solution at the air– solution or solution–air interface (10 mm and 4 mm from the interface, respectively). A single transparent glass model was used for uniformity in the width and size of the root canal.

Figure 2. Illustrations of the solution-air and air-solution systems

Sonic Parameters and Procedure.

A 22 mm polymer noncutting # 25, 0.04 (Dentsply) was mounted on an EndoActivator hand-piece (Dentsply Tulsa Dental Specialties, Tulsa, OK) set at high mode (190 Hz) and used to activate the irrigant. The models were subjected to active sonic irrigation. The tip of the sonic instrument was inserted into the solution at the air-solution (10 mm) or solution-air interface (4 mm), while the tip was inserted 2 mm shorter than the working length in the modified extracted tooth and activated passively without any filing motion. The hand-piece was fixed in a holder to ensure that the desired position was maintained. All the experiments with the different samples were performed by the same operator. (Figures to describe the procedure is needed)

Basic Visualization.

The process was recorded using a Phantom Miro 320S high-speed digital imaging system (Wayne, NJ, USA) at 25,000 frames per second with 320Å~240 pixels per image using a macro lens (60 mm, f/2.8; Nikon, Tokyo, Japan). The root canal model was illuminated using a Fibre-Lite LMI-6000 LED continuous light source (Dolan-Jenner Industries, Boxborough, MA, USA). Measurements were repeated three times for each sample.

RESULTS

Regardless of whether 17% EDTA or 3% NaOCl was used, the high-speed images and

video clearly showed that fluid streaming. The electrical sonic activated EndoActivator tip generated hydrodynamic and finally bubbles formation occurred during the sonic activation process in the solution-air system instead of in the restricted areas (the solution-air system and modified extracted tooth).

Activation in the glass model

Air-solution interface system

During the first 69 μ s, the sonic energy was transferred to the tip and the tip vibrated at a frequency of around 190 Hz, causing the solution surface (the air-solution interface) to move as a wave. After 75 μ s, macro and micro bubbles formed in the solution. Subsequently, accumulation of the energy during sonic activation at interface caused the micro and macro bubbles in the solution. Finally, the bubbles formation occurred only the vicinity of the tip (Figure 3A). (Supplemental Video S1 https://youtu.be/ut6VKWDGmj8). The life cycle of a bubble observed ranges from 250,000–700,000 μ s with predominantly 700 μ m in diameter. The end of the tip did not generate bubbles formation.

Solution-air interface system

A model with air trapped in the apical region was used, meaning there was a solution column and an air column. The energy of the tip vibration was not sufficiently enough to break the surface tension at the solution-air interface (Figure 3B). (Supplemental Video S2 https://youtu.be/rqzM0WmJUOY).

Activation in the modified extracted tooth (air-solution interface system)

No bubbles formation occurred during the activation process (Figure 3C). (Supplemental video S3 https://youtu.be/-BHdVIO3DkA).

Figure 3. Representative images showing the disruption of surface tension at the air solution (A) and solution-air systems (B) in glass models, (C) modified extracted human root canal, (D) Oscillation amplitude of EA tip in the air.

DISCUSSION

To be clinically relevant, in vitro studies should reproduce the clinical situation as much

as possible. Human dentin structure is not possible for direct visualization. All the in vitro models were made of transparent glass, and it is noted that glass may behave differently from root-canal-wall material. The results of the in vitro studies may therefore not reflect what actually happens in clinical settings. Avoiding the bias results of in vitro study that it does not mimic the real condition of clinical settings, the authors, in this study, solve this dilemma using modified extracted teeth as model, which more relevant to clinical condition. Comprehensive visualizations were acquired using a high-speed camera and the glass root canal models together with modified teeth, to allow the basic physical mechanism involved in cavitation in the solution during sonic activation to be studied. Ruddle claimed that cavitation occurs when a solution is activated using an electrical driven EndoActivator tip.⁸ According to the formula for the cavitation number Ca and the corresponding velocity threshold, which can be calculated using the equation $U = 2\pi f A$, it is not possible for cavitation to occur during an air or electrical sonic activation.^{13,14} It is not clear why bubbles occurred during activation using an electrical driven EndoActivator tip in solutions. Cavitation is important to the efficient cleaning of the root canal system.^{15,16} There is a clear need to improve our understanding of the mechanism involved in bubble formation in a solution activated using an electrical driven EndoActivator tip during root canal treatment. Two approaches were used, one using the air-solution interface system (in the coronal portion) and the other using the solution-air interface system (in the apical portion). The solution-air interface system represents air entrapment in the apical region. The process in the air-solution or solution-air systems can be observed closely using the modified extracted tooth and the glass canal model. The high-speed imaging method used enables the capture of images within intervals of milli-microseconds. Surprisingly, we clearly observed that the end of the vibrating tip did not generate the formation of bubbles. Furthermore, the tip vibrations disturbed the sonically activated solution interface. The vibrating activator tip transferred its energy to the solution and created waves, and these caused surface waves to form at the interfaces in both systems.

In the air-solution interface system, the energy from the vibrating tip (at a certain power setting) was able to change the specific internal energy of the air-solution system to another form of energy.¹⁷ The energy was then transferred to the air-solution interface as pressure waves, disrupting the interface and creating individual surface tension in the form of bubbles. In other words, accumulation of input energy (from the vibrating

tip) was used as kinetic energy for the vibrating tip to generate bubbles. This process set up a series of waves or oscillations at the air-solution interface. Finally, the formation of bubbles then occurred limited to the vicinity of the tip at around 75 µs post-EA tip activation. The life cycle of a bubble observed ranges from 250,000–700,000 µs with predominantly 700 µm in diameter. The authors assumed that the bubbles are not cavitation because it occurred below the threshold needed for cavitation.^{13,14} We have suggested that the vibration pattern of the tip at 190 Hz generated the first mode shape, which has single node and antinode. The node at the attachment and the antinode at the end of the tip, which vibrates freely as a cantilever beam model (back and forth linear movement). A mode shape is defined as a specific pattern of vibration executed by a mechanical system at a specific frequency.¹⁸ The back and forth linear motion of an electrical driven EndoActivator tip generated disruption of surface tension at the air–solution in the form of bubbles in the solution. The process involved the use of air (compressible fluid) and solution (incompressible fluid), which differ in density.¹⁹

The processes in the solution-air interface system showed that the accumulating kinetic energy reaching the surface of the solution (i.e., the solution-air interface) disturbed the surface and produced surface waves. However, the surface tension was just adequate to generate waves and acoustic streaming but not cavitation. We conclude that the energy transferred from the activator tip to the solution was not sufficient to disrupt the surface tension at interface. Surface tension is defined as the attractive force between molecules (internal energy) acting to decrease the surface area of a liquid. This produces a layer of surface molecules on the liquid that acts like a stretched membrane.²⁰ In particular, this internal energy limits the flow of the liquid into narrow canals. It suggests that the surface tension internal energy of the solution, acting at the solution-air interface, has an important role in preventing bubbles generation.²⁰ Furthermore, in restricted areas, due to the oscillation amplitude of the EA tip to be 5 mm (Figure 3D), the sonic driven tip will undergo much more wall contact, which inhibits the tip vibration and may reduce the efficient streaming of the irrigant. This phenomenon occurred in the restricted areas such as modified extracted tooth.¹¹ Although each image was captured in a different root canal model, the dynamics of pressure waves, bubble formation, and wave formation proved to be reproducible in this in vitro study.

To the best of our knowledge, this method of visualizing fluid dynamics and bubbles formation in real-time using real human tooth structures has not been used in any previous study. We believe that the results of this study improve our initial understanding of the basic mechanisms involved in the fluid dynamics in a solution used to treat root canals by means of an electrical driven EndoActivator tip. Further studies are required to investigate the natural frequencies of tip to improve its efficacy.

CONCLUSIONS

Regardless of the solution type, our results show that fluid streaming and bubbles formation occur in a solution when activated by an electrical driven EndoActivator tip at the air-solution interface, instead of at the solution-air interface and modified extracted tooth.

ACKNOWLEDGMENTS

The authors deny any conflict of interest related to this study. The authors would like to thank Professor Zainal Abidin PhD at the Institut Teknologi Bandung (Bandung, Indonesia), and Mr. Wowo Watumas at the Phantom Company for their helpful contributions to discussions.

REFERENCES

1. Bronnec F, Bouillaguet S, Machtou P. Ex vivo assessment of irrigant penetration and renewal during the final irrigation regimen. Int Endod J. 2010; 43(8): 663–72.

2. Huang TY, Gulabivala K, Ng YL. A bio-molecular film ex-vivo model to evaluate the influence of canal dimensions and irrigation variables on the efficacy of irrigation. Int Endod J. 2008; 41(1): 60–71.

3. Sabins RA, Johnson JD, Hellstein JW. A comparison of the cleaning efficacy of short-term sonic and ultrasonic passive irrigation after hand instrumentation in molar root canals. J Endod. 2003; 29(10): 674–8.

4. Tronstad L, Barnett F, Schwartzben L, Frasca P. Effectiveness and safety of a sonic vibratory endodontic instrument. Dent Traumatol. 1985; 1(2): 69–76.

5. Plotino G, Pameijer CH, Grande NM, Somma F. Ultrasonics in endodontics: a review of the literature. J Endod. 2007; 33(2): 81-95.

6. Gulabivala K, Ng YL, Gilbertson M, Eames I. The fluid mechanics of root canal

irrigation. Physiol Meas. 2010; 3(12): R49-84.

Weller RN, Brady JM, Bernier WE. Efficacy of ultrasonic cleaning. J Endod. 1980;
 6(9): 740–3.

8. Ruddle CJ. Hydrodynamic disinfection "Tsunami" endodontics. Int Dent SA. 2009; 11(4): 6–18.

9. de Gregorio C, Estevez R, Cisneros R, Heilborn C, Cohenca N. Effect of EDTA, sonic, and ultrasonic activation on the penetration of sodium hypochlorite into simulated lateral canals: an in vitro study. J Endod. 2009; 35(6): 891–5.

10. Caron G. Cleaning efficiency of the apical millimetres of curved canals using three different modalities of irrigant activation: an SEM study. Masters thesis, Paris VII University, Paris, France; 2007.

11. Swimberghe RCD, De Clercq A, De Moor RJG, Meire MA. Efficacy of sonically, ultrasonically and laser-activated irrigation in removing a biofilmmimicking hydrogel from an isthmus model. Int Endod J. 2019; 52(4): 515–23.

12. Jensen SA, Walker TL, Hutter JW, Nicoll BK. Comparison of the cleaning efficacy of passive sonic activation and passive ultrasonic activation after hand instrumentation in molar root canals. J Endod. 1999; 25(11): 735–8.

13. van der Sluis LWM, Versluis M, Wu MK, Wesselink PR. Passive ultrasonic irrigation of the root canal: a review of the literature. Int Endod J. 2007; 40(6): 415–26.
14. Macedo R, Verhaagen B, Rivas DF, Versluis M, Wesselink P, Van der Sluis L. Cavitation Measurement during Sonic and Ultrasonic Activated Irrigation. J Endod. 2014; 40(4): 580–3.

15. Matsumoto H, Yoshimine Y, Akamine A. Visualization of irrigant flow and cavitation induced by Er:YAG laser within a root canal model. J Endod. 2011; 37(6): 839–43.

16. De Moor RJG, Blanken J, Meire M, Verdaasdonk R. Laser induced explosive vapor and cavitation resulting in effective irrigation of the root canal. Part 2: evaluation of the efficacy. Lasers Surg Med. 2009; 41(7): 520–23.

 Cengel YA, Boles MA. Thermodynamics, an engineering approach. 7th edn. NY, Mc Graw Hill, 2013.

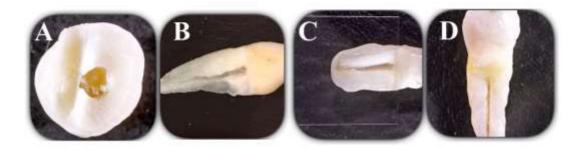
18. Thomson WT. Theory of Vibration with Applications. 4th edn. New Jersey, Prentice Hall, 1972.

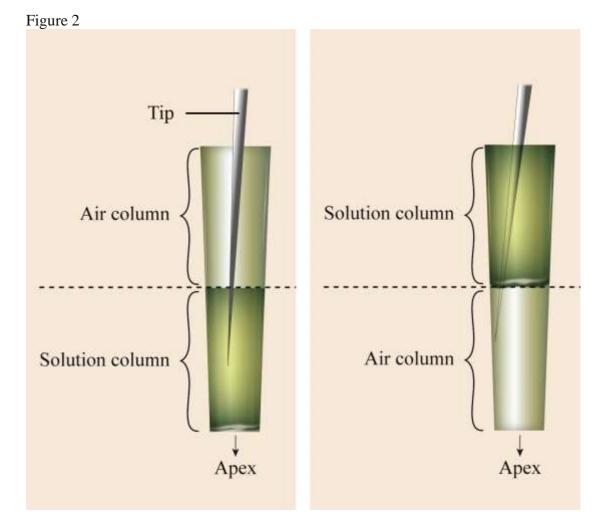
19. Peeters HH, Iskandar B, Suardita K, Suharto D. Visualization of Removal of

Trapped Air from the Apical Region of the Straight Root Canal Models Generating 2 phase Intermittent Counter Flow during Ultrasonically Activated Irrigation. J Endod. 2014; 40(6): 857-61.

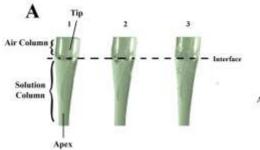
20. Crowe CT, Elger DF, Williams BC, Roberson JA. Engineering fluid mechanics, 9th edn. Wiley, New York, 2010.

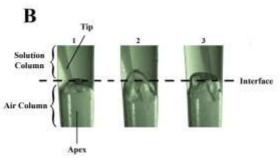
Figure 1



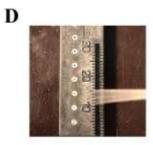












Comments to the Author:

Congratulations on your manuscript. It is an interesting study about activated irrigation using EndoActivator. There are some corrections and additions for improvement:

- 1. Please conduct a plagiarism check.
- 2. Research procedure/method: a schematic figure on how the process of observation along with photos of the utilities used during the experiment should be added. These figures can be combined for optimalization, figure 1 and 2 can be merged. A detailed pictures or photograph of the procedures on this research need to be added.
- 3. The result: Aside from the supplemented videos (YouTube), a detailed photos of the observation (result) regarding the bubbles must be added to better understand the bubble formation, it can be taken from the steps and please provide annotations and arrows in the photos to clarify the proposed results, including the photos taken both from the glass/acrylic model and extracted tooth for better viewing.
- 4. In the discussion, regarding the preparation of natural tooth with Balanced Force technique using Protaper hand file, what is your reasoning of using this technique and file? From the photo of the natural tooth, the root canal anatomy is somewhat not straight, why do you choose non straight canal? Please add your reasons in the discussion.
- 5. Most of the references are above 10 years (except reference number 11, 14, 17 no page provided, and 19), please replace with current references less than 10 years. Please refer the author guidelines in referencing. You may include your related previous publications (less than 10 years).

Dental Journal

Majalah Kedokteran Gigi

Dental Journal

(Majalah Kedokteran Gigi)

2022 June; 55(2): 71-75

Original article

Visualization of bubbles generation of electrical-driven EndoActivator tips during solutions activation in a root canal model and a modified extracted tooth: A pilot study

Harry Huiz Peeters¹, Elvira Theola Judith², Ketut Suardita³, Latief Mooduto⁴

¹Laser Research Center in Dentistry, Bandung, Indonesia

²Faculty of Dentistry, Universitas Maranatha, Bandung, Indonesia

³Faculty of Dentistry, IIK Bhakti Wiyata, Kediri, Indonesia

⁴Department of Conservative Dentistry, Faculty of Dental Medicine, Universitas Airlangga, Surabaya, Indonesia

ABSTRACT

Background: EndoActivator, a sonically-driven canal irrigation system (Dentsply Tulsa Dental Specialties, Tulsa, OK), has been developed for activating root canal irrigants, and has recently been released onto the market. **Purpose:** To obtain an initial understanding of bubbles generation of electrical EndoActivator tips during activation of the irrigant in a transparent root canal model and a modified extracted tooth. **Methods:** A modified extracted tooth and a straight glass model were filled with a solution containing 17% EDTA or 3% NaOCI. A medium activator tip 22-mm polymer noncutting #25, 0.04 file driven by an electrical sonic hand-piece at 190 Hz (highest level) induced pressure waves that produced macro- and micro-bubbles. The physical mechanisms involved were visualized using a Miro 320S high-speed imaging system (Phantom, Wayne, NJ, USA) with high temporal and spatial resolutions. The imaging system acquired images at 25,000 frames per second with 320x240 pixels per image, and attached a 60-mm f/2.8 macro lens (Nikon, Tokyo, Japan). **Results:** The end of the tip did not generate bubbles formation. Disruption of surface tension at the air–solution system in the glass canal model by an electrical sonic driven EndoActivator tip generated bubbles in the solution. However, it did not occur at the system of solution–air interfaces in the glass canal and modified extracted tooth. **Conclusion:** The physical mechanism of the solution activated by an electrical sonic driven EndoActivator tip in genering bubbles formation is because the surface tension at the air–solution system or modified extracted tooth. Better understanding of the physical mechanisms that relate specifically to the activation behaviour of EndoActivator tips in solutions is key to improving the cleaning mechanism that applies during root canal treatment.

Keywords: bubbles formation; EndoActivator; endodontics; sonic

Correspondence: Harry Huiz Peeters, Laser Research Center in Dentistry, Cihampelas 41 Bandung, West Java, 40174, Indonesia. Email: h2huiz@cbn.net.id

INTRODUCTION

Non-automated agitation of the irrigant can be produced using a file or a guttapercha cone that is inserted in the shaped and cleaned canal and subsequently moved periodically and rapidly by the operator.^{1,2} Electrical ultrasonic and sonic devices are examples of automated agitation.³ These devices use a fine non-cutting polymer or metal tip that is vibrated within the canal space, with different frequencies according to manufacturer's instruction. Sonically activated instruments are driven with a frequency range of 1,000–6,000 Hz and will generate a single node near the point at which the file is attached, as well as an antinode at the tip of the file.^{4–6} In ultrasonic solutions agitation, a tip oscillates at frequencies of 25 to 30 kHz in a pattern of motion consisting of nodes and antinodes along its length.⁷ EndoActivator, a sonically-driven canal irrigation system (Dentsply Tulsa Dental Specialties, Tulsa, OK, USA), has been developed for activating root canal irrigants, and has recently been released onto the market. It comprises a portable hand-piece with three modes (low, medium, and high) and 3 types of disposable

Dental Journal (Majalah Kedokteran Gigi) p-ISSN: 1978-3728; e-ISSN: 2442-9740. Accredited No. 158/E/KPT/2021. Open access under CC-BY-SA license. Available at https://e-journal.unair.ac.id/MKG/index DOI: 10.20473/j.djmkg.v55.i2.p71–75 flexible non-cutting polymer tips of different sizes. Its design allows for the safe activation of various irrigating solutions and could produce vigorous fluid dynamic in the canal space.⁸ Electrical devices driven sonically has been found to be effective for cleaning root canals in many studies.^{3,9,10} However, in the last decade, some conflicting results have been reported in the literature on the efficacy of the system and its superiority over other systems. Some researchers have found that sonically activated irrigation is less effective than ultrasonically activated irrigation, while others have suggested that the two methods produce comparable results.^{11,12} To our best knowledge, there have been no previous studies of the physical mechanism through which the electrical sonic driven EndoActivator tip generates bubbles during activation of irrigation solution. In the present study, the mechanism was investigated by acquiring real-time data using a modified extracted tooth and a transparent glass model of root canal in order to visualize the oscillation amplitude of an EndoActivator tip in the solution. Hence, more knowledge on the mechanism underlying the activation behaviour of the EndoActivator tips in the solutions is crucial to improve the outcomes of the root canal treatment.

MATERIALS AND METHODS

A single-rooted human mandibular first premolar with straight canal and mature apex, and a tooth length of 25 mm was selected. The tooth was not decoronated, and the intact crown (10 mm) acted as a reservoir for the irrigation solution. The preparation of the root canal was completed to the working length (WL) using a Pro Taper hand file (Dentsply Maillefer) up to a size of 25/0.06 as the master apical file (MAF) with the balanced force technique. The distal aspect of the root of the extracted tooth was carefully cut (from cervical to apex) using a fissure diamond bur (Dentsply, Tulsa, OK, USA) until the canal space was exposed (Figure 1A and B), then the surface was ground successively with 240-, P400, and 600-grit sandpaper to provide a smooth surface. A glass cover was then carefully glued to the surface without entering the canal, and this was covered with composite (Figure 1C and D). The apex was then sealed with composite resin and the outer surface of the tooth was covered with nail varnish, ensuring that there were no visible cracks, to simulate conditions within the root canal in vivo. The tooth was filled with the solution until it reached 5 mm below of the cervical level. The tip of the sonic instrument was inserted 2 mm shorter than the WL. A single modified extracted tooth was used for uniformity in the width and size of the root canal.

The conditions within a straight root canal were simulated and visualized using a glass model with artificial canal and pulp cavity (which acted as a reservoir). The model was a glass root canal model (Kimia Farma, Bandung, West Java, Indonesia), with canal inner diameter was 0.4 mm at the apex, crown height was 8 mm, crown diameter was 6 mm, a taper of 0.06 and an overall length of 25 mm. The canal was filled with a solution containing 17% EDTA or 3% NaOCl. The apex of the model was sealed with composite to allow the conditions within a root canal to be simulated. Two approaches were used, one used the air-solution interface (in the coronal portion) and the other used the solution-air interface (in the apical portion). The solution-air interface system represents air entrapment in the apical region (Figure 2). The models were filled with the solution until the level reached 5 mm below the coronal end (air-solution interface system), while in the solutionair interface system, the model was filled with solution starting 5 mm from the apex (air entrapment model). The tip of the sonic instrument was inserted into the solution at the air-solution or solution-air interface (10 mm and 4 mm from the interface, respectively). A single transparent glass model was used for uniformity in the width and size of the root canal.

A 22-mm polymer noncutting # 25, 0.04 file (Dentsply) was mounted on an EndoActivator hand-piece (Dentsply Tulsa Dental Specialties, Tulsa, OK) set at high mode (190 Hz) and used to activate the irrigant. The models were subjected to active sonic irrigation. The tip of the sonic instrument was inserted into the solution at the air–solution (10 mm) or solution–air interface (4 mm), while the tip was inserted 2 mm shorter than the working length in the modified extracted tooth and activated passively without any filing motion. The hand-piece was fixed in a holder to ensure that the desired position was maintained. All the experiments with the different samples were performed by the same operator.

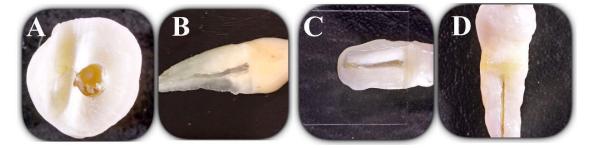


Figure 1. (A) A root sample with a longitudinal cut in the distal aspect of the root canal. (B) Exposed root canal space. (C) Exposed tooth with a glass cover (0.15 mm thick). (D) The glass cover was glued to the tooth and shaped to fit the shape of the root.

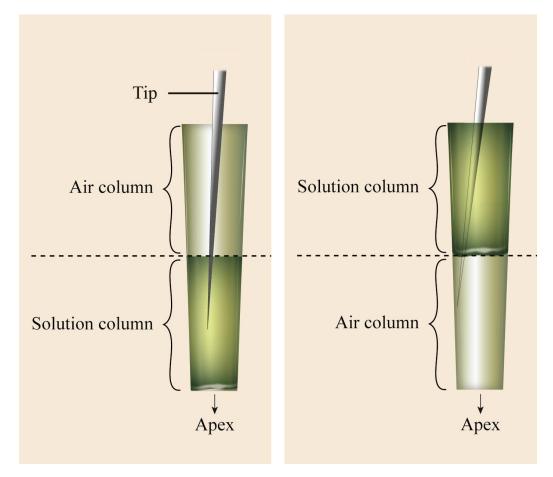


Figure 2. Illustrations of the solution-air and air-solution systems.

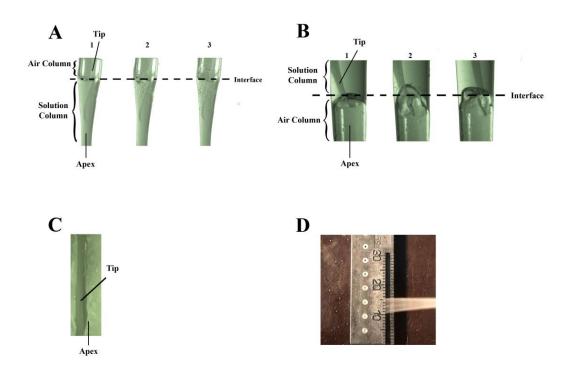


Figure 3. Representative images showing the disruption of surface tension at the air solution (A) and solution-air systems (B) in glass models, (C) modified extracted human root canal, (D) Oscillation amplitude of the EA tip in the air.

The process was recorded using a Phantom Miro 320S high-speed digital imaging system (Wayne, NJ, USA) at 25,000 frames per second with 320x240 pixels per image using a macro lens (60 mm, f/2.8; Nikon, Tokyo, Japan). The root canal model was illuminated using a Fibre-Lite LMI-6000 LED continuous light source (Dolan- Jenner Industries, Boxborough, MA, USA). Measurements were repeated three times for each sample.

RESULTS

Regardless of whether 17% EDTA or 3% NaOCl was used, the high-speed images and video clearly showed fluid streaming. The electrical sonic activated EndoActivator tip generated hydrodynamic and finally bubbles formation occurred during the sonic activation process in the solutionair system instead of in the restricted areas (the solution-air system and modified extracted tooth). Oscillation amplitude of the EA tip in the air was approximately 5 mm.

During the first 69 µs, the sonic energy was transferred to the tip and the tip vibrated at a frequency of approximately 190 Hz, causing the solution surface (the air-solution interface) to move as a wave. After 75 µs, macro and micro bubbles formed in the solution. Subsequently, accumulation of the energy during sonic activation at interface caused the micro and macro bubbles in the solution. Finally, the bubbles formation occurred only the vicinity of the tip (Figure 3A). (See supplemental video S1 at https://youtu. be/ut6VKWDGmj8). The life cycle of a bubble observed ranges from 250,000–700,000 µs with predominantly 700 µm in diameter. The end of the tip did not generate bubbles formation.

A model with air trapped in the apical region was used; thus, there was a solution column and an air column. The energy of the tip vibration was not sufficiently enough to break the surface tension at the solution–air interface (Figure 3B). (See supplemental video S2 at https://youtu.be/ rqzMOWmJUOY). No bubbles formation occurred during the activation process in the modified extracted tooth (airsolution interface system) (Figure 3C). (See supplement video S3 https://youtu.be/-BHdVIO3DkA).

DISCUSSION

To be clinically relevant, *in vitro* studies should reproduce the clinical situation as much as possible. Human dentin structure is not possible for direct visualization. All the *in vitro* models were made of transparent glass, and it is noted that glass may behave differently from rootcanal-wall material. The results of the *in vitro* studies may therefore not reflect what actually occurs in clinical settings. Avoiding the bias results of *in vitro* study that it does not mimic the real condition of clinical settings, the authors, in this study, solve this dilemma using modified extracted teeth as models, which more relevant to clinical condition. Comprehensive visualizations were acquired using a high-speed camera and the glass root canal models together with modified teeth, to allow the basic physical mechanism involved in bubbles formation in the solution during sonic activation to be studied. Ruddle⁸ claimed that cavitation occur when a solution is activated using an electrical-driven EndoActivator tip. According to the formula for the cavitation number Ca and the corresponding velocity threshold is 15 m/sec, the velocity of the file which can be calculated using the equation $U = 2\pi f A$ (with f being the file oscillation frequency an A is the amplitude of oscillation). The EA tip oscillating at a frequency of 190 Hz and oscillation amplitude of 5 mm oscillates with a velocity of 5.9 m/sec, it is not possible for cavitation to occur during an air or electrical sonic activation.^{13,14} It is not clear why bubbles occurred during activation using an electrical-driven EndoActivator tip in solutions. Cavitation is important to the efficient cleaning of the root canal system.^{15,16} There is a clear need to improve our understanding of the mechanism involved in bubble formation in a solution activated using an electricaldriven EndoActivator tip during root canal treatment. Two approaches were used, one using the air-solution interface system (in the coronal portion) and the other using the solution-air interface system (in the apical portion). The solution-air interface system represents air entrapment in the apical region. The process in the air-solution or solution-air systems can be observed closely using the modified extracted tooth and the glass canal model. The high-speed imaging method used enables the capture of images within intervals of mili-microseconds. Surprisingly, we clearly observed that the end of the vibrating tip did not generate the formation of bubbles. Furthermore, the tip vibrations disturbed the sonically activated solution interface. The vibrating activator tip transferred its energy to the solution and created waves, and these caused surface waves to form at the interfaces in both systems.

In the air-solution interface system, the energy from the vibrating tip (at a certain power setting) was able to change the specific internal energy of the air-solution system to another form of energy.¹⁷ The energy was then transferred to the air-solution interface as pressure waves, disrupting the interface and creating individual surface tension in the form of bubbles. In other words, accumulation of input energy (from the vibrating tip) was used as kinetic energy for the vibrating tip to generate bubbles. This process created a series of waves or oscillations at the air-solution interface. Finally, the formation of bubbles then occurred limited to the vicinity of the tip at around 75 µs post-EA tip activation. The life cycle of a bubble observed ranges from 250,000-700,000 µs with predominantly 700 µm in diameter. The authors have suggested that the bubbles are not cavitation because it occurred below the threshold needed for cavitation.^{13,14} The experiment showed that the vibration pattern of the tip at 190 Hz generated the first mode shape, which has single node and an antinode. The node at the attachment and the antinode at the end of the

tip, which vibrates freely as a cantilever beam model (back and forth linear movement). A mode shape is defined as a specific pattern of vibration executed by a mechanical system at a specific frequency.¹⁸ The back and forth linear motion/pendulum of an electrical-driven EndoActivator tip generated disruption of surface tension at the air–solution in the form of bubbles in the solution. The process involved the use of air (compressible fluid) and solution (incompressible fluid), which differ in density.¹⁹

The processes in the solution-air interface system showed that the accumulating kinetic energy reaching the surface of the solution (i.e., the solution-air interface) disturbed the surface and produced surface waves. However, the surface tension was just adequate to generate waves and streaming but not cavitation. We conclude that the energy transferred from the activator tip to the solution was not sufficient to disrupt the surface tension at the interface. Surface tension is defined as the attractive force between molecules (internal energy) acting to decrease the surface area of a liquid. This produces a layer of surface molecules on the liquid that acts like a stretched membrane.²⁰ In particular, this internal energy limits the flow of the liquid into narrow canals. It suggests that the surface tension internal energy of the solution, acting at the solution-air interface, has an important role in preventing bubbles generation.²⁰ Furthermore, in restricted areas, due to the oscillation amplitude of the EA tip to be 5 mm (Figure 3D), the sonic driven tip will make more wall contact, which inhibits the tip vibration and may reduce the efficient streaming of the irrigant. This phenomenon occurred in the restricted areas such as modified extracted tooth.¹¹ Although each image was captured in a different root canal model, the dynamics of pressure waves, bubble formation, and wave formation proved to be reproducible in this in vitro study.

To the best of our knowledge, this method of visualizing fluid dynamics and bubbles formation in real-time using real human tooth structures has not been used in any previous study. We believe that the results of this study improve our initial understanding of the basic mechanisms involved in the fluid dynamics in a solution used to treat root canals by means of an electrical-driven EndoActivator tip. Further studies are required to investigate the natural frequencies of the tip to improve its efficacy. Regardless of the solution type, our results show that fluid streaming and bubbles formation occurred in a solution when activated by an electrical-driven EndoActivator tip at the air-solution interface, instead of at the solution-air interface and modified extracted tooth.

ACKNOWLEDGMENTS

The authors deny any conflict of interest related to this study. The authors would like to thank Professor Zainal Abidin PhD at the Institut Teknologi Bandung (Bandung, Indonesia), and Mr. Wowo Watumas at the Phantom Company for their helpful contributions to discussions.

REFERENCES

- Andreani Y, Gad BT, Cocks TC, Harrison J, Keresztes ME, Pomfret JK, Rees EB, Ma D, Baloun BL, Rahimi M. Comparison of irrigant activation devices and conventional needle irrigation on smear layer and debris removal in curved canals. (Smear layer removal from irrigant activation using SEM). Aust Endod J. 2021; 47(2): 143–9.
- Andrabi SM-U-N, Kumar A, Mishra SK, Tewari RK, Alam S, Siddiqui S. Effect of manual dynamic activation on smear layer removal efficacy of ethylenediaminetetraacetic acid and SmearClear: an in vitro scanning electron microscopic study. Aust Endod J. 2013; 39(3): 131–6.
- Mancini M, Cerroni L, Iorio L, Armellin E, Conte G, Cianconi L. Smear layer removal and canal cleanliness using different irrigation systems (EndoActivator, EndoVac, and passive ultrasonic irrigation): field emission scanning electron microscopic evaluation in an in vitro study. J Endod. 2013; 39(11): 1456–60.
- Tronstad L, Barnett F, Schwartzben L, Frasca P. Effectiveness and safety of a sonic vibratory endodontic instrument. Endod Dent Traumatol. 1985; 1(2): 69–76.
- Plotino G, Pameijer CH, Grande NM, Somma F. Ultrasonics in endodontics: a review of the literature. J Endod. 2007; 33(2): 81–95.
- Gulabivala K, Ng Y-L, Gilbertson M, Eames I. The fluid mechanics of root canal irrigation. Physiol Meas. 2010; 31(12): R49-84.
- Weller RN, Brady JM, Bernier WE. Efficacy of ultrasonic cleaning. J Endod. 1980; 6(9): 740–3.
- Ruddle CJ. Hydrodynamic disinfection "tsunami" endodontics. Int Dent SA. 2009; 11(4): 6–18.
- Akman M, Akbulut MB, Aydınbelge HA, Belli S. Comparison of different irrigation activation regimens and conventional irrigation techniques for the removal of modified triple antibiotic paste from root canals. J Endod. 2015; 41(5): 720–4.
- Haupt F, Meinel M, Gunawardana A, Hülsmann M. Effectiveness of different activated irrigation techniques on debris and smear layer removal from curved root canals: a SEM evaluation. Aust Endod J. 2020; 46(1): 40–6.
- Swimberghe RCD, De Clercq A, De Moor RJG, Meire MA. Efficacy of sonically, ultrasonically and laser-activated irrigation in removing a biofilm-mimicking hydrogel from an isthmus model. Int Endod J. 2019; 52(4): 515–23.
- Rodrigues CT, EzEldeen M, Jacobs R, Lambrechts P, Alcalde MP, Hungaro Duarte MA. Cleaning efficacy and uncontrolled removal of dentin of two methods of irrigant activation in curved canals connected by an isthmus. Aust Endod J. 2021; 47(3): 631–8.
- van der Sluis LWM, Versluis M, Wu MK, Wesselink PR. Passive ultrasonic irrigation of the root canal: a review of the literature. Int Endod J. 2007; 40(6): 415–26.
- Macedo R, Verhaagen B, Rivas DF, Versluis M, Wesselink P, van der Sluis L. Cavitation measurement during sonic and ultrasonic activated irrigation. J Endod. 2014; 40(4): 580–3.
- Matsumoto H, Yoshimine Y, Akamine A. Visualization of irrigant flow and cavitation induced by Er:YAG laser within a root canal model. J Endod. 2011; 37(6): 839–43.
- Verhaagen B. Root canal cleaning: through cavitation and microstreaming. [Enschede, The Netherlands]: Enschede: University of Twente; 2012. p. 359.
- Çengel YA, Boles MA. Thermodynamics: an engineering approach. 7th ed. New York: McGraw-Hill; 2011. p. 978. (Cengel series in engineering thermal-fluid sciences).
- Scott AS, Fong E. Body structures & amp; functions. Thomson/ Delmar Learning; 2004. p. 495.
- Peeters HH, Iskandar B, Suardita K, Suharto D. Visualization of removal of trapped air from the apical region of the straight root canal models generating 2-phase intermittent counter flow during ultrasonically activated irrigation. J Endod. 2014; 40(6): 857–61.
- Elger DF, Williams BC, Crowe CT. Engineering fluid mechanics. 10th ed. New Jersey: John Wiley & Sons, Inc.; 2012. p. 688.