

The Effect of oral N-Acetylcystein on prevention of extensive tissue destruction in electrical burn injury

by Iswinarno Doso Saputro

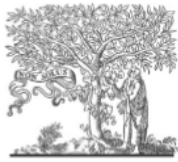
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The effect of oral N-acetylcystein on prevention of extensive tissue destruction in electrical burn injury

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ABSTRACT

Background: Electric burn patients usually suffer permanent injury and sequelae. Salvage of the zone of stasis is an important topic in the treatment of burn patients. N-Acetylcystein (NAC), as an antioxidant, has effect on the saving zone of stasis and extensive rhabdomyolysis. The aim of this study was therefore to evaluate the effect of oral NAC on tissue destruction indicators in an electric burn rat model.

Material and methods: An experimental study was conducted with thirty six male Wistar albino rats divided into 2 groups. Group A (n=18) and group B (n=18) were electrical burn injury groups without and with NAC therapy, respectively. The extent of burn wounds were evaluated by planimetry using a digital wound measuring device. Blood samples were obtained to analyze creatine kinase (CK) levels as a marker of extensive rhabdomyolysis on the first hour after electric injury (baseline) and on the 7th day to see the antioxidant effect of NAC. **Results:** A significant decrease in tissue destruction was seen by the necrotic area on day 7 in the NAC therapy group compared to the control group (mean $2.26 \pm 1.05 \text{ cm}^2$ versus mean $7.12 \pm 3.30 \text{ cm}^2$ respectively; $p=0.001$), which was confirmed by the level of serum CK (day 7: group A, mean $140 \pm 51 \text{ U/L}$ versus Group B, mean $102 \pm 6 \text{ U/L}$; $p=0.007$).

Conclusion: A decrease in electric burn necrotic area and tissue damage in the group using NAC treatment was demonstrated. NAC might have a beneficial effect in the treatment of electrical burns. Further experimental and clinical studies with NAC treatment are necessary to confirm these results.

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1. Introduction

Electrical burns occur in 3–5% of thermal burns patients [1,2], and cause serious injury in 1000–1500 people every year around the world with a mortality rate of 20–30%. Permanent injury and sequelae were seen in 74% of patients who survived [3,4].

Electrical burns, especially high-voltage injuries, often cause damage to the skin and deeper structures, resulting in large areas of necrosis. However, the severity of tissue damage depends on several factors, including the intensity of the electrical burn (high vs. low voltage), the current type (direct or indirect), the amount of electrical load, which body part is injured, the contact time, and the difference of resistance [5].

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Severe damage can lead to multi-organ failure (e.g. heart, kidney, nerves, eyes). Furthermore, injury in the extremities often lead to amputation, with the possibility of amputation of the affected fingers in low voltage electrical injuries.

A disturbance in the microvascular circulation is the main reason for severe tissue necrosis in electrical injury. The current treatment is still lacking in terms of prevention of tissue damage and organ damage due to electrical injury. Some studies report that antioxidant therapy is beneficial in overcoming the oxidative stress that develops during the wound healing process, particularly in electrical burns [6]. It has been shown that *N*-acetylcysteine (NAC) protects tissue damage caused by free radicals [7,8] by an increase in the levels of glutathione. NAC has already been used as a scavenger of oxygen free radicals, with many satisfactory results [6]. With oral antioxidant therapy, NAC is expected to prevent extensive tissue destruction from electrical burns, and thus reduce the morbidity and mortality rate.

2. Material and methods

This is a randomized controlled animal study that is performed in the Airlangga University Medical Faculty Biochemistry Experimental Animal Laboratory between June 2017 and July 2017. The Animal Care and Use Committee (ACUC) Faculty of Veterinary Medicine of Airlangga University Surabaya Indonesia approved this study protocol. The Ethical Clearance number is 756-KE.

2.1. Animals and experimental procedure

Thirty six male Wistar albino rats weighting between 350 and 400g were divided into 2 main experimental groups. Group A (n=18) and B (n=18) were electrical burn injury groups without and with NAC therapy, respectively. Two separate analyses were performed at different time points on 9 out of 18 rats from Group A and B at each time point. Biochemistry (creatin kinase levels) and necrotic areas at the 1st hour and at day 7 after electrical injury were evaluated. The animals were cared in standard laboratory conditions throughout the study.

2.2. Electrical burn injury

All animals were anesthetized with intramuscular injection of ketamine hydrochloride (50mg/kg) and xylazine hydrochloride (15mg/kg). Before applying the electrode, the left upper extremity and right lower extremity of the rats were shaved and ultrasound gel was used as a conductive material above the skin. A phase electrode was placed on the left upper extremity and the neutral electrode was placed on the right lower extremity. The extremities were exposed to 220V AC for 10s approximately 450-500mA current applied.

2.3. N-Acetylcystein (NAC) therapy

Oral *N*-acetylcystein effervescent tablet (600mg; Fluimucil, PT. Zambon Indonesia) with a therapeutic dose

(600mg/kg/day) was administered 15min after electrical burn injury.

Since we used male Wistar albino rats weighting between 350 and 400g, we dissolved the 600mg effervescent tablet into 5mL of water. Then, the solution was administered in concordance with a therapeutic dosage, about 1.5-2cc by an oral gavage 20G needle. The *N*-acetylcystein was given every day, 15min after injury, for in total 7 days. The dosage of NAC was determined in accordance with previous reports of therapeutic doses [9].

2.4. Biochemical evaluation

Creatine kinase (CK) can be used as a marker of tissue damage. Measurement of serum CK activity is still an important indicator of muscle cell necrosis and tissue damage due to disease or trauma. Based on the literature, progression of tissue damage can be assessed based on the increase in serum CK [10]. CK level measurements were performed with 3mL of blood specimen extracted from cardiac puncture. Blood was stored in an EDTA tube at 4-10°C temperature, and processed within 6h after blood extraction.

2.5. Evaluation of necrotic areas

Necrotic area in this study is the area of skin exposed to electrical burns, causing a necrotic zone [11]. This zone macroscopically appears as a bluish color, gray to black with clearly defined borders. For planometric measurements, we used Visitrak Digital (Smith & Nephew Medical LTD, England). We measured the zone using a template (transparent plastic sheet), and then interpolated in order to obtain Visitrak output broad zones within cm². The necrotic areas of the left upper and right lower extremities were measured.

2.6. Statistical analysis

Non-parametric tests were used, such as the Kruskal-Wallis Test, the Wilcoxon Signed Rank Test, and the Mann-Whitney Test, since both variables were not normally distributed.

The Kruskal Wallis Test was used to analyze the difference between all 4 groups (2 groups of necrotic areas and 2 groups of CK levels).

The Mann-Whitney Test was used to analyze the difference between unpaired groups. In this study, it was used to analyze the difference between a CK level control group day 0 and day 7, CK level treatment group day 0 and day 7, CK level control group day 0 and CK level treatment group day 0, CK level control group day 7 and CK level treatment group day 7, necrotic area control group day 0 and necrotic area treatment group day 0, and necrotic area control group day 7 and necrotic area treatment group day 7.

The Wilcoxon Signed Rank Test was used to analyze the difference between two paired groups. In this study, it was used to analyze the difference between necrotic area control group day 0 and day 7, and between necrotic area treatment group day 0 and day 7. This test was also used to analyze the difference between CK level control group day 0 and day 7, and between CK level treatment group day 0 and day 7.

3. Results

Nine mice from Group A at day 7 suffered significant more tissue necrosis with some compartment syndrome on their extremities (mean necrotic area $7.12 \pm 3.30 \text{ cm}^2$; Figs. 1 and 2) compared to nine mice from Group B (mean necrotic area $2.26 \pm 1.05 \text{ cm}^2$; $p=0.007$).

This significant difference of tissue necrosis between Group A and B at day 7 was also confirmed by the level of serum CK. Mean baseline value of serum CK at day 0 (1st hour) for Group A was $5244 \pm 4350 \text{ U/L}$ and Group B was $10898 \pm 6674 \text{ U/L}$ (Fig. 3). At day 7, Group A had a mean value of serum

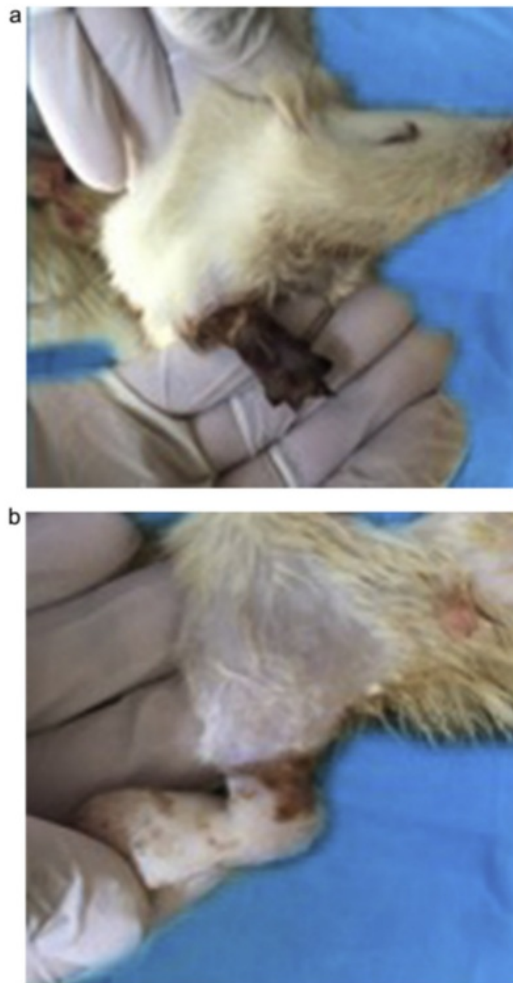


Fig. 1 – Clinical comparison of extremities between Group A and B at day 7.

(A) A significant severe necrotic area on the right front foot of a Group A mouse at day 7 (notice the compartment syndrome that it suffered resulting more devastating tissue necrosis).

(B) Small necrotic area on the left hind foot of a Group B mouse at day 7 (notice the burn injury was also circumferential without any signs of compartment syndrome).

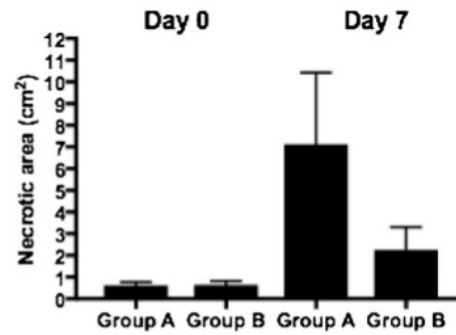


Fig. 2 – Mean necrotic area (cm^2) at day 0 and day 7 in group A compared to group B.

CK of $140 \pm 51 \text{ U/L}$ in comparison to $102 \pm 6 \text{ U/L}$ in Group B ($p=0.007$; Fig. 4).

4. Discussion

The effects of NAC have already been evaluated in several clinical studies, demonstrating that it has been used as a treatment modality for multiple diseases, such as cancer, cardiovascular diseases, diabetes [12], human immunodeficiency virus infections, liver and respiratory diseases [13-15]. In these diseases, oxidative stress is generated, which is defined by an imbalance between reactive oxygen species (ROS) and antioxidants. Also, in severe (electrical) burn patients, ROS and Reactive Nitrogen Species (RNS) are elevated in burned tissue. It has been shown that antioxidant therapy is beneficial in combating oxidative stress.

Sahib et al. showed that several antioxidants, including NAC, were useful in the treatment of patients with a burn injury, because it results in less wound infections and complications [16]. NAC, as an antioxidant, is a scavenger of free radicals as it interacts with ROS [17]. However, there are no reports comparing electric burn necrotic area and CK levels in severe electrical burns with or without NAC treatment. The beneficial effect of NAC in burn injury is confirmed by our study, in which a significant decrease in electric burn necrotic

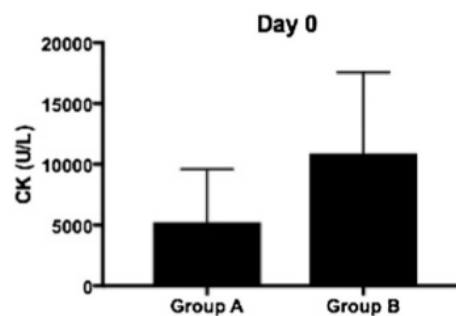


Fig. 3 – Mean value of serum CK at day 0 in group A compared to group B.

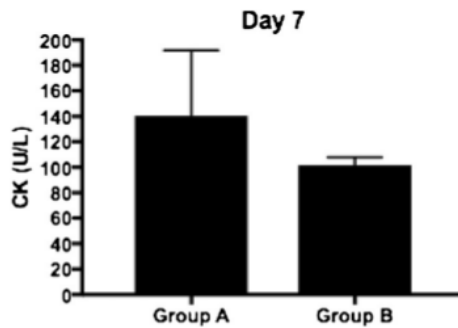


Fig. 4– Mean value of serum CK at day 7 in group A compared to group B.

area and tissue damage was seen in the group using NAC in an electric burn rat model. At the end of the 7th day, we found that the area of necrosis was prevented clearly by NAC in group B.

Furthermore, multiple animal studies were performed investigating the effect of NAC in burn wounds. One of these studies of Deniz *et al.* showed that NAC prevented tissue necrosis by saving the zone of stasis in a rat comb-burn model. They compared a group where NAC was administered orally or intraperitoneally with a control group, and evaluated the extent of burn wound and malondialdehyde levels. This study concluded that rats receiving NAC had a lower burn wound width. No significant differences were seen in malondialdehyde concentrations [18]. Another study of Tsai *et al.* also reported promising results regarding the use of NAC in a burn wound healing rat model. Better re-epithelialization was seen in wounds of rats treated with 3.0% NAC topically, and thus wound healing might be accelerated [19].

Csontos *et al.* analyzed the effect of NAC in a patient population ($n=30$). One group ($n=15$) received standard therapy, whereas in the other group ($n=15$) NAC was supplemented. Several mediators of oxidative stress and inflammation were measured. They showed that treatment with NAC resulted in a significant increase in glutathione on days 4-5, and significant lower inflammatory mediators, such as IL-6, IL-8, and IL-10. Thus, NAC leads to less oxidative stress and inflammation [20].

NAC is a precursor in glutathione formation. In burn injury, ROS is produced and several inflammatory cells cause tissue damage and finally multiple organ failure. Glutathione is elevated by NAC, and however, prevents cell damage caused by ROS.

NO-mediated vasodilation can be increased by NAC [18], thereby reducing the incidence of obstruction, ischemic thrombosis and ultimately necrosis of muscle tissue, which is confirmed in our experimental study showing decreased serum CK levels in Group B at day 7. Besides that, NAC has multiple advantages, such as easy applicable in oral form, effective, low costs, and safe.

In conclusion, in this experimental study, a significant decrease in electric burn necrotic area and CK levels was shown in the group using NAC treatment. Therefore, NAC might have a beneficial effect in the treatment of electrical

burn injury. Further experimental and clinical studies with NAC treatment are necessary to confirm these results.

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Conflict of interest statement

There are no editorial or financial conflicts of interest.

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