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**PENELITIAN BERJUDUL** : Quiscent Bone Marrow Mesenchymal Stem Cells  
Melalui Hypoxia Precondition Sebagai Upaya  
Mempertahankan Viabilitas Kultur Stem Cells

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## **Marrow Mesenchymal Stem Cell (MMSCs) Transplantation for the Improvement of Reproductive Function in Rat Testis Degeneration**

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**Abstract.** The purpose of this research was to obtain adaptive MMSCs for the therapy of testicular degeneration in rat. Adaptive MMSCs can be achieved by hypoxia precondition during in vitro culture. Hypoxia precondition in this research was conducted by applying 3% O<sub>2</sub> which was then compared with those cultured in normoxic 21% O<sub>2</sub>. The specific target of this research was divided into two experiments, Experiment 1 was the isolation and identification of mesenchymal stem cell from rat bone marrow. Identification was confirmed by the expression of CD 105<sup>+</sup> and CD45<sup>-</sup> cells. In Experiment 2 cellular MMSCs testiculomyoplasty were performed on male rats that have been preconditioned with testicle degeneration. Next, the success was identified in vivo based on the expression of spermatogonial stem cell (SSCs) and the improvement of testicle tissue. Transplantation results showed that under 3% O<sub>2</sub> concentration SSCs were expressed and testis tissue improvement occurred. Meanwhile at 21% O<sub>2</sub> concentration testis tissue improvement did not occur. The conclusion of this research showed that hypoxia precondition using 3% O<sub>2</sub> concentration very support MMSCs to remain adaptive after being transplanted to patient with spermatogenesis impairment, in addition to the testicle tissue improvement.

**Keywords:** *MMSCs, Testicular Degeneration, Testicle Tissue Improvement*

### **Introduction**

Testicular degeneration that lead to permanent damage of male main reproductive organ is normally followed by pathological remodelling (Schlatt *et al.*, 2002). Regeneration is merely possible to occur when normal spermatogonia and Sertoli cells still remaining in seminiferous tubules in a long period of time. Therefore, regeneration capacity to replace the degenerated cells is very limited (Hafez, 2000).

Transplantation of Marrow Mesenchymal Stem Cells (MMSCs) from bone marrow gave very promising solution in regenerating and returning the degenerating testicle tissue (Blanchard *et al.*, 1998). However, the minimum viability of the transplanted MMSCs in degenerated testicle caused a limited efficacies of this therapy (Kenichiro *et al.*, 2005; Tang *et al.*, 2005).

Hypoxia condition was needed during culture in order the stem cells remained adaptive and viable before and after transplantation (Safitri *et al.*, 2014). This condition was adjusted to mimic in vivo condition where the stem cells were produced. In testicle tissue, stem cells for spermatogenesis were in hypoxiacondition, i.e. under 1-6 % O<sub>2</sub> concentration (Wenger and Katschinski, 2005). In this research the concentration of O<sub>2</sub> applied was 3 % which was compared with conventional condition 21 % O<sub>2</sub>.

### **Research Methods**

#### **Procedure of RAT MSCs isolation and culture:**

Rat was premedicated and general anesthetized. MSCs from bone marrow was harvested by an aspiration at the middle of rat femur bone below the condylus. Aspirate contained rat MSCs from bone marrow was placed in heparinized tubes. Sample in tube was placed in thermos maintained at 4°C during transportation to be processed.

Sample was transferred into 15 ml sterile blue capped tubes and then the tube was rinsed twice with 5 ml sterile PBS and added up to a total volume of 10 ml. The diluted sample

was loaded over a same volume of Ficoll in a separate 15 ml tube. Centrifugation at 1600 rpm was performed for 15 minutes at room temperature. After centrifugation, the cells were collected from Ficoll-PBS interface using sterile pasteur pipette and transferred into a 15 ml tube. The cells were then resuspended in PBS up to a total volume of 15 ml. The tube was inverted gently 5 times to homogenize the suspension.

The suspension was then centrifuged again at 1600 rpm for 10 minutes. Supernatant and floating cells were discarded and cell pellet was resuspended in 6 ml of  $\alpha$  MEM media. Mononucleated cells were plated in 10 cm<sup>2</sup> plates at  $2 \times 10^7$  and incubated at 37°C in a humidified atmosphere containing 5% CO<sub>2</sub> for 24 hours to let the cells adhere. After 24 hours, media and non-adherent cells were discarded. Adherent cells were rinsed twice using 5 ml of PBS. Ten ml of fresh  $\alpha$  MEM media was then added into dish and the dish was returned into the incubator. Culture was observed daily under an inverted microscope. Every 4 days medium was changed, preceded by a rinse using 10 ml PBS then 10 ml of fresh  $\alpha$  MEM media were replaced. Culture was continued until approximately 75-80% confluence was attained. After confluence, cells were passaged into several dishes for subculture (Rantam *et al.*, 2009). Passage was conducted 3 times, then cells were divided into two hypoxic precondition treatments of 3% in hypoxic chamber inside a 5% CO<sub>2</sub> incubator while another treatment was the use of 21% oxygen (normoxia).

At the second day after hypoxic precondition treatments cells were analysed for the expression of surface marker CD105 and CD45 by flowcytometry. Flow cytometric analysis was performed using a flowcytometer (FACSCalibur) and CellQuest software. Antibodies used were monoclonal APC anti-mouse CD105 (Biolegend), dan PerCP-Cy5.5 anti-rabbit cross anti-rat CD45 (BD).

Two days after hypoxic preconditioning MMSCs were transplanted into rats that had been induced for testicular degeneration. The following ten days surgery was performed to collect testicle tissue. Testicle tissue improvement was observed through HE stained histopathological preparation.

### Results and Discussion

Flowcytometric analysis showed that under 3% O<sub>2</sub> concentration, the level of CD105 and CD44 cells in MMSCs culture were not altered (still undifferentiated). Meanwhile under 21% O<sub>2</sub> concentration, cells have experienced alteration (became differentiated), that was indicated by the decrease of CD105 and CD44 cells (Figure 1 - 3).

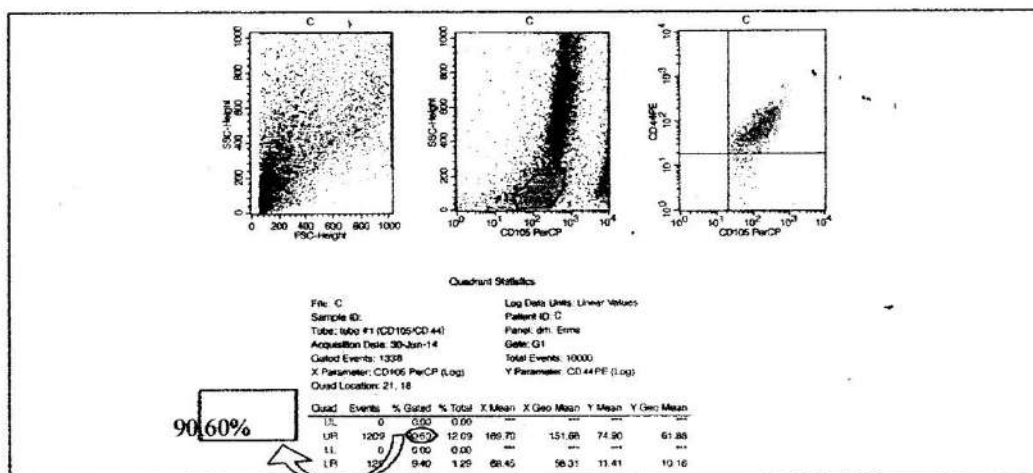


Fig. 1: Flowcytometric analysis of MSCs culture before hypoxic preconditioning (Control) showing positive expression of CD105 and CD44 (90,60%).



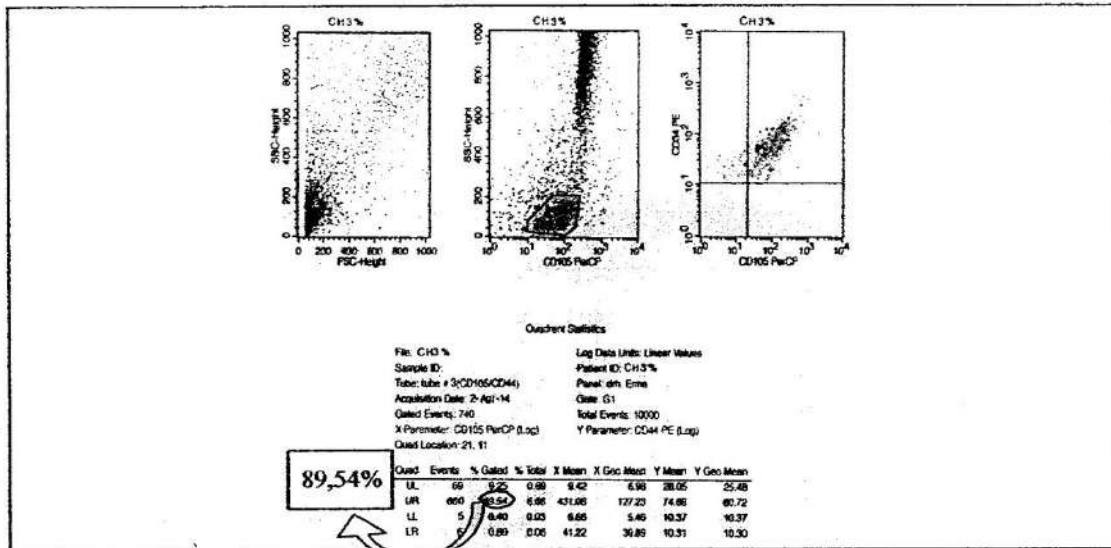


Fig. 2: Flowcytometric analysis of MSCs culture under hypoxic precondition (1% O<sub>2</sub> concentration) showing positive expression of CD105 and CD44 (89,54%).

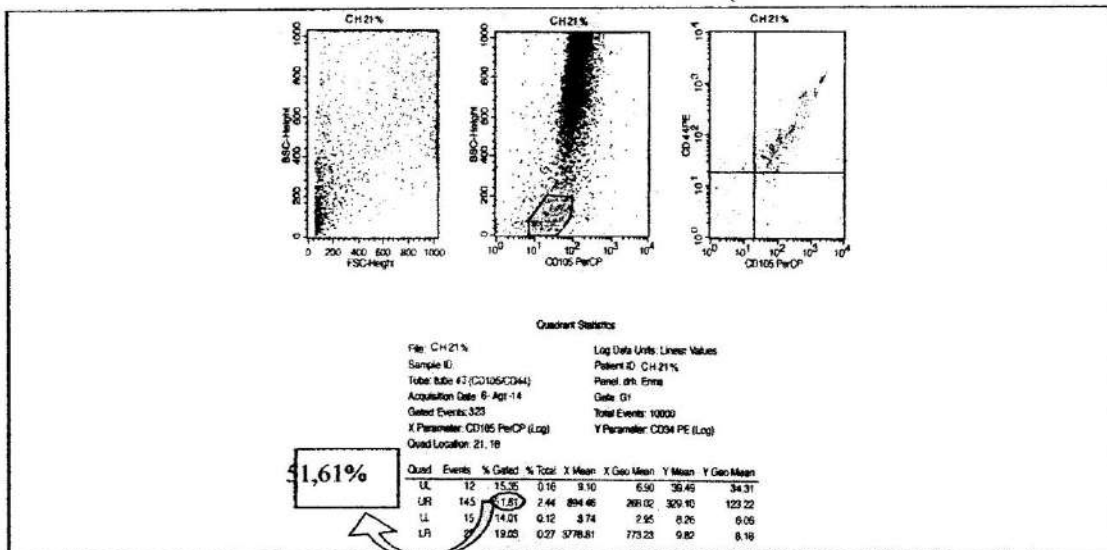


Fig. 3: Flowcytometric analysis of MSCs culture under 21% O<sub>2</sub> concentration (normoxia) showing positive expression of CD105 and CD44 (51,61%).

Histopathological observation showed that testicular tissue transplanted with MMSCs cultured under 3 % O<sub>2</sub> tension was regenerated, meanwhile testicular tissue transplanted with MMSCs cultured under 21 % O<sub>2</sub> tension was not regenerated.

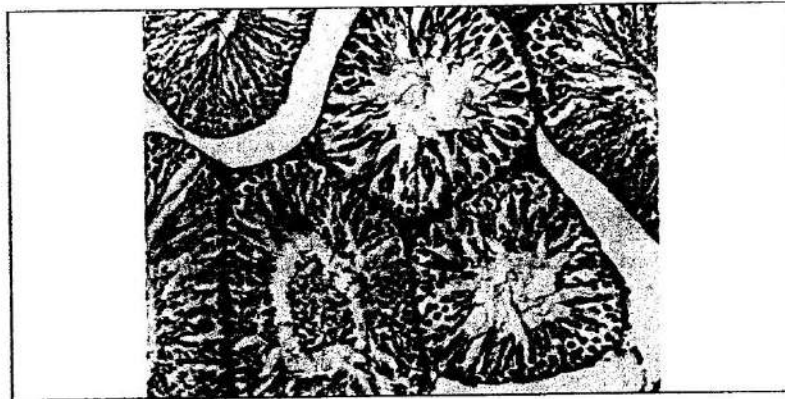


Fig. 4: Testicular tissue transplanted with MMSCs cultured under 3 % O<sub>2</sub> tension shows regeneration (200X)

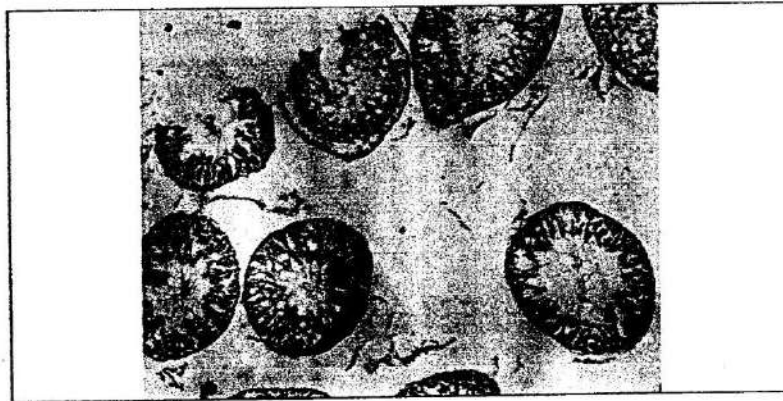


Fig. 5: Testicular tissue transplanted with MMSCs cultured under 21% O<sub>2</sub> tension shows no regeneration (200X)

Undifferentiated state of the stem cells was one of the main purposes in this research. Halim and co-workers (2010) stated that undifferentiation is one of the unique characteristics of stem cells that distinguish them from other cells of body organ. The existence of this unique undifferentiation made in vitro cultured stem cells capable to be viable longer compared to their progenitor or matured stem cells. This instance was based on researches conducted by Elliason et al. (2010) and Takubo (2011) revealed that for the transplanted stem cells viability to be maintained, the cultured stem cells have to be undifferentiated.

Undifferentiated state of stem cells, if fulfilled, further on will cause stem cells to be predicted to be able to differentiate into whatever cells needed when transplanted. This instance became the specialty of stem cells in vivo, that in the undifferentiated state, those cells can differentiate into more than one type of cells (multipotent/pluripotent). Therefore if undifferentiated state could be maintained in vitro, the expected capacity to differentiate into whatever cells needed upon transplantation will be fulfilled.

It can be said that stem cells inside the body are cells that have not had specific shapes and functions as in other cells of body organ (Halim *et al.*, 2010) such as myocardium cells to pulsate, neuron cells as the transmitter of nerve impulses and langerhans cells of pancreas to produce insulin (Guz *et al.*, 2001).

According to Lin *et al.* (2008), stem cells are different from the three cell types, stem cells have not had specific functions such as to pulsate, to transmit nerve impulses, producing hormones or other functions. Based on various researches it is said that stem cells population in a mature tissue inside the body showed an inactive cell population that start functioning in certain time and condition.

The existence of MMSCs *in vitro* in this research as inactive undifferentiated cells was aimed to keep regeneration continuity of tissue or organ composer. This instance can be done according to the undifferentiated stem cells capacity for later differentiation into body cells which are needed (Jones and Wagers, 2008). As the example ectoderm cells can differentiate into neuron cells (Halim *et al.*, 2010) or the endoderm cells such as hepatocyte cells (Schwartz *et al.*, 2002) and germline cells (spermatogonia, sertoli and Leydig cells) and germline tissues (seminiferous tubules) (Kilani, 2009).

#### **Conclusion**

The conclusion of this research showed that hypoxia precondition using 3% O<sub>2</sub> concentration very support MMSCs to remain adaptive after being transplanted to patient with spermatogenesis impairment, in addition to the testicle tissue improvement.

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