

eric priyo prasetyo <eric-p-p@fkg.unair.ac.id>

# Submission letter

2 messages

**Dental Journal (Majalah Kedokteran Gigi)** <dental\_journal@fkg.unair.ac.id> Fri, Fe To: eric priyo prasetyo <eric-p-p@fkg.unair.ac.id>, Eric Prasetyo <eppfkgunair@gmail.com>

Fri, Feb 11, 2022 at 11:25 AM

## Dear

Dr. Eric Priyo Prasetyo, drg., M.Kes., Sp.KG

Thank you for your article submission to Dental Journal (Majalah Kedokteran Gigi) and we gladly inform you that your article:

In-vitro antibacterial efficacy between calcium hydroxide-iodoform and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis* 

Authors: Eric Priyo Prasetyo,<sup>1</sup> Devi Eka Juniarti,<sup>1</sup> Galih Sampoerno,<sup>1</sup> Dian Agustin Wahjuningrum,<sup>1</sup> Ananta Tantri Budi,<sup>1</sup> Dyanita Hasri, <sup>2</sup> Evelyn Tjendronegoro<sup>3</sup>

The article is under review according to the provisions of the issuance of the Dental Journal (Majalah Kedokteran Gigi).

Certainty of revision or rejection of the article will be notified approximately one month after this notification.

If the article is accepted for publication the author is subject to administrative fees as follows:

- Proofreading fees of GBP 20\* per 1000 words.

(\*prices can change at any time, and the value of the rupiah follows the last exchange rate at that time and does not include transfer fee).

Please find the attached cover letter and copyright form, please fill it out and sign it by all authors and send it back by email.

Thank you for considering this journal as a venue for your work.

Best Regards, Chief Editor,

Muhammad Dimas Aditya Ari, drg., M.Kes., Sp.Pros

# Dental Journal (Majalah Kedokteran Gigi)

http://e-journal.unair.ac.id/MKG

Faculty of Dental Medicine, Universitas Airlangga

JI. Mayjen Prof. Dr. Moestopo 47 Surabaya 60132 INDONESIA Phone/Fax: +62 31 5039478

Cover\_letter\_n\_copyright\_form.docx
 16K

eric priyo prasetyo <eric-p-p@fkg.unair.ac.id> To: "Dental Journal (Majalah Kedokteran Gigi)" <dental\_journal@fkg.unair.ac.id> Fri, Feb 11, 2022 at 7:20 PM

Dear Chief Editor,

Thank you for your email regarding our manuscript submission, Hereby we attach the cover letter and copyright transfer form in the attachment.

Thank you once again.

Best regards, Dr. Eric Priyo Prasetyo, drg., M.Kes., Sp.KG(K). & team. [Quoted text hidden]

Cover\_letter\_n\_copyright\_form Eric CHI CHBS EF PG.pdf



eric priyo prasetyo <eric-p-p@fkg.unair.ac.id>

# Editor decision: revision required

3 messages

**Dental Journal (Majalah Kedokteran Gigi)** <dental\_journal@fkg.unair.ac.id> To: eric priyo prasetyo <eric-p-p@fkg.unair.ac.id>, Eric Prasetyo <eppfkgunair@gmail.com> Tue, Feb 22, 2022 at 9:53 AM

Dear

# Dr. Eric Priyo Prasetyo,drg.,M.Kes.,Sp.KG(K)

We have reached a decision regarding your submission to Dental Journal (Majalah Kedokteran Gigi),

# In-vitro antibacterial efficacy between calcium hydroxide-iodoform and calcium hydroxide-barium sulfate root canal dressings on Enterococcus faecalis and Porphyromonas gingivalis Our decision is to: Revise your manuscript

<u>Please revise according to the reviewer comments, highlight the text with color on the changes made and make a response letter (attached).</u> Please send the revised manuscript within a week (**3 March 2022**).

Determination of acceptance of the manuscript based on the revised results sent.

Best Regards, Chief Editor,

Muhammad Dimas Aditya Ari, drg., M.Kes., Sp.Pros

Dental Journal (Majalah Kedokteran Gigi) http://e-journal.unair.ac.id/MKG

Faculty of Dental Medicine, Universitas Airlangga JI. Mayjen Prof. Dr. Moestopo 47 Surabaya 60132 INDONESIA Phone/Fax: +62 31 5039478

4 attachments	
Response letter.docx 22K	
review 1 Eric Priyo article_In-vitro antibacterial efficacy between calcium (PA2). 160K	
review 1 Eric Priyo article_In-vitro antibacterial efficacy between calcium (PA1) 622K	pdf
review 1 Eric Priyo article_In-vitro antibacterial efficacy between calcium (PP)p	df

https://mail.google.com/mail/u/0/?ik=433d77c8df&view=pt&search=all&permthid=thread-f:1725430156694792447&simpl=msg-f:17254301566947924... 1/2

To: "Dental Journal (Majalah Kedokteran Gigi)" <dental journal@fkg.unair.ac.id>

Dear Dr. Muhammad Dimas Aditya Ari,

Thank you for your decision about the revisions for our manuscript. Please find the revised manuscript and response letter in the attachment. We hope the revisions will satisfy the reviewers and managing editor. Thank you once again for your consideration.

Best regards, Dr. Eric & team.

[Quoted text hidden]

## 

**Dental Journal (Majalah Kedokteran Gigi)** <dental\_journal@fkg.unair.ac.id> To: eric priyo prasetyo <eric-p-p@fkg.unair.ac.id> Tue, Mar 1, 2022 at 8:47 AM

Dear Dr. Eric Priyo Prasetyo,drg.,M.Kes.,Sp.KG(K)

Thank you for submitting a revised draft of your manuscript. The manuscript will be re-evaluated by reviewers based on the revisions you make.

Best wishes, Dental Journal (Majalah Kedokteran Gigi) http://e-journal.unair.ac.id/MKG

Faculty of Dental Medicine, Universitas Airlangga JI. Mayjen Prof. Dr. Moestopo 47 Surabaya 60132 INDONESIA Phone/Fax: +62 31 5039478

[Quoted text hidden]

# ASSESSMENT FORM OF ORIGINAL ARTICLE DENTAL JOURNAL (MAJALAH KEDOKTERAN GIGI)

Manuscript title: In-vitro antibacterial efficacy between calcium hydroxide-iodoform and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis* 

Receive date:

Send date:

	Please comments written in English		
	REVIEW	YES*	NO*
1. Ha	s this text ever been published on other media?		V
Co	mment:		
sever iodof	text ever been published on other media but, there have been ral studies to test the antibacterial efficacy of calcium hydroxide- orm for root canal dressing against Enterococcus bacteria he title appropriate, concise, clear, and describes the contribution of scientific		V
	velopment? (maximum 10 words, covering the variables studied)		
	mment:		
	e tithe too long, please make the short title likes this :		
	licium hydroxide-iodoform and calcium hydroxide-barium sulfate		
	tibacterial efficacy for root canal dressings.		
	search report:		
a)	Introduction includes the background clearly?	V	
,	Comment: Please add the more clear function of iodoform and barium sulfate		
	to increase the antibacterial efficacy		
b)	The purpose clearly?	V	
	Comment: The purpose is clear		
c)	Methods and research design in accordance with the purpose of the study?	V	
	<u>Comment</u> :		
	Please write the spesific nomenclature/strain of E.faecalis and P gingivalis		
	( E.faecalis ATCC, P.gingiivalis ATCC) that use in this research.		
d)	The research procedure is described precisely and in detail, thus ensuring	V	
	internal/external validity?		
	Comment: The procedure is clear and detail		
e)	The results can answer the research question?	V	
	Comment: The result can answer the research question		
f)	- The discussion does not repeat the results?		V

- Aligned with the scope of the study and compared with similar research		
results?		
- Explain the meaning of research results in answering the problem?		
<u>Comment</u> :		
1. There are several statement not necessary in discussion, i.e in paragraf 1		
and 3.		
2. Please compare this research with similar research result.		
g) References are aligned with the research material and use the literature of the		V
last 10 years?		
<u>Comment</u> :		
References aligned with the research material, but references number 23 :		
use journal at 2002 years.		
h) - The conclusion matches the title and the problem?	V	
- The research results contibute to the development of dentistry?		
- Perform synthesis base on similar research results that precede?		
<u>Comment</u> :		
Conclusions should be adjusted to the title and research objectives, the title mentions the difference but at the conclusion only mentions that calcium hydroxide-iodoform has a higher antimicrobial efficacy.		
i) References needs to be added/substacted**?		
<u>Comment</u> : References are already enough but Ithe old literature should be cha	nge	
4. Is there a section that needs to be added/summarized**?		
<u>Comment</u> : No.		

## Note:

1. \*) Put a check mark  $(\sqrt{})$ , \*\*) Cross the unnecessary ones

2. Correction can be made directly on the script

3. If the inquiry form is lacking, please write on the additional sheet

## **Recommendation for Editors**

[......] 1. The script can be published without changes.
 [V] 2. The manuscript can be published with corrections according to the direction of the Reviewer (suggestions for improvement please write directly to the script).
 <u>Comment</u>:
 [......] 3. The manuscript could not be published.
 <u>Reason</u>:

Date: Pebruary. 21, 2022

Reviewer,

[.]

# ASSESSMENT FORM OF ORIGINAL ARTICLE DENTAL JOURNAL (MAJALAH KEDOKTERAN GIGI)

Manuscript title: In-vitro antibacterial efficacy between calcium hydroxide-iodoform and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis* 

# Send date:21<sup>th</sup>February 2022 Please comments written in English

Receive date: 11<sup>th</sup>February 2022

YES\*

NO\*

 $\sqrt{}$ 

	Please comments written in English
	REVIEW
1.	Has this text ever been published on other media?
	Comment:
	This text never been published on other media
2.	Is the title appropriate, concise, clear, and describes the contribution

Thi	s text never been published on other media		
2. Is t	he title appropriate, concise, clear, and describes the contribution of scientific	$\checkmark$	
dev	velopment? (maximum 10 words, covering the variables studied)		
<u>Co</u>	<u>mment</u> :		
Th	e title is appropriate, concise, cler and describes the contribution of scientific		
dev	velopment		
3. Re	search report:		
a)	Introduction includes the background clearly?	$\checkmark$	
	Comment:		
	Introduction includes the background clearly		
b)	The purpose clearly?	$\checkmark$	
	Comment:		
	The purpose is clear		
c)	Methods and research design in accordance with the purpose of the study?	$\checkmark$	
	Comment:		
	The methods and research design has been appropiate with the purpose of		
	the study		
d)	The research procedure is described precisely and in detail, thus ensuring	$\checkmark$	
	internal/external validity?		
	Comment:		
	The research procedure has described precisely, and covers internal/external		
	validity		
e)	The results can answer the research question?	$\checkmark$	
	Comment:		
L			

e discussion does not repeat the results? igned with the scope of the study and compared with similar research sults? plain the meaning of research results in answering the problem? <u>mment</u> : The discussion does not repeat the results, has been aligned the scope of the study and has been compared with similar research ilts discussion has explained the meaning of reseach results in answering problem erences are aligned with the research material and use the literature of the	√	
igned with the scope of the study and compared with similar research sults? plain the meaning of research results in answering the problem? <u>mment</u> : The discussion does not repeat the results, has been aligned the scope of the study and has been compared with similar research its discussion has explained the meaning of reseach results in answering problem erences are aligned with the research material and use the literature of the		
sults? plain the meaning of research results in answering the problem? <u>mment</u> : The discussion does not repeat the results, has been aligned the scope of the study and has been compared with similar research ilts discussion has explained the meaning of reseach results in answering problem erences are aligned with the research material and use the literature of the	√	
<u>mment</u> : The discussion does not repeat the results, has been aligned the scope of the study and has been compared with similar research ilts discussion has explained the meaning of reseach results in answering problem erences are aligned with the research material and use the literature of the	1	
the scope of the study and has been compared with similar research lits discussion has explained the meaning of reseach results in answering problem erences are aligned with the research material and use the literature of the	1	
ilts discussion has explained the meaning of reseach results in answering problem erences are aligned with the research material and use the literature of the	√	
discussion has explained the meaning of reseach results in answering problem erences are aligned with the research material and use the literature of the	V	
problem erences are aligned with the research material and use the literature of the	V	
erences are aligned with the research material and use the literature of the	1	
	$\checkmark$	
10 years?		
nment: References are aligned with the research material and use the		
ature of the last 10 years		
e conclusion matches the title and the problem?	$\checkmark$	
e research results contibute to the development of dentistry?		
erform synthesis base on similar research results that precede?		
<u>ıment</u> :		
conclusion has matched the title and the problem, but need more		
cription for clinical		
ds		
research results can contribute to the development of dentistry,		
escially endodontic, but need further research , both in situ and in vivo		
escially endodontic, but need further research , both in situ and in vivo earch		
-		
earch		
earch conclusion has perform synthesis base on similar research results that		
es	onclusion has perform synthesis base on similar research results that	onclusion has perform synthesis base on similar research results that

# Note:

- 1. \*) Put a check mark ( $\sqrt{}$ ), \*\*) Cross the unnecessary ones 2. Correction can be made directly on the script
- 3. If the inquiry form is lacking, please write on the additional sheet

# **Recommendation for Editors**

		ublished with corrections according to the dire tent please write direcly to the script).	ection of the Reviewer
	.Please	see	to
	manuscript		
[] 3.	The manuscript could not	be published.	
	<u>Reason</u> :		

.

Date: .20th February 2020

Reviewer,

#### Original article

In-vitro antibacterial efficacy between calcium hydroxideiodoform and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis* 

#### ABSTRACT

Background: A successful endodontic treatment is inseparable from the right choice of root canal dressing. The right choice of medicaments would result in patient satisfaction. Enterococcus faecalis (E. faecalis) and Porphyromonas gingivalis (P. gingivalis) are usually found in failed root canal treatments. Calcium hydroxide is a gold standard dressing which create an alkaline environment in the root canal and have a bactericidal effect. Commercially, there are calcium hydroxide dressings with supporting additions, including calcium hydroxideiodoform (CH-Iodoform) and Calcium hydroxide-barium sulfate (CH-Barium Sulfate). Purpose: This study aimed to determine the antibacterial efficacy of CH-Iodoform and CH-Barium Sulfate on E. faecalis and P. gingivalis. Methods: CH-Iodoform and CH-Barium Sulfate was obtained commercially. E. faecalis and P. gingivalis were obtained from stock culture taken from the root canal of failed endodontic treatment. E. faecalis and P. gingivalis were cultured in petri dishes and for each bacterium, 12 wells were made in the media, 6 wells were used for CH-Iodoform group and 6 wells were used for CH-Barium Sulfate group. CH-Iodoform and CH-Barium Sulfate were deployed in the wells in E. faecalis and P. gingivalis cultured medium in the petri dishes. After incubation, the inhibition zone diameters were measured. Independent t-test was used for analysis and significance level was set at 5%. Results: There is a significant antibacterial efficacy between CH-Iodoform and CH-Barium Sulfate on E. faecalis and P. gingivalis (p=0.00001). Conclusion: CH-Iodoform has a better antibacterial efficacy than CH-Barium Sulfate on both E. faecalis and P. gingivalis.

*Keywords:* Enterococcus faecalis; Porphyromonas gingivalis; calcium hydroxide; iodoform; barium sulfate; patient satisfaction.

#### INTRODUCTION

The right choice of root canal dressing to eliminate bacteria in the root canal is important for a successful endodontic treatment. A successful endodontic treatment would result in patient's satisfaction. Many attempts were conducted to increase the success of root canal treatment, including finding the efficient instrumentation, effective cleaning,<sup>1,2</sup> antibacterial dressing and irrigation materials.<sup>3,4</sup>

Root canal treatment has a high success rate, but in some cases, there are failures. Isolated bacteria from root canal treatment failures and the prevalence are about 45.8-77% caused by *E. faecalis* and 28.17% caused by *P. gingivalis* in the root canal system.<sup>5</sup> Both microorganisms are among the ones that survive disinfecting protocol.<sup>6</sup>

*E. faecalis* can invade dentine tubules and resistence and periapical abnormalities which occur several months or years after root canal treatment.<sup>7</sup> *P. gingivalis* can survive in the extraradicular area mostly in the area approximate with root surface, and responsible for periodontitis and peri-radicular lesions.<sup>8</sup> *P. gingivalis* also contribute to root canal treatment failures, through its byproduct, lipopolysaccharide which effect biological processes, inflammation and tissue destruction.<sup>9</sup>

Different substances have been added to calcium hydroxide to increase its efficacy. The added substances are distilled water, saline, propylene glycol, chlorhexidine, glycerin, iodoform, barium sulfate, corticosteroids, antibiotics, anesthetic solution, methyl cellulose, and detergents. Iodoform has been added to calcium hydroxide to work with different bacterial characteristic.<sup>10</sup> Barium sulfate is added to calcium hydroxide aside from its antibacterial effect, it functions to increase radiopacity.<sup>11</sup>

Root canal treatment failure may happen even after dressing with calcium hydroxide. In this research, the efficacy of calcium hydroxide addition with iodoform (CH-Iodoform) and barium sulfate (CH-Barium Sulfate) was analyzed on *E. faecalis* and *P. gingivalis*. Based on this background, the authors would like to purposely compare the antibacterial efficacy between CH-Iodoform and CH-Barium Sulfate root canal dressing on *E. faecalis* and *P. gingivalis*.

Commented [Ta1]: ...protocol6.

**Commented [Ta2]:** ...can spread to preriradicular which causes the formation of lesi periradiculer after root canal treatment<sup>7</sup>.

Commented [Ta3]: periradiculer

Commented [Ta4]: ..lesion<sup>8</sup>

**Commented [Ta5]:** ...if done incorrectly, depending on right instrumentation and irrigation to remove smear layer anorganic and organic

#### MATERIALS AND METHODS

This is an experimental laboratory study, conducted in Conservative Dentistry Section, Universitas Airlangga Dental Hospital and Microbiology Laboratory, Research Center, Faculty of Dental Medicine, Universitas Airlangga. Materials used in this research were commercially available calcium hydroxide dressings with Iodoform (Meta Biomed, Korea) and Barium Sulfate (Meta Biomed, Korea). *Enterococcus faecalis* and *Porphyromonas gingivalis* bacteria used in this study were stock bacteria previously cultured from failed endodontic treatment.

The method used in this study was agar diffusion method using Mueller Hinton (MH) agar and Brain Heart Infusion (BHI) broth. The method is according to Alharthi et al. (2021) and Balouiri et al. (2016) with modifications.<sup>12,13</sup> The media were allocated for 4 groups of experiments. In the first group, 12 wells were prepared for CH-Iodoform (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*), and in the second group, 12 wells were prepared for CH-Barium Sulfate dressing (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*).

Both *E. faecalis* and *P. gingivalis* bacterial culture from stock was moved into separate reaction tube, each containing BHI broth and stirred. Incubation was done for both culture at 37°C for 24 hours in anaerobic condition. After 24 hours, *E. faecalis* and *P. gingivalis* bacterial culture in BHI broth was taken 0.5 ml using micro pipette, and poured into another reaction tube containing BHI broth until equal with 0.5 McFarland standard of turbidity.

The bacterial cultures were taken from BHI broth using sterile cotton swab and swabbed on the surface of each MH agar allocated for *E. faecalis* and *P gingivalis*. Antibacterial test was conducted by making the wells for the tested dressing materials (CH-Iodoform and CH-Barium Sulfate). The samples were incubated at  $37^{\circ}$ C for 48 hours in anaerobic condition.

After 48 hours, measurements were conducted on antibacterial efficacy through inhibition zone measurement using a Vernier Digital Caliper (Mitutoyo, Japan). The clear zones of inhibition around the wells were measured, revealing no bacterial growth. Data of measurements on inhibition zone diameters (in millimeters) were collected for each sample wells.

Inhibition zones data were analyzed statistically and significance level was set at 5%. Data normality was tested using Shapiro Wilk test. The significance was tested using independent t-test.

Commented [Ta6]: ... from patients who failed endodotic treatment

Commented [Ta7]: This Method just for P. gingivalis

Commented [Ta8]: ..need explain how the modification is conducted

#### RESULTS

The number of replications (n) for each treatment group was 6. Mean and standard deviation of inhibition zone diameter of CH-Iodoform and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* are shown in Table 1. The mean of *E. faecalis* inhibition zone from CH-Iodoform was 11.8125 mm and CH-Barium Sulfate was 6.3750 mm. The mean of *P. gingivalis* inhibition zone from CH-Iodoform was 12.7875 mm and CH-Barium Sulfate was 6.6750 mm.

#### Table 1:

Mean and standard deviation from inhibition zone diameter of CH-Iodoform and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* 

Groups	n	E. faecalis	P. gingivalis
		Mean <u>+</u> SD	Mean <u>+</u> SD
CH-Iodoform	6	11.8125 <u>+</u> 1.32001	12.7875 <u>+</u> 1.34961
CH-Barium Sulfate	6	6.3750 <u>+</u> 0.19494	6.6750 <u>+</u> 0.51865

n = replication

SD = standard deviation

 Table 2: Significance between inhibition zone diameter of CH-Iodoform and CH-Barium

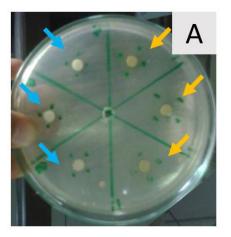
 Sulfate on *E. faecalis* and *P. gingivalis*

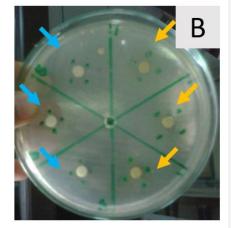
Groups	E. faecalis	P. gingivalis	
	CH-Iodoform	CH-Iodoform	
E. faecalis	0.00001*	-	
CH-Barium Sulfate			
P. gingivalis	-	0.00001*	
CH-Barium Sulfate			

Independent t-test was used in this study to check the significance between CH-Iodoform group and CH-Barium Sulfate group on *E. faecalis* and *P. gingivalis*. The significance between inhibition zone diameter of CH-Iodoform and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* growth is shown in Table 2. We found a significant difference between the two treatment groups on both bacteria.

The CH-Iodoform group has higher antibacterial efficacy with wider antibacterial clear zone in both bacteria (p = 0.00001). CH-Barium Sulfate group has lower antibacterial efficacy with narrower antibacterial clear zone in both bacteria. There is no significant difference of antibacterial activity of CH-Iodoform between *E. faecalis* and *P. gingivalis* (p = 0.11726). There is also no significant difference of antibacterial activity of CH-Barium Sulfate between *E. faecalis* and *P. gingivalis* (p = 0.10712).

Commented [Ta9]: ???





#### Figure 1.

Agar diffusion assay containing *E. faecalis* (A) and *P. gingivalis* (B). The green dots indicate the inhibition area of each well. CH-Iodoform is pointed by yellow arrows, and CH-Barium Sulfate is pointed by blue arrows.

#### DISCUSSION

*Enterococcus faecalis* and *Porphyromonas gingivalis* were used in this study because these bacteria are opportunistic, living in facultative anaerobic condition, found in pathogenic state, and associated with failed root canal treatment.<sup>5</sup> These bacteria also have a major role in persistent root canal infections, can survive in the root canals and resistant to commonly used intracanal dressings.<sup>14</sup>

Antibacterial efficacy of CH-Iodoform and CH-Barium Sulfate dressing materials on *E. faecalis* and *P. gingivalis* growths were experimentally checked with agar diffusion method and inhibition clear zone were measured on the media. Inhibition zone is a laboratory calculation method to check whether a material have the ability to inhibit bacterial growth.<sup>13</sup> The diameter of clear zones would describe the strength of such material to inhibit bacterial growth, the stronger this material inhibit, the larger the clear zones would appear.

In this study Mueller Hinton agar was used because this media can grow *E. faecalis* and *P. gingivalis* actively and sensitive to drugs effect. Inhibition zones appeared on both groups of CH-Iodoform and CH-Barium Sulfate. This showed that each of these dressing materials have the ability to inhibit both *E. faecalis* and *P. gingivalis*.

The antibacterial properties of both root canal dressing materials mainly come from calcium hydroxide, which mechanism involve hydroxyl ion to kill bacteria by protein denaturation, cytoplasmic membrane and DNA destruction will physically inhibit this bacterial growth in the root canal systems.<sup>15</sup> Antibacterial activity is connected with alkali formation which can destruct lipid, the protein structure of bacteria and nucleic acid.<sup>16</sup> Direct contact of hydroxyl ion in alkaline pH with cytoplasmic membrane will destruct the hydrogen chain of protein polypeptide. Contact of hydroxyl ion with DNA will result in replication inhibition, causing lethal mutation. DNA is sensitive to temperature and pH change. In alkali condition DNA structure and function will break and leading to bacterial cell death. Alkali condition from calcium hydroxide would impact on the surrounding tissues, including healing, anti-inflammation<sup>17</sup> and cytotoxic which lead to apoptosis.<sup>18,19</sup>

The result of this study showed a significant difference between CH-Iodoform and CH-Barium Sulfate. The ability of CH-Iodoform to inhibit *E. faecalis* and *P. gingivalis* growths are significantly greater than CH-Barium Sulfate root canal dressing. In CH-Iodoform dressing, the iodoform substance will release iodine with high reactivity to promote protein oxidation. Iodoform functions as disinfectant and infection control. Iodoform and calcium hydroxide can diffuse into dentinal tubules for added disinfection and eventually will stimulate tissue repair.

**Commented [Ta10]:** It is clear CH-lodoform to inhibit E.faecalis and P. gingivalis because CH with ions OH can change the surrounding tissue to alkali conditions, lodoform will release iodine with high reactivity to promote protein oxidation. Thus the combination CH and lodoform will strengthen each other.

Commented [Ta11]: ...and eliminate the bacteri

In CH-Barium Sulfate, barium sulfate can diffuse into dentinal tubule and also perform antibacterial activity. In this study the antimicrobial strength is not as good as CH-Iodoform. This may be caused by the form and structure of barium sulfate used in the mixture. Previous study on antimicrobial of barium sulfate showed that barium sulfate in micron particulate form would have potent antimicrobial properties.<sup>20</sup> Barium sulfate is generally used for its radiopacity effect on radiographic examination.<sup>11</sup>

*E. faecalis* and *P. gingivalis* can be killed with high pH level. High pH or an extreme alkaline environment would disturb the survival of most bacteria.<sup>21,22</sup> However, there are some studies stated that pH of higher than 11.5 is required for potent disinfection.<sup>23</sup> This study showed inhibition zone of CH-Iodoform is two folds higher than CH-Barium Sulfate, this may be caused by the combination of these two materials, calcium hydroxide and iodoform would create synergistic effect on the antimicrobial activity to inhibit *E. faecalis* growth. Even though this dressing material won't eliminate *E. faecalis or P. gingivalis*, the material is expected to weaken the bacteria, and eventually body defense mechanism will be able eliminate these bacteria and their byproducts.

Many researches have been done to found novel, biocompatible, antimicrobial and regenerating root canal dressing materials in order to support successful root canal treatment and enhance patient's satisfaction. The success of root canal treatment aside from the right choice of dressing, there are other factors such as the root canal systems complexity, diffusion ability of dressing materials within the dentinal tubules, and the effect on mixed cultured bacteria or biofilm formation, and bacterial resistance, which need to be evaluated in further studies. As there are many microorganisms involved in failed root canal treatments, <sup>24-26</sup> even after biomechanical and chemical instrumentations during treatment, individual and personalized assessment of these bacteria or biofilm is needed for ideal treatment. In conclusion, calcium hydroxide-Iodoform root canal dressing has higher antimicrobial efficacy on both *E. faecalis* and *P. gingivalis*. However, further studies need to be done regarding the effect on periodontal and periapical tissues with more complexities.

#### ACKNOWLEDGEMENT

The authors would like to thank the Publication Center, Dental Hospital, and Research Center, Faculty of Dental Medicine, Universitas Airlangga for the given facility. **Commented [Ta12]:** If we add the barium sulfat to CH, must be considered how the nature of the material as the flow rate etc

**Commented [Ta13]:** The limitations of this research just done as in vitro study. There are many factors to consider if used the material directly on teeth, both in situ and in vivo study.

#### REFERENCES

- Santoso CMA, Samadi K, Prasetyo EP, Wahjuningrum DA. Root canal cleanliness between mangosteen peel extract irrigant and NaOCl 2.5%. Conservative Dentistry Journal. 2020; 10(1): 40-3. doi: 10.20473/cdj.v10i1.2020.40-43.
- Juniarti DE, Kusumaningsih T, Soetojo A, Prasetyo EP, Sunur YK. Antibacterial activity and phytochemical analysis of ethanolic purple leaf extract (*Graptophyllum Pictum L. griff*) on *Lactobacillus acidophilus*. Mal J Med Health Sci. 2021; 17(Suppl2): 71-3.
- Astuti RHN, Samadi K, Prasetyo EP. Antibacterial activity of *Averrhoa bilimbi Linn*. leaf extract against Enterococcus faecalis. Conservative Dentistry Journal. 2016; 6(2): 93-8. doi: 10.20473/cdj.v6i2.2016.93-98.
- Harseno S, Mooduto L, Prasetyo EP. Antibacterial potency of kedondong Bangkok leaves extract (*Spondias dulcis Forst.*) against Enterococcus faecalis bacteria. Conservative Dentistry Journal. 2016; 6(2): 110-6. doi: 10.20473/cdj.v6i2.2016.110-116.
- Prada I, Mico-Munoz P, Giner-Lluesma T, Mico-Martinez P, Collado-Castellano N, Manzano-Saiz A. Influence of microbiology on endodontic failure. Literature review. Med Oral Patol Oral Cir Bucal. 2019; 24(3): e364-72. doi: 10.4317/medoral.22907.
- Del Fabbro M, Samaranayake LP, Lolato A, Weinstein T, Taschieri S. Analysis of the secondary endodontic lesions focusing on the extraradicular microorganisms: an overview. J Investig Clin Dent. 2014; 5(4): 245-54. doi: 10.1111/jicd.12045.
- Kaiwar A, Nadig G, Hegde J, Lekha S. Assessment of antimicrobial activity of endodontic sealers on Enterococcus faecalis: an in vitro study. World J Dent. 2012; 3(1): 26-31. doi: 10.5005/jp-journals-10015-1123.
- Noguchi N, Noiri Y, Narimatsu M, Ebisu S. Identification and localization of extraradicular biofilm-forming bacteria associated with refractory endodontic pathogens. Appl Environ Microbiol. 2005; 71:8738-43. doi: 10.1128/AEM.71.12.8738-8743.2005.
- Kuntjoro M, Prasetyo EP, Cahyani F, Kamadjaja MJK, Hendrijantini N, Laksono H, Rahmania PN, Ariestania V, Nugraha AP, Ihsan IS, Dinaryanti A, Rantam FA. Lipopolysaccharide's cytotoxicity on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0048. doi:

10.1590/pboci.2020.153.

- Estrela C, Estrela CRDA, Hollanda ACB, Decurcio DDA, Pecora JD. Influence of iodoform on antimicrobial potential of calcium hydroxide. J Appl Oral Sci. 2006; 14(1): 33-7. doi: 10.1590/S1678-77572006000100007.
- Orucoglu H, Cobankara FK. Effect of unintentionally extruded calcium hydroxide paste including barium sulfate as a radiopaquing agent in treatment of teeth with periapical lesions: Report of a case. J Endod. 2008; 34(7): 888-91. doi: :10.1016/j.joen.2008.04.012.
- 12. Alharthi SS, Binshabaib M, Almasoud NS, Shawky HA, Aabed KF, Alomar TS, Albrekan AB, Alfaifi AJ, Melaibari AA. Myrrh mixed with silver nanoparticles demonstrates superior antimicrobial activity against *Porphyromonas gingivalis* compared to myrrh and silver nanoparticles alone. Saudi Dent J. 2021; 33: 890-6. doi: 10.1016/j.sdentj.2021.09.009.
- 13. Balouiri M, Sadiki M, Ibnsouda SK. Methods for in vitro evaluating antimicrobial activity: a review. J Pharm Anal. 2016; 6(2): 71-9. doi: 10.1016/j.jpha.2015.11.005.
- Jhamb S, Singla R, Kaur A, Sharma J, Bhushan J. An in vitro determination of antibacterial effect of silver nanoparticles gel as an intracanal medicament in combination with other medicaments against Enterococcus faecalis. J Conserv Dent. 2019; 22: 479-82. doi: 10.4103/JCD.JCD\_113\_20.
- Mohammadi Z, Dummer PMH. Properties and applications of calcium hydroxide in endodontics and dental traumatology. Int Endod J. 2011; 44(8): 697-730. doi: 10.1111/j.1365-2591.2011.01886.x.
- Athanassiadis B, Abbott PV, Walsh LJ. The use of calcium hydroxide, antibiotics, and biocides as antimicrobial medicaments in Endodontics. Australian Dental Journal. 2007; 52(1): 64-82. doi: 10.1111/j.1834-7819.2007.tb00527.x.
- 17. Prasetyo EP, Kuntjoro M, Cahyani F, Goenharto S, Saraswati W, Juniarti DE, Hendrijantini N, Hariyani N, Nugraha AP, Rantam FA. Calcium hydroxide upregulates interleukin-10 expression in time dependent exposure and induces osteogenic differentiation of human umbilical cord mesenchymal stem cells. Int J Pharm Res. 2021; 13(1): 140-5. Doi: 10.31838/ijpr/2021.13.01.023.
- Prasetyo EP, Widjiastuti I, Cahyani F, Kuntjoro M, Hendrijantini N, Hariyani N, Winoto ER, Nugraha AP, Goenharto S, Susilowati H, Hendrianto E, Rantam FA. Cytotoxicity of calcium hydroxide on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0044. doi:

10.1590/pboci.2020.141.

- Prasetyo EP, Kuntjoro M, Goenharto S, Juniarti DE, Cahyani F, Hendrijantini N, Nugraha AP, Hariyani N, Rantam FA. Calcium hydroxide increases human umbilical cord mesenchymal stem cells expressions of apoptotic protease-activating factor-1, caspase-3 and caspase-9. Clinical, cosmetic and investigational dentistry. 2021;13: 59-65. doi: 10.2147/CCIDE.S284240.
- Aninwene II G, Stout D, Yang Z, Webster TJ. Nano-BaSO4: a novel antimicrobial additive to pellethane. Int J Nanomedicine. 2013;8(1):1197-1205. doi: 10.2147/IJN.S40300.
- Jhajharia K, Parolia A, Shetty KV, Mehta LK. Biofilm in endodontics: a review. J Int Soc Prev Community Dent. 2015; 5: 1-12. doi: 10.4103/2231-0762.151956.
- Zancan RF, Vivan RR, Lopes MRM, Weckwerth PH, Andrade FB, Ponce JB, Duarte MAH. Antimicrobial activity and physicochemical properties of calcium hydroxide pastes used as intracanal medication. J Endod. 2016; 42(12): 1822-8. doi: 101016/j.joen.2016.08.017.
- Evans M, Davies JK, Sundqvist G, Figdor D. Mechanisms involved in the resistance of Enterococcus faecalis to calcium hydroxide. Int Endod J. 2002; 35(3): 221-8. doi: 10.1046/j.1365-2591.2002.00504.x.
- Dudeja PG, Dudeja KK, Srivastava D, Grover S. Microorganisms in periradicular tissues: Do they exist? A perennial controversy. J Oral Maxillofac Pathol. 2015; 19(3): 356-63. doi: 10.4103/0973-029X.174612.
- 25. Pereira RS, Rodrigues VAA, Furtado WT, Gueiros S, Pereira GS, Avila-Campos MJ. Microbial analysis of root canal and periradicular lesion associated to teeth with endodontic failure. Anaerobe. 2017; 48: 12-8. doi: 10.1016/j.anaerobe.2017.06.016.
- Siqueira JF, Antunes HS, Rocas IN, Rachid CTCC, Alves FRF. Microbiome in the apical root canal system of teeth with post-treatment apical periodontitis. PLoS ONE. 2016; 11(9): e0162887. doi: 10.1371/journal.pone.0162887.

Commented [Ta14]:

Commented [Ta15]: Page?

# ASSESSMENT FORM FOR MANAGING EDITOR (EDITING LANGUAGE AND FORMATTING SCRIPT)

Manuscript Title: In-vitro antibacterial efficacy between calcium hydroxide-iodoform and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis* 

Send date:

Receive date:

## Please comments written in English

	REVIEW	COMMENT
FO	RMAT	
ls f	the length of the script sufficient?	Yes
-	(10-12 pages, one and a half space, A4 format, 12 pt Times New Roman font)	
•	Sections of the contens on the script are proportional (The discussion is longer than the introduction	Yes
Tit	le	
•	In accordance with the problem, purpose and contain the main variable	Yes/
	Not too long (maximum of 10 words) with the first word starting with a capital letter	Yes
Ab	stract	
	A concise (maximum 250 words), single space, structured in one paragraph	Yes
•	Keywords correspond to major variables/ main concept in English	Yes
•	Keywords must be provided 3-5 words and/or a phrase and separated by a semicolon (;)	No
	The Abstract in Research Reports should consist of a single paragraph containing <b>Background:</b> , <b>Purpose:</b> , <b>Methods:</b> , <b>Results:</b> and <b>Conclusion:</b> written in bold type	Yes
Re	ferences	
•	Vancouver superscript	Yes
Fiç	gures and Tables	
•	In accordance with the style of the journal (the truth, the completeness of the title and description/ legend) and included reference	No
•	Number of figure/ tables in the research report and literature review maximum 4	No
•	Number of figure/ tables in the case report maximum 8	Yes
-	Figures/ tables are written separately with text	Yes

	1	
Re	ferences (Vancouver superscript)	
-	In accordance with the style of the journal (Vancouver superscript sistem)	Yes
•	The last 10 years	Yes
•	Primary reference ≥ 70% (journals, books, patent document)	Yes
•	Volume/ number and journal page already listed	Yes
•	Editions, Publisher, City and book pages are listed	Yes
•	In sequence on the text	Yes
•	Author name written all	Yes
	Consistency of the author's name increase	Yes
•	From the internet include website address and access time	Yes
•	Abbreviate journal title according to the dental index and medical index	Yes
La	nguage	
•	Not enumerative	No
•	No typos	No
•	Standard spelling	No
•	Standard sentence (subject, predicate, object)	No
•	One paragraph, one subject (more than two sentences)	No
	ORIGINAL ARTICLE	
FO	RMAT	
•	Systematic manuscript of the original research consists of the introduction, materials and methods, results, discussion ended conclusion, and references.	Yes
Int	roduction	
•	Empirical/ theoretical background	There is
•	Problems/ goals	There is
•	Purpose of the study	There is
Ма	terials and Methods	
•	Design (type, time, place of the study)	No
•	Sampling technique	No

Editing language and formatting script form Dental Journal (Majalah Kedokteran Gigi)

• [	Data analysis	No
Resu	lts	
• E	Exposure data	No
• /	Analyze results	No
Discu	ussion	
• [	Discussion does not repeat the results	No
• /	Aligned with the scope of the study and compared with similar research results?	No
• E	explain the meaning of research results and answer the problem	Yes
• C	Conclusion	There is
• S	Suggesstions	No

Note:

- 1. \*) Cross the unnecessary ones.
- 2. If there is no match between the author and the editor should be reconciled to obtain a solution.

# **Recommendation from Managing Editor**

[......] 1. The manuscript can be processed without repair by the author.

[v] 2. The manuscript can be processed by repair by the author, thet is on the part: this paper needs major improvement

in the quality aspect, please kindly check the comment box to boost the quality. Thank you

[.....] 3. The manuscript was rejected, because .....

.....

Date: 21 February 2022 Managing Editor,

Sign

#### Original article

In-vitro antibacterial efficacy between calcium hydroxideiodoform and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis <u>in</u> <u>vitro</u>*  Commented [Office1]: Over all, this paper needs Grammar correction, English editing and proofreading, please attached and send the certificate of language editing along with this manuscript

Formatted: Font: Italic

#### ABSTRACT

Background: A successful endodontic treatment is inseparable from the right choice of root canal dressing. The right choice of medicaments would result in patient satisfaction. Enterococcus faecalis (E. faecalis) and Porphyromonas gingivalis (P. gingivalis) are usually found in failed root canal treatments. Calcium hydroxide is a gold standard dressing which create an alkaline environment in the root canal and have a bactericidal effect. Commercially, there are calcium hydroxide dressings with supporting additions, including calcium hydroxide-iodoform (CH-Iodoform) and Calcium hydroxide-barium sulfate (CH-Barium Sulfate). Purpose: This study aimed to determine the antibacterial efficacy of CH-Iodoform and CH-Barium Sulfate on E. faecalis and P. gingivalis. Methods: CH-Iodoform and CH-Barium Sulfate was obtained commercially. E. faecalis and P. gingivalis were obtained from stock culture taken from the root canal of failed endodontic treatment, E. faecalis and P. gingivalis were cultured in petri dishes and for each bacterium, 12 wells were made in the media, 6 wells were used for CH-Iodoform group and 6 wells were used for CH-Barium Sulfate group. CH-Iodoform and CH-Barium Sulfate were deployed in the wells in E. faecalis and P. gingivalis cultured medium in the petri dishes. After incubation, the inhibition zone diameters were measured. Independent t-test was used for analysis and significance level was set at 5%. Results: There is a significant antibacterial efficacy between CH-Iodoform and CH-Barium Sulfate on E. faecalis and P. gingivalis (p=0.00001; p<0.05). Conclusion: CH-Iodoform has a better antibacterial efficacy than CH-Barium Sulfate on both E. faecalis and P. gingivalis.

**Keywords:** Enterococcus faecalis; Porphyromonas gingivalis; calcium hydroxide; iodoform; barium sulfate; patient satisfaction.

Commented [Office2]: Why they add CH iodoform and CHbarium sulfate? This sentence unclear

**Commented [Office3]:** compare the antibacterial efficacy between CH-Iodoform and CH-Barium Sulfate root canal dressing on *E. faecalis* and *P. gingivalis.* 

Commented [Office4]: Please add the design and setting of this study

**Commented [Office5]:** How determine the isolated bacteria was E.faecalis or P.gingivalis?

**Commented [Office6]:** The inhibition zone using diffusion method is not adequate, please add more methods such as MIC or MBC

Commented [Office7]: The conclusion and the objective of the study did not match

Commented [Office8]: Please follow the MesH keywords

#### **INTRODUCTION**

The right choice of root canal dressing to eliminate bacteria in the root canal is important for a successful endodontic treatment. A successful endodontic treatment would result in patient's satisfaction. Many attempts were conducted to increase the success of root canal treatment, including finding the efficient instrumentation, effective cleaning,<sup>1,2</sup> antibacterial dressing and irrigation materials.<sup>3,4</sup>

Root canal treatment has a high success rate, but in some cases, there are failures. Isolated bacteria from root canal treatment failures and the prevalence are about 45.8-77% caused by *E. faecalis* and 28.17% caused by *P. gingivalis* in the root canal system.<sup>5</sup> Both microorganisms are among the ones that survive disinfecting protocol.<sup>6</sup>

*E. faecalis* can invade dentine tubules and unreachable from chemo-mechanical preparation and dressings. This may cause unseen persistence and periapical abnormalities which occur several months or years after root canal treatment.<sup>7</sup> *P. gingivalis* can survive in the extra-radicular area mostly in the area approximate with root surface, and responsible for periodontitis and peri-radicular lesions.<sup>8</sup> *P. gingivalis* also contribute to root canal treatment failures, through its byproduct, lipopolysaccharide which effect biological processes, inflammation and tissue destruction.<sup>9</sup>

Different substances have been added to calcium hydroxide to increase its efficacy. The added substances are distilled water, saline, propylene glycol, chlorhexidine, glycerin, iodoform, barium sulfate, corticosteroids, antibiotics, anesthetic solution, methyl cellulose, and detergents. Iodoform has been added to calcium hydroxide to work with different bacterial characteristic.<sup>10</sup> Barium sulfate is added to calcium hydroxide aside from its antibacterial effect, it functions to increase radiopacity.<sup>11</sup>

Root canal treatment failure may happen even after dressing with calcium hydroxide. In this research, the efficacy of calcium hydroxide addition with iodoform (CH-Iodoform) and barium sulfate (CH-Barium Sulfate) was analyzed on *E. faecalis* and *P. gingivalis*. Based on this background, the authors would like to purposely compare the antibacterial efficacy between CH-Iodoform and CH-Barium Sulfate root canal dressing on *E. faecalis* and *P. gingivalis*.

Commented [Office9]: Please state the abbreviation first Commented [Office10]: Please state the abbreviation first

#### MATERIALS AND METHODS

This is an experimental laboratory study, conducted in Conservative Dentistry Section, Universitas Airlangga Dental Hospital and Microbiology Laboratory, Research Center, Faculty of Dental Medicine, Universitas Airlangga. Materials used in this research were commercially available calcium hydroxide dressings with Iodoform (Meta Biomed, Korea) and Barium Sulfate (Meta Biomed, Korea). *Enterococcus faecalis* and *Porphyromonas gingivalis* bacteria used in this study were stock bacteria previously cultured from failed endodontic treatment.

The method used in this study was agar diffusion method using Mueller Hinton (MH) agar and Brain Heart Infusion (BHI) broth. The method is according to Alharthi et al. (2021) and Balouiri et al. (2016) with modifications.<sup>12,13</sup> The media were allocated for 4 groups of experiments. In the first group, 12 wells were prepared for CH-Iodoform (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*), and in the second group, 12 wells were prepared for CH-Barium Sulfate dressing (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*).

Both *E. faecalis* and *P. gingivalis* bacterial culture from stock was moved into separate reaction tube, each containing BHI broth and stirred. Incubation was done for both culture at 37°C for 24 hours in anaerobic condition. After 24 hours, *E. faecalis* and *P. gingivalis* bacterial culture in BHI broth was taken 0.5 ml using micro pipette, and poured into another reaction tube containing BHI broth until equal with 0.5 McFarland standard of turbidity.

The bacterial cultures were taken from BHI broth using sterile cotton swab and swabbed on the surface of each MH agar allocated for *E. faecalis* and *P gingivalis*. Antibacterial test was conducted by making the wells for the tested dressing materials (CH-Iodoform and CH-Barium Sulfate). The samples were incubated at  $37^{\circ}$ C for 48 hours in anaerobic condition.

After 48 hours, measurements were conducted on antibacterial efficacy through inhibition zone measurement using a Vernier Digital Caliper (Mitutoyo, Japan). The clear zones of inhibition around the wells were measured, revealing no bacterial growth. Data of measurements on inhibition zone diameters (in millimeters) were collected for each sample wells.

Inhibition zones data were analyzed statistically and significance level was set at 5%. Data normality was tested using Shapiro Wilk test. The significance was tested using independent t-test.

Commented [Office11]: Please add the ethical clearance number and statement clearly

**Commented [Office12]:** How determine the isolated bacteria was E.faecalis or P.gingivalis? please provide this kind of information

Commented [Office13]: What kind of modification, please state specificly

**Commented [Office14]:** How determine the sample size and how to select the sample?

Commented [Office15]: The inhibition zone using diffusion method is not adequate, please add more methods such as MIC or MBC

**Commented [Office16]:** Please add the statistical analysis software that have been used

#### RESULTS

The number of replications (n) for each treatment group was 6. Mean and standard deviation of inhibition zone diameter of CH-Iodoform and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* are shown in Table 1. The mean of *E. faecalis* inhibition zone from CH-Iodoform was 11.8125 mm and CH-Barium Sulfate was 6.3750 mm. The mean of *P. gingivalis* inhibition zone from CH-Iodoform was 12.7875 mm and CH-Barium Sulfate was 6.6750 mm.

#### Table 1:

Mean and standard deviation from inhibition zone diameter of CH-Iodoform and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* 

Groups	Groups n		P. gingivalis
		Mean <u>+</u> SD	Mean <u>+</u> SD
CH-Iodoform	6	11.8125 <u>+</u> 1.32001	12.7875 <u>+</u> 1.34961
CH-Barium Sulfate	6	6.3750 <u>+</u> 0.19494	6.6750 <u>+</u> 0.51865
n = replication			

SD = standard deviation

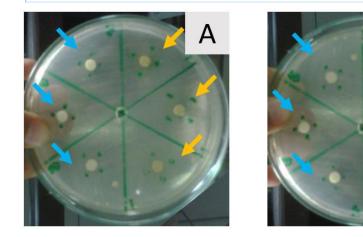
# Table 2: Significance between inhibition zone diameter of CH-Iodoform and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis*

Groups	E. faecalis	P. gingivalis	
	<b>CH-Iodoform</b>	CH-Iodoform	
E. faecalis	0.00001*	-	
<b>CH-Barium Sulfate</b>			
P. gingivalis	-	0.00001*	
<b>CH-Barium Sulfate</b>			
Statistically significant			

Commented [Office17]: Please combine table 1 and table 2 into concise table

Independent t-test was used in this study to check the significance between CH-Iodoform group and CH-Barium Sulfate group on *E. faecalis* and *P. gingivalis*. The significance between inhibition zone diameter of CH-Iodoform and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* growth is shown in Table 2. We found a significant difference between the two treatment groups on both bacteria.

The CH-Iodoform group has higher antibacterial efficacy with wider antibacterial clear zone in both bacteria (p = 0.00001; p<0.05). CH-Barium Sulfate group has lower antibacterial efficacy with narrower antibacterial clear zone in both bacteria. There is no significant difference of antibacterial activity of CH-Iodoform between *E. faecalis* and *P. gingivalis* (p = 0.11726; p<0.05). There is also no significant difference of antibacterial activity of CH-Barium Sulfate between *E. faecalis* and *P. gingivalis* (p = 0.10712; p<0.05).



#### Figure 1.

Agar diffusion assay containing *E. faecalis* (A) and *P. gingivalis* (B). The green dots indicate the inhibition area of each well. CH-Iodoform is pointed by yellow arrows, and CH-Barium Sulfate is pointed by blue arrows.

**Commented [Office18]:** The figure is not per instruction and not following the standard of the journal, please revise

B

#### DISCUSSION

*Enterococcus faecalis* and *Porphyromonas gingivalis* were used in this study because these bacteria are opportunistic, living in facultative anaerobic condition, found in pathogenic state, and associated with failed root canal treatment.<sup>5</sup> These bacteria also have a major role in persistent root canal infections, can survive in the root canals and resistant to commonly used intracanal dressings.<sup>14</sup>

Antibacterial efficacy of CH-Iodoform and CH-Barium Sulfate dressing materials on *E. faecalis* and *P. gingivalis* growths were experimentally checked with agar diffusion method and inhibition clear zone were measured on the media. Inhibition zone is a laboratory calculation method to check whether a material have the ability to inhibit bacterial growth.<sup>13</sup> The diameter of clear zones would describe the strength of such material to inhibit bacterial growth, the stronger this material inhibit, the larger the clear zones would appear.

In this study Mueller Hinton agar was used because this media can grow *E. faecalis* and *P. gingivalis* actively and sensitive to drugs effect. Inhibition zones appeared on both groups of CH-Iodoform and CH-Barium Sulfate. This showed that each of these dressing materials have the ability to inhibit both *E. faecalis* and *P. gingivalis*.

The antibacterial properties of both root canal dressing materials mainly come from calcium hydroxide, which mechanism involve hydroxyl ion to kill bacteria by protein denaturation, cytoplasmic membrane and DNA destruction will physically inhibit this bacterial growth in the root canal systems.<sup>15</sup> Antibacterial activity is connected with alkali formation which can destruct lipid, the protein structure of bacteria and nucleic acid.<sup>16</sup> Direct contact of hydroxyl ion in alkaline pH with cytoplasmic membrane will destruct the hydrogen chain of protein polypeptide. Contact of hydroxyl ion with DNA will result in replication inhibition, causing lethal mutation. DNA is sensitive to temperature and pH change. In alkali condition DNA structure and function will break and leading to bacterial cell death. Alkali condition from calcium hydroxide would impact on the surrounding tissues, including healing, anti-inflammation<sup>17</sup> and cytotoxic which lead to apoptosis.<sup>18,19</sup>

The result of this study showed a significant difference between CH-Iodoform and CH-Barium Sulfate. The ability of CH-Iodoform to inhibit *E. faecalis* and *P. gingivalis* growths are significantly greater than CH-Barium Sulfate root canal dressing. In CH-Iodoform dressing, the iodoform substance will release iodine with high reactivity to promote protein oxidation. Iodoform functions as disinfectant and infection control. Iodoform and calcium hydroxide can diffuse into dentinal tubules for added disinfection and eventually will stimulate tissue repair.

In CH-Barium Sulfate, barium sulfate can diffuse into dentinal tubule and also perform antibacterial activity. In this study the antimicrobial strength is not as good as CH-Iodoform. This may be caused by the form and structure of barium sulfate used in the mixture. Previous study on antimicrobial of barium sulfate showed that barium sulfate in micron particulate form would have potent antimicrobial properties.<sup>20</sup> Barium sulfate is generally used for its radiopacity effect on radiographic examination.<sup>11</sup>

*E. faecalis* and *P. gingivalis* can be killed with high pH level. High pH or an extreme alkaline environment would disturb the survival of most bacteria.<sup>21,22</sup> However, there are some studies stated that pH of higher than 11.5 is required for potent disinfection.<sup>23</sup> This study showed inhibition zone of CH-Iodoform is two folds higher than CH-Barium Sulfate, this may be caused by the combination of these two materials, calcium hydroxide and iodoform would create synergistic effect on the antimicrobial activity to inhibit *E. faecalis* growth. Even though this dressing material won't eliminate *E. faecalis or P. gingivalis*, the material is expected to weaken the bacteria, and eventually body defense mechanism will be able eliminate these bacteria and their byproducts.

Many researches have been done to found novel, biocompatible, antimicrobial and regenerating root canal dressing materials in order to support successful root canal treatment and enhance patient's satisfaction. The success of root canal treatment aside from the right choice of dressing, there are other factors such as the root canal systems complexity, diffusion ability of dressing materials within the dentinal tubules, and the effect on mixed cultured bacteria or biofilm formation, and bacterial resistance, which need to be evaluated in further studies. As there are many microorganisms involved in failed root canal treatments, <sup>24-26</sup> even after biomechanical and chemical instrumentations during treatment, individual and personalized assessment of these bacteria or biofilm is needed for ideal treatment.

In conclusion, calcium hydroxide-Iodoform root canal dressing has higher antimicrobial efficacy on both *E. faecalis* and *P. gingivalis*. However, further studies need to be done regarding the effect on periodontal and periapical tissues with more complexities.

#### ACKNOWLEDGEMENT

The authors would like to thank the Publication Center, Dental Hospital, and Research Center, Faculty of Dental Medicine, Universitas Airlangga for the given facility. Commented [Office19]: Please add the limitation of this study

#### REFERENCES

- Santoso CMA, Samadi K, Prasetyo EP, Wahjuningrum DA. Root canal cleanliness between mangosteen peel extract irrigant and NaOCl 2.5%. Conservative Dentistry Journal. 2020; 10(1): 40-3. doi: 10.20473/cdj.v10i1.2020.40-43.
- Juniarti DE, Kusumaningsih T, Soetojo A, Prasetyo EP, Sunur YK. Antibacterial activity and phytochemical analysis of ethanolic purple leaf extract (*Graptophyllum Pictum L. griff*) on *Lactobacillus acidophilus*. Mal J Med Health Sci. 2021; 17(Suppl2): 71-3.
- Astuti RHN, Samadi K, Prasetyo EP. Antibacterial activity of *Averrhoa bilimbi Linn*. leaf extract against Enterococcus faecalis. Conservative Dentistry Journal. 2016; 6(2): 93-8. doi: 10.20473/cdj.v6i2.2016.93-98.
- Harseno S, Mooduto L, Prasetyo EP. Antibacterial potency of kedondong Bangkok leaves extract (*Spondias dulcis Forst.*) against Enterococcus faecalis bacteria. Conservative Dentistry Journal. 2016; 6(2): 110-6. doi: 10.20473/cdj.v6i2.2016.110-116.
- Prada I, Mico-Munoz P, Giner-Lluesma T, Mico-Martinez P, Collado-Castellano N, Manzano-Saiz A. Influence of microbiology on endodontic failure. Literature review. Med Oral Patol Oral Cir Bucal. 2019; 24(3): e364-72. doi: 10.4317/medoral.22907.
- Del Fabbro M, Samaranayake LP, Lolato A, Weinstein T, Taschieri S. Analysis of the secondary endodontic lesions focusing on the extraradicular microorganisms: an overview. J Investig Clin Dent. 2014; 5(4): 245-54. doi: 10.1111/jicd.12045.
- Kaiwar A, Nadig G, Hegde J, Lekha S. Assessment of antimicrobial activity of endodontic sealers on Enterococcus faecalis: an in vitro study. World J Dent. 2012; 3(1): 26-31. doi: 10.5005/jp-journals-10015-1123.
- Noguchi N, Noiri Y, Narimatsu M, Ebisu S. Identification and localization of extraradicular biofilm-forming bacteria associated with refractory endodontic pathogens. Appl Environ Microbiol. 2005; 71:8738-43. doi: 10.1128/AEM.71.12.8738-8743.2005.
- Kuntjoro M, Prasetyo EP, Cahyani F, Kamadjaja MJK, Hendrijantini N, Laksono H, Rahmania PN, Ariestania V, Nugraha AP, Ihsan IS, Dinaryanti A, Rantam FA. Lipopolysaccharide's cytotoxicity on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0048. doi:

10.1590/pboci.2020.153.

- Estrela C, Estrela CRDA, Hollanda ACB, Decurcio DDA, Pecora JD. Influence of iodoform on antimicrobial potential of calcium hydroxide. J Appl Oral Sci. 2006; 14(1): 33-7. doi: 10.1590/S1678-77572006000100007.
- Orucoglu H, Cobankara FK. Effect of unintentionally extruded calcium hydroxide paste including barium sulfate as a radiopaquing agent in treatment of teeth with periapical lesions: Report of a case. J Endod. 2008; 34(7): 888-91. doi: :10.1016/j.joen.2008.04.012.
- 12. Alharthi SS, Binshabaib M, Almasoud NS, Shawky HA, Aabed KF, Alomar TS, Albrekan AB, Alfaifi AJ, Melaibari AA. Myrrh mixed with silver nanoparticles demonstrates superior antimicrobial activity against *Porphyromonas gingivalis* compared to myrrh and silver nanoparticles alone. Saudi Dent J. 2021; 33: 890-6. doi: 10.1016/j.sdentj.2021.09.009.
- 13. Balouiri M, Sadiki M, Ibnsouda SK. Methods for in vitro evaluating antimicrobial activity: a review. J Pharm Anal. 2016; 6(2): 71-9. doi: 10.1016/j.jpha.2015.11.005.
- Jhamb S, Singla R, Kaur A, Sharma J, Bhushan J. An in vitro determination of antibacterial effect of silver nanoparticles gel as an intracanal medicament in combination with other medicaments against Enterococcus faecalis. J Conserv Dent. 2019; 22: 479-82. doi: 10.4103/JCD.JCD\_113\_20.
- Mohammadi Z, Dummer PMH. Properties and applications of calcium hydroxide in endodontics and dental traumatology. Int Endod J. 2011; 44(8): 697-730. doi: 10.1111/j.1365-2591.2011.01886.x.
- Athanassiadis B, Abbott PV, Walsh LJ. The use of calcium hydroxide, antibiotics, and biocides as antimicrobial medicaments in Endodontics. Australian Dental Journal. 2007; 52(1): 64-82. doi: 10.1111/j.1834-7819.2007.tb00527.x.
- 17. Prasetyo EP, Kuntjoro M, Cahyani F, Goenharto S, Saraswati W, Juniarti DE, Hendrijantini N, Hariyani N, Nugraha AP, Rantam FA. Calcium hydroxide upregulates interleukin-10 expression in time dependent exposure and induces osteogenic differentiation of human umbilical cord mesenchymal stem cells. Int J Pharm Res. 2021; 13(1): 140-5. Doi: 10.31838/ijpr/2021.13.01.023.
- Prasetyo EP, Widjiastuti I, Cahyani F, Kuntjoro M, Hendrijantini N, Hariyani N, Winoto ER, Nugraha AP, Goenharto S, Susilowati H, Hendrianto E, Rantam FA. Cytotoxicity of calcium hydroxide on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0044. doi:

10.1590/pboci.2020.141.

- Prasetyo EP, Kuntjoro M, Goenharto S, Juniarti DE, Cahyani F, Hendrijantini N, Nugraha AP, Hariyani N, Rantam FA. Calcium hydroxide increases human umbilical cord mesenchymal stem cells expressions of apoptotic protease-activating factor-1, caspase-3 and caspase-9. Clinical, cosmetic and investigational dentistry. 2021;13: 59-65. doi: 10.2147/CCIDE.S284240.
- Aninwene II G, Stout D, Yang Z, Webster TJ. Nano-BaSO4: a novel antimicrobial additive to pellethane. Int J Nanomedicine. 2013;8(1):1197-1205. doi: 10.2147/IJN.S40300.
- Jhajharia K, Parolia A, Shetty KV, Mehta LK. Biofilm in endodontics: a review. J Int Soc Prev Community Dent. 2015; 5: 1-12. doi: 10.4103/2231-0762.151956.
- Zancan RF, Vivan RR, Lopes MRM, Weckwerth PH, Andrade FB, Ponce JB, Duarte MAH. Antimicrobial activity and physicochemical properties of calcium hydroxide pastes used as intracanal medication. J Endod. 2016; 42(12): 1822-8. doi: 101016/j.joen.2016.08.017.
- Evans M, Davies JK, Sundqvist G, Figdor D. Mechanisms involved in the resistance of Enterococcus faecalis to calcium hydroxide. Int Endod J. 2002; 35(3): 221-8. doi: 10.1046/j.1365-2591.2002.00504.x.
- Dudeja PG, Dudeja KK, Srivastava D, Grover S. Microorganisms in periradicular tissues: Do they exist? A perennial controversy. J Oral Maxillofac Pathol. 2015; 19(3): 356-63. doi: 10.4103/0973-029X.174612.
- 25. Pereira RS, Rodrigues VAA, Furtado WT, Gueiros S, Pereira GS, Avila-Campos MJ. Microbial analysis of root canal and periradicular lesion associated to teeth with endodontic failure. Anaerobe. 2017; 48: 12-8. doi: 10.1016/j.anaerobe.2017.06.016.
- 26. Siqueira JF, Antunes HS, Rocas IN, Rachid CTCC, Alves FRF. Microbiome in the apical root canal system of teeth with post-treatment apical periodontitis. PLoS ONE. 2016; 11(9): e0162887. doi: 10.1371/journal.pone.0162887.

Dear Editor of DJMKG,

Thank you for giving me the opportunity to submit a revised draft of my manuscript titled **Antibacterial efficacy between calcium hydroxide-iodophors and calcium hydroxide-barium sulfate root canal dressings on** *Enterococcus faecalis* and *Porphyromonas gingivalis in-vitro* to Dental Journal (Majalah Kedokteran Gigi). We appreciate the time and effort that you and the reviewers have dedicated to providing your valuable feedback on our manuscript. We are grateful to the reviewers for their insightful comments on our manuscript. We have been able to incorporate changes to reflect most of the suggestions provided by the reviewers. We have highlighted the changes within the manuscript.

Here is a point-by-point response to the reviewers' comments and concerns.

No.	Reviewer comments	Before revision	Revision	Notes	
	Revision-1				
	Reviewer 1 (RV1)				
1.	Reference number 23 to be replaced	Reference no 23	Replaced with reference number 6	Page: 6 Paragraph: 2	
2.	Conclusion adjusted to the title	none	Added conclusion: calcium hydroxide-Iodoform root canal dressing has different (higher) antimicrobial efficacy on both <i>E. faecalis</i> and <i>P.</i> <i>gingivalis</i> .	Page: 6 Paragraph: 3	
	Reviewer 2 (RV2)	· . · ·	· · · · ·		
1.	Correction on page 2 paragraph 3	resistance and periapical abnormalities	can spread to peri-radicular which causes the formation of peri-radicular lesions after root canal treatment.	Page: 2 Paragraph: 3	
2.	Addition on page 2 paragraph 5	none	If done correctly, depending on the right instrumentation and irrigation to remove	Page: 2 Paragraph: 5	
			inorganic and organic smear layers.		
3.	Addition on page 3 paragraph 1	none	patients who failed endodontic treatment	Page: 3 Paragraph: 1	
4.	Explain about the modification	none	Modifications on the sum and position of the wells in the plates.	Page: 3 Paragraph: 2	
5.	Correction on page 5 paragraph 2	ThereisnosignificancedifferenceE.faecalisandP.gingivalis.	The sentences were removed for clarity	Page: 4 Paragraph: 2	
6.	Addition on page 6 paragraph 5	none	Thus, the combination of CH and Iodoform will synergistically strengthen each other. eliminate the bacteria.	Page: 5 Paragraph: 5	

7.	Addition on page 7	none	The addition of this material	Page: 6
	paragraph 1	none	must consider its properties as it will affect the dressing's consistency and application.	Paragraph: 1
8.	Limitations of the study. Addition on page 7 paragraph 3	none	There are limitations, as this is only an in-vitro study and there are many factors to consider, both in-situ and in- vivo.	Page: 6 Paragraph: 3
	Managing Editor (ME)			
1.	Title correction	In-vitro antibacterial efficacy between calcium	Antibacterial efficacy between calcium hydroxide- iodoform and calcium hydroxide-barium sulfate	Page: 1 Paragraph: title
		hydroxide- iodoform and calcium hydroxide-barium sulfate root canal dressings on <i>Enterococcus</i> <i>faecalis</i> and <i>Porphyromonas</i> <i>gingivalis</i>	root canal dressings on Enterococcus faecalis and Porphyromonas gingivalis in- vitro	
2.	Purpose and conclusion	To determine	To compare the antibacterial efficacy	Page: 1 Paragraph: abstract
3.	lodoform to be adjusted according to MeSH	iodoform	Iodophors (replaced all)	Page: all Paragraph: all
4.	Full and abbreviation of <i>E. faecalis</i> and <i>P. gingivalis</i>	none	Enterococcus faecalis (E. faecalis and Porphyromonas gingivalis (P. gingivalis)	Page: 2 Paragraph: 2
5.	Add the ethical clearance number	none	Ethical Clearance Commission (166/KKEPK.FKG)	Page: 3 Paragraph: 1
6.	Bacterial source	none	Stock bacteria of Research Center obtained from patients who failed endodontic treatment	
7.	Method modification	none	Modifications on the sum and position of the wells in the plates.	Page: 3 Paragraph: 2
8.	Sample size	none	According to methods used frequently in microbiology laboratory.	
9.	MIC and MBC	none	MIC and MBC were not conducted because this study use commercially available	

				[]
			products, not a basic	
			substance or extracts.	
10.	Statistical analysis	none	SPSS 20.0 for Windows (SPSS	Page: 3
	software		Inc., Chicago, Illinois, USA)	Paragraph: 6
			was used in this study.	
11.	Tables and Figure	In the text	The tables and figure have	Page 10 and 11
			been moved at the end, after	
			the references	
12.	Limitation of the study	none	There are limitations, as this is	Page: 7
			only an in-vitro study and	Paragraph: 3
			there are many factors to	
			consider, both in-situ and in-	
			vivo.	
	Revision-2			
	Reviewer			
1.				Page:
				Paragraph:
2.				Page:
				Paragraph:
3.				Page:
				Paragraph:
4.				
5.				

In addition to the above comments, all spelling and grammatical errors pointed out by the reviewers have been corrected. We look forward to hearing from you in due time regarding our submission and to respond to any further questions and comments you may have.

For improvement about the language editing, layout, tables and figure please arrange accordingly, thank you.

Sincerely,

z,

Eric Priyo Prasetyo Date: February 25, 2022.

# Antibacterial efficacy between calcium hydroxide-iodophors and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis in-vitro*

# Eric Priyo Prasetyo,<sup>1</sup> Devi Eka Juniarti,<sup>1</sup> Galih Sampoerno,<sup>1</sup> Dian Agustin Wahjuningrum,<sup>1</sup> Ananta Tantri Budi,<sup>1</sup> Dyanita Hasri,<sup>2</sup> Evelyn Tjendronegoro<sup>3</sup>

<sup>1</sup>Department of Conservative Dentistry, Faculty of Dental Medicine, Universitas Airlangga, Surabaya, Indonesia. <sup>2</sup>Dentistry Program, Faculty of Dental Medicine, Universitas Airlangga, Surabaya, Indonesia.

<sup>3</sup>Healthcare and Research, Irvine Medical Center, University of California, California, United States of America.

#### **ABSTRACT**

**Background:** A successful endodontic treatment is inseparable from the right choice of root canal dressing. The right choice of medicaments would result in patient satisfaction. Enterococcus faecalis (E. faecalis) and Porphyromonas gingivalis (P. gingivalis) are usually found in failed root canal treatments. Calcium hydroxide is a gold standard dressing which create an alkaline environment in the root canal and have a bactericidal effect. Commercially, there are calcium hydroxide dressings with supporting additions, including calcium hydroxide-iodophors (CH-Iodophors) and Calcium hydroxide-barium sulfate (CH-Barium Sulfate). **Purpose:** This study aimed to compare the antibacterial efficacy of CH-Iodophors and CH-Barium Sulfate on E. faecalis and P. gingivalis. Methods: CH-Iodophors and CH-Barium Sulfate was obtained commercially. E. faecalis and P. gingivalis were obtained from stock culture taken from the root canal of failed endodontic treatment. E. faecalis and P. gingivalis were cultured in petri dishes and for each bacterium, 12 wells were made in the media, 6 wells were used for CH-Iodophors group and 6 wells were used for CH-Barium Sulfate group. CH-Iodophors and CH-Barium Sulfate were deployed in the wells in E. faecalis and P. gingivalis cultured medium in the petri dishes. After incubation, the inhibition zone diameters were measured. Independent t-test was used for analysis and significance level was set at 5%. Results: There is a significant antibacterial efficacy between CH-Iodophors and CH-Barium Sulfate on E. faecalis and P. gingivalis (p=0.00001). Conclusion: CH-Iodophors has a higher antibacterial efficacy than CH-Barium Sulfate on both E. faecalis and P. gingivalis.

*Keywords:* Enterococcus faecalis; Porphyromonas gingivalis; calcium hydroxide; *iodophors*; barium sulfate; patient satisfaction.

Correspondence: Eric Priyo Prasetyo, Department of Conservative Dentistry, Faculty of Dental Medicine, Universitas Airlangga, Jalan Mayjend. Prof. Dr. Moestopo 47, Surabaya, Indonesia, Email address: eric-p-p@fkg.unair.ac.id.

# **INTRODUCTION**

The right choice of root canal dressing to eliminate bacteria in the root canal is important for a successful endodontic treatment. A successful endodontic treatment would result in patient's satisfaction. Many attempts were conducted to increase the success of root canal treatment, including finding the efficient instrumentation, effective cleaning,<sup>1,2</sup> antibacterial dressing and irrigation materials.<sup>3-5</sup>

Root canal treatment has a high success rate, but in some cases, there are failures. Isolated bacteria from root canal treatment failures and the prevalence are about 45.8-77% caused by *Enterococcus faecalis (E. faecalis)* and 28.17% caused by *Porphyromonas gingivalis (P. gingivalis)* in the root canal system.<sup>6</sup> Both microorganisms are among the ones that survive disinfecting protocol.<sup>7</sup>

*E. faecalis* can invade dentine tubules and can spread into peri-radicular which causes the formation of peri-radicular lesions after root canal treatment.<sup>8</sup> *P. gingivalis* can survive in the extra-radicular area mostly in the area approximate with root surface, and responsible for periodontitis and peri-radicular lesions.<sup>9</sup> *P. gingivalis* also contribute to root canal treatment failures, through its byproduct, lipopolysaccharide which effect biological processes, inflammation and tissue destruction.<sup>10</sup>

Different substances have been added to calcium hydroxide to increase its efficacy. The added substances are distilled water, saline, propylene glycol, chlorhexidine, glycerin, iodophors, barium sulfate, corticosteroids, antibiotics, anesthetic solution, methyl cellulose, and detergents. Iodophors has been added to calcium hydroxide to work with different bacterial characteristic.<sup>11</sup> Barium sulfate is added to calcium hydroxide aside from its antibacterial effect, it functions to increase radiopacity.<sup>12</sup>

Root canal treatment failure may happen even after dressing with calcium hydroxide if done incorrectly, depending on the right instrumentation and irrigation to remove inorganic and organic smear layers. In this research, the efficacy of calcium hydroxide addition with iodophors (CH-Iodophors) and barium sulfate (CH-Barium Sulfate) was analyzed on *E. faecalis* and *P. gingivalis*. Based on this background, the authors would like to purposely compare the antibacterial efficacy between CH-Iodophors and CH-Barium Sulfate root canal dressing on *E. faecalis* and *P. gingivalis*.

## **MATERIALS AND METHODS**

This is an experimental laboratory study, conducted in Conservative Dentistry Section, Universitas Airlangga Dental Hospital and Microbiology Laboratory, Research Center, Faculty of Dental Medicine, Universitas Airlangga. This study was ethically approved by Universitas Airlangga Faculty of Dental Medicine Health Research Ethical Clearance Commission (166/KKEPK.FKG). Materials used in this research were commercially available calcium hydroxide dressings with Iodophors (Meta Biomed, Korea) and Barium Sulfate (Meta Biomed, Korea). *Enterococcus faecalis* and *Porphyromonas gingivalis* bacteria used in this study were stock bacteria previously cultured from patients who failed endodontic treatment.

The method used in this study was agar diffusion method using Mueller Hinton (MH) agar and Brain Heart Infusion (BHI) broth. The method is according to Alharthi et al. (2021) and Balouiri et al. (2016) with modifications on the sum and position of the wells in the plates.<sup>13,14</sup> The media were allocated for 4 groups of experiments. In the first group, 12 wells were prepared for CH-Iodophors (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*), and in the second group, 12 wells were prepared for CH-Barium Sulfate dressing (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*).

Both *E. faecalis* and *P. gingivalis* bacterial culture from stock was moved into separate reaction tube, each containing BHI broth and stirred. Incubation was done for both culture at 37°C for 24 hours in anaerobic condition. After 24 hours, *E. faecalis* and *P. gingivalis* bacterial culture in BHI broth was taken 0.5 ml using micro pipette, and poured into another reaction tube containing BHI broth until equal with 0.5 McFarland standard of turbidity.

The bacterial cultures were taken from BHI broth using sterile cotton swab and swabbed on the surface of each MH agar allocated for *E. faecalis* and *P gingivalis*. Antibacterial test was conducted by making the wells for the tested dressing materials (CH-Iodophors and CH-Barium Sulfate). The samples were incubated at 37°C for 48 hours in anaerobic condition. After 48 hours, measurements were conducted on antibacterial efficacy through inhibition zone measurement using a Vernier Digital Caliper (Mitutoyo, Japan). The clear zones of inhibition around the wells were measured, revealing no bacterial growth. Data of measurements on inhibition zone diameters (in millimeters) were collected for each sample wells.

Inhibition zones data were analyzed statistically and significance level was set at 5%. SPSS 20.0 for Windows (SPSS Inc., Chicago, Illinois, USA) was used in this study. Data

normality was tested using Shapiro Wilk test. The significance was tested using independent ttest.

# RESULTS

The number of replications (n) for each treatment group was 6. Mean and standard deviation of inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* are shown in Table 1. The mean of *E. faecalis* inhibition zone from CH-Iodophors was 11.8125 mm and CH-Barium Sulfate was 6.3750 mm. The mean of *P. gingivalis* inhibition zone from CH-Iodophors was 12.7875 mm and CH-Barium Sulfate was 6.6750 mm.

Independent t-test was used in this study to check the significance between CH-Iodophors group and CH-Barium Sulfate group on *E. faecalis* and *P. gingivalis*. The significance between inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* growth is shown in Table 2. We found a significant difference between the two treatment groups on both bacteria. The CH-Iodophors group has higher antibacterial efficacy with wider antibacterial clear zone in both bacteria (p = 0.00001). CH-Barium Sulfate group has lower antibacterial efficacy with narrower antibacterial clear zone in both bacteria.

# DISCUSSION

*Enterococcus faecalis* and *Porphyromonas gingivalis* were used in this study because these bacteria are opportunistic, living in facultative anaerobic condition, found in pathogenic state, and associated with failed root canal treatment.<sup>6</sup> These bacteria also have a major role in persistent root canal infections, can survive in the root canals and resistant to commonly used intracanal dressings.<sup>15</sup>

Antibacterial efficacy of CH-Iodophors and CH-Barium Sulfate dressing materials on *E*. *faecalis* and *P. gingivalis* growths were experimentally checked with agar diffusion method and inhibition clear zone were measured on the media. Inhibition zone is a laboratory calculation method to check whether a material have the ability to inhibit bacterial growth.<sup>14</sup> The diameter of clear zones would describe the strength of such material to inhibit bacterial growth, the stronger this material inhibit, the larger the clear zones would appear.

In this study Mueller Hinton agar was used because this media can grow *E. faecalis* and *P. gingivalis* actively and sensitive to drugs effect. Inhibition zones appeared on both groups of CH-Iodophors and CH-Barium Sulfate. This showed that each of these dressing materials have the ability to inhibit both *E. faecalis* and *P. gingivalis*.

The antibacterial properties of both root canal dressing materials mainly come from calcium hydroxide, which mechanism involve hydroxyl ion to kill bacteria by protein denaturation, cytoplasmic membrane and DNA destruction will physically inhibit this bacterial growth in the root canal systems.<sup>16</sup> Antibacterial activity is connected with alkali formation which can destruct lipid, the protein structure of bacteria and nucleic acid.<sup>17</sup> Direct contact of hydroxyl ion in alkaline pH with cytoplasmic membrane will destruct the hydrogen chain of protein polypeptide. Contact of hydroxyl ion with DNA will result in replication inhibition, causing lethal mutation. DNA is sensitive to temperature and pH change. In alkali condition DNA structure and function will break and leading to bacterial cell death. Alkali condition from calcium hydroxide would impact on the surrounding tissues, including healing, anti-inflammation<sup>18</sup> and cytotoxic which lead to apoptosis.<sup>19,20</sup>

The result of this study showed a significant difference between CH-Iodophors and CH-Barium Sulfate. The ability of CH-Iodophors to inhibit *E. faecalis* and *P. gingivalis* growths are significantly greater than CH-Barium Sulfate root canal dressing. In CH-Iodophors dressing, the iodophors substance will release iodine with high reactivity to promote protein oxidation. Iodophors functions as disinfectant and infection control. Thus, the combination of

CH and Iodophors will synergistically strengthen each other. Iodophors and calcium hydroxide can diffuse into dentinal tubules for added disinfection and eliminate the bacteria.

In CH-Barium Sulfate, barium sulfate can diffuse into dentinal tubule and also perform antibacterial activity. In this study the antimicrobial strength is not as good as CH-Iodophors. This may be caused by the form and structure of barium sulfate used in the mixture. Previous study on antimicrobial of barium sulfate showed that barium sulfate in micron particulate form would have potent antimicrobial properties.<sup>21</sup> Barium sulfate is generally used for its radiopacity effect on radiographic examination.<sup>12</sup> The addition of this material must consider its properties as it will affect the dressing's consistency and application.

*E. faecalis* and *P. gingivalis* can be killed with high pH level. High pH or an extreme alkaline environment would disturb the survival of most bacteria.<sup>22,23</sup> However, there are some studies stated that pH of higher than 11.5 is required for potent disinfection.<sup>6</sup> This study showed inhibition zone of CH-Iodophors is two folds higher than CH-Barium Sulfate, this may be caused by the combination of these two materials, calcium hydroxide and iodophors would create synergistic effect on the antimicrobial activity to inhibit *E. faecalis* growth. Even though this dressing material won't eliminate *E. faecalis or P. gingivalis*, the material is expected to weaken the bacteria, and eventually body defense mechanism will be able eliminate these bacteria and their byproducts.

Many researches have been done to found novel, biocompatible, antimicrobial and regenerating root canal dressing materials in order to support successful root canal treatment and enhance patient's satisfaction. The success of root canal treatment aside from the right choice of dressing, there are other factors such as the root canal systems complexity, diffusion ability of dressing materials within the dentinal tubules, and the effect on mixed cultured bacteria or biofilm formation, and bacterial resistance, which need to be evaluated in further studies. As there are many microorganisms involved in failed root canal treatments, <sup>24-26</sup> even after biomechanical and chemical instrumentations during treatment, individual and personalized assessment of these bacteria or biofilm is needed for ideal treatment. There are limitations, as this is only an in-vitro study and there are many factors to consider, both in-situ and in-vivo. In conclusion, calcium hydroxide-Iodophors root canal dressing has different (higher) antimicrobial efficacy on both *E. faecalis* and *P. gingivalis*. However, further studies need to be done regarding the effect on periodontal and periapical tissues with more complexities.

## ACKNOWLEDGEMENT

The authors would like to thank the Publication Center, Dental Hospital, and Research Center, Faculty of Dental Medicine, Universitas Airlangga for the given facility.

# REFERENCES

- Santoso CMA, Samadi K, Prasetyo EP, Wahjuningrum DA. Root canal cleanliness between mangosteen peel extract irrigant and NaOCl 2.5%. Conservative Dentistry Journal. 2020; 10(1): 40-3. doi: 10.20473/cdj.v10i1.2020.40-43.
- Juniarti DE, Kusumaningsih T, Soetojo A, Prasetyo EP, Sunur YK. Antibacterial activity and phytochemical analysis of ethanolic purple leaf extract (*Graptophyllum Pictum L. griff*) on *Lactobacillus acidophilus*. Mal J Med Health Sci. 2021; 17(Suppl2): 71-3.
- Astuti RHN, Samadi K, Prasetyo EP. Antibacterial activity of *Averrhoa bilimbi Linn*. leaf extract against Enterococcus faecalis. Conservative Dentistry Journal. 2016; 6(2): 93-8. doi: 10.20473/cdj.v6i2.2016.93-98.
- Prasetyo EP, Saraswati W, Goenharto S, Wahjuningrum D, Mooduto L, Rosidin RF, Tjendronegoro E. White pomegranate (*Punica granatum*) peels extract bactericidal potency on *Enterococcus faecalis*. Conservative Dentistry Journal. 2021; 11(2): 84-8. doi: 10.20473/cdj.v11i2.2021.84-88.
- Harseno S, Mooduto L, Prasetyo EP. Antibacterial potency of kedondong Bangkok leaves extract (*Spondias dulcis Forst.*) against Enterococcus faecalis bacteria. Conservative Dentistry Journal. 2016; 6(2): 110-6. doi: 10.20473/cdj.v6i2.2016.110-116.
- Prada I, Mico-Munoz P, Giner-Lluesma T, Mico-Martinez P, Collado-Castellano N, Manzano-Saiz A. Influence of microbiology on endodontic failure. Literature review. Med Oral Patol Oral Cir Bucal. 2019; 24(3): e364-72. doi: 10.4317/medoral.22907.
- Del Fabbro M, Samaranayake LP, Lolato A, Weinstein T, Taschieri S. Analysis of the secondary endodontic lesions focusing on the extraradicular microorganisms: an overview. J Investig Clin Dent. 2014; 5(4): 245-54. doi: 10.1111/jicd.12045.
- 8. Kaiwar A, Nadig G, Hegde J, Lekha S. Assessment of antimicrobial activity of endodontic sealers on Enterococcus faecalis: an in vitro study. World J Dent. 2012;

3(1): 26-31. doi: 10.5005/jp-journals-10015-1123.

- Noguchi N, Noiri Y, Narimatsu M, Ebisu S. Identification and localization of extraradicular biofilm-forming bacteria associated with refractory endodontic pathogens. Appl Environ Microbiol. 2005; 71:8738-43. doi: 10.1128/AEM.71.12.8738-8743.2005.
- Kuntjoro M, Prasetyo EP, Cahyani F, Kamadjaja MJK, Hendrijantini N, Laksono H, Rahmania PN, Ariestania V, Nugraha AP, Ihsan IS, Dinaryanti A, Rantam FA. Lipopolysaccharide's cytotoxicity on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0048. doi: 10.1590/pboci.2020.153.
- Estrela C, Estrela CRDA, Hollanda ACB, Decurcio DDA, Pecora JD. Influence of iodophors on antimicrobial potential of calcium hydroxide. J Appl Oral Sci. 2006; 14(1): 33-7. doi: 10.1590/S1678-77572006000100007.
- Orucoglu H, Cobankara FK. Effect of unintentionally extruded calcium hydroxide paste including barium sulfate as a radiopaquing agent in treatment of teeth with periapical lesions: Report of a case. J Endod. 2008; 34(7): 888-91. doi: :10.1016/j.joen.2008.04.012.
- 13. Alharthi SS, Binshabaib M, Almasoud NS, Shawky HA, Aabed KF, Alomar TS, Albrekan AB, Alfaifi AJ, Melaibari AA. Myrrh mixed with silver nanoparticles demonstrates superior antimicrobial activity against *Porphyromonas gingivalis* compared to myrrh and silver nanoparticles alone. Saudi Dent J. 2021; 33: 890-6. doi: 10.1016/j.sdentj.2021.09.009.
- 14. Balouiri M, Sadiki M, Ibnsouda SK. Methods for in vitro evaluating antimicrobial activity: a review. J Pharm Anal. 2016; 6(2): 71-9. doi: 10.1016/j.jpha.2015.11.005.
- 15. Jhamb S, Singla R, Kaur A, Sharma J, Bhushan J. An in vitro determination of antibacterial effect of silver nanoparticles gel as an intracanal medicament in combination with other medicaments against Enterococcus faecalis. J Conserv Dent. 2019; 22: 479-82. doi: 10.4103/JCD.JCD\_113\_20.
- Mohammadi Z, Dummer PMH. Properties and applications of calcium hydroxide in endodontics and dental traumatology. Int Endod J. 2011; 44(8): 697-730. doi: 10.1111/j.1365-2591.2011.01886.x.
- Athanassiadis B, Abbott PV, Walsh LJ. The use of calcium hydroxide, antibiotics, and biocides as antimicrobial medicaments in Endodontics. Australian Dental Journal. 2007; 52(1): 64-82. doi: 10.1111/j.1834-7819.2007.tb00527.x.

- 18. Prasetyo EP, Kuntjoro M, Cahyani F, Goenharto S, Saraswati W, Juniarti DE, Hendrijantini N, Hariyani N, Nugraha AP, Rantam FA. Calcium hydroxide upregulates interleukin-10 expression in time dependent exposure and induces osteogenic differentiation of human umbilical cord mesenchymal stem cells. Int J Pharm Res. 2021; 13(1): 140-5. Doi: 10.31838/ijpr/2021.13.01.023.
- Prasetyo EP, Widjiastuti I, Cahyani F, Kuntjoro M, Hendrijantini N, Hariyani N, Winoto ER, Nugraha AP, Goenharto S, Susilowati H, Hendrianto E, Rantam FA. Cytotoxicity of calcium hydroxide on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0044. doi: 10.1590/pboci.2020.141.
- 20. Prasetyo EP, Kuntjoro M, Goenharto S, Juniarti DE, Cahyani F, Hendrijantini N, Nugraha AP, Hariyani N, Rantam FA. Calcium hydroxide increases human umbilical cord mesenchymal stem cells expressions of apoptotic protease-activating factor-1, caspase-3 and caspase-9. Clinical, cosmetic and investigational dentistry. 2021;13: 59-65. doi: 10.2147/CCIDE.S284240.
- Aninwene II G, Stout D, Yang Z, Webster TJ. Nano-BaSO4: a novel antimicrobial additive to pellethane. Int J Nanomedicine. 2013;8(1):1197-1205. doi: 10.2147/IJN.S40300.
- Jhajharia K, Parolia A, Shetty KV, Mehta LK. Biofilm in endodontics: a review. J Int Soc Prev Community Dent. 2015; 5: 1-12. doi: 10.4103/2231-0762.151956.
- 23. Zancan RF, Vivan RR, Lopes MRM, Weckwerth PH, Andrade FB, Ponce JB, Duarte MAH. Antimicrobial activity and physicochemical properties of calcium hydroxide pastes used as intracanal medication. J Endod. 2016; 42(12): 1822-8. doi: 101016/j.joen.2016.08.017.
- Dudeja PG, Dudeja KK, Srivastava D, Grover S. Microorganisms in periradicular tissues: Do they exist? A perennial controversy. J Oral Maxillofac Pathol. 2015; 19(3): 356-63. doi: 10.4103/0973-029X.174612.
- 25. Pereira RS, Rodrigues VAA, Furtado WT, Gueiros S, Pereira GS, Avila-Campos MJ. Microbial analysis of root canal and periradicular lesion associated to teeth with endodontic failure. Anaerobe. 2017; 48: 12-8. doi: 10.1016/j.anaerobe.2017.06.016.
- 26. Siqueira JF, Antunes HS, Rocas IN, Rachid CTCC, Alves FRF. Microbiome in the apical root canal system of teeth with post-treatment apical periodontitis. PLoS ONE. 2016; 11(9): e0162887. doi: 10.1371/journal.pone.0162887.

# Table 1:

Mean and standard deviation from inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* 

Groups	n	E. faecalis	P. gingivalis	
		Mean <u>+</u> SD	Mean <u>+</u> SD	
CH-Iodophors	6	$11.8125 \pm 1.32001$	12.7875 <u>+</u> 1.34961	
CH-Barium Sulfate	6	6.3750 <u>+</u> 0.19494	6.6750 <u>+</u> 0.51865	

n = replication

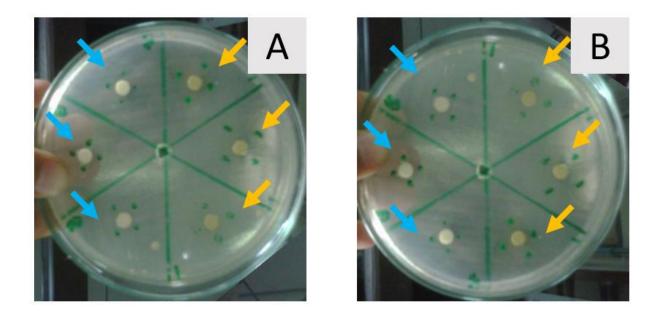
SD = standard deviation

 Table 2:
 Significance between inhibition zone diameter of CH-Iodophors and CH-Barium

 Sulfate on *E. faecalis* and *P. gingivalis*

Groups	E. faecalis	P. gingivalis
	<b>CH-Iodophors</b>	CH-Iodophors
E. faecalis	0.00001*	-
CH-Barium Sulfate		
P. gingivalis	-	0.00001*
CH-Barium Sulfate		

\*Statistically significant



# Figure 1.

Agar diffusion assay containing *E. faecalis* (A) and *P. gingivalis* (B). The green dots indicate the inhibition area of each well. CH-Iodophors is pointed by yellow arrows, and CH-Barium Sulfate is pointed by blue arrows.



eric priyo prasetyo <eric-p-p@fkg.unair.ac.id>

# Editor decision: second revision required

3 messages

**Dental Journal (Majalah Kedokteran Gigi)** <dental\_journal@fkg.unair.ac.id> To: eric priyo prasetyo <eric-p-p@fkg.unair.ac.id> Tue, Mar 8, 2022 at 10:44 AM

Dear **Dr Eric** 

Please find the attached second review file from the reviewer.

Please revise according to the comments and highlight the text with color on the changes made. Revisions, please send back no later than March 14, 2022. Thank you.

Regards,

Muhammad Dimas Aditya Ari

Dental Journal (Majalah Kedokteran Gigi) http://e-journal.unair.ac.id/MKG

Faculty of Dental Medicine, Universitas Airlangga Jl. Mayjen Prof. Dr. Moestopo 47 Surabaya 60132 INDONESIA Phone/Fax: +62 31 5039478

review 2 Eric Priyo article\_In-vitro antibacterial efficacy between calcium (BE).pdf 392K

eric priyo prasetyo <eric-p-p@fkg.unair.ac.id> To: "Dental Journal (Majalah Kedokteran Gigi)" <dental\_journal@fkg.unair.ac.id> Wed, Mar 9, 2022 at 1:07 AM

Dear Chief Editor,

Here is the 2nd revision and response letter. Please find them in the attachments. Thank you.

Sincerely, Dr. Eric & team. [Quoted text hidden]

2 attachments

Response letter review Eric et al revision 2.pdf 181K

[DJMKG] Original article template CHI CHBS EF PG Eric et al Rev 2.docx 690K

**Dental Journal (Majalah Kedokteran Gigi)** <dental\_journal@fkg.unair.ac.id> To: eric priyo prasetyo <eric-p-p@fkg.unair.ac.id> Wed, Mar 9, 2022 at 8:55 AM

Dear Drg Eric Thank you for submitting a revised draft of your manuscript. The manuscript will be re-evaluated by reviewers based on the revisions you make.

Best wishes, Dental Journal (Majalah Kedokteran Gigi) http://e-journal.unair.ac.id/MKG

Faculty of Dental Medicine, Universitas Airlangga Jl. Mayjen Prof. Dr. Moestopo 47 Surabaya 60132 INDONESIA Phone/Fax: +62 31 5039478

[Quoted text hidden]

### Original article

Antibacterial efficacy between calcium hydroxide-iodophors and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis in-vitro* 

### ABSTRACT

Background: A successful endodontic treatment is inseparable from the right choice of root canal dressing. The right choice of medicaments would result in patient satisfaction. Enterococcus faecalis (E. faecalis) and Porphyromonas gingivalis (P. gingivalis) are usually found in failed root canal treatments. Calcium hydroxide is a gold standard dressing which create an alkaline environment in the root canal and have a bactericidal effect. Commercially, there are calcium hydroxide dressings with supporting additions, including calcium hydroxide-iodophors (CH-Iodophors) and Calcium hydroxide-barium sulfate (CH-Barium Sulfate). Purpose: This study aimed to compare the antibacterial efficacy of CH-Iodophors and CH-Barium Sulfate on E. faecalis and P. gingivalis. Methods: CH-Iodophors and CH-Barium Sulfate was obtained commercially. E. faecalis and P. gingivalis were obtained from stock culture taken from the root canal of failed endodontic treatment, E. faecalis and P. gingivalis were cultured in petri dishes and for each bacterium, 12 wells were made in the media, 6 wells were used for CH-Iodophors group and 6 wells were used for CH-Barium Sulfate group. CH-Iodophors and CH-Barium Sulfate were deployed in the wells in E. faecalis and P. gingivalis cultured medium in the petri dishes. After incubation, the inhibition zone diameters were measured. Independent t-test was used for analysis and significance level was set at 5%. Results: There is a significant antibacterial efficacy between CH-Iodophors and CH-Barium Sulfate on E. faecalis and P. gingivalis (p=0.00001). Conclusion: CH-Iodophors has a higher antibacterial efficacy than CH-Barium Sulfate on both E. faecalis and P. gingivalis.

**Keywords:** Enterococcus faecalis; Porphyromonas gingivalis; calcium hydroxide; iodophors; barium sulfate; patient satisfaction.

### INTRODUCTION

The right choice of root canal dressing to eliminate bacteria in the root canal is important for a successful endodontic treatment. A successful endodontic treatment would result in patient's satisfaction. Many attempts were conducted to increase the success of root canal treatment, including finding the efficient instrumentation, effective cleaning,<sup>1,2</sup> antibacterial dressing and irrigation materials.<sup>3-5</sup>

**Commented [BNR2]:** managing editor's review has not been revised: why they add CH iodoform and CHbarium sulfate? This sentence unclear

Commented [BNR3]: managing editor's review has not been revised:

-compare the antibacterial efficacy between CH-lodoform and CH-Barium Sulfate root canal dressing on E. faecalis and P. gingivalis.

-Please add the design and setting of this study

-How determine the isolated bacteria was E.faecalis or P.gingivalis?

Root canal treatment has a high success rate, but in some cases, there are failures. Isolated bacteria from root canal treatment failures and the prevalence are about 45.8-77% caused by *Enterococcus faecalis (E. faecalis)* and 28.17% caused by *Porphyromonas gingivalis (P. gingivalis)* in the root canal system.<sup>6</sup> Both microorganisms are among the ones that survive disinfecting protocol.<sup>7</sup>

*E. faecalis* can invade dentine tubules and can spread into peri-radicular which causes the formation of peri-radicular lesions after root canal treatment.<sup>8</sup> *P. gingivalis* can survive in the extra-radicular area mostly in the area approximate with root surface, and responsible for periodontitis and peri-radicular lesions.<sup>9</sup> *P. gingivalis* also contribute to root canal treatment failures, through its byproduct, lipopolysaccharide which effect biological processes, inflammation and tissue destruction.<sup>10</sup>

Different substances have been added to calcium hydroxide to increase its efficacy. The added substances are distilled water, saline, propylene glycol, chlorhexidine, glycerin, iodophors, barium sulfate, corticosteroids, antibiotics, anesthetic solution, methyl cellulose, and detergents. Iodophors has been added to calcium hydroxide to work with different bacterial characteristic.<sup>11</sup> Barium sulfate is added to calcium hydroxide aside from its antibacterial effect, it functions to increase radiopacity.<sup>12</sup>

Root canal treatment failure may happen even after dressing with calcium hydroxide if done incorrectly, depending on the right instrumentation and irrigation to remove inorganic and organic smear layers. In this research, the efficacy of calcium hydroxide addition with iodophors (CH-Iodophors) and barium sulfate (CH-Barium Sulfate) was analyzed on *E. faecalis* and *P. gingivalis*. Based on this background, the authors would like to purposely compare the antibacterial efficacy between CH-Iodophors and CH-Barium Sulfate root canal dressing on *E. faecalis* and *P. gingivalis*.

#### MATERIALS AND METHODS

This is an experimental laboratory study, conducted in Conservative Dentistry Section, Universitas Airlangga Dental Hospital and Microbiology Laboratory, Research Center, Faculty of Dental Medicine, Universitas Airlangga. This study was ethically approved by Universitas Airlangga Faculty of Dental Medicine Health Research Ethical Clearance Commission (166/KKEPK.FKG). Materials used in this research were commercially available calcium hydroxide dressings with Iodophors (Meta Biomed, Korea) and Barium Sulfate (Meta Biomed, Korea). *Enterococcus faecalis* and *Porphyromonas gingivalis* bacteria used in this study were stock bacteria previously cultured from patients who failed endodontic treatment.

The method used in this study was agar diffusion method using Mueller Hinton (MH) agar and Brain Heart Infusion (BHI) broth. The method is according to Alharthi et al. (2021) and Balouiri et al. (2016) with modifications on the sum and position of the wells in the plates.<sup>13,14</sup> The media were allocated for 4 groups of experiments. In the first group, 12 wells were prepared for CH-Iodophors (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*), and in the second group, 12 wells were prepared for CH-Barium Sulfate dressing (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*).

Both *E. faecalis* and *P. gingivalis* bacterial culture from stock was moved into separate reaction tube, each containing BHI broth and stirred. Incubation was done for both culture at 37°C for 24 hours in anaerobic condition. After 24 hours, *E. faecalis* and *P. gingivalis* bacterial culture in BHI broth was taken 0.5 ml using micro pipette, and poured into another reaction tube containing BHI broth until equal with 0.5 McFarland standard of turbidity.

The bacterial cultures were taken from BHI broth using sterile cotton swab and swabbed on the surface of each MH agar allocated for *E. faecalis* and *P gingivalis*. Antibacterial test was conducted by making the wells for the tested dressing materials (CH-Iodophors and CH-Barium Sulfate). The samples were incubated at 37°C for 48 hours in anaerobic condition. After 48 hours, measurements were conducted on antibacterial efficacy through inhibition zone measurement using a Vernier Digital Caliper (Mitutoyo, Japan). The clear zones of inhibition around the wells were measured, revealing no bacterial growth. Data of measurements on inhibition zone diameters (in millimeters) were collected for each sample wells.

Inhibition zones data were analyzed statistically and significance level was set at 5%. SPSS 20.0 for Windows (SPSS Inc., Chicago, Illinois, USA) was used in this study. Data normality was tested using Shapiro Wilk test. The significance was tested using independent ttest.

### RESULTS

The number of replications (n) for each treatment group was 6. Mean and standard deviation of inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* are shown in Table 1. The mean of *E. faecalis* inhibition zone from CH-Iodophors was 11.8125 mm and CH-Barium Sulfate was 6.3750 mm. The mean of *P. gingivalis* inhibition zone from CH-Iodophors was 12.7875 mm and CH-Barium Sulfate was 6.6750 mm.

Independent t-test was used in this study to check the significance between CH-Iodophors group and CH-Barium Sulfate group on *E. faecalis* and *P. gingivalis*. The significance between inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* growth is shown in Table 2. We found a significant difference between the two treatment groups on both bacteria. The CH-Iodophors group has higher antibacterial efficacy with wider antibacterial clear zone in both bacteria (p = 0.00001). CH-Barium Sulfate group has lower antibacterial efficacy with narrower antibacterial clear zone in both bacteria.

#### DISCUSSION

*Enterococcus faecalis* and *Porphyromonas gingivalis* were used in this study because these bacteria are opportunistic, living in facultative anaerobic condition, found in pathogenic state, and associated with failed root canal treatment.<sup>6</sup> These bacteria also have a major role in persistent root canal infections, can survive in the root canals and resistant to commonly used intracanal dressings.<sup>15</sup>

Antibacterial efficacy of CH-Iodophors and CH-Barium Sulfate dressing materials on *E. faecalis* and *P. gingivalis* growths were experimentally checked with agar diffusion method and inhibition clear zone were measured on the media. Inhibition zone is a laboratory calculation method to check whether a material have the ability to inhibit bacterial growth.<sup>14</sup> The diameter of clear zones would describe the strength of such material to inhibit bacterial growth, the stronger this material inhibit, the larger the clear zones would appear.

In this study Mueller Hinton agar was used because this media can grow *E. faecalis* and *P. gingivalis* actively and sensitive to drugs effect. Inhibition zones appeared on both groups of CH-Iodophors and CH-Barium Sulfate. This showed that each of these dressing materials have the ability to inhibit both *E. faecalis* and *P. gingivalis*.

The antibacterial properties of both root canal dressing materials mainly come from calcium hydroxide, which mechanism involve hydroxyl ion to kill bacteria by protein denaturation, cytoplasmic membrane and DNA destruction will physically inhibit this bacterial growth in the root canal systems.<sup>16</sup> Antibacterial activity is connected with alkali formation which can destruct lipid, the protein structure of bacteria and nucleic acid.<sup>17</sup> Direct contact of hydroxyl ion in alkaline pH with cytoplasmic membrane will destruct the hydrogen chain of protein polypeptide. Contact of hydroxyl ion with DNA will result in replication inhibition, causing lethal mutation. DNA is sensitive to temperature and pH change. In alkali condition DNA structure and function will break and leading to bacterial cell death. Alkali condition from calcium hydroxide would impact on the surrounding tissues, including healing, anti-inflammation<sup>18</sup> and cytotoxic which lead to apoptosis.<sup>19,20</sup>

The result of this study showed a significant difference between CH-Iodophors and CH-Barium Sulfate. The ability of CH-Iodophors to inhibit *E. faecalis* and *P. gingivalis* growths are significantly greater than CH-Barium Sulfate root canal dressing. In CH-Iodophors dressing, the iodophors substance will release iodine with high reactivity to promote protein oxidation. Iodophors functions as disinfectant and infection control. Thus, the combination of

CH and Iodophors will synergistically strengthen each other. Iodophors and calcium hydroxide can diffuse into dentinal tubules for added disinfection and eliminate the bacteria.

In CH-Barium Sulfate, barium sulfate can diffuse into dentinal tubule and also perform antibacterial activity. In this study the antimicrobial strength is not as good as CH-Iodophors. This may be caused by the form and structure of barium sulfate used in the mixture. Previous study on antimicrobial of barium sulfate showed that barium sulfate in micron particulate form would have potent antimicrobial properties.<sup>21</sup> Barium sulfate is generally used for its radiopacity effect on radiographic examination.<sup>12</sup> The addition of this material must consider its properties as it will affect the dressing's consistency and application.

*E. faecalis* and *P. gingivalis* can be killed with high pH level. High pH or an extreme alkaline environment would disturb the survival of most bacteria.<sup>22,23</sup> However, there are some studies stated that pH of higher than 11.5 is required for potent disinfection.<sup>6</sup> This study showed inhibition zone of CH-Iodophors is two folds higher than CH-Barium Sulfate, this may be caused by the combination of these two materials, calcium hydroxide and iodophors would create synergistic effect on the antimicrobial activity to inhibit *E. faecalis* growth. Even though this dressing material won't eliminate *E. faecalis or P. gingivalis*, the material is expected to weaken the bacteria, and eventually body defense mechanism will be able eliminate these bacteria and their byproducts.

Many researches have been done to found novel, biocompatible, antimicrobial and regenerating root canal dressing materials in order to support successful root canal treatment and enhance patient's satisfaction. The success of root canal treatment aside from the right choice of dressing, there are other factors such as the root canal systems complexity, diffusion ability of dressing materials within the dentinal tubules, and the effect on mixed cultured bacteria or biofilm formation, and bacterial resistance, which need to be evaluated in further studies. As there are many microorganisms involved in failed root canal treatments, <sup>24-26</sup> even after biomechanical and chemical instrumentations during treatment, individual and personalized assessment of these bacteria or biofilm is needed for ideal treatment. There are limitations, as this is only an in-vitro study and there are many factors to consider, both in-situ and in-vivo. In conclusion, calcium hydroxide-Iodophors root canal dressing has different (higher) antimicrobial efficacy on both *E. faecalis* and *P. gingivalis*. However, further studies need to be done regarding the effect on periodontal and periapical tissues with more complexities.

### ACKNOWLEDGEMENT

The authors would like to thank the Publication Center, Dental Hospital, and Research Center, Faculty of Dental Medicine, Universitas Airlangga for the given facility.

### REFERENCES

- Santoso CMA, Samadi K, Prasetyo EP, Wahjuningrum DA. Root canal cleanliness between mangosteen peel extract irrigant and NaOCl 2.5%. Conservative Dentistry Journal. 2020; 10(1): 40-3. doi: 10.20473/cdj.v10i1.2020.40-43.
- Juniarti DE, Kusumaningsih T, Soetojo A, Prasetyo EP, Sunur YK. Antibacterial activity and phytochemical analysis of ethanolic purple leaf extract (*Graptophyllum Pictum L. griff*) on *Lactobacillus acidophilus*. Mal J Med Health Sci. 2021; 17(Suppl2): 71-3.
- Astuti RHN, Samadi K, Prasetyo EP. Antibacterial activity of *Averrhoa bilimbi Linn*. leaf extract against Enterococcus faecalis. Conservative Dentistry Journal. 2016; 6(2): 93-8. doi: 10.20473/cdj.v6i2.2016.93-98.
- Prasetyo EP, Saraswati W, Goenharto S, Wahjuningrum D, Mooduto L, Rosidin RF, Tjendronegoro E. White pomegranate (*Punica granatum*) peels extract bactericidal potency on *Enterococcus faecalis*. Conservative Dentistry Journal. 2021; 11(2): 84-8. doi: 10.20473/cdj.v11i2.2021.84-88.
- Harseno S, Mooduto L, Prasetyo EP. Antibacterial potency of kedondong Bangkok leaves extract (*Spondias dulcis Forst.*) against Enterococcus faecalis bacteria. Conservative Dentistry Journal. 2016; 6(2): 110-6. doi: 10.20473/cdj.v6i2.2016.110-116.
- Prada I, Mico-Munoz P, Giner-Lluesma T, Mico-Martinez P, Collado-Castellano N, Manzano-Saiz A. Influence of microbiology on endodontic failure. Literature review. Med Oral Patol Oral Cir Bucal. 2019; 24(3): e364-72. doi: 10.4317/medoral.22907.
- Del Fabbro M, Samaranayake LP, Lolato A, Weinstein T, Taschieri S. Analysis of the secondary endodontic lesions focusing on the extraradicular microorganisms: an overview. J Investig Clin Dent. 2014; 5(4): 245-54. doi: 10.1111/jicd.12045.
- 8. Kaiwar A, Nadig G, Hegde J, Lekha S. Assessment of antimicrobial activity of endodontic sealers on Enterococcus faecalis: an in vitro study. World J Dent. 2012;

### Commented [BNR4]: the old literature should be change

3(1): 26-31. doi: 10.5005/jp-journals-10015-1123.

- Noguchi N, Noiri Y, Narimatsu M, Ebisu S. Identification and localization of extraradicular biofilm-forming bacteria associated with refractory endodontic pathogens. Appl Environ Microbiol. 2005; 71:8738-43. doi: 10.1128/AEM.71.12.8738-8743.2005.
- Kuntjoro M, Prasetyo EP, Cahyani F, Kamadjaja MJK, Hendrijantini N, Laksono H, Rahmania PN, Ariestania V, Nugraha AP, Ihsan IS, Dinaryanti A, Rantam FA. Lipopolysaccharide's cytotoxicity on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0048. doi: 10.1590/pboci.2020.153.
- Estrela C, Estrela CRDA, Hollanda ACB, Decurcio DDA, Pecora JD. Influence of iodophors on antimicrobial potential of calcium hydroxide. J Appl Oral Sci. 2006; 14(1): 33-7. doi: 10.1590/S1678-77572006000100007.
- Orucoglu H, Cobankara FK. Effect of unintentionally extruded calcium hydroxide paste including barium sulfate as a radiopaquing agent in treatment of teeth with periapical lesions: Report of a case. J Endod. 2008; 34(7): 888-91. doi: :10.1016/j.joen.2008.04.012.
- 13. Alharthi SS, Binshabaib M, Almasoud NS, Shawky HA, Aabed KF, Alomar TS, Albrekan AB, Alfaifi AJ, Melaibari AA. Myrrh mixed with silver nanoparticles demonstrates superior antimicrobial activity against *Porphyromonas gingivalis* compared to myrrh and silver nanoparticles alone. Saudi Dent J. 2021; 33: 890-6. doi: 10.1016/j.sdentj.2021.09.009.
- 14. Balouiri M, Sadiki M, Ibnsouda SK. Methods for in vitro evaluating antimicrobial activity: a review. J Pharm Anal. 2016; 6(2): 71-9. doi: 10.1016/j.jpha.2015.11.005.
- 15. Jhamb S, Singla R, Kaur A, Sharma J, Bhushan J. An in vitro determination of antibacterial effect of silver nanoparticles gel as an intracanal medicament in combination with other medicaments against Enterococcus faecalis. J Conserv Dent. 2019; 22: 479-82. doi: 10.4103/JCD.JCD\_113\_20.
- Mohammadi Z, Dummer PMH. Properties and applications of calcium hydroxide in endodontics and dental traumatology. Int Endod J. 2011; 44(8): 697-730. doi: 10.1111/j.1365-2591.2011.01886.x.
- Athanassiadis B, Abbott PV, Walsh LJ. The use of calcium hydroxide, antibiotics, and biocides as antimicrobial medicaments in Endodontics. Australian Dental Journal. 2007; 52(1): 64-82. doi: 10.1111/j.1834-7819.2007.tb00527.x.

- 18. Prasetyo EP, Kuntjoro M, Cahyani F, Goenharto S, Saraswati W, Juniarti DE, Hendrijantini N, Hariyani N, Nugraha AP, Rantam FA. Calcium hydroxide upregulates interleukin-10 expression in time dependent exposure and induces osteogenic differentiation of human umbilical cord mesenchymal stem cells. Int J Pharm Res. 2021; 13(1): 140-5. Doi: 10.31838/ijpr/2021.13.01.023.
- Prasetyo EP, Widjiastuti I, Cahyani F, Kuntjoro M, Hendrijantini N, Hariyani N, Winoto ER, Nugraha AP, Goenharto S, Susilowati H, Hendrianto E, Rantam FA. Cytotoxicity of calcium hydroxide on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0044. doi: 10.1590/pboci.2020.141.
- 20. Prasetyo EP, Kuntjoro M, Goenharto S, Juniarti DE, Cahyani F, Hendrijantini N, Nugraha AP, Hariyani N, Rantam FA. Calcium hydroxide increases human umbilical cord mesenchymal stem cells expressions of apoptotic protease-activating factor-1, caspase-3 and caspase-9. Clinical, cosmetic and investigational dentistry. 2021;13: 59-65. doi: 10.2147/CCIDE.S284240.
- Aninwene II G, Stout D, Yang Z, Webster TJ. Nano-BaSO4: a novel antimicrobial additive to pellethane. Int J Nanomedicine. 2013;8(1):1197-1205. doi: 10.2147/IJN.S40300.
- Jhajharia K, Parolia A, Shetty KV, Mehta LK. Biofilm in endodontics: a review. J Int Soc Prev Community Dent. 2015; 5: 1-12. doi: 10.4103/2231-0762.151956.
- Zancan RF, Vivan RR, Lopes MRM, Weckwerth PH, Andrade FB, Ponce JB, Duarte MAH. Antimicrobial activity and physicochemical properties of calcium hydroxide pastes used as intracanal medication. J Endod. 2016; 42(12): 1822-8. doi: 101016/j.joen.2016.08.017.
- Dudeja PG, Dudeja KK, Srivastava D, Grover S. Microorganisms in periradicular tissues: Do they exist? A perennial controversy. J Oral Maxillofac Pathol. 2015; 19(3): 356-63. doi: 10.4103/0973-029X.174612.
- Pereira RS, Rodrigues VAA, Furtado WT, Gueiros S, Pereira GS, Avila-Campos MJ. Microbial analysis of root canal and periradicular lesion associated to teeth with endodontic failure. Anaerobe. 2017; 48: 12-8. doi: 10.1016/j.anaerobe.2017.06.016.
- Siqueira JF, Antunes HS, Rocas IN, Rachid CTCC, Alves FRF. Microbiome in the apical root canal system of teeth with post-treatment apical periodontitis. PLoS ONE. 2016; 11(9): e0162887. doi: 10.1371/journal.pone.0162887.

Table 1:

Mean and standard deviation from inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* 

Groups	n	E. faecalis	P. gingivalis
		Mean <u>+</u> SD	Mean <u>+</u> SD
CH-Iodophors	6	11.8125 <u>+</u> 1.32001	12.7875 <u>+</u> 1.34961
CH-Barium Sulfate	6	6.3750 <u>+</u> 0.19494	6.6750 <u>+</u> 0.51865

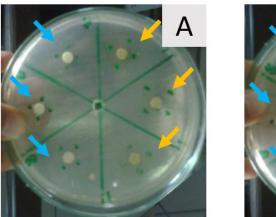
n = replication

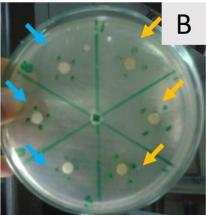
SD = standard deviation

# Table 2: Significance between inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis*

E. faecalis	P. gingivalis	
<b>CH-Iodophors</b>	<b>CH-Iodophors</b>	
0.00001*	-	
-	0.00001*	
	CH-Iodophors	

**Commented [BNR5]:** managing editor's review has not been revised : Please combine table 1 and table 2 into concise table





# Figure 1.

Agar diffusion assay containing *E. faecalis* (A) and *P. gingivalis* (B). The green dots indicate the inhibition area of each well. CH-Iodophors is pointed by yellow arrows, and CH-Barium Sulfate is pointed by blue arrows.

**Commented [BNR6]:** managing editor's review has not been revised : The figure is not per instruction and not following the standard of the journal, please revise Dear Editor of DJMKG,

Thank you for giving me the opportunity to submit a 2<sup>nd</sup> revision draft of my manuscript titled **Antibacterial efficacy between calcium hydroxide-iodophors and calcium hydroxide-barium sulfate root canal dressings on** *Enterococcus faecalis* and *Porphyromonas gingivalis in-vitro* to Dental Journal (Majalah Kedokteran Gigi). We appreciate the time and effort that you and the reviewers have dedicated to providing your valuable feedback on our manuscript. We are grateful to the reviewers for their insightful comments on our manuscript. We have been able to incorporate changes to reflect most of the suggestions provided by the reviewers. We have highlighted the changes within the manuscript.

Here is a point-by-point response to the reviewers' comments and concerns.

No.	Reviewer comments	Before revision	Revision	Notes
		Revision-2	• •	
	Managing Editor (ME)			
1.	Managing editor's review has not been revised: why they add CH iodoform and Ch barium sulfate? This sentence unclear	Already explained, it is a commercially available medicament. Explanation is available in introduction and discussion.	Already explained, it is a commercially available medicament. Explanation is available in introduction and discussion.	Page: 1 Paragraph: Abstract
2.	A. Compare the antibacterial efficacy between CH-lodoform and CH-Barium Sulfate root canal dressing on E. faecalis and P. gingivalis. B. Please add the design and setting of this study	A. of B. This study is an in vitro.	A. replaced B. Already available in the title, abstract and text, explained in detail in the	Page: 1 Paragraph: Abstract
	C. How determine the isolated bacteria was E. faecalis or P. gingivalis?	C. These bacteria are cultured and isolated by the microbiology laboratory of Research Center.	materials and methods. C. Already available in the title, abstract and text, explained in detail in the materials and methods.	
3.	The old literature should be change	Reference no 9. Reference no 10, 11, 16.	Reference is removed. Reference is replaced.	Page: 7 Paragraph: References
4.	Please combine table 1 and table 2 into concise table	2 tables	1 table, combined.	Page: 10 Paragraph: Table
5.	The figure is not per instruction and not following the standard of the journal, please revise	The figure is missing in the text.	The figure is mentioned in the text (Results).	Page: 11 Paragraph: Figure

In addition to the above comments, all spelling and grammatical errors pointed out by the reviewers have been corrected. We look forward to hearing from you in due time regarding our submission and to respond to any further questions and comments you may have.

For improvement about the language editing, layout, tables and figure please arrange accordingly, thank you.

Sincerely,

Eric Priyo Prasetyo Date: March 8, 2022.

# Antibacterial efficacy between calcium hydroxide-iodophors and calcium hydroxide-barium sulfate root canal dressings on *Enterococcus faecalis* and *Porphyromonas gingivalis in-vitro*

# Eric Priyo Prasetyo,<sup>1</sup> Devi Eka Juniarti,<sup>1</sup> Galih Sampoerno,<sup>1</sup> Dian Agustin Wahjuningrum,<sup>1</sup> Ananta Tantri Budi,<sup>1</sup> Dyanita Hasri,<sup>2</sup> Evelyn Tjendronegoro<sup>3</sup>

<sup>1</sup>Department of Conservative Dentistry, Faculty of Dental Medicine, Universitas Airlangga, Surabaya, Indonesia. <sup>2</sup>Dentistry Program, Faculty of Dental Medicine, Universitas Airlangga, Surabaya, Indonesia.

<sup>3</sup>Healthcare and Research, Irvine Medical Center, University of California, California, United States of America.

### **ABSTRACT**

**Background:** A successful endodontic treatment is inseparable from the right choice of root canal dressing. The right choice of medicaments would result in patient satisfaction. Enterococcus faecalis (E. faecalis) and Porphyromonas gingivalis (P. gingivalis) are usually found in failed root canal treatments. Calcium hydroxide is a gold standard dressing which create an alkaline environment in the root canal and have a bactericidal effect. Commercially, there are calcium hydroxide dressings with supporting additions, including calcium hydroxide-iodophors (CH-Iodophors) and Calcium hydroxide-barium sulfate (CH-Barium Sulfate). **Purpose:** This study aimed to compare the antibacterial efficacy between CH-Iodophors and CH-Barium Sulfate root canal dressing on E. faecalis and P. gingivalis. Methods: CH-Iodophors and CH-Barium Sulfate was obtained commercially. E. faecalis and P. gingivalis were obtained from stock culture taken from the root canal of failed endodontic treatment. E. faecalis and P. gingivalis were cultured in petri dishes and for each bacterium, 12 wells were made in the media, 6 wells were used for CH-Iodophors group and 6 wells were used for CH-Barium Sulfate group. CH-Iodophors and CH-Barium Sulfate were deployed in the wells in E. faecalis and P. gingivalis cultured medium in the petri dishes. After incubation, the inhibition zone diameters were measured. Independent t-test was used for analysis and significance level was set at 5%. Results: There is a significant antibacterial efficacy between CH-Iodophors and CH-Barium Sulfate on E. faecalis and P. gingivalis (p=0.00001). Conclusion: CH-Iodophors has a higher antibacterial efficacy than CH-Barium Sulfate on both E. faecalis and P. gingivalis.

*Keywords:* Enterococcus faecalis; Porphyromonas gingivalis; calcium hydroxide; iodophors; barium sulfate; patient satisfaction.

Correspondence: Eric Priyo Prasetyo, Department of Conservative Dentistry, Faculty of Dental Medicine, Universitas Airlangga, Jalan Mayjend. Prof. Dr. Moestopo 47, Surabaya, Indonesia, Email address: eric-p-p@fkg.unair.ac.id.

# **INTRODUCTION**

The right choice of root canal dressing to eliminate bacteria in the root canal is important for a successful endodontic treatment. A successful endodontic treatment would result in patient's satisfaction. Many attempts were conducted to increase the success of root canal treatment, including finding the efficient instrumentation, effective cleaning,<sup>1,2</sup> antibacterial dressing and irrigation materials.<sup>3-5</sup>

Root canal treatment has a high success rate, but in some cases, there are failures. Isolated bacteria from root canal treatment failures and the prevalence are about 45.8-77% caused by *Enterococcus faecalis (E. faecalis)* and 28.17% caused by *Porphyromonas gingivalis (P. gingivalis)* in the root canal system.<sup>6</sup> Both microorganisms are among the ones that survive disinfecting protocol.<sup>7</sup>

*E. faecalis* can invade dentine tubules and can spread into peri-radicular which causes the formation of peri-radicular lesions after root canal treatment.<sup>8</sup> *P. gingivalis* can survive in the extra-radicular area mostly in the area approximate with root surface, and responsible for periodontitis and peri-radicular lesions.<sup>6</sup> *P. gingivalis* also contribute to root canal treatment failures, through its byproduct, lipopolysaccharide which effect biological processes, inflammation and tissue destruction.<sup>9</sup>

Different substances have been added to calcium hydroxide to increase its efficacy. The added substances are distilled water, saline, propylene glycol, chlorhexidine, glycerin, iodophors, barium sulfate, corticosteroids, antibiotics, anesthetic solution, methyl cellulose, and detergents. Iodophors has been added to calcium hydroxide to work with different bacterial characteristic.<sup>10</sup> Barium sulfate is added to calcium hydroxide aside from its antibacterial effect, it functions to increase radiopacity.<sup>11</sup>

Root canal treatment failure may happen even after dressing with calcium hydroxide if done incorrectly, depending on the right instrumentation and irrigation to remove inorganic and organic smear layers. In this research, the efficacy of calcium hydroxide addition with iodophors (CH-Iodophors) and barium sulfate (CH-Barium Sulfate) was analyzed on *E. faecalis* and *P. gingivalis*. Based on this background, the authors would like to purposely compare the antibacterial efficacy between CH-Iodophors and CH-Barium Sulfate root canal dressing on *E. faecalis* and *P. gingivalis*.

## **MATERIALS AND METHODS**

This is an experimental laboratory study, conducted in Conservative Dentistry Section, Universitas Airlangga Dental Hospital and Microbiology Laboratory, Research Center, Faculty of Dental Medicine, Universitas Airlangga. This study was ethically approved by Universitas Airlangga Faculty of Dental Medicine Health Research Ethical Clearance Commission (166/KKEPK.FKG). Materials used in this research were commercially available calcium hydroxide dressings with Iodophors (Meta Biomed, Korea) and Barium Sulfate (Meta Biomed, Korea). *Enterococcus faecalis* and *Porphyromonas gingivalis* bacteria used in this study were stock bacteria previously cultured from patients who failed endodontic treatment.

The method used in this study was agar diffusion method using Mueller Hinton (MH) agar and Brain Heart Infusion (BHI) broth. The method is according to Alharthi et al. (2021) and Balouiri et al. (2016) with modifications on the sum and position of the wells in the plates.<sup>12,13</sup> The media were allocated for 4 groups of experiments. In the first group, 12 wells were prepared for CH-Iodophors (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*), and in the second group, 12 wells were prepared for CH-Barium Sulfate dressing (6 wells with *E. faecalis* and 6 wells with *P. gingivalis*).

Both *E. faecalis* and *P. gingivalis* bacterial culture from stock was moved into separate reaction tube, each containing BHI broth and stirred. Incubation was done for both culture at 37°C for 24 hours in anaerobic condition. After 24 hours, *E. faecalis* and *P. gingivalis* bacterial culture in BHI broth was taken 0.5 ml using micro pipette, and poured into another reaction tube containing BHI broth until equal with 0.5 McFarland standard of turbidity.

The bacterial cultures were taken from BHI broth using sterile cotton swab and swabbed on the surface of each MH agar allocated for *E. faecalis* and *P gingivalis*. Antibacterial test was conducted by making the wells for the tested dressing materials (CH-Iodophors and CH-Barium Sulfate). The samples were incubated at 37°C for 48 hours in anaerobic condition. After 48 hours, measurements were conducted on antibacterial efficacy through inhibition zone measurement using a Vernier Digital Caliper (Mitutoyo, Japan). The clear zones of inhibition around the wells were measured, revealing no bacterial growth. Data of measurements on inhibition zone diameters (in millimeters) were collected for each sample wells.

Inhibition zones data were analyzed statistically and significance level was set at 5%. SPSS 20.0 for Windows (SPSS Inc., Chicago, Illinois, USA) was used in this study. Data

normality was tested using Shapiro Wilk test. The significance was tested using independent ttest.

# RESULTS

The number of replications (n) for each treatment group was 6. Mean and standard deviation of inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* are shown in Table 1. The mean of *E. faecalis* inhibition zone from CH-Iodophors was 11.8125 mm and CH-Barium Sulfate was 6.3750 mm. The mean of *P. gingivalis* inhibition zone from CH-Iodophors was 12.7875 mm and CH-Barium Sulfate was 6.6750 mm.

Independent t-test was used in this study to check the significance between CH-Iodophors group and CH-Barium Sulfate group on *E. faecalis* and *P. gingivalis*. The significance between inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* growth is shown in Table 1. We found a significant difference between the two treatment groups on both bacteria. The CH-Iodophors group has higher antibacterial efficacy with wider antibacterial clear zone in both bacteria (p = 0.00001). CH-Barium Sulfate group has lower antibacterial efficacy with narrower antibacterial clear zone in both bacteria.

## DISCUSSION

*Enterococcus faecalis* and *Porphyromonas gingivalis* were used in this study because these bacteria are opportunistic, living in facultative anaerobic condition, found in pathogenic state, and associated with failed root canal treatment.<sup>6</sup> These bacteria also have a major role in persistent root canal infections, can survive in the root canals and resistant to commonly used intracanal dressings.<sup>14</sup>

Antibacterial efficacy of CH-Iodophors and CH-Barium Sulfate dressing materials on *E*. *faecalis* and *P. gingivalis* growths were experimentally checked with agar diffusion method and inhibition clear zone were measured on the media. Inhibition zone is a laboratory calculation method to check whether a material have the ability to inhibit bacterial growth.<sup>13</sup> The diameter of clear zones would describe the strength of such material to inhibit bacterial growth, the stronger this material inhibit, the larger the clear zones would appear.

In this study Mueller Hinton agar was used because this media can grow *E. faecalis* and *P. gingivalis* actively and sensitive to drugs effect. Inhibition zones appeared on both groups of CH-Iodophors and CH-Barium Sulfate. This showed that each of these dressing materials have the ability to inhibit both *E. faecalis* and *P. gingivalis*.

The antibacterial properties of both root canal dressing materials mainly come from calcium hydroxide, which mechanism involve hydroxyl ion to kill bacteria by protein denaturation, cytoplasmic membrane and DNA destruction will physically inhibit this bacterial growth in the root canal systems.<sup>15</sup> Antibacterial activity is connected with alkali formation which can destruct lipid, the protein structure of bacteria and nucleic acid.<sup>16</sup> Direct contact of hydroxyl ion in alkaline pH with cytoplasmic membrane will destruct the hydrogen chain of protein polypeptide. Contact of hydroxyl ion with DNA will result in replication inhibition, causing lethal mutation. DNA is sensitive to temperature and pH change. In alkali condition DNA structure and function will break and leading to bacterial cell death. Alkali condition from calcium hydroxide would impact on the surrounding tissues, including healing, anti-inflammation<sup>17</sup> and cytotoxic which lead to apoptosis.<sup>18,19</sup>

The result of this study showed a significant difference between CH-Iodophors and CH-Barium Sulfate. The ability of CH-Iodophors to inhibit *E. faecalis* and *P. gingivalis* growths are significantly greater than CH-Barium Sulfate root canal dressing. In CH-Iodophors dressing, the iodophors substance will release iodine with high reactivity to promote protein oxidation. Iodophors functions as disinfectant and infection control. Thus, the combination of

CH and Iodophors will synergistically strengthen each other. Iodophors and calcium hydroxide can diffuse into dentinal tubules for added disinfection and eliminate the bacteria.

In CH-Barium Sulfate, barium sulfate can diffuse into dentinal tubule and also perform antibacterial activity. In this study the antimicrobial strength is not as good as CH-Iodophors. This may be caused by the form and structure of barium sulfate used in the mixture. Previous study on antimicrobial of barium sulfate showed that barium sulfate in micron particulate form would have potent antimicrobial properties.<sup>20</sup> Barium sulfate is generally used for its radiopacity effect on radiographic examination.<sup>11</sup> The addition of this material must consider its properties as it will affect the dressing's consistency and application.

*E. faecalis* and *P. gingivalis* can be killed with high pH level. High pH or an extreme alkaline environment would disturb the survival of most bacteria.<sup>21,22</sup> However, there are some studies stated that pH of higher than 11.5 is required for potent disinfection.<sup>6</sup> This study showed inhibition zone of CH-Iodophors is two folds higher than CH-Barium Sulfate, this may be caused by the combination of these two materials, calcium hydroxide and iodophors would create synergistic effect on the antimicrobial activity to inhibit *E. faecalis* growth. Even though this dressing material won't eliminate *E. faecalis or P. gingivalis*, the material is expected to weaken the bacteria, and eventually body defense mechanism will be able eliminate these bacteria and their byproducts.

Many researches have been done to found novel, biocompatible, antimicrobial and regenerating root canal dressing materials in order to support successful root canal treatment and enhance patient's satisfaction. The success of root canal treatment aside from the right choice of dressing, there are other factors such as the root canal systems complexity, diffusion ability of dressing materials within the dentinal tubules, and the effect on mixed cultured bacteria or biofilm formation, and bacterial resistance, which need to be evaluated in further studies. As there are many microorganisms involved in failed root canal treatments, <sup>23-25</sup> even after biomechanical and chemical instrumentations during treatment, individual and personalized assessment of these bacteria or biofilm is needed for ideal treatment. There are limitations, as this is only an in-vitro study and there are many factors to consider, both in-situ and in-vivo. In conclusion, calcium hydroxide-Iodophors root canal dressing has different (higher) antimicrobial efficacy on both *E. faecalis* and *P. gingivalis*. However, further studies need to be done regarding the effect on periodontal and periapical tissues with more complexities.

# ACKNOWLEDGEMENT

The authors would like to thank the Publication Center, Dental Hospital, and Research Center, Faculty of Dental Medicine, Universitas Airlangga for the given facility.

# REFERENCES

- Santoso CMA, Samadi K, Prasetyo EP, Wahjuningrum DA. Root canal cleanliness between mangosteen peel extract irrigant and NaOCl 2.5%. Conservative Dentistry Journal. 2020; 10(1): 40-3. doi: 10.20473/cdj.v10i1.2020.40-43.
- Juniarti DE, Kusumaningsih T, Soetojo A, Prasetyo EP, Sunur YK. Antibacterial activity and phytochemical analysis of ethanolic purple leaf extract (*Graptophyllum Pictum L. griff*) on *Lactobacillus acidophilus*. Mal J Med Health Sci. 2021; 17(Suppl2): 71-3.
- Astuti RHN, Samadi K, Prasetyo EP. Antibacterial activity of *Averrhoa bilimbi Linn*. leaf extract against Enterococcus faecalis. Conservative Dentistry Journal. 2016; 6(2): 93-8. doi: 10.20473/cdj.v6i2.2016.93-98.
- Prasetyo EP, Saraswati W, Goenharto S, Wahjuningrum D, Mooduto L, Rosidin RF, Tjendronegoro E. White pomegranate (*Punica granatum*) peels extract bactericidal potency on *Enterococcus faecalis*. Conservative Dentistry Journal. 2021; 11(2): 84-8. doi: 10.20473/cdj.v11i2.2021.84-88.
- Harseno S, Mooduto L, Prasetyo EP. Antibacterial potency of kedondong Bangkok leaves extract (*Spondias dulcis Forst.*) against Enterococcus faecalis bacteria. Conservative Dentistry Journal. 2016; 6(2): 110-6. doi: 10.20473/cdj.v6i2.2016.110-116.
- Prada I, Mico-Munoz P, Giner-Lluesma T, Mico-Martinez P, Collado-Castellano N, Manzano-Saiz A. Influence of microbiology on endodontic failure. Literature review. Med Oral Patol Oral Cir Bucal. 2019; 24(3): e364-72. doi: 10.4317/medoral.22907.
- Del Fabbro M, Samaranayake LP, Lolato A, Weinstein T, Taschieri S. Analysis of the secondary endodontic lesions focusing on the extraradicular microorganisms: an overview. J Investig Clin Dent. 2014; 5(4): 245-54. doi: 10.1111/jicd.12045.
- Kaiwar A, Nadig G, Hegde J, Lekha S. Assessment of antimicrobial activity of endodontic sealers on Enterococcus faecalis: an in vitro study. World J Dent. 2012; 3(1): 26-31. doi: 10.5005/jp-journals-10015-1123.

- Kuntjoro M, Prasetyo EP, Cahyani F, Kamadjaja MJK, Hendrijantini N, Laksono H, Rahmania PN, Ariestania V, Nugraha AP, Ihsan IS, Dinaryanti A, Rantam FA. Lipopolysaccharide's cytotoxicity on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0048. doi: 10.1590/pboci.2020.153.
- 10. Najjar RS, Alamoudi NM, El-Housseiny AA, Al-Tuwirqi AA, Sabbagh HJ. A comparison of calcium hydroxide/iodoform paste and zinc oxide eugenol as root filling materials for pulpectomy in primary teeth: a systematic review and meta-analysis. Clin Exp Dent Res. 2019; 5: 294-310. doi: 10.1002/cre2.173.
- Ba-Hattab R, Al-Jamie M, Aldreib H, Alessa L, Alonazi M. Calcium hydroxide in endodontics: an overview. Open Journal of Stomatology. 2016; 6(12): 274-89. doi: :10.4236/ojst.2016.612033.
- 12. Alharthi SS, Binshabaib M, Almasoud NS, Shawky HA, Aabed KF, Alomar TS, Albrekan AB, Alfaifi AJ, Melaibari AA. Myrrh mixed with silver nanoparticles demonstrates superior antimicrobial activity against *Porphyromonas gingivalis* compared to myrrh and silver nanoparticles alone. Saudi Dent J. 2021; 33: 890-6. doi: 10.1016/j.sdentj.2021.09.009.
- 13. Balouiri M, Sadiki M, Ibnsouda SK. Methods for in vitro evaluating antimicrobial activity: a review. J Pharm Anal. 2016; 6(2): 71-9. doi: 10.1016/j.jpha.2015.11.005.
- 14. Jhamb S, Singla R, Kaur A, Sharma J, Bhushan J. An in vitro determination of antibacterial effect of silver nanoparticles gel as an intracanal medicament in combination with other medicaments against Enterococcus faecalis. J Conserv Dent. 2019; 22: 479-82. doi: 10.4103/JCD.JCD\_113\_20.
- 15. Mohammadi Z, Dummer PMH. Properties and applications of calcium hydroxide in endodontics and dental traumatology. Int Endod J. 2011; 44(8): 697-730. doi: 10.1111/j.1365-2591.2011.01886.x.
- 16. Sharma G, Ahmed HMA, Zilm PS, Rossi-Fedele G. Antimicrobial properties of calcium hydroxide dressing when used for long-term application: a systematic review. Aust Endod J. 2018; 44: 60-5. doi: 10.1111/aej.12216.
- 17. Prasetyo EP, Kuntjoro M, Cahyani F, Goenharto S, Saraswati W, Juniarti DE, Hendrijantini N, Hariyani N, Nugraha AP, Rantam FA. Calcium hydroxide upregulates interleukin-10 expression in time dependent exposure and induces osteogenic differentiation of human umbilical cord mesenchymal stem cells. Int J Pharm Res. 2021; 13(1): 140-5. Doi: 10.31838/ijpr/2021.13.01.023.

- Prasetyo EP, Widjiastuti I, Cahyani F, Kuntjoro M, Hendrijantini N, Hariyani N, Winoto ER, Nugraha AP, Goenharto S, Susilowati H, Hendrianto E, Rantam FA. Cytotoxicity of calcium hydroxide on human umbilical cord mesenchymal stem cells. Pesqui Bras Odontopediatria Clin Integr. 2020; 20: e0044. doi: 10.1590/pboci.2020.141.
- 19. Prasetyo EP, Kuntjoro M, Goenharto S, Juniarti DE, Cahyani F, Hendrijantini N, Nugraha AP, Hariyani N, Rantam FA. Calcium hydroxide increases human umbilical cord mesenchymal stem cells expressions of apoptotic protease-activating factor-1, caspase-3 and caspase-9. Clinical, cosmetic and investigational dentistry. 2021;13: 59-65. doi: 10.2147/CCIDE.S284240.
- 20. Aninwene II G, Stout D, Yang Z, Webster TJ. Nano-BaSO4: a novel antimicrobial additive to pellethane. Int J Nanomedicine. 2013;8(1):1197-1205. doi: 10.2147/IJN.S40300.
- Jhajharia K, Parolia A, Shetty KV, Mehta LK. Biofilm in endodontics: a review. J Int Soc Prev Community Dent. 2015; 5: 1-12. doi: 10.4103/2231-0762.151956.
- 22. Zancan RF, Vivan RR, Lopes MRM, Weckwerth PH, Andrade FB, Ponce JB, Duarte MAH. Antimicrobial activity and physicochemical properties of calcium hydroxide pastes used as intracanal medication. J Endod. 2016; 42(12): 1822-8. doi: 101016/j.joen.2016.08.017.
- Dudeja PG, Dudeja KK, Srivastava D, Grover S. Microorganisms in periradicular tissues: Do they exist? A perennial controversy. J Oral Maxillofac Pathol. 2015; 19(3): 356-63. doi: 10.4103/0973-029X.174612.
- 24. Pereira RS, Rodrigues VAA, Furtado WT, Gueiros S, Pereira GS, Avila-Campos MJ. Microbial analysis of root canal and periradicular lesion associated to teeth with endodontic failure. Anaerobe. 2017; 48: 12-8. doi: 10.1016/j.anaerobe.2017.06.016.
- 25. Siqueira JF, Antunes HS, Rocas IN, Rachid CTCC, Alves FRF. Microbiome in the apical root canal system of teeth with post-treatment apical periodontitis. PLoS ONE. 2016; 11(9): e0162887. doi: 10.1371/journal.pone.0162887.

# Table 1:

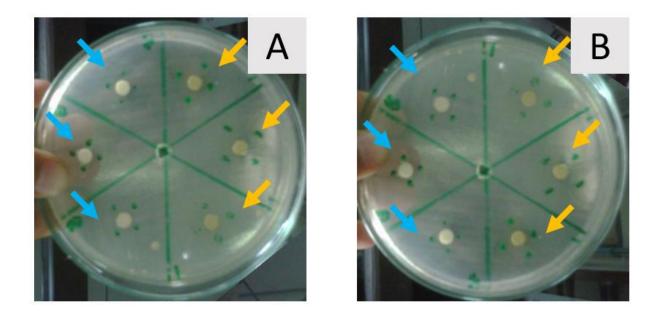
Mean, standard deviation, and significance from inhibition zone diameter of CH-Iodophors and CH-Barium Sulfate on *E. faecalis* and *P. gingivalis* 

Groups	n	<mark>E. faecalis</mark>	<mark>P value</mark>	P. gingivalis	<mark>P value</mark>
		<mark>Mean <u>+</u> SD</mark>		<mark>Mean <u>+</u> SD</mark>	
CH-Iodophors	<mark>6</mark>	<u>11.8125 ± 1.32001</u>	<mark>0.00001*</mark>	12.7875 <u>+</u> 1.34961	<mark>0.00001*</mark>
CH-Barium Sulfate	<mark>6</mark>	<mark>6.3750 <u>+</u> 0.19494</mark>		<u>6.6750 <u>+</u> 0.51865</u>	

n = replication

SD = standard deviation

\*= statistically significant



# Figure 1.

Agar diffusion assay containing *E. faecalis* (A) and *P. gingivalis* (B). The green dots indicate the inhibition area of each well. CH-Iodophors is pointed by yellow arrows, and CH-Barium Sulfate is pointed by blue arrows.



eric priyo prasetyo <eric-p-p@fkg.unair.ac.id>

# Acceptance letter

**Dental Journal (Majalah Kedokteran Gigi)** <dental\_journal@fkg.unair.ac.id> Wed, Mar 16, 2022 at 9:30 AM To: eric priyo prasetyo <eric-p-p@fkg.unair.ac.id>, Eric Prasetyo <eppfkgunair@gmail.com>

Dear

# Eric Priyo Prasetyo,

Faculty of Dental Medicine, Universitas Airlangga Surabaya, Indonesia

We have reached a decision regarding your submission to Dental Journal (Majalah Kedokteran Gigi), Effects of tooth preparation on the microleakage of fissure sealantAntibacterial efficacy between calcium hydroxideiodophors and calcium hydroxide-barium sulfate root canal dressings on Enterococcus faecalis and Porphyromonas gingivalis in-vitro.

Authors: Eric Priyo Prasetyo, Devi Eka Juniarti, Galih Sampoerno, Dian Agustin Wahjuningrum, Ananta Tantri Budi, Dyanita Hasri, Evelyn Tjendronegoro

Our decision is to: Accept your manuscript

It will be published by Dental Journal (Majalah Kedokteran Gigi) on volume 55, issue 2 – 2022. Articles will go through the process of copyediting (including plagiarism check), proofreading, layouting and publishing. In the attachment files we provide the copyediting file and plagiarism check. **Manuscript is in the proofreading stage. We'll let you know when it's finished.** 

Thank you for your submission. Your next manuscript is very welcome. Best Regards,

Muhammad Dimas Aditya Ari

Dental Journal (Majalah Kedokteran Gigi)

http://e-journal.unair.ac.id/MKG

Faculty of Dental Medicine, Universitas Airlangga JI. Mayjen Prof. Dr. Moestopo 47 Surabaya 60132 INDONESIA Phone/Fax: +62 31 5039478

2 attachments

Eric (EDIT).docx 684K

Turnitin 11%\_Eric.pdf