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Woro Hastuti Satyantini <worohastuti79@gmail.com>  
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Mar 6, 2020, 1:13 AM

Assalamualaikum,  
Rozi, ini revisi terakhir, yaitu perbaikan di penulisan vaname shrimp yang seharusnya ditulis white shrimp, penulisan perlakuan masih ada yang tertulis P0,P1, P2, P3. Corresponding author yang tidak sesuai.  
Terima kasih.

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**Rozi S.Pi.,M.Biotech.** <rozi@fpk.unair.ac.id> to me Thu, Mar 19, 2020, 1:21 PM

Dr. Woro Hastuti Satyantini, Ir., M.Si:

Thank you for reviewing the copyediting of your manuscript, "Application of Probiotics for Organic Matter and Enhancement of Growth Performance in White Shrimp (*Litopenaeus vannamei*)," for Jurnal Ilmiah Perikanan dan Kelautan. We look forward to publishing this work.

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Noted with thanks. Thanks a lot. Well received with thanks.



## Research Article

### Application of Probiotics for Organic Matter and Enhancement of Growth Performance in Vaname Shrimp (*Litopenaeus vannamei*)

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#### Abstract

Intensive shrimp farming has raised environmental issues due to aquaculture pollution, which is caused by uneaten feed, feces, organic matter, and toxic compounds (ammonia and H<sub>2</sub>S). One way to solve this issue is by using probiotics. Thus, this study aimed at investigating the effect of probiotic administration (*Pseudomonas* sp. and *Bacillus* sp.) at varying concentrations on total organic matter in the rearing water, feed conversion ratio, feed efficiency, and growth rate of vaname shrimp (*Litopenaeus vannamei*). This study used a Completely Randomized Design consisting of four treatments: T<sub>0</sub> (without the probiotic supplementations), T<sub>1</sub> (probiotics at 10<sup>6</sup> CFU/ml), T<sub>2</sub> (probiotics at 10<sup>7</sup> CFU/mL), T<sub>3</sub> (probiotics at 10<sup>8</sup> CFU/mL) with three replicates each. The results showed that the administration of probiotics at different concentrations had a significant effect on the total amount of organic matter, feed conversion ratio, feed efficiency, and specific growth rate of vaname shrimp. Administration of probiotics at a concentration of 10<sup>7</sup> CFU/ml had the lowest organic matter in the rearing water, 138 mg/L. While probiotic administration at 10<sup>8</sup> CFU/ml gave the lowest feed conversion ratio (1.22), the highest feed efficiency (81.57%) and the best growth rate (2.25%/day). This research suggests that the administration of 10<sup>7</sup>-10<sup>8</sup> CFU/ml probiotics could be applied to reduce organic content in the rearing water and improve the growth performances of vaname shrimp.

Keywords:

**Keywords:**  
*Bacillus* sp.  
*Pseudomonas* sp.  
organic matter  
growth performances  
vaname shrimp.

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## 1. Introduction

Shrimp production could be increased by an intensive aquaculture system, in which more inputs, such as shrimp larvae and feed are required (Ekasari, 2009). However, the intensive aquaculture system has negative impacts, such as increasing the amount of organic matter, food waste and feces, increasing the density of phytoplankton, more toxic compounds such as  $\text{NH}_3$  and  $\text{H}_2\text{S}$  and increasing disease transmission in cultured organisms (Herdianti et al., 2015). Environmental factors, especially water quality determine the success of shrimp farming. Prevention of water quality deterioration can be achieved by giving probiotic from the beginning of the culture period to degrade organic material in the pond (Irianto, 2003). The application of probiotics in aquaculture has been used to maintain rearing water quality in several fish species such as tilapia (Wang et al., 2008), snapper (Rengpipat et al., 2008), and vaname shrimp (Kumar et al., 2016). Furthermore, Yuvaraj and Karthik (2015) stated that the latest effort to improve water quality in aquaculture is the application of probiotics and or enzymes in shrimp ponds.

*Pseudomonas* sp. and *Bacillus* sp. are bacterial decomposers (biodegradation) or what is called probiotic bacteria. According to Edy and Busono (2000), microorganisms that have a capacity to decompose organic materials in rearing water could be classified as products of biotechnology. These types of bacteria were generally obtained from nature and had been selected due to their capacity to help the process of purification of organic materials in the water. Bacteria could be used to remove unwanted components in the aquaculture media, such as ammonium. The main strength and driving force for the operation of this system is the intensive growth of heterotrophic bacteria (Avnimelech, 2006).

Yudiarti et al. (2010) documented that the administration of probiotics could reduce the total population of *Vibrio*, total ammonia levels, and organic matter. According to Aslamyah (2011), the application of bioremediation probiotic *Bacillus* sp. with a concentration of 0.5 ppm per week in the culture medium could improve the water quality and the activity of the  $\alpha$ -amylase and protease enzymes, as well as could increase the survival and growth of vaname shrimp. A study by Eissa et al. (2014) showed that the administration of *Pseudomonas fluorescens* at a concentration of  $10^8$  CFU/g could protect cultured organisms against pathogenic bacteria in tilapia. Linggarjati et al. (2013) reported that the use of *Bacillus* with a density of  $10^6$  CFU/ml reduced the content of ammonia ( $\text{NH}_3$ ), nitrite ( $\text{NO}_2^-$ ),

and the total *Vibrio* sp. While Wang and Gu (2010) also showed that the administration of probiotic *Lactobacillus acidophilus* RS058, *Rhodopseudomonas palustris* GH642, and *Bacillus coagulans* NJ105 respectively in vaname shrimp gave daily growth and specific growth

rates higher than controls (without probiotics). The results in the study of Cha et al. (2013) showed that the administration of *B. subtilis* had a beneficial effect on growth performance, non-specific immunity, and disease resistance in flounder fish and maintenance water quality.

Acknowledging the information above, this study was aimed to investigate the effect of probiotic administration (*Pseudomonas* sp. and *Bacillus* sp.) at the different concentration on the amount of organic matter in rearing water and growth performance of vaname shrimp (*Litopenaeus vannamei*).

## 2. Materials and Methods

### Place and Time

This research was performed in the Laboratory of Fish Health Management, the Brackish Water Aquaculture Development Centre, Jepara, Indonesia from September 2018 to February 2019.

### Shrimp

Vaname shrimps (DOC 35, 8-9 cm length, and weighing 5-7 g) were collected from pond farmers around the Brackish Water Development Centre, Jepara. The shrimps were in healthy condition and not infected by a disease such as *Vibrio*.

### Probiotic Isolates

Three probiotics (*Pseudomonas* sp., *Bacillus* sp. SDB<sub>14</sub>, dan *Bacillus* sp. SDB<sub>16</sub>) originated from sediment of shrimp ponds and were not toxic. Each bacterial isolate was isolated separately using sterile Tryptic soya Broth (TSB) and incubated at 37°C for 24 h.

### Experimental Design

The present study used completely randomized design (CRD) with four treatments T0 (no probiotic addition), T1 (probiotic at a concentration of  $10^6$  CFU/ml), T2 (probiotic at  $10^7$ CFU/ml), T3 (probiotic at  $10^8$  CFU/ml) and each treatment had three replicates.

### Probiotic Preparation

*Pseudomonas* sp., *Bacillus* sp. SDB14, and *Bacillus* sp. SDB16 were cultured in a modified culture media. The modified media were made by preparing 50 ml of sterile *tofu*-waste liquid, 25 ml molasses, 6 g glucose solution, and 15 g NaCl dissolved in 1L distilled water.

Then, 100 g skim milk was dissolved in 1L distilled water and sterilized by heating in a water bath at a temperature of 70-80 ° C for 10 minutes. After the sterilization process was completed, a 10% skim milk solution was added into 900 ml of the modified culture media. There-

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after, the probiotic isolates were isolated from *Pseudomonas* sp., *Bacillus* sp. SDB14 and *Bacillus* sp. SDB16 of  $10^5$  CFU / ml each.

*Probiotic Administration*

The administration of probiotics was conducted through rearing water twice a week with a dose of 1 ml/liter and adjusted according to the treatments  $10^6$  CFU/ml,  $10^7$  CFU/ml, and  $10^8$  CFU / ml.

*Culturing of Vaname Shrimp*

The vaname shrimps were cultured in brackish water with 15 ppt salinity (Suharyadi, 2011), and fed with commercial pellet containing 35% protein content (Nur, 2011). The feeding was performed four times a day (Widagdo, 2011), with the feeding rate at 3% BW per day. The feces secreted by vaname shrimps was siphoned off every seven days.

*2.10 Data Analysis*

Data were analyzed using Analysis of Variance (ANOVA) to determine the effect of each treatment (Kusriningrum, 2008). When the ANOVA test results were significantly different, the Duncan test (DMRT) was performed to determine the difference among treatments.

**3. Result and Discussion**

The main source of organic matter in aquaculture pond is organic fertilizer, feces, uneaten feed, phytoplankton, zooplankton and benthos, zooplankton, and benthos (Boyd, 2016). The measurement of organic matter is very important in this present study to investigate the role of administered probiotics to reduce organic matter content in the shrimp aquaculture system and contribute to the growth performance of shrimps.

**Table 1.** Organic-matter content in rearing water of vaname shrimp aquaculture after administration of probiotics.

Treatment	Organic content in the rearing water (mg/l±SD)				
	Day-0	Day-7	Day-14	Day-21	Day-28
T0	88,71±0,00	156,73 <sup>a</sup> ±13,68	172,49 <sup>a</sup> ± 8,40	170,45 <sup>ab</sup> ±19,27	177,33 <sup>ab</sup> ±21,77
T1	78,87±0,00	152,69 <sup>a</sup> ±39,00	138,57 <sup>ab</sup> ±31,36	194,94 <sup>a</sup> ±24,47	212,70 <sup>a</sup> ±13,17
T2	85,44±0,00	127,57 <sup>a</sup> ±13,95	129,85 <sup>a</sup> ±8,88	139,10 <sup>b</sup> ±23,94	138,68 <sup>b</sup> ±33,06
T3	89,31±0,00	150,66 <sup>a</sup> ±25,44	163,77 <sup>ab</sup> ±8,88	148,80 <sup>ab</sup> ±38,35	170,63 <sup>ab</sup> ±25,93

T0 (a treatment control without probiotic administration), T1 (probiotic at concentration of  $10^6$  CFU/ml), T2 (probiotics with concentration of  $10^7$  CFU/ml), T3 (probiotic  $10^8$  CFU/ml), and SD = Standard Deviation. Superscript with different letters indicated there are significant differences (p<0.05).

*Measured Parameters*

The main parameters observed in this study were organic material using the titrimetry method, feed conversion ratio (FCR) (Standen et al., 2015), feed efficiency (EP) (Effendi, 2003; Dahlan et al., 2017), and specific growth rate (SGR) (Standen et al., 2015). While supporting parameters were temperature, pH, and dissolved oxygen (Dissolved Oxygen).

*Measurements of Observed Parameter*

The amount of organic matter was measured by using a titration method (SNI 06-6989.22-2004). Body-weight of vaname shrimp was measured using a ruler, and the total weight was measured using an analytical scale, and both parameters were measured every seven days. The feed conversion ratio and feed efficiency were calculated at the end of the study. While pH was measured by pH pen, DO with YSI DO meter, salinity with a refractometer, and the temperature with a thermometer. Water quality measurements were read out every day for 28 days.

The result of organic content measurement during this study is presented in Table 1.

There were no significant differences in the total organic matter of the shrimp’s rearing water among all treatments (T0, T1, T2, and T3), which were observed on day 7 (p>0.05).

On day 14<sup>th</sup>, organic matter on the rearing water of T0 was not significantly different (p> 0.05) from the organic matter in rearing water of T1 and T3. However, total organic matter in the rearing water of T1 was significantly different from that in T2 (p <0.05). On the 21<sup>st</sup> day, the results showed that organic matter in T1 was significantly different (p <0.05) from that of T2 but not significantly different from those of T0 and T3, (p> 0.05). On day 28, organic content in the control water (T0) was significantly different from T2 (p <0.05), but organic matter in T2 (138.68 mg / l) and was not significantly different from T3 (170.63 mg / l) and T0 (177.33 mg / l) (p> 0.05).

The total amount of organic matter had increased in the rearing water without probiotics administration (T0) until the end of the culture period. While the rearing water with the addition of probiotics showed lower values than the control (T0) except in T1 (probiotic at a concentration of 10<sup>6</sup> CFU/ml). Organic content in T2 was lower compared to T1 (10<sup>6</sup> CFU/ml) and T3 (10<sup>8</sup> CFU/ml) until the end of culture. These results are in agreement with the result of Yuvaraj and Karthik (2015), in which mineralization of organic matter and getting rid of unwanted waste compounds can be performed by microorganisms. According to Boyd (2016), the decomposition of organic material by microbes depends on several factors, including water temperature, pH, DO and composition of the organic material itself. Bacteria and other decomposing microorganisms degrade organic matter quickly at temperatures ranging from 30 - 35°C. Water temperature measured in the present study was below 30°C (Table 5), and this caused low performance of probiotic bacteria in decomposing the organic matter. This could be seen from the measurement results of organic matter at the end of the culturing period which was ranging from 138.68 - 212.0 mg / l.

On the day 28<sup>th</sup> day, the lowest organic matter was found in the rearing water of T2, 138.68 mg / l, where the results were still below the maximum limit of organic matter for vaname shrimp culture. This is consistent with the statement of Adiwijaya et al. (2008) in which states that the maximum limit for organic matter is <150.00 mg / l. According to Putra et al., (2014), organic matter content in the culture media with the range of 41.14-162.45 mg / l and produced ammonia ~ 0,000 mg / l. Thus, it is suggested that probiotic bacteria with a density of 10<sup>7</sup> CFU / ml (T2) could be degrade and reduce total organic matter in shrimp rearing water.

On day 28<sup>th</sup>, organic matter at T0 treatment was lower than that of T1 treatment. This is due to the number of shrimps in the fourth week in the T0 tank was lower than the number of shrimps in T1. As a consequence, the amount of feed given in the T0 was lower, which resulted in lower amount of organic matter formed in T0 than in T1. Whereas the high organic matter in the T1 was due to the administered probiotic bacteria was unable to degrade the organic material produced by vaname shrimp. However, not all of the administered probiotics with higher density were able to degrade organic matter optimally. This was indicated by the rearing water in T3 (10<sup>8</sup> CFU/ml) where the amount of organic matter showed no significant difference than that of T0 (p <0.05). This result might be due to probiotic consortium in rearing water of T3 to compete with each other for obtaining nutrients.

Setiawati and Hudaidah (2013) explained that the high administration of probiotic bacteria could lead to competition in obtaining nutrients or substrates, which could be inhibit bacterial growth activities. This condition causes a decrease in the performance of probiotic bacteria in degrading organic matter. Furthermore, Boyd (2016) stated that organic matter containing higher nitrogen is easier to be broken down than organic matter with low nitrogen content. This might be due to the fact that the probiotic microorganisms required more nitrogen to produce their cells.

The conversion ratio (FCR) of shrimps in all treatments was lower than the control (T0) and was significantly different among treatments (p <0.05). FCR in treatment T3 (1.22) was significantly different (p <0.05) from T1 (1.75) and T2 (1.87). In this study, the lowest FCR value was recorded at Treatment 3 (1.22).

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**Table 2.** The feed conversion ratio of white shrimp (*Litopenaeus vannamei*) supplemented with probiotic at different concentrations.

Treatments	Feed Conversion Ratio			Average±SD
	Replications			
	I	II	III	
T0	2.58	2.74	2.50	2.60 <sup>a</sup> ±0.12
T1	1.99	1.61	1.65	1.75 <sup>b</sup> ±0.21
T2	1.46	2.11	2.04	1.87 <sup>b</sup> ±0.35
T3	1.22	1.22	1.23	1.22 <sup>c</sup> ±0.01

T0 (without probiotic administration). T1 (probiotic administration at 10<sup>6</sup> CFU/ml). T2 (probiotic administration at 10<sup>7</sup> CFU/ml). T3 (probiotic administration of 10<sup>8</sup> CFU/ml). SD = Standard Deviation. Superscripts with different letters indicate different in FCR values (p<0.05).

Uneaten feed, shrimp feces, and dead plankton are the main elements in the formation of organic matter in the rearing water. Microbes break down organic matter resulting in the production of new bacterial biomass, ~ 40-60% of the organic material that is metabolized (Avnimelech, 2006).

The FCR was calculated from the total amount of feed consumed divided by the weight gain of shrimp during the culture period. The smaller the value of the FCR, the better the feed is utilized in shrimp growth. The average FCR of vaname shrimp feed is presented in Table 2.

**Table 3.** Feed efficiency of vaname shrimp (*Litopenaus vannamei*) supplemented with probiotics at three different concentrations

Perlakuan	Feed Efficiency (%)			Mean±SD
	Treatment			
	I	II	III	
P0	38.67	36.47	39.85	38.36 <sup>c</sup> ±1.72
P1	50.02	62.1	60.4	57.50 <sup>b</sup> ±6.53
P2	68.48	47.38	48.83	54.90 <sup>b</sup> ±11.78
P3	82.01	81.28	81.43	81.57 <sup>a</sup> ±0.38

T0 (without probiotic administration). T1 (probiotic administration at 10<sup>6</sup> CFU/ml). T2 (probiotic administration at 10<sup>7</sup> CFU/ml). T3 (probiotic administration of 10<sup>8</sup> CFU/ml). SD = Standard Deviation. Superscripts with different letters indicate different in FCR values (p<0.05).

**Table 4.** The specific growth rate of vaname shrimp (*Litopenaus vannamei*) reared in probiotic-treated rearing water at three different concentrations.

Treatments	Specific Growth Rate (%BW/day±SD)			
	Day-7	Day-14	Day-21	Day-28
T0	0.71 <sup>b</sup> ±0.44	1.50 <sup>a</sup> ±0.23	1.41 <sup>b</sup> ±0.21	1.14 <sup>c</sup> ±0.09
T1	1.40 <sup>a</sup> ±0.22	1.73 <sup>a</sup> ±0.25	1.84 <sup>ab</sup> ±0.20	1.45 <sup>bc</sup> ±0.26
T2	0.91 <sup>ab</sup> ±0.24	1.95 <sup>a</sup> ±0.65	2.08 <sup>a</sup> ±0.55	1.71 <sup>b</sup> ±0.44
T3	1.35 <sup>ab</sup> ±0.37	1.73 <sup>a</sup> ±0.14	1.99 <sup>ab</sup> ±0.18	2.25 <sup>a</sup> ±0.03

T0 (without probiotic administration). T1 (probiotic administration at 10<sup>6</sup> CFU/ml). T2 (probiotic administration at 10<sup>7</sup> CFU/ml). T3 (probiotic administration of 10<sup>8</sup> CFU/ml). SD = Standard Deviation. Superscripts with different letters indicate different in FCR values (p<0.05).

The feed efficiency of vaname shrimp ranged from 38.36 to 81.57%. Feed efficiency is the opposite of the feed conversion ratio. Feed efficiency is the percentage increase in body weight divided by the amount of food consumed. The higher the value of feed efficiency, the lower the value of feed conversion ratio, and vice versa. Higher value of feed conversion results in lower value of feed efficiency. The feed efficiency of shrimp in T0 (38.36%) was significantly different from shrimp in P1 (57.50%), P2 (54.90%), and P3 (81.57%) (p <0.05). While feed efficiency in shrimps receiving P3 was significantly different shrimps from P1 and P2. (p <0.05). The feed efficiency of vaname shrimp receiving probiotic supplementation could be seen in Table 3.

The highest feed efficiency obtained in the present study was 81.57% in shrimps with T3 treatment, while the lowest in control (T0), which was 38.36%. p<0.05. These results show that the probiotic supplementation improved the feed efficiency of vaname shrimp. This result might indicate that the probiotic bacteria are used by the vaname shrimp as a protein source for growth, or the probiotic bacteria produce extracellular enzymes that help in shrimp digestion.

According to Pahlawi et al. (2019) *Pseudomonas* sp. could produce protease, lipase, cellulase, and amylase enzymes. Whereas *Bacillus* sp. had been reported to secrete extracellular enzymes such as protease, amylase, and lipase (Karigar and Rao, 2011) hence it could help the digesting process that takes place in the stomach. These enzymes help hydrolyze complex molecular nutrients such as breaking down carbohydrates, proteins and fats into simpler molecules such as glucose, amino acids, and fatty acids. Furthermore, Putra (2010) stated that the extracellular enzymes helped to hydrolyze complex molecular nutrients into simpler molecules to facilitate the process of digestion and absorption in the digestive tract of fish.

The specific growth rate of vaname shrimp during the culture period increased overtime. At day-28, the specific growth rate of shrimps in the control group (P0) was significantly different from shrimps at P2 and P3 but was not significantly different from shrimps in P1 (p> 0.05). While the specific growth rate of shrimps at P3 was significantly different from the specific growth rate of shrimp at P1 and P2 (p <0.05). The average specific growth rate of vaname shrimp supplemented with probiotic at the different concentrations is presented in Table 4.

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The specific growth rate of vaname shrimp calculated on day-28 in P3 treatment. (2.25%BW/day) was significantly higher than the vaname shrimp in control (T0; 1.14%BW/day). This result indicated that the administration of probiotics at 10<sup>8</sup> CFU/ml (T3) improved feed digestion in the shrimp through extracellular enzyme production. The production of extracellular enzymes by the probiotics increased the digestibility of protein, carbohydrates, and fat so that the feed utilization and growth of the shrimp were higher than the control shrimps. This result is supported by a research result of Djauhari et al. (2016) in which the addition of probiotics at 10<sup>6</sup>-10<sup>10</sup> CFU / g of feed reduced the amount of feed needed for carp growth. In addition, Wang and Gu (2010) reported that the addition of *Lactobacillus acidophilus* RS058, *Rhodospseudomonas palustris* GH642 and *Bacillus coagulans* NJ105 as additives to rearing water increased the growth performance and immune response of vaname shrimp. Similarly, Setyono et al. (2019) explained that the application of biofloc and probiotics could increase the specific growth rate of vaname shrimp by 4.90%.

Water quality parameters, including pH, temperature, DO (*Dissolved Oxygen*), and salinity measured during the culture period is presented in Table 5.

**Table 5.** Water quality of shrimps' rearing water measured during 28 days

Water quality parameters	Average Values
Temperature	24.5 - 26.8°C
DO ( <i>Dissolved Oxygen</i> )	4.92-5.78 ppm
pH	7.1-8.0
Salinity	26-30 ppt

The average temperature of the shrimps' rearing water during the culturing period ranged from 24.5-26.8 ° C. The temperature was slightly below the tolerance limit for vaname shrimp, which is 26-35°C (WWF-Indonesia Fisheries Team, 2014). This is due to the cold weather of the rainy season therefore tends to cause the ambient temperature to decrease. The pH level ranged from 7.1 to 8.0, which was within the tolerance limits for vaname shrimp ranging from 7.0-8.5 (WWF-Indonesia Fisheries Team, 2014). Dissolved oxygen levels during the study ranged from 4.92 to 5.78 ppm, which was a safe range for vaname shrimp (4.4 to 5.4 ppm) as reported by Thakur and Lin (2002). The measured salinity during the study was between 26-30 ppt. The salinity is still within the tolerance limits for vaname shrimp. According to Sulistinarto (2008) salinity tolerance of vaname shrimp ranges from 5 to 35 ppt.

#### 4. Conclusion

In conclusion, the administration of probiotics (*Pseudomonas* sp. and *Bacillus* sp.) at different concentrations significantly affected the amount of organic matter, feed conversion ratio, feed efficiency, and specific growth rate of vaname shrimps. The lowest organic matter (138.68 mg/l) was obtained from probiotic administration at a concentration of 10<sup>7</sup> CFU/ml (T2). While the administration of 10<sup>8</sup> CFU/ml (T3) had the lowest feed conversion ratio (1.22), highest feed efficiency (81.57 %), and the best specific growth rate of vaname shrimp (2.25% BW/day).

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#### Authors' Contributions

All authors have discussed the results and contributed from the start to the final manuscript; MJ, WHS: was responsible for the overall data collection and writing the initial until final draft of this paper. DDN, RZ: responsible for some data collection, particularly secondary data, and discussion of findings. MJ: besides being involved in a few field works, was also involved in the discussion of this paper.

#### Conflict of Interest

The authors do not have conflicts of interest.

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