

TROPICAL JOURNAL OF
**NATURAL
PRODUCT
RESEARCH**

TJNPR

www.tjnpr.org VOL. 11(6) 1, JULY 2017

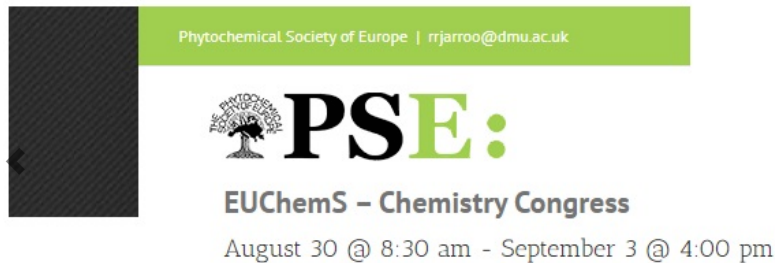


**Tropical Journal of
NATURAL PRODUCT RESEARCH**

This journal is indexed in Open J-Gate, Crossref, Creative Commons, Index Copernicus, Google Scholar.

All published papers are DOI assigned.

Official Journal of Natural Product Research Group
University of Benin



(https://phytochemicalsociety.org/?tribe_events=phytochemicals-in-nutrition-and-health)

Editorial Board

Editor-in-Chief: Professor Abiodun Falodun, Faculty of Pharmacy, University of Benin, Nigeria.

Associate Editors:

Professor Dr. Dr. Peter Langer, Institute of Organic Chemistry, University of Rostock, (Germany)

Professor Frederick O. Ekhaize, Microbiology, University of Benin, Nigeria.

Editorial Assistant:

Dr. Erharuyi Osayemwenre, Faculty of Pharmacy, University of Benin, Nigeria.

Board Members

Professor Ikhlas A. Khan, National Center for Natural Product Research, Mississippi (USA)

Professor Nosa Egiebor, College of Environmental Science & Forestry, State University of New York

Professor Samuel Qiu, South China Botanical Gardens, Chinese Academy of Sciences (China)

Professor Xavier Barril, De Fisicoquimica-Facultat De Farmacia Universitat de Barcelona, (Spain)

Professor Abiodun Ogundiani, Pharmaceutical Chemistry, OAU, Ile-Ife, (Nigeria)

Professor Thomas Kodadek The Scripps Research Institute, Scripps Florida (USA)

Professor Anthony B Ebeigbe, Physiology, College of Medical Sciences, University of Benin, (Nigeria)

Professor Eric KI Omogbai, Pharmacology and Toxicology, University of Benin, Nigeria

Professor Dr. Udo Kragl, Institute of Organic Chemistry, University of Rostock, (Germany)

Professor Cyril O. Usifoh, Faculty of Pharmacy, University of Benin, Nigeria.

Professor Azuka C Opara, Clinical Pharmacy & Pharmacy Practice, University of Benin, (Nigeria).

Professor Ikhide G. Imumorin, Biological Sciences, Georgia Institute of Technology, Atlanta (USA)

Professor Mark T. Hamann, Medical University College, South Carolina (USA)

Professor Barbara Nebe, University of Rostock, (Germany)

Professor Anthony I. Okoh, University of Fort Hare, Alice (South Africa)

Professor Dr. M. Iqbal Choudhary, HEJ, University of Karachi, (Pakistan)

Professor Omoanghe S. Isikhuemhen, North Carolina A&T State University, (USA)

Professor Ezekiel Green, University of Johannesburg, (South Africa)

Professor John Igoli, Strathclyde Institute of Pharmacy and Biomedical Sciences, UK

Professor Peter Akah, Pharmacology & Toxicology, University of Nigeria, Nigeria

Professor H.A.B. Coker, Faculty of Pharmacy, University of Lagos (Nigeria)

Dr Kingsly Agho, School of Science and Health, Western Sydney University (Australia)

Dr. E. Igbinosa, Microbiology, University of Benin, Nigeria

Dr. Pius Fasinu, School of Pharmacy, Campbell University (USA)

Professor Larry A Walker, National Center for Natural Products Research, Mississippi, USA

Prof. Dr. Alireza Heidari, Faculty of Chemistry, California South University (CSU), Irvine, California, USA

Professor Iyere O Onoagbe, Faculty of Life Sciences, University of Benin, (Nigeria)

Professor Broderick Eribo, Department of Biology, Howard University, Washington DC, (USA)

Professor Simon Gibbons, School of Pharmacy, University College London, UK

Professor Masashi Mizuno, Laboratory of Food & Nutritional Chemistry, Kobe University, Japan

Dr Edwin Uwagie-Ero(DVM, M.Sc., Ph.D., FCVSN), Associate Professor, Department of VETERINARY SURGERY, University of Benin

Dr Petra Obioma Nnamani, Drug Delivery & Nanomedicines Research Group, Department of Pharmaceutics, University of Nigeria.



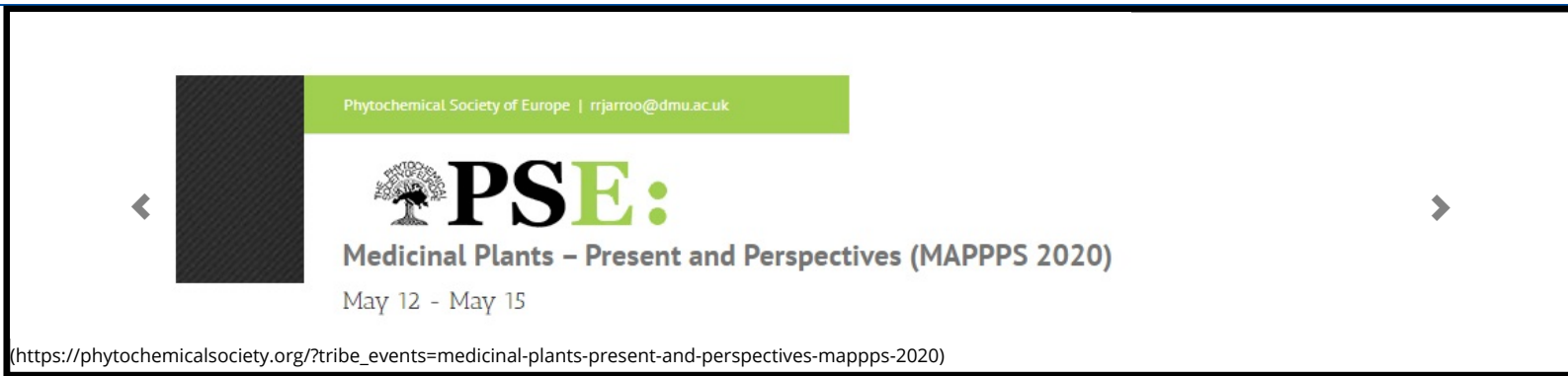
Submit a Manuscript (submitmanuscript.aspx)

The image shows the cover of the Tropical Journal of Natural Product Research (TJNPR). The cover has a green header with the journal title in white and green text. Below the title is the website address www.tjnpr.org. The main body of the cover features two photographs of lush green plants. At the bottom, there is a green footer with the text 'Official Journal of Natural Product Research Group University of Benin'. A small logo for 'NATURAL PRODUCT RESEARCH TJNPR' is also present.

The image displays a ScimagoJR performance metrics box for the Tropical Journal of Natural Product Research. It shows the journal is in the Q3 quartile for Pharmaceutical Science, which is the best quartile. The SJR 2019 score is 0.13. The box is powered by scimagojr.com.

(<https://www.scimagojr.com/journalsearch.php?>

q=21100933230&tip=sid&exact=no)



Vol. 4(10) pp. 653 - 849: October 2020

02 November 2020

Reviews

Pharmacology of Herbal Remedies for Urinary Tract Infection in Western Countries (viewarticle.aspx?articleid=858)

Sadeq A. G. Kaabi, and Baidaa M. Ali

<https://doi.org/10.26538/tjnpr/v4i10.1> (<https://doi.org/10.26538/tjnpr/v4i10.1>)

pages 653 - 660 HTML (viewarticle.aspx?articleid=858) PDF (img/manuscript_858_1-TJNPR-2020-M095A Galley Proof.pdf)

Microsphere-Based Drug Delivery to Alveolar Macrophages - a Review (viewarticle.aspx?articleid=860)

Herlina Ekapatama, Mahardian Rahmadi, Dewi M. Hariyadi

<https://doi.org/10.26538/tjnpr/v4i10.2> (<https://doi.org/10.26538/tjnpr/v4i10.2>)

pages 661 - 671 HTML (viewarticle.aspx?articleid=860) PDF (img/manuscript_860_2-TJNPR-2020-M195A Galley Proof.pdf)

Present and Future Potential of Antiparasitic Activity of *Opuntia ficus-indica* (viewarticle.aspx?articleid=893)

Wafaa M. Hikal, Hussein A.H. Said-Al Ahl, Kirill G. Tkachenko

<https://doi.org/10.26538/tjnpr/v4i10.3> (<https://doi.org/10.26538/tjnpr/v4i10.3>)

pages 672 - 679 HTML (viewarticle.aspx?articleid=893) PDF (img/manuscript_893_3-TJNPR-2020-M229A Galley Proof.pdf)

Original Research Articles

Novel Phage Cocktail for the Treatment of Bacteria Causing Chronic Suppurative Otitis Media (viewarticle.aspx?articleid=862)

Sadeq A. G. Kaabi, Hadeel K. Musafar, Saba T. Hashim, Zahraa K. Raheem

<https://doi.org/10.26538/tjnpr/v4i10.4> (<https://doi.org/10.26538/tjnpr/v4i10.4>)

pages 680 - 686 HTML (viewarticle.aspx?articleid=862) PDF (img/manuscript_862_4-TJNPR-2020-M086A Galley Proof.pdf)

Antimicrobial, Antifungal and HRBC Membrane Hemolysis and Membrane Stabilization Properties of Various Extracts of *Justicia gendarussa* (viewarticle.aspx?articleid=895)

Amin M. Mir, Muhammad W. Ashraf, Himani Himani, Bilal A. Mir

<https://doi.org/10.26538/tjnpr/v4i10.5> (<https://doi.org/10.26538/tjnpr/v4i10.5>)

pages 687 - 690 HTML (<viewarticle.aspx?articleid=895>) PDF (img/manuscript_895_5-TJNPR-2020-M103A Galley Proof.pdf)

Oral and Acute Dermal Toxicity with *Passiflora edulis* Sims Aqueous Extract in Sprague-Dawley Rats (<viewarticle.aspx?articleid=894>)

Dina K. Sari, Liza M. Sari, Rudy Heryanto

<https://doi.org/10.26538/tjnpr/v4i10.6> (<https://doi.org/10.26538/tjnpr/v4i10.6>)

pages 691 - 694 HTML (<viewarticle.aspx?articleid=894>) PDF (img/manuscript_894_6-TJNPR-2020-M117A Galley Proof.pdf)

Antibacterial Activity of Cough Suppressant Functional Drink from Betel Leaves (*Piper sarmentosum*), Lime (*Citrus aurantifolia*) and Honey (<viewarticle.aspx?articleid=865>)

Hardoko Hardoko, Marcella Jessica, Yuniwati Halim

<https://doi.org/10.26538/tjnpr/v4i10.7> (<https://doi.org/10.26538/tjnpr/v4i10.7>)

pages 695 - 702 HTML (<viewarticle.aspx?articleid=865>) PDF (img/manuscript_865_7-TJNPR-2020-M124A Galley Proof .pdf)

In Vitro Cytotoxic Activity of Constituents of the Aerial Parts of *Glycosmis parviflora* (<viewarticle.aspx?articleid=866>)

Phu Q. D. Nguyen, Hoai T. Nguyen, Linh T. K. Nguyen, Hung Q. Vo, Anh T. Le, Thao T. Do, Duc V. Ho

<https://doi.org/10.26538/tjnpr/v4i10.8> (<https://doi.org/10.26538/tjnpr/v4i10.8>)

pages 703 - 707 HTML (<viewarticle.aspx?articleid=866>) PDF (img/manuscript_866_8-TJNPR-2020-M137A Galley Proof .pdf)

Aerial Parts of *Euphorbia hirta* L. in Polar and Non-Polar Solvents: Phytochemical, Antioxidant and Glucose Uptake Studies for Potential Source of Adjunct Drug for Diabetes (<viewarticle.aspx?articleid=867>)

Levent B. B. Aquino, Marjette Y. U. Barbaza1#, Judson L. T. Ramos1, Mon-Juan Lee, Shun-Yao Ko, Chieh-Lun Hsieh, Kathlia A. De Castro-Cruz, Po-Wei Tsai

<https://doi.org/10.26538/tjnpr/v4i10.9> (<https://doi.org/10.26538/tjnpr/v4i10.9>)

pages 708 - 713 HTML (<viewarticle.aspx?articleid=867>) PDF (img/manuscript_867_9-TJNPR-2020-M151A Galley Proof .pdf)

Green Silver Nanoparticles Based on the Chemical Constituents of *Glinus lotoides* L.: *In Vitro* Anticancer and Antiviral Evaluation (<viewarticle.aspx?articleid=868>)

Mai M. Farid, Mahmoud Emam, Reda S. Mohammed, Sameh R. Hussein, Mona M. Marzouk

<https://doi.org/10.26538/tjnpr/v4i10.10> (<https://doi.org/10.26538/tjnpr/v4i10.10>)

pages 714 - 721 HTML (<viewarticle.aspx?articleid=868>) PDF (img/manuscript_868_10-TJNPR-2020-M158A Galley Proof.pdf)

Artemisia judaica Attenuates Hyperglycaemia-Mediated Oxidative Stress and Cardiac Injury in Streptozotocin-Induced Diabetic Rats (<viewarticle.aspx?articleid=869>)

Wesam al-Amarat,

<https://doi.org/10.26538/tjnpr/v4i10.11> (<https://doi.org/10.26538/tjnpr/v4i10.11>)

pages 722 - 727 HTML (<viewarticle.aspx?articleid=869>) PDF (img/manuscript_869_11-TJNPR-2020-M197A Galley Proof.pdf)

A Potent Anti-Ageing and Immunomodulatory Activity of Apricot Seed Standardized Extract and its Major Compound; Amygdalin (<viewarticle.aspx?articleid=870>)

Amer Ramadan, Gehan Kamel, Aya A. Shokry, Riham A. El-Shiekh

<https://doi.org/10.26538/tjnpr/v4i10.12> (<https://doi.org/10.26538/tjnpr/v4i10.12>)

pages 728 - 733 HTML (<viewarticle.aspx?articleid=870>) PDF (img/manuscript_870_12-TJNPR-2020-M205A Galley Proof.pdf)

Antihypertensive Effects of TCT: "Thanh Can Thang", a Vietnamese Tradition Medicine Remedy, in a Cortisone Acetate Rat Model (viewarticle.aspx?articleid=872)

Quoc B. Pham, Hong P. Le, Hong H. Nguyen, Minh T. Doan, Thai H. Pham, Hoang N. Nguyen, Manh H. Tran, Thanh Ha T. Nguyen
<https://doi.org/10.26538/tjnpr/v4i10.13> (<https://doi.org/10.26538/tjnpr/v4i10.13>)

pages 734 - 738 HTML (viewarticle.aspx?articleid=872) PDF (img/manuscript_872_13-TJNPR-2020-M208A2 Galley Proof.pdf)

Hesperidin and Myricetin Attenuated Non-Alcoholic Fatty Liver Disease (NAFLD) in HepG2 Cells (viewarticle.aspx?articleid=873)

Nadta Sukkasem, Waranya Chatuphonprasert, Kanokwan Jarukamjorn
<https://doi.org/10.26538/tjnpr/v4i10.14> (<https://doi.org/10.26538/tjnpr/v4i10.14>)

pages 739 - 747 HTML (viewarticle.aspx?articleid=873) PDF (img/manuscript_873_14-TJNPR-2020-M249A Galley Proof .pdf)

Comparative Botanical and Genetic Diversity Study of Different Cultivars of *Sesamum indicum* L. Cultivated in Egypt (viewarticle.aspx?articleid=876)

Seham S. El-Hawary, Rehab M. S. Ashour, Mahitab H. El-Bishbishy, Mona M. Okba, Heba A. Hassan
<https://doi.org/10.26538/tjnpr/v4i10.15> (<https://doi.org/10.26538/tjnpr/v4i10.15>)

pages 748 - 755 HTML (viewarticle.aspx?articleid=876) PDF (img/manuscript_876_15-TJNPR-2020-M250A Galley Proof.pdf)

Qualitative and Quantitative Phytochemicals Studies of Ethanol Stem bark Extracts of *Isobberlinia doka* Craib & Stapf and *Isobberlinia tomentosa* (Harms) Craib & Stapf (viewarticle.aspx?articleid=877)

Bello Hadiza, Katsayal A. Umar, Ambi A. Aminu, Abubakar Y. Bashir
<https://doi.org/10.26538/tjnpr/v4i10.16> (<https://doi.org/10.26538/tjnpr/v4i10.16>)

pages 756 - 764 HTML (viewarticle.aspx?articleid=877) PDF (img/manuscript_877_16-TJNPR-2020-M048A Galley Proof .pdf)

Hepatocurative Effects of Alpha Lipoic Acid in 5-Fluorouracil-Induced Toxicity Involves Activation of TNF Alpha and NF-?? Signaling Pathway (viewarticle.aspx?articleid=878)

Jamal-Deen I. Tiiba, Mohammed G. Magaji, Nuhu M. Danjuma, Abdullahi B. Sallau
<https://doi.org/10.26538/tjnpr/v4i10.17> (<https://doi.org/10.26538/tjnpr/v4i10.17>)

pages 765 - 770 HTML (viewarticle.aspx?articleid=878) PDF (img/manuscript_878_17-TJNPR-2020-M089A Galley Proof.pdf)

Platinum-Based Anticancer Chemotherapy Tissue Toxicity: The Use of Carrot and Tomato Supplements as a Dietary Intervention in *Rattus norvegicus* (viewarticle.aspx?articleid=879)

Idris Z. Sadiq, Fatima S. Abubakar, Maryam Ibrahim, Godwin O. Adejo, Ali S. Idoko, Faith Afolayan, Kenneth Akhabue
<https://doi.org/10.26538/tjnpr/v4i10.18> (<https://doi.org/10.26538/tjnpr/v4i10.18>)

pages 771 - 776 HTML (viewarticle.aspx?articleid=879) PDF (img/manuscript_879_18-TJNPR-2020-M091A Galley Proof.pdf)

In-vitro Evaluation of the Trypanocidal Activity of *Andrographis paniculata* Against *Trypanosoma brucei brucei* (viewarticle.aspx?articleid=880)

Zakari Ladan, Timothy O. Olanrewaju, Dominic B. Maikaje, Rolayo T. Emmanuel, Levi A. Apinega, Timothy J. Isaiah, Nathaniel G. Isaiah, Peter M. Waziri
<https://doi.org/10.26538/tjnpr/v4i10.19> (<https://doi.org/10.26538/tjnpr/v4i10.19>)

pages 777 - 783 HTML (viewarticle.aspx?articleid=880) PDF (img/manuscript_880_19-TJNPR-2020-M134A Galley Proof.pdf)

Anti-Ulcerogenic Effect of Unripe Plantain (*Musa paradisiaca*) Pulp on Indomethacin-Induced Ulcer in Wistar Rat (viewarticle.aspx?articleid=881)

Augustine C. Madueke, Diane O. Ugwu, Valentine O. Nwanelo, Assumpta C. Anosike, Rapheal C. Anajemba, Pearl A. Onoh
<https://doi.org/10.26538/tjnpr/v4i10.20> (<https://doi.org/10.26538/tjnpr/v4i10.20>)

Effect of Methanol Leaf Extract of *Duranta erecta* in Rats Induced with Benign Prostatic Hyperplasia (viewarticle.aspx?articleid=882)

Chioma A. Anosike, Christopher C. Ugwu, Amaechi L. Ogara, Chiedozie S. Ozioko, Martins O Ougofor

<https://doi.org/10.26538/tjnpr/v4i10.21> (<https://doi.org/10.26538/tjnpr/v4i10.21>)

pages 790 - 795 HTML (viewarticle.aspx?articleid=882) PDF (img/manuscript_882_21-TJNPR-2020-M153A Galley Proof.pdf)

Aju-Mbaise Decoction Improves Haematological and Kidney Markers in High-Fat Diet-Fed Wistar Rats (viewarticle.aspx?articleid=883)

Nene O. Uchendu, Emeka G. Anaduaka, Valentine O. Nwanelo, Wilfred I. Ugwuoke, Kenneth O. Okoye, Micheal N. Okonkwo, Onyinye F. Nsofor, James Ugwu

<https://doi.org/10.26538/tjnpr/v4i10.22> (<https://doi.org/10.26538/tjnpr/v4i10.22>)

pages 796 - 800 HTML (viewarticle.aspx?articleid=883) PDF (img/manuscript_883_22-TJNPR-2020-M154A Galley Proof.pdf)

Alpha-Amylase Inhibitory Activity of Extract Combination of *Morinda citrifolia* L. Fruit, *Trigonella foenum-graecum* and *Nigella sativa* L. Seeds Using *In vitro* and *In Vivo* (viewarticle.aspx?articleid=885)

Musfiroh Ida, Muhtadi Ahmad, Apriliani N. Dwi

<https://doi.org/10.26538/tjnpr/v4i10.23> (<https://doi.org/10.26538/tjnpr/v4i10.23>)

pages 801 - 805 HTML (viewarticle.aspx?articleid=885) PDF (img/manuscript_885_23-TJNPR-2020-M160A Galley Proof.pdf)

Evaluation of the Potential Mechanisms of Anti-Inflammatory Activities of *Fagara zanthoxyloides* Lam. Leave Extract in Wistar Rats (viewarticle.aspx?articleid=886)

Osmund C. Enechi, Emmanuel S. Okeke, Ndidi E. Nkwoemeka, Nicodemus E. Nwankwo, Izuchukwu Agogbua, Benneth M. Ebere

<https://doi.org/10.26538/tjnpr/v4i10.24> (<https://doi.org/10.26538/tjnpr/v4i10.24>)

pages 806 - 811 HTML (viewarticle.aspx?articleid=886) PDF (img/manuscript_886_24-TJNPR-2020-M169A Galley Proof.pdf)

Membrane Stabilization, Albumin Denaturation, Protease Inhibition, and Antioxidant Activity as Possible Mechanisms for the Anti-Inflammatory Effects of Flavonoid-Rich Extract of *Peltophorum pteroca* (viewarticle.aspx?articleid=887)

Osmund C. Enechi, Emmanuel S. Okeke, Nicodemus E. Nwankwo, John E. Nweze, Chukwuebuka P. Obilor, Christopher I. Okoye, Ogochukwu E. Awoh

<https://doi.org/10.26538/tjnpr/v4i10.25> (<https://doi.org/10.26538/tjnpr/v4i10.25>)

pages 812 - 816 HTML (viewarticle.aspx?articleid=887) PDF (img/manuscript_887_25-TJNPR-2020-M191A Galley Proof.pdf)

Liquid Chromatography-Mass Spectrometric Analyses of Potential Antioxidant Constituents from *Zanthoxylum zanthoxyloides* Leaves: Probing into the Role of Alkaloids (viewarticle.aspx?articleid=888)

Thecla O. Ayoka, Ngwu Nwachukwun, Aloysius C. Ene, Chidi U. Igwe, Charles O. Nnadi

<https://doi.org/10.26538/tjnpr/v4i10.26> (<https://doi.org/10.26538/tjnpr/v4i10.26>)

pages 817 - 823 HTML (viewarticle.aspx?articleid=888) PDF (img/manuscript_888_26-TJNPR-2020-M200A Galley Proof.pdf)

Methanol Extract of *Ficus platyphylla* Stem Bark Inhibits Cholinesterase Enzyme on Catfish (viewarticle.aspx?articleid=889)

Ibrahim M. Hassan, and Abdullahi I. Ja'afaru

<https://doi.org/10.26538/tjnpr/v4i10.27> (<https://doi.org/10.26538/tjnpr/v4i10.27>)

pages 824 - 830 HTML (viewarticle.aspx?articleid=889) PDF (img/manuscript_889_27-TJNPR-2020-M201A Galley Proof.pdf)

Evaluation of *Zingiber officinale* Rosc. and *Ocimum basilicum* L. Essential Oils-Loaded Gel Base for the Treatment of Oral Candidiasis (viewarticle.aspx?articleid=890)

Evaluation of Proximate, Mineral, Anti-Nutrients and Phytochemical Constituents of Indigenous Beans (Cajanus cajan, Sphenostylis stenocarpa and Phaseolus lunatus) (<viewarticle.aspx?articleid=891>)

Wale A. Ojewumi, and Omowumi J. F. Sanusi

<https://doi.org/10.26538/tjnpr/v4i10.29> (<https://doi.org/10.26538/tjnpr/v4i10.29>)

Comparative Anticonvulsant Activity of Leaf, Stem Bark and Root Bark Extracts of Bombax costatum Pellegr. and Vuillet in Acute Models of Epilepsy (<viewarticle.aspx?articleid=892>)

Abdullahi B. Nazifi, Aliyu Ahmed, Fatima I. Hassan, Nuhu Mohammed, Ahmed A. Danbala, Saidi Odoma

<https://doi.org/10.26538/tjnpr/v4i10.30> (<https://doi.org/10.26538/tjnpr/v4i10.30>)



Submit a Manuscript (<submitmanuscript.aspx>)

The image shows the cover of the Tropical Journal of Natural Product Research (TJNPR). The cover features a green background with a white box containing the journal's title in green and black text. Below the title, there are two photographs: one of a lush green forest scene and another of a close-up of green plants. At the bottom of the cover, there is a green banner with white text that reads "Official Journal of Natural Product Research Group University of Benin".

The image shows a Scimago SJR 2019 index badge for the Tropical Journal of Natural Product Research. The badge is divided into two sections. The top section is orange and contains the text "Q3 Pharmaceutical Science" and "best quartile". The bottom section is white and contains the text "SJR 2019 0.13" and "powered by scimagojr.com".

(<https://www.scimagojr.com/journalsearch.php?>

<q=21100933230&tip=sid&exact=no>)

ISSN: 2616-0684 (Print)

ISSN: 2616-0692 (Online)

DOI: 10.26538/tjnpr (<https://tjnpr.org/>)

Index Copernicus Value (ICV) for 2017: 59.83 (<https://journals.indexcopernicus.com/search/form?search=Tropical%20Journal%20of%20Natural%20Product%20Research>)

Indexing & Abstracting

**Microsphere-Based Drug Delivery to Alveolar Macrophages - a Review**Herlina Ekapratama¹, Mahardian Rahmadi², Dewi M. Hariyadi^{1*}¹Department of Pharmaceutics, Faculty of Pharmacy, Universitas Airlangga, Campus C Mulyorejo, Surabaya 60115, Indonesia²Department of Clinical Pharmacy, Faculty of Pharmacy, Universitas Airlangga, Campus C Jl.Mulyorejo 60115, Surabaya, Indonesia

ARTICLE INFO

ABSTRACT

Article history:

Received 01 September 2020

Revised 06 October 2020

Accepted 22 October 2020

Published online 02 November 2020

Copyright: © 2020 Ekapratama *et al.* This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

The lungs have a large surface area and high permeability, hence pulmonary delivery systems provide both local and systemic therapeutic effects. Pulmonary delivery system has been selected by many researchers because the route of administration is not invasive, has low metabolic activity, controlled environment for systemic absorption and avoids first pass metabolism. Alveolar macrophages are the first defense in the lung tissue to fight airborne pollutant, other foreign particle and pathogen by phagocytosis mechanism. Alveolar macrophages play an important role in the process of activation of the adaptive immunity including in inflammation and cancer diseases. Drug targeting to alveolar macrophages can achieve improvement in efficacy of therapeutic treatment for medical conditions including tumor, cancer, inflammation and infection. Respiratory infection-causing bacteria such as tuberculosis and pneumonia are able to survive in alveolar macrophages and they turn macrophages become a reservoir. This presents the challenge of making macrophages as targets in pulmonary delivery system because most of drugs do not reach the macrophages level effectively. To achieve this goal, the use of carrier particles in either micro-sized or nano-sized technology is the right choice. This review focuses on the influences of various physicochemical properties of microspheres carrier include particle size, aerosolisation property, morphology surface charge, surface properties and hydrophilicity on their uptake by alveolar macrophages either enhance macrophages uptake or decrease macrophages uptake. Making macrophage a target of treatment especially for infectious diseases is a promising strategy to improve the efficacy of treatment although in its development there are still many challenges.

Keywords: Microspheres, Inhalation, Alveolar macrophage, Macrophage uptake, Physicochemical properties.

Introduction

Lungs have a complex but coordinated system to eliminate inhaled pathogenic and pollutant particles. Pulmonary contact with pathogenic particles has the potential to cause respiratory disturbances, so the process of eliminating foreign particles must be ensured to continue functioning normally. Pulmonary delivery system becomes the choice of drug delivery, for example in the provision of inhaled antibiotics aimed at several diseases such as tuberculosis and pneumonia. This delivery route is also intended for the treatment of pulmonary hypertension¹ and the administration of paclitaxel and doxorubicin in the treatment of lung cancer.² In several studies that have been carried out, inhalation delivery system is also intended to have a systemic effect, for example insulin delivery,³ delivery of anti-nerve growth factor hormone,⁴ and antithrombotic therapy.⁵ Lungs' natural defense mechanism to fight pollutants and potentially pathogenic particles is a complex system and involves several processes such as mucociliary cleansing, the release of anti-pathogenic endogenous proteins, and the presence of leukocyte responses that occur in the lungs.⁶ Alveolar macrophages are the first defense

responsible for the process of fusion and elimination of pathogenic particles and pollutants that enter through the respiratory system. Alveolar macrophages have immunoglobulin, mannose, and some special receptors responsible for the phagocytosis process of inhaled foreign particles. Several studies have shown that in certain circumstances, there is a change in the cleaning function and phagocytosis by alveolar macrophages that can initiate the emergence of several diseases such as asthma, cancer, atherosclerosis, idiopathic pulmonary fibrosis, and infection.⁷⁻¹⁰ In certain cases, it is also found that some bacteria like *Toxoplasma gondii* and several species of *Leishmania*, *Mycobacterium tuberculosis*, and *Listeria monocytogenes* are able to survive and avoid the mechanism of phagocytosis by macrophages.¹¹

Passive targeted system to deliver several drugs made in the nano system or microparticles to target infected macrophages becomes the right choice to handle several cases of infection caused by some of these bacteria. Furthermore, an active targeting system can also be carried out by giving selective ligands to the drug carrier particles. The success of inhalation delivery to achieve the target of alveolar macrophages depends on the optimization in terms of pharmaceuticals, such as the design of drug release and the increase in drug residence time in the target area. At present, preparation formulated for inhalation purpose tends to be in the forms of liquid mist and particles given as dry powder. These formulation tend to form particulates which when recognized by alveolar macrophages are phagocytosed. When formulating a preparation with the aim of targeting macrophages, it is necessary to understand what conditions are needed to facilitate particle uptake by macrophages with the aim of increasing

*Corresponding author. E mail: dewi-m-h@ff.unair.ac.id
Tel: + 62 812 32238383

Citation: Ekapratama H, Rahmadi M, Hariyadi DM. Microsphere-Based Drug Delivery to Alveolar Macrophages - a Review. Trop J Nat Prod Res. 2020; 4(10):661-671. doi.org/10.26538/tjnpr/v4i10.2

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

the residence time of drugs in macrophages to formulation approaches that will determine the characteristics obtained by macrophages which also affect particle uptake by macrophages. The following section will explain the use of microspheres with various forming polymers for targeted purposes on alveolar macrophages with the aim of increasing drug residence time in the lung area and increasing drug uptake by alveolar macrophages.

Microspheres system is an ideal system for targeted delivery in phagocytic cells such as macrophages and dendritic cells. In vitro and in vivo tests administration of hepatitis C vaccine microspheres can trigger a mediation of immune cell response by inducing CD8+ T cells expression and increasing the number of CD8+ T cells in mice significantly.¹² Furthermore, microspheres without drugs can stimulate innate immunity which will trigger bacillary killing in macrophages. The use of certain polymers with certain polysaccharide and carbohydrate groups in the microsphere system at the same time can act as ligands that are easily recognized by receptors on macrophages. Drug release profiles from microspheres that are easy to modify are also what make this system widely developed. Drug release from microspheres is controlled by two factors, namely drug dissolved from the polymer or the micro spherical polymer matrix degradation process. In this case, the use of polymers plays a fairly large role.

Targeted Delivery System on Alveolar Macrophages

Pulmonary delivery systems provides many advantages compared to conventional delivery systems, one of which is the rapid onset of work and is able to deliver drugs locally to the target, minimizing drug dosage so as to reduce the toxic effects of drugs and increase the index of drug therapy.¹³ The lungs have a short diffusion pathway from the respiratory tract to the systemic circulation and an increase in blood flow makes the lung the pathway for drug entry into the systemic pathway.

The upper airway is covered by thin mucus that serves to protect the tissue and capture and clean the particles that pass through it. In the deeper part there is an alveolar. Alveolar contains a variety of proteins and lipids that act as barriers to transport several molecules. Along with alveolar, the tight junction along epithelial cells also acts as a major barrier in the transport process. Protein transporters play an important role in the process of delivering drugs either through the mechanism of active absorption or passive diffusion, depending on the nature and structure of the drug delivered. Another important aspect in this area is the mechanism of clearance of molecules by macrophages which need to be considered in the process of transporting drugs to the lungs. Molecules that can cross the barrier will be inhaled by cells and absorbed into the systemic circulation or can also undergo phagocytosis by macrophages. Drug molecules can be absorbed more efficiently from the lungs compared to other non-invasive routes. The mechanism of deposition and uptake of particles in the lungs is shown in Figure 1.

Furthermore, the process of deposition of particles in the lung including the uptake mechanism by macrophages is strongly influenced by particle size. Particles with a size of 1-3 μm will experience uptake by macrophages (with a diameter of each cell 15-22 μm) better than particles with a diameter of 6 μm , whereas particles with a size of 0.26 μm are able to avoid phagocytosis.¹³ Smaller particles will interact with non-phagocytic cells in the epithelium and initiate endocytosis regulated by clathrin-coated and caveola. Nano-scale particles are more likely to be delivered to the systemic circulation. Inspirational expansion and expulsion of pulmonary alveoli can trigger the opening and closing of the caveola. The opening process itself can reach sizes of 40 and 100 nm which can allow macromolecular components such as proteins to pass through the alveolar-capillary barrier.¹⁵ This shows the mechanism and location of deposition of particles in the lung differ depending on the size of the particle itself as illustrated in Figure 2.

Phagocytosis is the primary mechanism for the process of taking particles by macrophages. Macrophages are one type of phagocytic cells that are responsible for cytokine secretion and help in delivering messages against the occurrence of pathogenic infections that will produce an immune response.¹⁶ Macrophages produce antibacterial

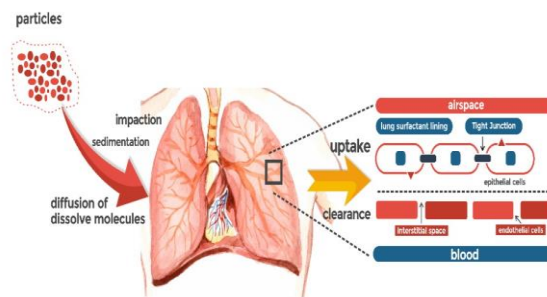


Figure 1: Mechanism of deposition and uptake of particles in the lungs.¹⁴

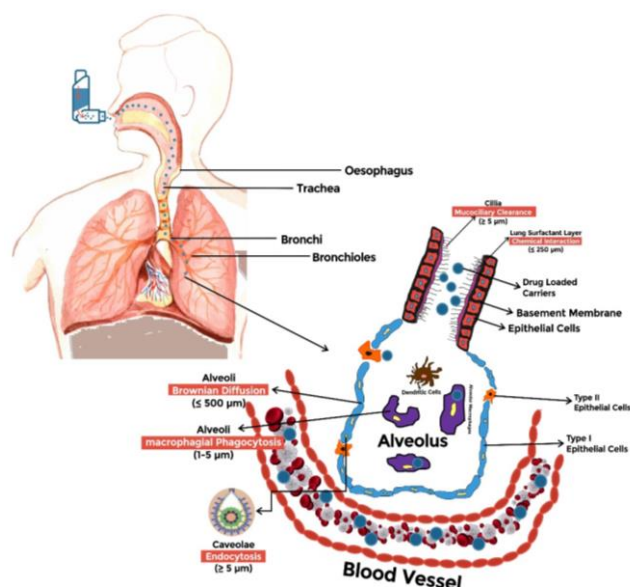


Figure 2: Relationship between particle size and route of distribution of particles, clearance mechanisms and absorption.¹⁵

substances, such as nitric oxide and cationic proteins that contribute to the destruction of microorganisms.¹⁷ In some cases, macrophages are not able to damage the components of pathogenic microorganisms because it is known that some of these microorganisms can avoid the fusion mechanism by lysosomes and phagosomes or by inhibiting the antimicrobial response in phagolysosomes. Not only have a survival strategy for macrophages, some bacteria also obtain some nutrients in the macrophages to carry out intracellular replication.¹⁸ This phenomenon makes macrophages a target of treatment for several pathological conditions. Alveolar macrophages are the most dominant part in alveoli. The combination of alveolar macrophages with epithelial cells, dendritic cells, and lymphocyte T cells found in the alveoli provide a good defense in the respiratory tract through several receptors and cytokines/chemokines that can control the immune system to clear pathogens from the lungs.¹⁹

The targeting on macrophages can increase the efficacy of treatment in tumor genesis, inflammation and infection therapy. The delivery system for particulate drugs both nano or microparticles, liposomes, micelles, polymeric conjugates, and dendrimers has been used for drug delivery via the pulmonary route including to target alveolar macrophages. One of the microparticles used is microspheres. In addition to having particle size that allows entry into the respiratory system and inner lungs, microspheres also have a great tendency to be phagocytosed by macrophages, so as to increase drug influx in macrophages. The particulate system has also been formulated in such a way as to increase its specific side to bind to the target, provide

sustained release, achieve deposition in the inner lung, and increase the bioavailability of the drug it carries. Although it is also well known that the lungs have a mucociliary cleansing mechanism, the possibility of particles to get trapped in the mucous layer and undergo phagocytosis by alveolar macrophages can eliminate drugs encapsulated in the particles or cut the residence time of the drug which has an impact on the limited efficacy of the drug. However, in some pathological conditions such as in inflammatory and infectious conditions, making macrophages as a target of treatment can increase the therapeutic effect. This depends on the purpose of therapy, whether aimed at increasing or avoiding the uptake by macrophages.⁶

Parameters of Microparticles Targeted on Alveolar Macrophage

To design particles so that they can enter the respiratory tract and reach pulmonary area that is rich in alveolar macrophages, there are several factors related to the nature of the particles that have to be considered.

Particle Size

In preparation intended for inhalation systems, particle size plays a role in determining the deposition mechanism and deposition location in the lungs. Particle size also determines the success of uptake by macrophages. Larger spherical polystyrene particles (3 μm) have been reported to be taken up more slowly compared to 1.5 μm sized particles, suggesting that higher energy requirement for membrane deformation of large particles could abate uptake.²⁰ This relates to the activation of the complement pathway.²¹ Particles of different sizes provoked different responses to macrophages. Larger particles tend to interact with tissues, while smaller particles (<200nm) tend to circulate on veins and flow along lymphatic, providing better antigen presentation.²² Particle size also determines the path of endocytosis. In general, particles larger than 1 micron will be analyzed by phagocytosis and smaller particles of 0.2-1 micron through endocytosis.²³ Particles ranging from 1-3 μm are the most easily phagocytosed size by alveolar macrophages, whereas particles > 10 μm or <0.2 μm are able to escape phagocytosis.²⁴ Particles with a diameter of 1-6 μm show higher uptake compared to larger particles.²⁵ The effect of particle size on the deposition mechanism in the lungs and the process of entry into macrophages are summarized in Table 1. The effect of particle size on the success of uptake by macrophages has been carried out on polystyrene microspheres with sizes of 0.2, 0.5, 1.0, 6.0 and 10 μm , respectively. The parameter of successful uptake of particles by macrophages is seen from the superoxide levels produced by macrophages, where the higher the superoxide levels produced indicates the more particles are uptake by macrophages. Particles with sizes 1.0 and 6.0 μm showed the highest levels of superoxide, followed by particles with sizes 0.5, 10.0 and 0.2 μm that had superoxide levels similar to those of buffer phosphate solution control.²⁹ Based on the research, it was concluded that there was an increase in particle uptake by macrophages for particles with sizes above 1 μm and below 6 μm . Similar research was also carried out by Hirota.³⁰ PLGA-rifampicin microspheres was made with sizes 1.0, 3.0, 6.0 and 10 μm respectively. The number of particles successfully phagocytosed by macrophage depends heavily on the particle size of the microsphere. Microspheres measuring 3 and 6 μm are more effective than particles measuring 1 and 10 μm . From the results of this study it is also possible that particle size 3 - 6 μm is the right size to obtain optimum phagocytosis activity.³⁰ Some examples of particle size parameters successfully taken by particles by macrophages are summarized in Table 2.

Properties of Aerosolization

Particles with a size that is too small are likely to come out again with carbon dioxide in the expiration process, so besides having a small size, the particles must have a certain weight. This value is known as mass median aerodynamic diameter (MMAD). A good MMAD is 1-6 μm ,²⁵ while another study reports MMAD ranges from 1-5 μm to be deposited in the lungs, but particles smaller than 3 μm are easier to reach the respiratory system.³⁵ To be able to target macrophages, the particles made must be ensured to be able to enter the respiratory tract particularly into the inner lungs, especially on the alveoli. Besides by

synchronizing with the milling, ball milling, and spray drying techniques,^{36, 37} to improve the aerosolization properties of a given particulate system in the form of dry powder inhalation (DPI), it also uses other excipients such as lactose and mannose with certain particle sizes. Du *et al.*³⁸ conducted an evaluation of the effects of lactose and granule lactose administration within a certain size to the aerosolization of salbutamol DPI. The size, rough or smooth surface of lactose, the density and flow properties of lactose as a carrier contribute to aerosol dispersion performance. In that study, it was concluded that redispersion decrease or increase not only related to particle size, but also other properties of the lactose used.³⁸ The instruments used to evaluate the aerodynamic properties of particles include Twin Stage Impactor (TSI), Next Generator Impactor (NGI) and Anderson Cascade Impactor (ACI). TSI has a 'throat' angle followed by two chambers to hold the particles (stage I and stage II). The first stage and 'throat' will hold larger particles, then finer particles will be accommodated on the second stage.³¹ A number of particles successfully deposited on stage II on the instrument show the success of drug deposition stated in fine particle fraction (FPF). The airflow velocity used in TSI is generally 60 ± 5 L/min. The airflow velocity and vacuum regulation on the pump contained in the device will determine the location of particle deposition on the device. After the aspiration process, the TSI instrument is released and each chamber of the instrument is rinsed with phosphate buffer saline and then measured to obtain the number of drugs deposited on each stage quantitatively. This procedure is also carried out if measurements are carried out using NGI. Furthermore, the drug that is stored in the inhaler, capsules, and adapters is also cleaned and dissolved in the acetate buffer. The MMAD value is determined by looking at the deposition of particles at different stages on NGI.²⁴ The size and shape of the particle affects the fine particle fraction (FPF) value. The addition of leucine to microparticles is able to increase FPF 4.3 to 6.9 times higher than microparticles without leucine. Leucine was useful as a natural antiadherent amino acid to improve the deagglomeration of particles prepared using spray drying method.³⁹ MMAD also evaluated using eight stages of ACI.⁴⁰ Microspheres are inserted into the tool as many as 5-6 cycles with a flow rate of 28.3 L/min.⁶ Carrier system includes polymeric liposomes, nanocarrier system with cyclodextrin or with the use of gelatin, micelles, dendrimers, and other various carrier systems such as microspheres and nanosphere also used to improve the flow properties of the drug. Microspheres are spherical particles of less than 200 μm of size which are used as a carrier system for delivery to various work targets in the body. Microspheres with the aim of inhalation must have an MMAD value of 3 μm to obtain optimal delivery in the lungs and can be captured by alveolar macrophages.²⁵

Morphology

The shape of the particles affects the process of internalization or the process of avoidance of particles by macrophages. This geometry shape determines the initiation of contact with macrophages and the subsequent phagocytosis process. Macrophages internalize foreign particles through the process of phagocytosis, a process in which particles attach to macrophages and then engulfment by the plasma membrane. The process of attachment of particles with different geometries is very dependent on the shape and size of the particles. Particles with different geometry shapes are able to provide at least one side to contact with macrophages and trigger phagocytosis. The shape of the particles influences the process of phagocytosis by macrophages, and furthermore, separately the shape of the particles influences the attachment and internalization of the particles. Particles with high attachment values will reduce the percent of particles internalized to macrophages. In oblate particle, it shows high attachment and internalization, so the number of particles phagocytosis is also high.⁴¹ The research was conducted on three particle forms, namely prolate ellipsoid (major axis 0.35 - 2.5 μm , minor axis 0.2 - 2 μm), oblate ellipsoid (major axis 0.35 μm -2.5 μm , minor axis 0.2 - 2 μm) and spheres (radius: 0.26 - 1.8 μm). Research that has been done shows that particle shape also influences the internalization process or the avoidance process of particles by macrophages.⁴² This geometry shape determines the initiation of

contact with macrophages and the subsequent phagocytosis process. Tests carried out on particles with different shapes, namely spherical, oval, ellipse, to rectangular plates. The result is that the elongated particles tend not to be taken by mouse peritoneal macrophages, while spherical particles tend to be more easily taken by macrophages. Furthermore, spherical particles tend to be able to avoid macrophages and increase anticancer activity.⁴³ Effect of particle morphology on the number of particles that are phagocytes showed in Table 3.

Surface Charge

It is known that macrophage cells have sialic acid on the surface, which makes the surface of macrophage cells negatively charged.⁴⁴ This leads the researchers that the particle surface charge plays a role in determining the success of particle uptake by macrophages. The surface charge of a particle determines the stability and interaction of particles with phagocytic cells. Positive charged particles are widely used for intracellular delivery because of their ability to interact with cell membranes which mostly have negative charges. Positively charged nanoparticles, with many more positively moieties than amynoglicosides are typically trapped in mucus.⁴⁵ So that they are able to increase uptake by cells and enhance immune responses. Hwang *et al.*³¹ used hyaluronic acid to increase the uptake of microspheres by macrophage cells by up to two times. Hyaluronic acid has a negative charge, besides that the use of hyaluronic acid can increase the mucoadhesive properties of particles.³¹ Hyaluronic acid itself is able to act as a ligand that is recognized by the CD44 protein, so the use of this material can increase the selectivity of CD44 receptors that are overexpressed in tissue that is inflamed.⁴⁶ Positively charged particles have a deficiency in acceptance related to toxicity because they can trigger the formation of ROS and induce apoptosis.⁴⁷ This makes the focus shift to negatively charged or uncharged particles that are more physiologically compatible. Particle composition in addition to affecting the physicochemical character also affects the process of particle recognition by macrophages. Some lipid groups such as phosphatidylserin and phosphatidylglycerol can be detected by macrophages because they have a negative charge. Particles made from functionalized alginates produce negatively charged particles.⁴⁸ Particles with a negative charge provide several advantages for reducing bioadhesive with plasma proteins and decreasing the speed of particles to be taken up by non-specific cells. Particle charge and macrophage delivery can be seen in Table 4.

Surface Properties of Particles

Particle rigidity affects the ability of particles to be taken up by macrophages. Phagocytosis is a process that depends on actin which is affected by the target's mechanical properties. Harder and stiffer polyacrylamide particles can be internalized into cells more efficiently than softer particles. Particle rigidity is generally responsible for the reception and interaction of particles with macrophages.⁵² Although it has lower entrapment efficiency, porous particles will release the drug faster than nonporous particles. In the process of deposition in the lungs, an increase in porosity and a decrease in the density of microspheres close to the size of the particle geometry trigger a decrease in aerodynamic diameter. Meanwhile, in the uptake process by macrophages, spherical particles without pores or spherical particles with low pores are more effective in experiencing uptakes by macrophages.⁵³ The most nonporous particles are deposited in the nasal cavity, while the porous particles are most deposited in the nasal cavity or bronchi and there is the least deposited in the pharynx and trachea. Porous particles with a geometry diameter of 5 μm tend to be deposited into the bronchial and alveolar regions, whereas particles with larger geometry tend to be left in the nasal cavity and trachea.⁵⁴ The difference in deposition properties between porous and unpredictable particles is influenced by differences in the degree of cohesiveness of each particle.⁵⁵ From various studies conducted, a conclusion is drawn that porous particles can be deposited more deeply on the inside, but tend to avoid phagocytosis by macrophages. Some examples of porous particles given by the inhalation route are presented in Table 5. Particle surfaces can be modified to increase uptake by macrophages by utilizing receptors on the surface of macrophages including Fc, manosil, galactosil, lipoprotein, and

fibrinocetin receptors. Some examples of ligands used include peptides, antibody, and polysaccharide-based polymers. Polysaccharide based polymers are also used as ligands for delivery to macrophages. Carrageenan, a polymer with sulfated sugar groups other than fucoidan and ulvan, is used as a ligand for macrophage delivery.⁵⁸ Several examples of other ligands used include bovine serum albumin and O-steroyl amylopectin (O-SAP) which are used in the manufacture of targeted in macrophages.⁵⁹ Receptors on macrophages can be targeted for active targeting so that further research is expected to develop specific ligands for various macrophage receptors for more efficient delivery. Use of ligands in particle surface is shown in Table 6.

Hydrophilicity

The lipophil hydrophilic nature of a particulate also affects the uptake of particles by macrophages. Particle coating with lipophilic material facilitates the process of particle recognition by macrophages, while coating with hydrophilic material such as polyethylene glycol (PEG) allows particles to survive the process of opsonization by serum proteins, inhibits hepatic clearance, and decreases the chance of particles to be recognized by macrophages. The use of appropriate polymers can affect the hydrophilic and lipophilic nature of particles and modulate their uptake by macrophages. Polymers such as poloxamer and poloxamine can make particles tend to be hydrophilic and prevent particles from being taken up by macrophages.⁶² Surfactant protein in the lungs is included in collagen-lectin family surfactant protein-A (SP-A) and surfactant protein-D (SP-D) which have specific receptors on the surface of macrophages. This protein can be used as an opsonin for targeted delivery on macrophages. In contrast, surfactant phospholipids which are a major component in pulmonary surfactants show inhibitory activity on particle uptake by macrophages.⁶³ Furthermore, coating the particles with DPPC that is the main lipid component in surfactants decreases the uptake of particles by cell macrophages NR8383.⁶⁴ Factors of physical and chemical characteristics that can reduce and increase particle uptake by macrophages are summarized in Table 7.

Polymer Characteristics as Factor for Macrophage Targeting

In general, microspheres are made by using polymers, both natural polymers such as chitosan, gelatin, alginate, and carrageenan, as well as with synthetic polymers such as Poly-Lactic-co-Glycolic-Acid (PLGA), Poly Lactic Acid (PLA), and Poly- ϵ -Caprolacone (PEC). The chemical physical properties of the polymer determine how the drug is deposited in the lungs. Both synthetic and natural polymers, and hydrophilic and hydrophobic choices must be adapted to the purpose of development. For the target system in macrophages, hydrophobic polymers have a greater chance of being phagocytosed by macrophages, but polymers with polysaccharide groups such as chitosan, alginate, and carrageenan also have the ability to interact with human receptors and activate the phagocytic mechanism.²⁴ The choice of polymer affects many things for the drug delivery, where the polymer used determines drug release and drug accumulation on the target. This is because the polymer determines the shape and size and the charge of the particles or system produced. The shape, size, and load become the parameters that need to be considered in the inhalation delivery system. The concentration of the polymer used in making a system affects the pattern of drug release from the system. Microspheres consisting of 50% polymers give burst effect in the release test compared to over-the-counter drugs. This can be caused either by the chemical interaction between the drug and the polymer or because of the nature of the microspheres that form an amorphous particle.¹ The use of polymers and crosslinker with different concentrations also affect the pattern of drug release from the system. Slower release is obtained by increasing the polymer ratio and crosslinker.⁶⁵ To obtain the microspheres system with controlled release also carried out with a combination of polymers. Kolesnyk⁶⁶ makes microspheres with a combination of alginate-kappa carrageenan with the use of CaCl₂ as the crosslinker. The difference in the comparison of alginate to κ -carrageenan provides a different release profile.⁶⁶ Polymer characteristics as carrier for targeting macrophage as shown in Table 8.

Table 1: Mechanisms of deposition and endocytosis pathways based on particle size

Particle SizeT (μm)	Deposition	Mechanism/ Endocytosis	Site of particle deposition in the lung	References
5 – 9 (slow inhalation)	Inertial impaction		Large airways include oropharynx, trachea and bronchi	15
3 – 6 (fast inhalation)	Inertial impaction		Large airways (trachea and bronchi)	
1 – 5	Gravitational sedimentation		Smaller airways	
<0.5	Brownian diffusion		Alveoli	
0.5 – 1	Brownian motion		Alveoli	
5 – 10	Impaction		Primary bronchi	26, 28
1 – 5	Sedimentation		Secondary bronchi	
1 – 3	Sedimentation		Bronchioles	
0.5 – 1	Brownian motion		Alveoli	
<0.2 – 1	Endocytosis		-	
<200	Clathrin-coated		-	26, 27
>500	Caveola mediated		-	
>1	Phagocytosis		-	
<0.2 – 1	Endocytosis		-	

Table 2: Example of particle size at the target of macrophage delivery

Delivery System	Drug	Particle Size (μm)	Particle Target	References
Polymeric Microparticles	Isoniazid	$4.1 \pm 0.57 \mu\text{m}$	Phagocytosed particles and concentration of INH in macrophages increased $8.28 \pm 0.3\%$ compared to the administration of free INH of $1.74\% \pm 0.69$	6
Microspheres	Rifampicin	1 – 6 μm	Particles are effectively phagocytosed by macrophages through the process of mediated scavenge receptors	25
Microspheres	Ofloxacin	2-5 μm	An increase of uptake of Microspheres with hyaluronates 1.7 times compared with microspheres without hyaluronates up to 2.1 times higher than free ofloxacin solutions	31
Microspheres	Ofloxacin	1 – 6 μm	Uptake of particles by macrophages increased up to 3.5 times compared with free ofloxacin	32
Microspheres	Isoniazid	$3.54 \pm 3.14 \mu\text{m}$	Particles undergo uptake by macrophages. Under fluorescent macrographs, microspheres are seen in the intracellular region and even the nucleus	33
Microspheres	Rifampicin	1-4 μm ($\sim 3 \mu\text{m}$)	As compared with free rifampicin, microspheres significantly more rifampicin in PLGA MS was uptaken by macrpahages at different time point.	34

Table 3: Effect of particle morphology on the number of particles that are phagocytes

Particle Morphology	Internalization (%)	Attachment (relative to spheres)	Phagocytosed (relative to spheres)	References
<i>Prolate ellipsoid</i>	52%	3.8	0.6	41
<i>Oblate ellipsoid</i>	86%	2.5	2.7	
<i>Spheres</i>	70%	1	1	
	Diameter of Initial Spheres		Particle attached per cell	42
<i>Spheres</i>		0.5	1.7	
		1	2.1	
		3	3.0	
<i>Rods</i>		0.5	3.5	
		1	3.1	
		3	1.3	
<i>Oblate ellipsoid</i>		0.5	2.7	
		1	3.7	
		3	0.5	

Table 4: Relationship between particle charge and macrophage delivery

Delivery System	Drug	Charge	Target of macrophage	References
Microspheres	Lysine hydrochloride, manose	Negative	The presence of free amino acids makes the system negatively charged and can be used by macrophages. After 15 minutes of administration, the particles have reached the cytosol through the mechanism of endocytosis.	33
Solid lipid nanoparticle (SLN)	Mannose	Negative	Negatively charged particles increase cell uptake through a charged scavenger receptor. SLN increases the process of endocytosis by macrophages. Microparticles are detected in the cytoplasm.	49
Nanostructured lipid carrier	Tuftstin	Negative	Nanoparticles with tuftsin peptide components are significantly internalized compared to nanoparticles without tuftsin.	50
Microspheres	Mannosylated gelatin	Positive	Mannosylated gelatin microsphere uptake by macrophages is higher than that of microspheres without mannosylated gelatin. This is related to the interaction of mannose groups with surface receptors on macrophages.	51

Table 5: Porous particle of inhalation delivery

Delivery System	Drug	Particle Porosity	Target Parameter	References
Microparticle	Lysozyme	Highly porous	Particles can be deposited in the trachea and inner lung. Particles with pores can avoid phagocytosis by macrophages, whereas particles without pores can quickly experience uptake by macrophages.	28
Microparticle	-	Porous particle (5 – 10 µm)	Porous particles with geometric diameters >3 µm are able to reach the lung alveoli region (stage 6-8 in ACI) and are able to avoid phagocytosis. Even particles with a geometry diameter of 5 µm tend to be more able to reach the inner lung than particles with a geometry diameter of 10 µm.	54
Microspheres	L-lactic acid	Nonporous Microporous (0.2 – 2 nm)	Porous particles with a geometrical size of 5-10 µm with lower MMAD (<3 µm) have good aerosolization properties,	55

		Mesoporous (2 – 50 nm) Macroporous (>50 nm)	are able to avoid phagocytosis and are deposited in the inner lung. Maximum uptake occurs in nonporous particles.	
Porous particle	Rifampicin	Porous size of 4 µm	In vivo test showed that rifampicin in the form of porous particle (PPs) is more effective for delivering drugs reaching the alveoli than in the form of free powder. PPs can avoid the mechanism of clearance in the respiratory tract.	56
Particulate	Meloxicam	Large Porous Particle (LPPs) (>5 µm)	Large porous parts (LPPs) have a higher deposit fraction compared to nonporous particles despite having the same MMAD value (2.55 µm). In aerodynamic testing using ACI, LPPs has EF (Emitted Fraction) EF and (Fine Particle Fraction) FPF higher than nonporous particle >85.4% and >65.8%	57

Table 6: Use of ligand in particle surface

Delivery System	Drug	Ligand	Target Parameter	References
Nanostructured lipid carrier	Rifampicin	Peptide tuftsin	Selectively, tuftsin recognizes infected surface receptors of macrophages, thereby increasing uptake by macrophages. furthermore, tuftsin increases the antimicrobial activity of rifampicin	50
Microspheres	Isoniazid	Mannose	Microsphere with mannose selectively experiences uptake and can reach phagolysosome vesicles on macrophages. Formulasi can maintain therapeutic drug concentration use despite a decrease in clinical dose	51
Nanoparticle	Licoris	Mannose	Formulations with mannose have increased uptake due to the interaction between mannose and mannose receptors on macrophages.	60
Microparticle	Budesonide	Phospholipid 1,2-distearoyl-sn-glycero-3-phosphoethanolamine (DSPE), Dipalmitoylphosphatidylcholine (DPPC), Dipalmitoylphosphatidylglycerols (DPPG).	The use of phospholipids can increase macrophage uptake. Where in the same comparison DSPE is the most effective phospholipid, followed by DPPG and DPPC	61

Table 7: Particle characteristic factors that affect macrophage uptake

Parameter of particle	Characteristics of particles to increase uptake by macrophages	Characteristics of particles to reduce uptake by macrophages
Size	Particle sizes are 100-200 nm and 1-6 µm	Particles sizes are <1 µm and > 6 µm
Surface morphology	Spherical particles	Elongated, branched, and filamentous particles
Surface properties	Modified particle surface with mannose, SP-A and SP-D, O-SAP, maleylated bovine serum albumin (MBSA), and tuftsin	The particle surface is modified with PEG, poloxamer, and poloxamin.

	Modified particle surface with mannose, SP-A and SP-D, O-SAP, (MBSA), and tuftsin	The particle surface is modified with PEG, poloxamer, and poloxamin.
Charge	Very positive or negative charged particles	Particles with a charge tend to be neutral
Rigidity	Rigid and non-porous particles	Fragile and porous particles
Hydrophilicity	Insoluble and hydrophobic particles	Dissolved and hydrophilic particles

Table 8: Polymer characteristics on targeted delivery of macrophages

Polymer	Role of polymers in the delivery of macrophages	References
PLGA	<ul style="list-style-type: none"> - Can be used for targeted delivery of macrophages even though macrophages do not have specific receptors for PLGA. - The introduction of PLGA particles by macrophages is determined by the proportion of lactic and glycolic acid in PLGA and the molecular weight of the copolymer used. - The degradation rate of the polymer is difficult to control. PLGA degradation causes changes in the lung environment to become more acidic which can interfere with the stability of peptides and proteins which will have an effect on its therapeutic effect. 	2, 30, 32, 67, 68
PEC	PEC is hydrophobic polymer which is able to activate the phagocytic process of macrophages. PEC will produce particles with porosity that are good enough to be used by macrophages.	6, 69
PLA	<ul style="list-style-type: none"> - PLA is a hydrophobic polymer that is also capable of activating the process of phagocytosis by macrophages. - The different route of administration gives different immune expression which might influence the efficacy of drug delivery. - The lactate produced by PLA degradation also causes the lung area to become more acidic which will affect the stability of the particular drug it delivers. 	41
Alginate	<ul style="list-style-type: none"> - Alginate is composed of manuronic acid which can be a specific ligand for TLR-2 and TLR-4 receptors found in infected macrophages. - Manuronic acid influences the innate immune response that is responsible for the activation process of the bactericidal effect on host cells. Manuronic acid also plays a role in increasing the activity of macrophage phagocytosis. 	36, 53, 70, 71
Carragenan	The presence of sulfate in carrageenan makes this polymer negatively charged as an alternative to targeted systems in macrophages, where carrageenan will bind to CysD in the extracellular region of the cell. To increase the stability and efficiency of its entrapment, carrageenan is combined with other polymers such as chitosan and alginate.	58, 72, 73
Gelatine	Gelatine has low antigenic properties and has an active group that can bind to the human receptors on macrophages. Gelatine can be modified to increase the tendency of microparticles to be taken up by macrophages. The presence of a free -NH ₂ group on gelatin provides a side to be conjugated with mannose to deliver the drug more effectively.	51, 60, 74
Chitosan	<ul style="list-style-type: none"> - Positive charge of amino groups of chitosan can increase contact time between particles and negatively charged respiratory tract mucosa. - Chitosan with different molecular weights gives different cell uptake and trap efficiency. - Chitosan interacts with macrophage manosa receptors that trigger phagocytosis followed by degradation of lysozyme and N-acetyl-β-D-glucosamidase in phagosomes. - Chitosan able to activate macrophages by increasing the production of proinflammatory cytokines such as TNF-α, IL-1β and IL-6 and decreasing the release of anti-inflammatory cytokines IL-10. 	32, 75, 76, 77

Conclusion

To sum up, the various advantages of inhalation delivery system make many researchers compete to obtain an effective formulation by making various modifications both to the carrier system and to the excipient. Modifications are made to obtain an appropriate size for the inhalation process with good aerosolization properties and enhancement of drug loading carrier system and to obtain the desired release profile. Furthermore, alveolar macrophages are potential targets for more efficient drug delivery, especially for handling respiratory tract infections and diseases. The microspheres carrying system has been modified in such a way as to increase uptake by alveolar macrophages, either by utilizing a passive target system or by adding ligands to the surface of the microspheres. The chemical and physical properties of particulate systems, such as particle size, shape, surface, charge, and other properties directly and indirectly contribute to the increase or decrease in particle uptake by macrophages. This goes back to the initial purpose of providing therapy. This will have an impact on the pattern of drug distribution in the respiratory tract which will determine the point of drug accumulation in the respiratory system.

Conflict of interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

Acknowledgements

Authors would like to thank to Directorate of General Higher Education (DRPM DIKTI) for grant and Faculty of Pharmacy Universitas Airlangga for research support and facilities.

References

- Rashid J, Patel B, Nozik-Grayck E, McMurtry IF, Stenmark KR, Ahsan F. Inhaled sildenafil as an alternative to oral sildenafil in the treatment of pulmonary arterial hypertension (PAH). *J Cont Rel.* 2017; 250:96-106.
- Jyoti K, Pandey RS, Kush P, Kaushik D, Jain UK, Madan J. Inhalable bioresponsive chitosan microspheres of doxorubicin and soluble curcumin augmented drug delivery in lung cancer cells. *Int J Biol Macromol.* 2017; 98:50-58.
- Lin XF, Kankala RK, Tang N, Xu P-Y, Hao, L-Z, Yang D-Y, Wang S-B, Zhang YS, Chen AZ. Supercritical Fluid-Assisted Porous Microspheres for Efficient Delivery of Insulin and Inhalation Therapy of Diabetes. *Adv. Healthcare Mater.* 2019; 8:1800910.
- She W, Mei Z, Zhao H, Li G, Lin Y. Nebulized inhalation of Anti Nerve Growth Factor Microspheres Inhibits Airway remodeling in an Ovalbumin-induced Rat Asthma Model. *J Aerosol Pulm Drug Deliv.* 2019; 32(2):70-77.
- Yildiz-Pekoz A and Ozsoy Y. Inhaled Heparin: Therapeutic Efficacy And Recent Formulations. *J Aerosol Med Pulm Drug Deliv.* 2017; 30(3):143-156.
- Parikh R, Dalwadi S, Aboti P, Patel L. Inhaled microparticles of antitubercular antibiotic for *in vitro* and *in vivo* alveolar macrophage targeting and activation of phagocytosis. *The J of Antibio.* 2014; (67):387-394.
- Klinkert K, Whelan D, Clover Aj, Leblond AL, Kumar AH, Caplice, NM. Selective M2 macrophage depletion leads to prolonged inflammation in surgical wounds. *Eur Surg Res.* 2017; 58:109-120.
- Aldawsari HM, Gorain B, Alhakamy NA, Md S. Role of therapeutic agents on repolarisation of tumour-associated macrophage to halt lung cancer progression. *J Drug Target.* 2020; 28(2):166-175.
- Zhang L, Wang Y, Wu G. Macrophages: friend or foe in idiopathic pulmonary fibrosis?. *Respir Res.* 2018; 19:170.
- Swirski FK, Robins CS, Nahedroff M. Development and function of arterial and cardiac macrophages. *Trends Immunol.* 2016; 37:32-40.
- Jayachandran R, Dasgupta SB, Pieters J. Surviving the macrophage: tools and tricks employed by Mycobacterium tuberculosis. *Curr Top Microbiol Immunol.* 2014; 374:189-209.
- Roopngam P, Liu K, Mei L, Zheng Y, Zhu X, Tsai HI, Huang L. Hepatitis C virus E2 protein encapsulation into poly d, l-lactic-co-glycolide microspheres could induce mice cytotoxic T-cell response. *Int J Nanomed.* 2016; 11:5361-5370.
- Loira-Pastoriza C, Todorof J, Vanveber R. Delivery Strategies for sustained drug release in the lung. *Adv Drug Deliv Rev.* 2014; 75:81-91.
- Paranjpe M and Müller-Goymann CC. Nanoparticle-mediated pulmonary drug delivery: a review. *Int J Mol Sci.* 2014; 8;15(4):5852-73.
- Dhand C, Molamma PP, Oger W, Beuerman R, Lakshminarayanan, Neeraj D, Seeram R. Role of size of drug delivery carriers for pulmonary and intravenous administration with emphasis on cancer therapeutics and lung-targeted drug delivery. *Review. RSC Adv.* 2014; 4:32673-32689.
- Weiss G and Schaible UE. Macrophage defense mechanisms against intracellular bacteria. *Immunol Rev.* 2015; 264:182-203.
- Mosaiab T, Farr DC, Keifel MJ, Houston TA. Carbohydrate-based nanocarriers and their application to target macrophages and deliver antimicrobial agent. *Adv Drug Deliv Rev.* 2019; (15):94-129.
- Sprenger M, Kasper L, Hensel M, Hube B, Metabolic adaptation of intracellular bacteria and fungi to macrophages. *Int J Med Microbiol.* 2018; 308(1):215-227.
- Morales-Nebreda L, Misharin AV, Perlman H, Budinger GR. The heterogeneity of lung macrophages in the susceptibility to disease. *Eur Resp Rev.* 2015; 24:505-509.
- Garapaty A and Champion JA. Tunable particles alter macrophage uptake based on combinatorial effects of physical properties. *Bioengin & Trans Med.* 2017:92-101.
- Nimje N, Agarwal A, Saraogi GK, Lariya N, Rai G, Agrawal H, Agrawal GP. Mannosylated nanoparticulate carriers of rifabutin for alveolar targeting. *J Drug Target.* 2009; 17:777-787.
- Liu Y, Hardie J, Zhang X, Rotello VM. Effects of engineered nanoparticles on the innate immune system. *Sem Immunol.* 2017; 34:25-32.
- Geiser M. Update on macrophage clearance of inhaled micro- and nanoparticles. *J Aerosol Med Pulm Drug Deliv.* 2010; 23:207-217.
- El-Sherbiny IM, Villanueva DG, Herrera D, Smyth HD. Overcoming lung clearance mechanisms for controlled release drug delivery. *Cont Pulm Drug Deliv.* Springer. 2011. 101-126 p.
- Hirota K, Hasegawa T, Nakajima T, Inagawa H, Kohchi C, Soma G, Makino K, Terada H. Delivery of rifampicin-PLGA microspheres into alveolar macrophages is promising for treatment of tuberculosis. *J Cont Rel.* 2010; 142:339-346.
- Costa A, Pinheiro M, Magalhães J. The formulation of nanomedicines for treating tuberculosis. *Adv Drug Deliv Rev.* 2016; 102:102-115.
- Iversen TG, Skotland T, Sandvig K. Endocytosis and intracellular transport of nanoparticles: Present knowledge

- and need for future studies. *Nano Today*. 2011; 6(2):176-185.
28. Yang Y, Bajaj N, Xu P, Ohn K, Tsifansky MD, Yeo Y. Development of highly porous large PLGA microparticle for pulmonary drug delivery. *Biomater*. 2009; 30(10): 1947-1953.
 29. Makino K, Nobuko Y, Kazue H, Nobuyuki H, Hiroyuki O, Hiroshi T. Phagocytic uptake of polystyrene microspheres by alveolar macrophages: effects of the size and surface properties of the microspheres. *Coll Surf B: Biointer*. 2003; 27:33-39.
 30. Hirota K, Taizo H, Hideyuki H, Fuminori I, Hiroyuki I, Chie K, Gen-Ichiro S, Kimiko M, Hiroshi Terada. Optimum conditions for efficient phagocytosis of rifampicin-loaded PLGA microspheres by alveolar macrophages. *J Cont Rel*. 2007; 119:69-76.
 31. Hwang SM, Kim DD, Chung SJ. Delivery of ofloxacin to the lung and alveolar macrophages via hyaluronan microspheres for the treatment of tuberculosis. *J Cont Rel*. 2008; 129:100-106.
 32. Park JH, Hyo-Eon J, Dae-Duk K, Suk-Jae C, Won-Sik S, Chang-Koo S. Chitosan microspheres as an alveolar macrophage delivery system of ofloxacin via pulmonary inhalation. *Int J Pharm*. 2013; 441:562-569.
 33. Tiwari S, Chaturvedi AP, Tripathi YB, M Brameshwar. Microspheres based on mannosylated lysine-co-sodium alginate for macrophage-specific delivery of isoniazid. *Carb Pol*. 2012; 87(2):1575-1582.
 34. Zhiqiang L, Xia L, Bingshui X, Cuimi D, Jiangxue L, Xuhui Z, Xiqin Y, Wenhao D, Heather J, Heqiu Z, Xiaoyan F. A novel and simple preparative method for uniform-sized PLGA microspheres: Preliminary application in antitubercular drug delivery. *Coll Surf B: Biointer*. 2016; 145:679-687.
 35. Højby N. Recent advances in the treatment of *Pseudomonas aeruginosa* infections in cystic fibrosis. *BMC Med*. 2012; 9:32-38.
 36. Lakio S, Morton DAV, Ralph AP, Lambert P. Optimizing aerosolization of a high-dose L-arginine powder for pulmonary delivery. *AJPS*. 2015; 10(6):528-540.
 37. Luinstra M, Grasmeijer F, Hagedoorn P, Moes JR, Frijlink HW, Boer AH. A levodopa dry powder inhaler for the treatment of Parkinson's disease patients in off periods. *Eur J Pharm Biopharm*. 2015; 97:22-29.
 38. Du P, Du J, Smyth HD. Evaluation of Granulated Lactose as a Carrier for DPI Formulations 1: Effect of Granule Size. *AAPS*. 2016; 15(6):1417-1428.
 39. Takeuchi I, Yoshihiro T, Yuki T, Kazuhiro O, Kimiko M. Effects of L-leucine on PLGA microparticles for pulmonary administration prepared using spray drying: fine particle fraction and phagocytotic ratio of alveolar macrophages. *Coll Surf A: Phys Eng Asp*. 2018; 537:411-417.
 40. David CJ, Patel RB, Mitchell JP. Discriminating Ability of Abbreviated Impactor Measurement Approach (AIM) to Detect Changes in Mass Median Aerodynamic Diameter (MMAD) of an Albuterol/Salbutamol pMDI Aerosol. *AAPS*. 2017; 18:3296-3306.
 41. Sharma D, Valenta DT, Altman Y, Harvey S, Xie H, Mitragotri S, Smith JW. Polymer Particle shape independently influences binding and internalization by macrophages. *J Cont Rel*. 2010; 147(3):408-412.
 42. Chikaura H, Nakashima Y, Fujiwara Y, Komohara Y, Takeya M, Nakanishi Y. Effect of particle size on biological response by human monocyte-derived macrophages. *Biosurf Biotri*. 2016; 2(1):18-25.
 43. Yoo JW and Mitragotri S. Polymer particles that switch shape in response to a stimulus. In *Proc Natl Acad Sci*. 2010; 107(25):1125-11210.
 44. Pricer WE and Ashwell G. The binding of desialylated glycoproteins by plasma membranes of rat liver. *J Biol Chem*. 1971; 246:4825-4833.
 45. Witten J and Ribbeck K. The particle in the spider's web: transport through biological hydrogels. *Nanoscale*. 2017; 9:8080-8095.
 46. Dosio F, Arpicco S, Stella B, Fattal E. Hyaluronic acid for anticancer drug and nucleic acid delivery. *Adv Drug Deliv Rev*. 2016; 97:204-236.
 47. Wei X, Shao B, He Z. Cationic nanocarriers induce cell necrosis through impairment of Na⁺/K⁺-ATPase and cause subsequent inflammatory response. *Cell Res*. 2016; 25(2):237.
 48. Zhang C, Gaona S, Ju Z, Huijuan S, Jinfeng N, Shengbin S, Pingsheng H, Yanming W, Weiwei W, Chen L, Deling K. Targeted antigen delivery to dendritic cell via functionalized alginate nanoparticles for cancer immunotherapy. *J Cont Rel*. 2017; 256:170-181.
 49. Eleonora M, Luca C, Cecilia R, Eliana L, Maria AC, Francesca B, Eleonora T, Valentina I. Surface engineering of Solid Lipid Nanoparticle assemblies by methyl α -D-mannopyranoside for the active targeting to macrophages in anti-tuberculosis inhalation therapy. *Int J Pharm*. 2017; 528: (1-2):440-451.
 50. Carneiro SP, Carvalho KV, Soares RD, Martin C, Andrade MHG, Duarte RS, Santos OH. Functionalized rifampicin-loaded nanostructured lipid carrier enhance macrophages uptake and antimycobacteril activity. *Coll Surf B: Biointer*. 2019; 175:306-313.
 51. Tiwari S, Chaturvedi AP, Tripathi YB, Mishra B. Macrophage-specific targeting of isoniazid through mannosylated gelatin microspheres. *AAPS*. 2011; 12(3):900-908.
 52. Beningo KA and Yu-Li W. Fc-receptor-mediated phagocytosis is regulated by mechanical properties of the target. *J Cell Sci*. 2002; 15(115):849-56.
 53. Sharma A, Vagashiya K, Verma RK. Inhalable microspheres with hierarchical pore size for tuning the release of biotherapeutic in lungs. *Microp Mesop Mat*. 2016; 235:195-203.
 54. Nishimura S, Takami T, Murakami Y. Porous PLGA microparticle formed by 'one-step' emulsification for pulmonary drug delivery: the surface morphology and the aerodynamic properties. *Coll and surf B: Biointer*. 2017; 159:318-326.
 55. Baldeli A and Vehring R. Analysis of cohesion forces between monodisperse microparticle with rough surface. *Coll Surf. A : Phys Eng Asp*. 2016; 506:179-189.
 56. Contreras LG, Sung J, Ibrahim M, Elbert K, Edwards D, Hickey A. Pharmacokinetics of inhaled rifampicin porous particle for tuberculosis treatment: insight into rifampicin absorption from the lungs of guinea pigs. *Mol Pharm*. 2015; 12(8):2642-2650.
 57. Chvatal A, Rita A, Petra P, Gabor K, Orsolya JL, Piroška S, Elias F and Nicolas T. Formulation and comparison of spray dried non-porous and large porous particles containing meloxicam for pulmonary drug delivery. *Int J Pharm*. 2019; 559:68-75.
 58. Paula GA, Benevides NM, Cunha AP, de Oliveira AV, Pinto AMB, Morais JPS, Azeredo HMC. Development and characterization of edible films from mixtures of κ -carrageenan, ι -carrageenan, and alginate. *Food Hydrocoll*. 2015; 47:140-145.
 59. Ramaiah B, Nagaraja, SH, Kapanigowda, UG. High azithromycin concentration in lungs by way of bovine serum albumin microspheres as targeted drug delivery: lung targeting efficiency in albino mice. *DARU J Pharm Sci*. 2016; 24:14.
 60. Viswnathan V, Mehta H, Pharande R, Bannaliker A, Gupta P, Gupta U, Mukne A. Mannosylated gelatin nanoparticles of licorice for use in tuberculosis: Formulation, in vitro

- evaluation, *in vitro* cell uptake, *in vivo* pharmacokinetics and *in vivo* anti-tubercular efficacy. *J Drug Deliv Sci Tech.* 2018; 45:225-263.
61. Li Z, Zheng H, Li X, Su J, Qin L, Sun Y, Guo C, Moritz B, Moehwald M, Chen L, Zhang Y, Mao S. Phospholipid-modified poly(-co-glycolide) microparticles for tuning the interaction with macrophages: *in vitro* and *in vivo* assessment. *Eur J Pharm Biopharm.* 2019; 143:70-79.
 62. Hamilton R, Thakur SA, Holian A. Silica binding and toxicity in alveolar macrophages. *Free Rad Biol Med.* 2008; 44(7):1246-1258.
 63. Sano H and Kuroki Y. The lung collectins, SP-A and SP-D, modulate pulmonary innate immunity. *Mol Immunol.* 2005; 42:279-287.
 64. Jones BG, Dickinson PA, Gumbleton M, Kellaway IW. The inhibition of phagocytosis of respirable microspheres by alveolar and peritoneal macrophages. *Int J Pharm.* 2002; 236:65-79.
 65. Ventura CA, Tommasini S, Crupi E, Giannone I, Cardile V, Musumeci T, Puglisi G. Chitosan microspheres for intrapulmonary administration of moxifloxacin: Interaction with biomembrane models and *in vitro* permeation studies. *Eur J Pharm Biopharm.* 2008; 68(2):235-244.
 66. Kolensyk I, Konovalova V, Burhan A. Alginate/ κ -carrageenan microspheres and their application for protein drug controlled release. *Chem Chem Tech.* 2015; 9(4):485-492.
 67. Gaspar MC, Alberto AC, Pais-João JS, Julien B, Jean CO. Development of levofloxacin-loaded PLGA microspheres of suitable properties for sustained pulmonary release. *Int J Pharm.* 2019; 556:117-127.
 68. Bitencourt C, da Silva LB, Pereira PA, Gelfuso G.M, Faccioli L.H. Microspheres prepared with different copolymers of poly(lactic-glycolic acid) (PLGA) or with chitosan cause distinct effects on macrophages. *Coll Surf B Biointer.* 2015; 136: 678-686.
 69. Gizem RT, Burcu D, Mijde E, Asuman B. Design of ciprofloxacin-loaded nano-and microcomposite particles for dry powder inhaler formulations: preparation, *in vitro* characterisation, and antimicrobial efficacy. *J Microencaps Micro Nano Carr.* 2018; 35(6), 533-547.
 70. West AP, Brodsky IE, Rahner C, Woo DK, Erdjument-Bromage H, Tempst P. TLR signalling augments macrophage bactericidal activity through mitochondrial ROS. *Nature.* 2011; 472:476-480.
 71. Vaghasiya K, Eram A, Sharma A. Alginate Microspheres Elicit Innate M1-Inflammatory Response in Macrophages Leading to Bacillary Killing. *AAPS.* 2019; 20:241.
 72. Elbi S, Nimal TR, Rajan VK, Baranwal G, Biswas R, Jayakumar R, Sathianarayanan S. Fucoidan coated ciprofloxacin loaded chitosannanoparticles for the treatment of intracellular and biofilm infections of Salmonella. *Coll Surf. B: Biointer.* 2017; 160:40-47.
 73. Abdelghany SM, Alkhalaf H, Al Khatib. Carageenan-stabilised chitosan alginate nanoparticles loaded with ethionamide for the treatment of tuberculosis. *J Drug Deliv Sci Tech.* 2017; 39:442-449.
 74. Mura S, Hillaireau H, Nicolas J, Kerdine-Römer S, Le DB, Deloménie, CN, Pallardy M, Tsapis N, Fattal E. Biodegradable nanoparticles meet the bronchial airway barrier: how surface properties affect their interaction with mucus and epithelial cells. *Biomacromol.* 2011; 12:4136-4143.
 75. Bagre A, Narendra KL, Mohan LK. Therapeutic Management of Pulmonary Tuberculosis by Mannosylated Chitosan Ascorbate Microspheres: Preparation and Characterization. *J Drug Deliv Ther.* 2019; 9(3):13-25.
 76. Gaspar MC, Sousa AP, Cardoso O, Murtinho D, Serra MS, FTewes, Olivier JC. Optimization of levofloxacin-loaded crosslinked chitosan microspheres for inhaled aerosol therapy. *Eur J of Pharm and Biopharm.* 2015; 96:65-75.
 77. Oliveira PM, Matos BN, Pereira PAT, Gratieri T, Faccioli LH, Marcilio SS, Filho C, Gelfuso GM. Microparticles prepared with 50-190 kDa chitosan as promising non-toxic carriers for pulmonary delivery of isoniazid. *Carb Pol.* 2017; 174:427-431.

Oxygen Transport Whitepaper

Download the whitepaper to learn more

GE Healthcare

DOWNLOAD

Tropical Journal of Natural Product Research

COUNTRY

Nigeria



Universities and research institutions in Nigeria

SUBJECT AREA AND CATEGORY

Biochemistry, Genetics and Molecular Biology
Biochemistry
Molecular Medicine

Chemistry
Analytical Chemistry

Medicine
Complementary and Alternative Medicine

Pharmacology, Toxicology and Pharmaceutics
Drug Discovery
Pharmaceutical Science
Pharmacology

PUBLISHER

Faculty of Pharmacy, University of Benin

H-INDEX

2

PUBLICATION TYPE

Journals

ISSN

26160692, 26160684

COVERAGE

2017-2020

INFORMATION

[Homepage](#)

[How to publish in this journal](#)

[Contact](#)

Submit your EV Article

Open Access EV Journal

Open Access journal pushing research on extracellular vesicles, microvesicles, and exosomes.

onlinelibrary.wiley.com

SCOPE

The Tropical Journal of Natural Product Research is open access, peer-reviewed international journal aimed at making important contributions in the field of Natural Product Research, Pharmaceutical and Natural Sciences. The journal covers all aspects of Pharmaceutical research, chemistry and biochemistry of naturally occurring compounds, ethnomedicine, ethnopharmacology, pharmacognosy, biomedical research, biotechnology and related disciplines. The journal welcome submissions from a broad spectrum of scientific endeavour involving biological evaluation of natural substances of plant, microbial and animal origin against different disease targets, processes or therapeutic strategies that can lead to or assist in the prevention and management of chronic and infectious diseases, clinical therapeutics, Isolation and characterization of metabolites, structure elucidation, synthesis and experimental biosynthesis of natural Product as well as developments of methods in these areas. Research papers in the fields of chemistry-biology boundary, e.g. fermentation chemistry, plant tissue culture investigations etc. are also welcomed. Although the journal focuses mainly on original research articles, timely, concise and focused reviews on recent progress in active areas of natural Product is also encouraged.

 Join the conversation about this journal

Open Access EV Journal

Submit your abstract

onlinelibrary.wiley.com

OPEN



Quartiles



Roulette Terbaru 2021

Sebuah permainan kecil yang bisa dimainkan kapan saja, dimana saja datang dan bersantai

777slot.io

B



1 **Pharmacognosy Journal**

IND

76%
similarity

2 **Journal of Traditional and Complementary Medicine**

NLD

76%
similarity

3 **Pharmacologyonline**

ITA

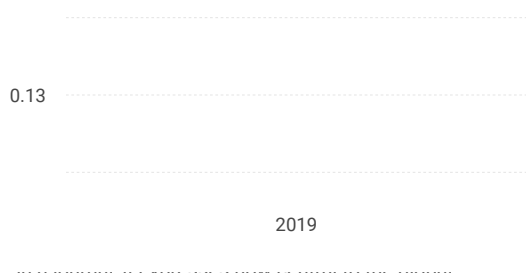
75%
similarity

4 **Pharmacognosy Research**

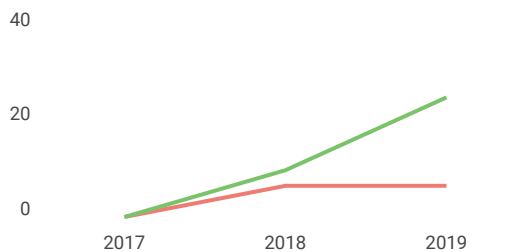
IND

73%
similarity

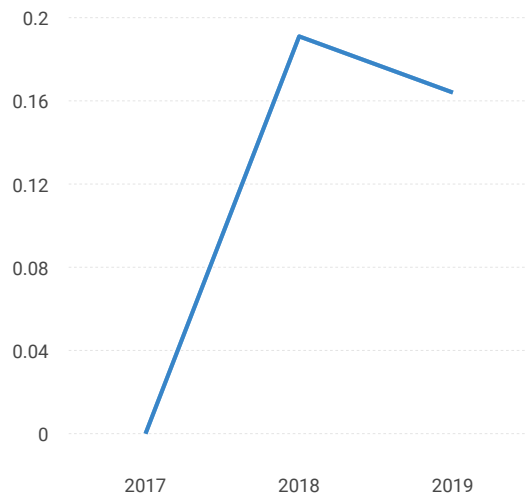
SJR



Total Cites Self-Cites

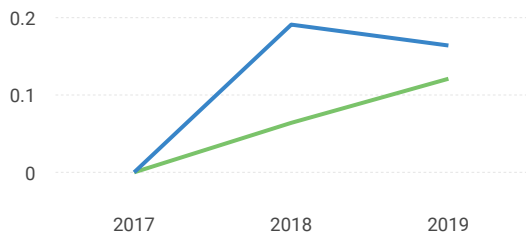


Citations per document

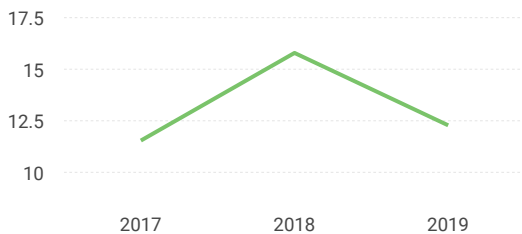


External Cites per Doc

Cites per Doc



% International Collaboration



Citable documents

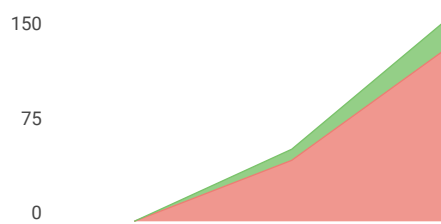
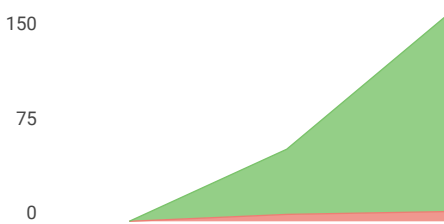
Non-citable documents



Cited documents

Uncited documents





Tropical Journal of Natural Product Research

Q3 Pharmaceutical Science
best quartile

SJR 2019
0.13

powered by scimagojr.com

← Show this widget in your own website

Just copy the code below and paste within your html code:

```
<a href="https://www.scimaç
```

New Open Access Journal - Online Only

Browse the new issue from Open Access journal Function. academic.oup.com

Metrics based on Scopus® data as of April 2020

Leave a comment

Name

Email

(will not be published)

I'm not a robot

reCAPTCHA
Privacy - Terms

Submit

The users of Scimago Journal & Country Rank have the possibility to dialogue through comments linked to a specific journal. The purpose is to have a forum in which general doubts about the processes of publication in the journal, experiences and other issues derived from the publication of papers are resolved. For topics on particular articles, maintain the dialogue through the usual channels with your editor.

Developed by:



Powered by:



Follow us on [@ScimagoJR](#)

Scimago Lab, Copyright 2007-2020. Data Source: Scopus®

EST MODUS IN REBUS

Horatio (Satire 1, 1, 106)
