

**Research (Published online: 08-12-2021)**

**1. Antioxidant activity and protective effect of propolis against carbon tetrachloride-induced liver and kidney injury by modulation of oxidative parameters**

Redouan El-Haskoury, Noori Al-Waili, Zeineb Kamoun, Mohamed Makni, Ahmed Al-Waili and Badiaa Lyoussi  
Veterinary World, 14(12): 3076-3083

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/1.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/1.pdf>)

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**Research (Published online: 08-12-2021)**

**2. Development of a rapid and reliable high-performance liquid chromatography method for determination of water-soluble vitamins in veterinary feed premix**

Md. Zahangir Hosain, S. M. Shariful Islam and Md. Mostofa Kamal  
Veterinary World, 14(12): 3084-3090

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/2.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/2.pdf>)

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**Research (Published online: 09-12-2021)**

**3. Improving the knowledge of high school students about zoonotic diseases from pets in Medellín-Colombia**

Natalia Uribe Corrales, Karen Velásquez Giraldo, Catalina María Saldarriaga Garcés and Angie Lorena Navarro Giraldo  
Veterinary World, 14(12): 3091-3098

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/3.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/3.pdf>)

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**Research (Published online: 13-12-2021)**

**4. Antimicrobial resistance profile of extended-spectrum beta-lactamases, adenosine-monophosphate-cyclic, and carbapenemase-producing Gram-negative bacteria isolated from domestic animals**

Jôiciglecia Pereira dos Santos, Valesca Ferreira Machado de Souza, Marcos Wilker da Conceição Santos, Juliany Nunes dos Santos, Natilene Silva dos Santos, Angélica Prado de Oliveira, Valquíria Tatiele da Silva Rodrigues, Ianei Oliveira Carneiro and Layze Cilmar Alves da Silva Vieira  
Veterinary World, 14(12): 3099-3104

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/4.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/4.pdf>)

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**Research (Published online: 14-12-2021)**

**5. Development of multiplex reverse transcription-polymerase chain reaction for differentiation of strains of infectious bursal disease virus and primary screening of the virus in Thailand**

Nataya Charoenvisal

Veterinary World, 14(12): 3105-3110

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/5.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/5.pdf>)

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**Research (Published online: 14-12-2021)**

**6. Sociodemographic, income, and environmental characteristics of individuals displaying animal and object hoarding behavior in a major city in South Brazil: A cross-sectional study**

Graziela Ribeiro da Cunha, Camila Marinelli Martins, Maysa Pellizzaro, Christina Pettan-Brewer and Alexander Welker Biondo

Veterinary World, 14(12): 3111-3118

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/6.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/6.pdf>)

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**Research (Published online: 15-12-2021)**

**7. Validation of single-step genomic predictions using the linear regression method for milk yield and heat tolerance in a Thai-Holstein population**

Piriyaporn Sungkhapreecha, Ignacy Misztal, Jorge Hidalgo, Daniela Lourenco, Sayan Buaban, Vibuntita Chankitisakul and Wuttigrai Boonkum

Veterinary World, 14(12): 3119-3125

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/7.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/7.pdf>)

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**Research (Published online: 19-12-2021)**

**8. Detection of *invA* virulence gene of multidrug-resistant *Salmonella* species isolated from the cloacal swab of broiler chickens in Blitar district, East Java, Indonesia**

Freshindy Marissa Wibisono, Hayyun Durrotul Faridah, Freshinta Jellia Wibisono, Wiwiek Tyasningsih, Mustofa Helmi Effendi, Adiana Mutamsari Witaningrum and Emmanuel Nnabuike Ugbo

Veterinary World, 14(12): 3126-3131

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/8.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/8.pdf>)

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**Research (Published online: 20-12-2021)**

**9. Apigenin induces oxidative stress in mouse Sertoli TM4 cells**

Sathaporn Jittapalapong, Thapanee Poompoung and Samak Sutjarit  
Veterinary World, 14(12): 3132-3137

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/9.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/9.pdf>)

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**Research (Published online: 20-12-2021)**

**10. Detection of microbial contamination in chicken meat from local markets in Surabaya, East Java, Indonesia**

Dhandy Koesoemo Wardhana, Ajeng Erika Prihastuti Haskito, Muhammad Thohawi Elziyad Purnama, Devi Ayu Safitri and Suwaibatul Annisa  
Veterinary World, 14(12): 3138-3143

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/10.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/10.pdf>)

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**Research (Published online: 24-12-2021)**

**11. The first report of the seroprevalence of antibodies against *Bartonella* spp. in water buffaloes (*Bubalus bubalis*) from South Thailand**

Sumalee Boonmar, Phirabhat Saengsawang, Watcharapong Mitsuwan, Decha Panjai, Kamchai Kidsin, Chalutwan Sansamur and Ittidet Wichianrat  
Veterinary World, 14(12): 3144-3148

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/11.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/11.pdf>)

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**Research (Published online: 25-12-2021)**

**12. Diversity of gastrointestinal parasites in sympatric mammals in Moukalaba-Doudou National Park, Gabon**

Serge-Ely Dibakou, Ulrich Maloueki, Barthélémy Ngoubangoye, Larson Boundenga, Stephan Ntie, Thierry-Audrey Tsoumbou, Cyr Moussadji, Rina Obame Zang, Dikenane Kombila and Didier Basset  
Veterinary World, 14(12): 3149-3155

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/12.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/12.pdf>)

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**Research (Published online: 25-12-2021)**

**13. Establishment of semen collection technique using electroejaculator and semen cryopreservation of Javan leopard (*Panthera pardus melas* Cuvier, 1809)**

Bongot Huaso Mulia, Ardyta Widiyanti, Jansen Manansang, Dedi Rahmat Setiadi, Vincentia Trisna Yoelinda, Taufiq Purna Nugraha, Ni Wayan Kurniani Karja and Raden Iis Arifiantini  
Veterinary World, 14(12): 3156-3163

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/13.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/13.pdf>)

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**Research (Published online: 26-12-2021)**

**14. Impact of L-carnitine supplementation on the *in vitro* developmental competence and cryotolerance of buffalo embryos**

Mohamed M. M. El-Sokary, Al-Shimaa Al-H. H. El-Naby, Amal R. Abd El Hameed, Karima Gh. M. Mahmoud and T. H. Scholkamy  
Veterinary World, 14(12): 3164-3169

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/14.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/14.pdf>)

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**Research (Published online: 26-12-2021)**

**15. Analysis of culling reasons during the breeding cycle and lifetime performance: The strategy to remove crossbred Landrace and Large White sows under tropical climate**

Ratchadaporn Boripun, Watcharapong Mitsuwan, Pawinee Kulnanan, Thotsapol Thomrongsuwannakij and Warangkana Kitpipit  
Veterinary World, 14(12): 3170-3174

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/15.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/15.pdf>)

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**Research (Published online: 28-12-2021)**

**16. *In silico* molecular docking and *in vitro* analysis of ethanolic extract *Ocimum sanctum* Linn.: Inhibitory and apoptotic effects against non-small cell lung cancer**

Ulayatul Kustiati, T. S. Dewi Ratih, N. Dwi Aris Agung, Dwi Liliek Kusindarta and Hevi Wihadmadyatami  
Veterinary World, 14(12): 3175-3187

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/16.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/16.pdf>)

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**Research (Published online: 28-12-2021)**

**17. Effects of xylazine and adrenaline combinations: Preliminary clinical application for non-surgical protocols of nephrosplenic entrapment in horses**

Worakij Cherdchutham, Patskit Sukhong, Kanchanog Sae-oueng, Nithisphat Supanwinijkul, Kittanai Wiangnak, Jirayut Srimuang, Tawanhathai Apichaimongkonkun, Sarocha Limratchapong and Soontaree Petchdee  
Veterinary World, 14(12): 3188-3193

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/17.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/17.pdf>)

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**Research (Published online: 29-12-2021)**

**18. Validation of Brix refractometer to estimate immunoglobulin G concentration in goat colostrum**

Chollada Buranakarl, Sumpun Thammacharoen, Morakot Nuntapaitoon,  
Sapon Semsirboon and Kazuo Katoh  
Veterinary World, 14(12): 3194-3199

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/18.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/18.pdf>)

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**Research (Published online: 29-12-2021)**

**19. Therapeutic effect of biosynthetic gold nanoparticles on multidrug-resistant *Escherichia coli* and *Salmonella* species isolated from ruminants**

Abeer M. Abdalhamed, Alaa A. Ghazy, Eman S. Ibrahim, Amany A. Arafa and  
Gamil S. G. Zeedan  
Veterinary World, 14(12): 3200-3210

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/19.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/19.pdf>)

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**Research (Published online: 30-12-2021)**

**20. Comparison of enzyme-linked immunosorbent assay and Fassis<sup>®</sup> bovine immunoglobulin G (IgG) immunoassay for quantification of bovine IgG in neonatal calf serum**

Marian Hampe, Stefanie Söllner-Donat, Klaus Failing and Axel Wehrend  
Veterinary World, 14(12): 3211-3215

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/20.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/20.pdf>)

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**Research (Published online: 30-12-2021)**

**21. Prevalence and antibiotic resistance of *Salmonella* isolated from poultry and its environment in the Mekong Delta, Vietnam**

Thuan K. Nguyen, Lam T. Nguyen, Trang T. H. Chau, Tam T. Nguyen, Bich N.  
Tran, Takahide Taniguchi, Hideki Hayashidani and Khai T. L. Ly  
Veterinary World, 14(12): 3216-3223

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/21.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/21.pdf>)

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**Research (Published online: 31-12-2021)**

**22. Melanocortin-4 receptor and leptin as genes for the selection of superior Madrasin cattle**

Budi Utomo, Rimayanti Rimayanti, Indah Norma Triana and Amaq Fadholly  
Veterinary World, 14(12): 3224-3228

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/22.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/22.pdf>)

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**Research** (Published online: 31-12-2021)

**23. The self-nanoemulsifying drug delivery system of *Petiveria alliacea* extract reduced the homeostatic model assessment-insulin resistance value, interleukin-6, and tumor necrosis factor- $\alpha$  level in diabetic rat models**

Arifa Mustika, Nurmawati Fatimah and **Gadis Meinar Sari**

Veterinary World, 14(12): 3229-3234

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/23.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/23.pdf>)

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**Research** (Published online: 31-12-2021)

**24. Culture of spermatogonial stem cells and use of surrogate sires as a breeding technology to propagate superior genetics in livestock production: A systematic review**

Wilkister Nakami, Ambrose Ng'eno Kipyegon, James Nguhiu-Mwangi, Christian Tiambo and Stephen Kemp

Veterinary World, 14(12): 3235-3248

Abstract (<http://www.veterinaryworld.org/Vol.14/December-2021/24.html>)

PDF (<http://www.veterinaryworld.org/Vol.14/December-2021/24.pdf>)

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Interest area: Brucellosis - Microbiology - Veterinary Medicine - Veterinary Public Health - Zoonoses

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Interest area: Bacteriology - Clinical Microbiology - Infectious Diseases - Veterinary Medicine

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Interest area: Poultry Husbandry

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Interest area: Poultry Husbandry - Poultry Medicine

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Interest area: Dairy Science - Epidemiology - Food Science - Food Technology - Food Law - One Health - Parasitology - Meat Inspection - Pathogens - Foodborne Diseases - Food Toxicology - Veterinary Public Health - Wildlife Diseases - Zoonoses

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Interest area: Biology - Ethnoveterinary - Parasitology - Veterinary Medicine - Veterinary Public Health

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Interest area: Biology - Animal Nutrition

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Interest area: Large Animal Medicine - Surgery - Veterinary Medicine

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Interest area: Animal Science - Antimicrobial resistance

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Interest area: Biotechnology - Genetics - Veterinary Medicine

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Interest area: Pharmacology - Toxicology - Physiology - Cardiology

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Interest area: Zoonotic parasitic diseases - Parasite phylogeny - Zoology - Parasitology

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Interest area: Classical swine fever - African swine fever - Pseudorabies - Innate and adaptive immunity - Virus-host interactions - Pathogenesis - Epidemiology - Vaccines - Diagnostic assays - Probiotics

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Interest area: Food - Food science - Food Technology - Food borne diseases - Antibiotic resistance - One Health - Veterinary Public Health

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Interest area: Internal Medicine - Spectrophotometry - Ultrasonography - Parasitological analysis - Blood gas analysis - Metabolic profiling - Veterinary Medicine - Large Animal Medicine - Equine Medicine - Mastitis

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Interest area: Clinical and Anatomic Pathology - Oncology - Fisheries with special interest in bacterial diseases of fishes and other diseases associated with aquaculture management - Diseases of small and large ruminants - Laboratory animal medicine - Diseases of Dogs, horses and pigs as well as non-infectious diseases such as Diabetes and stress-induced pathologies

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Interest area: Antimicrobial resistance - Infectious Diseases - Poultry Medicine - Veterinary Medicine - Veterinary Microbiology and Parasitology - Veterinary Public Health - Veterinary Science - Meta-analysis - Phylogenetic analysis

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Interest area: Antimicrobial resistance - Antibiotic growth promoters in poultry and their alternatives - Phytochemicals - Oxidative stress and antioxidants - Mycotoxin toxicity and amelioration - Selenium and selenoproteins

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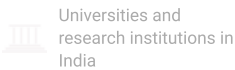
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**SUBJECT AREA AND CATEGORY**

Veterinary  
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**PUBLISHER**

Veterinary World

**H-INDEX**

**35**

**PUBLICATION TYPE**

Journals

**ISSN**

09728988, 22310916

**COVERAGE**

2008-2021

**INFORMATION**

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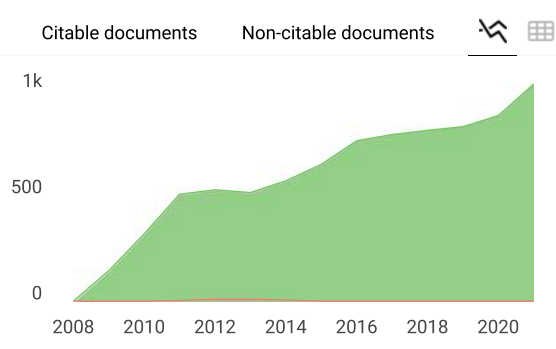
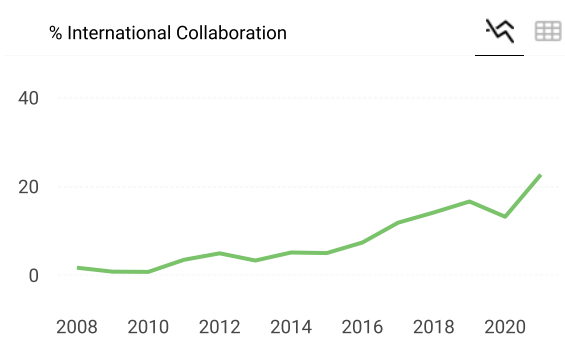
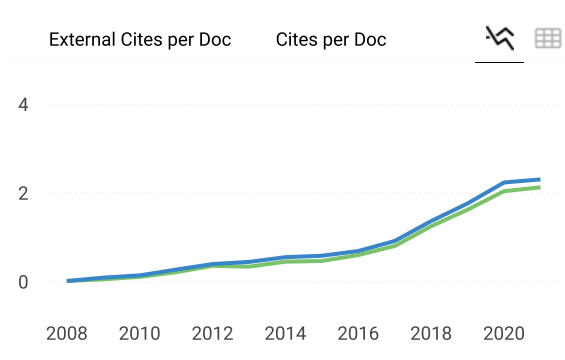
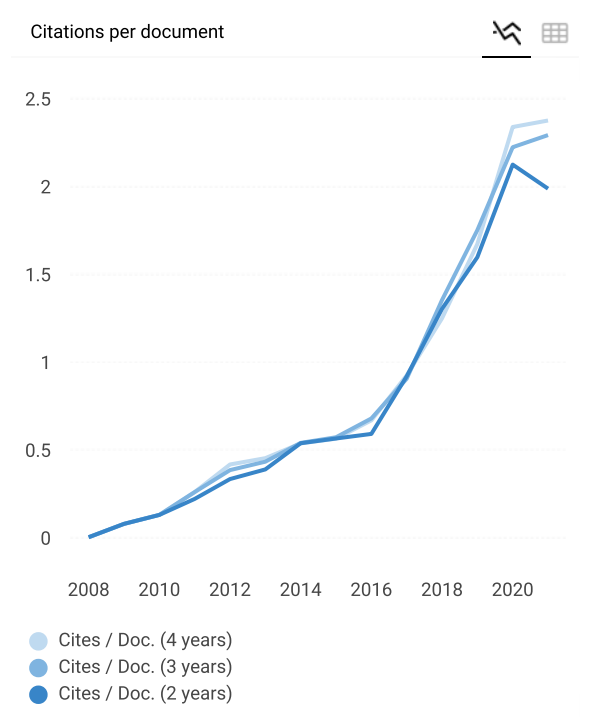
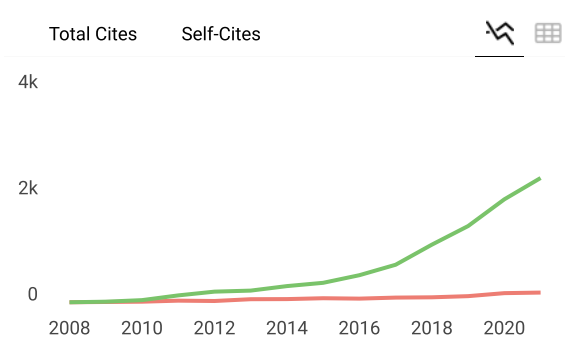
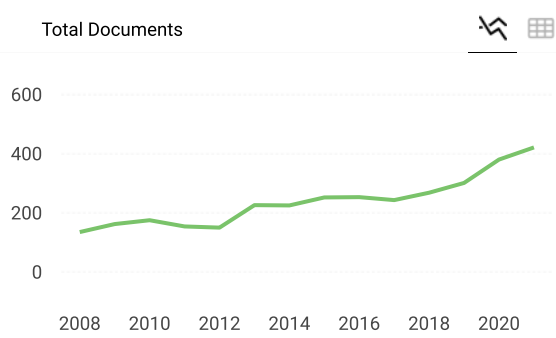
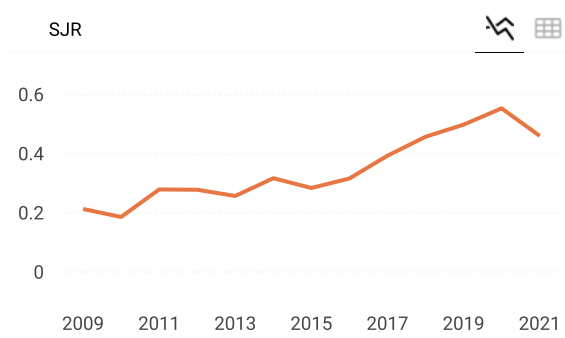
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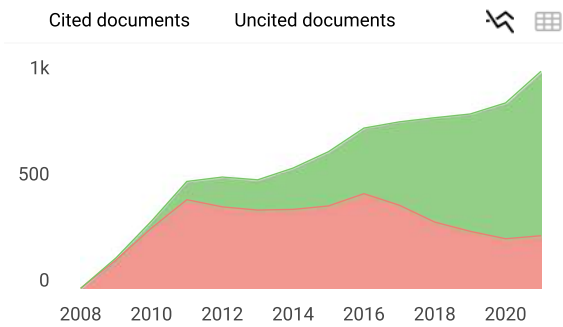
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Thankyou.

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**Melanie Ortiz** 5 months ago

Dear Otto, Thank you for contacting us.

As you probably already know, our data come from Scopus, they annually send us an update of the data. This update is sent to us around April / May every year.

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# The self-nanoemulsifying drug delivery system of *Petiveria alliacea* extract reduced the homeostatic model assessment-insulin resistance value, interleukin-6, and tumor necrosis factor- $\alpha$ level in diabetic rat models

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**Received:** 10-07-2021, **Accepted:** 17-11-2021, **Published online:** 31-12-2021

**doi:** [www.doi.org/10.14202/vetworld.2021.3229-3234](http://www.doi.org/10.14202/vetworld.2021.3229-3234) **How to cite this article:** Mustika A, Fatimah N, Sari GM (2021) The self-nanoemulsifying drug delivery system of *Petiveria alliacea* extract reduced the homeostatic model assessment-insulin resistance value, interleukin-6, and tumor necrosis factor- $\alpha$  level in diabetic rat models, *Veterinary World*, 14(12): 3229-3234.

## Abstract

**Background and Aim:** Metaflammation plays a significant role in the pathogenesis, development, and complication of diabetes mellitus (DM). This inflammation is associated with insulin resistance. Therefore, the inflammatory pathways have been targeted for pharmacological treatment. *Petiveria alliacea* can decrease blood glucose levels and has anti-inflammatory and antioxidant activities; however, there are still insufficient data regarding its efficacy for the treatment of DM. This study aimed to investigate the effect of the self-nanoemulsifying drug delivery system (SNEDDS) of *P. alliacea* leaf extract on the homeostatic model assessment (HOMA)-insulin resistance (IR) value and interleukin (IL)-6 and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) levels in a streptozotocin (STZ)-induced diabetic rat model.

**Materials and Methods:** Thirty-five diabetic rat models were randomly divided into five groups. The first group received the SNEDDS of *P. alliacea* leaf extract at a dose of 50 mg/kg body weight (BW), the second group received it at a dose of 100 mg/kg BW, the third group received it at a dose of 200 mg/kg BW, the fourth group received 18 mg of metformin, and the fifth group only received the SNEDDS formula. The treatment was administered once a day, orally, for 14 days. On the 15<sup>th</sup> day after treatment, the rats were sacrificed to obtain blood samples for cardiac examination. The IL-6, TNF- $\alpha$ , and insulin levels in the serum were measured using the enzyme-linked immunosorbent assay method. The HOMA-IR value was calculated using a formula.

**Results:** The mean IL-6 and TNF- $\alpha$  levels were low in the group that received the SNEDDS of *P. alliacea* leaf extract. There was no significant difference in the insulin level in all treatment and control groups. However, a significant difference in the HOMA-IR value was noted between the group that received the SNEDDS of *P. alliacea* leaf extract and metformin and the group that did not receive treatment ( $p < 0.05$ ).

**Conclusion:** The SNEDDS of *P. alliacea* leaf extract reduced the HOMA-IR value and suppressed the TNF- $\alpha$  and IL-6 levels in the STZ-induced diabetic rat model.

**Keywords:** diabetes, homeostatic model assessment-insulin resistance, tumor necrosis factor- $\alpha$ , interleukin-6, nanoemulsifying, *Petiveria alliacea*.

## Introduction

Diabetes mellitus (DM) is a multifactorial metabolic disorder affecting the glucose status of the human body [1,2]. There is a crucial link between metabolic disorder and inflammation, leading to metaflammation. Metaflammation is a systemic and chronic low-grade inflammation associated with excess nutrients and energy. Previous studies [3,4] have demonstrated that diabetes, both types 1 DM (T1DM) and 2 (T2DM), is an inflammatory disease. T1DM occurs

due to cell-mediated autoimmunity damage to pancreatic  $\beta$ -cells. T2DM has been regarded as a metabolic disease, so that the metabolic determinant is the main pathogenetic factor. However, more recent research has begun to focus on low-grade inflammation as a significant factor in the pathogenesis, development, and complication of T2DM [4,5]. T2DM is characterized by decreased insulin secretion and increased insulin resistance (IR), associated with chronic inflammation [6,7]. Evidence shows that the adipose tissue in diabetic patients secretes various inflammatory cytokines, such as tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interleukin (IL)-6. Increased inflammatory cytokines can predispose the liver to activate the I $\kappa$ B kinase (IKK) activity. Activated IKK inhibits the expression of insulin receptor 1 (IRS-1) substrate, resulting in an increase in IR [8]. Long-term exposure to hyperglycemia leads to an imbalance in oxidative stress, endoplasmic reticulum

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stress, hypoxia, amyloid and lipid deposition, lipotoxicity, and glucotoxicity. This process can stimulate an immune response to secrete various inflammatory mediators and is associated with systemic IR. Macrophages migrate to the adipose tissue, liver, and muscle in diabetics and obese patients. The macrophages produce pro-inflammatory factors, such as nitric oxide, reactive oxygen species, prostaglandin E<sub>2</sub>, TNF- $\alpha$ , IL-1 $\beta$ , and IL-6, which induce inflammation. They also increase complications such as atherosclerosis, impaired lung function, and cardiovascular disease [9,10]. As evidence suggests that inflammation plays a significant role in its pathogenesis, T2DM is now redefined as an immune disorder [4,11]. Therefore, the inflammatory pathways involved in T2DM have been targeted for pharmacological manipulation to suppress cytokine inflammation and combat IR.

*Petiveria alliacea* is a perennial plant widely found in Indonesia and empirically used as a folk medicine to treat various diseases, such as diabetes, bleeding cough, anti-inflammation, and immunomodulatory. Several studies revealed that *P. alliacea* has anti-inflammatory [12-15] and antioxidant activities [16]. All parts of *P. alliacea* are available for use in treatment from oxidative damage of cells [17]. Mustika *et al* [18]. showed that the extract of *P. alliacea* leaves decreased the blood glucose level in the T2DM mouse model and increased the 5'-adenosine monophosphate-activated protein kinase (AMPK) expression in the liver [16]. These studies provide hope that *P. alliacea* may be used to control the inflammation in hyperglycemic conditions, thereby reducing IR. One of the disadvantages of *P. alliacea* leaf extract is that it contains complex active compounds, which may compete in the absorption process, resulting in a decline in the absorption of active compounds in the gastrointestinal tract [19,20]. Therefore, to overcome this problem, *P. alliacea* extract formulated in the form of a self-emulsifying drug delivery system (SNEDDS) is made [21]. The evaluation of the SNEDDS formulation of *P. alliacea* leaf extract on the diabetes treatment has not yet been clearly elucidated.

Therefore, this study aimed to investigate the effect of the SNEDDS of *P. alliacea* leaf extract on the homeostatic model assessment (HOMA-IR) value and IL-6 and TNF- $\alpha$  levels in the streptozotocin (STZ)-induced diabetic rat model.

## Materials and Methods

### Ethical approval

This study was approved by the Animal Care and Use Committee of the Faculty of Veterinary Medicine, Universitas Airlangga, Indonesia (No. 2. KE-092.05.2018).

### Study period and location

This study was conducted for 5 months (May-September 2019). Male Wistar rats were reared in the Laboratory Animal of Pharmacology Department, Faculty of Medicine, Universitas Airlangga. The

SNEDDS of *P. alliacea* leaf extract preparation was carried out at the Pharmacology Laboratory, Faculty of Medicine, Universitas Airlangga. ELISA of insulin, IL-6, and TNF- $\alpha$  were performed at the Institute of Tropical Diseases, Universitas Airlangga, Surabaya.

### Plant material

*P. alliacea* leaves were collected from Balai Materia Medika, Batu, Indonesia. Subsequently, 1000 g of *P. alliacea* leaf powder was extracted with 70% ethanol solvent. The extraction was performed by maceration for 3 days. The SNEDDS formulation was formed from virgin coconut oil, Tween 80, and propylene glycol. The SNEDDS of *P. alliacea* leaf extract was made by adding *P. alliacea* leaf extract to the SNEDDS formulation [21].

### Chemical

About 70% alcohol, 10% sucrose solution, virgin coconut oil, Tween 80, and polyethylene glycol were purchased from Brataco Co., Ltd. (Indonesia). The enzyme-linked immunosorbent assay (ELISA) kits for IL-6 (catalog no.: EK306/2), TNF- $\alpha$  (catalog no.: EK382/2), and insulin were purchased from Multi Sciences. STZ and metformin were purchased from Sigma-Aldrich® (Singapore) and Kimia Farma® (Indonesia), respectively.

### Animal models

Male *Rattus norvegicus* strain Wistar rats aged 3-4 months old were used in the experiment. The rats weighed 150-200 g and were in good health. They were obtained from the Pharmacological Department, Faculty of Medicine, Universitas Airlangga.

The experimental animals were adapted to the environment for 7 days and fed *ad libitum*. The diabetic rat models were created by inducing these rats with STZ [22-25]. The dose of STZ was 50 mg/kg body weight (BW) and administered by intraperitoneal injection. The rats were given 10% sucrose solution during the first night after induction to avoid sudden hypoglycemia post-injection. Three days after induction, the rats were checked for blood glucose levels using a glucometer. The rats were declared to have diabetes if their blood glucose levels were  $\geq 200$  mg/dL and were called the diabetic rat model [18,26-28].

### Experimental treatment

Thirty-five diabetic rat models were randomly divided into five groups. The first group received the SNEDDS of *P. alliacea* leaf extract at a dose of 50 mg/kg BW, the second group received it at a dose of 100 mg/kg BW, the third group received it at a dose of 200 mg/kg BW, the fourth group received 18 mg of metformin, and the fifth group only received the SNEDDS formula.

The treatment was administered once a day, orally, for 14 days. Before and after treatment, the rats were weighed. The rats were sacrificed on the 15<sup>th</sup> day after therapy to obtain blood samples for cardiac

examination. The IL-6, TNF- $\alpha$ , and insulin levels in the serum were measured using the ELISA method.

**Cytokines and insulin examination**

Serum samples were collected from all groups and diluted according to the manufacturer’s instructions in the kit. The IL-6, TNF- $\alpha$ , and insulin levels were measured by the ELISA kit (Multi Sciences).

**HOMA-IR value**

The HOMA-IR value was calculated using the formula from Qu *et al.* [29], that is, fasting insulin level ( $\mu$ IU/mL) multiplied by the fasting blood glucose level (mmol/L)/22.5. The HOMA-IR value is used to determine IR. The HOMA-IR values were categorized as follows:  $\leq 2.60$ , normal; 2.60-3.80, borderline; and  $>3.80$ , a high correlation with IR [29-31].

**Statistical analysis**

Statistical analysis was performed using the SPSS Statistics for Windows, Version 17.0. (IBM Corp., NY, USA). The values were analyzed by one-way analysis of variance followed by the least significant difference *post hoc* multiple comparison test.  $p < 0.05$  was considered statistically significant.

**Results**

**The effect of STZ administration on the blood glucose level**

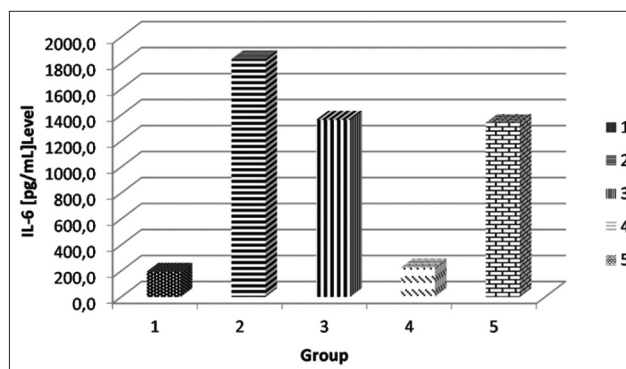
The mean blood glucose levels in rats before and 3 days after STZ administration were 104.5 and 397.2 mg/d, respectively. These data suggest that STZ administration can increase the blood glucose level in rats.

**The effect of the SNEDDS of *P. alliacea* leaf extract on the IL-6 and TNF- $\alpha$  levels in the serum diabetic rat model**

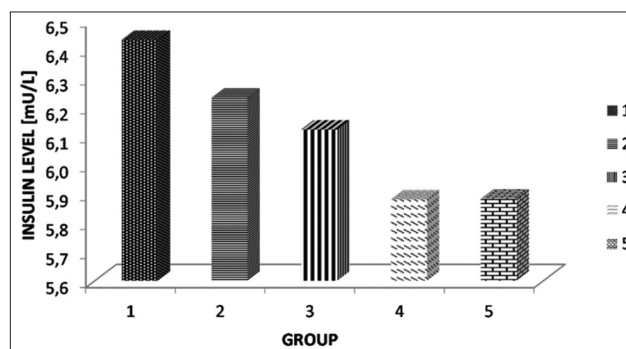
The results of the IL-6 levels in the diabetic rat serum are shown in Figure-1. The results of the TNF- $\alpha$  levels in the diabetic rat serum are shown in Table-1. The statistical analysis showed significant differences in the mean TNF- $\alpha$  level between the groups that received either the SNEDDS of *P. alliacea* leaf extract or metformin treatment and the groups that did not receive treatment ( $p < 0.05$ ). The IL-6 level also showed a significant difference between the group that received the SNEDDS of *P. alliacea* leaf extract at a dose of 50 mg/kg BW and metformin and the group that did not receive treatment ( $p < 0.05$ ).

**The effect of the SNEDDS of *P. alliacea* leaf extract on the serum insulin level and HOMA-IR in the diabetic rat model**

The mean insulin level in the serum diabetic rat model is shown in Figure-2, and the mean HOMA-IR value is shown in Figure-3. Based on the HOMA-IR value in the untreated group (6.8), it showed that induction using STZ can cause IR. In the group that received metformin therapy, the HOMA-IR value was 3.3, indicating a borderline level, while in the group that received the SNEDDS of *P. alliacea* leaf extract



**Figure-1:** The mean interleukin (IL)-6 level in the serum diabetic rat model. Group 1 received the self-nanoemulsifying drug delivery system (SNEDDS) of *Petiveria alliacea* leaf extract at a dose of 50 mg/kg body weight (BW), Group 2 received it at a dose of 100 mg/kg BW, Group 3 received it at a dose of 200 mg/kg BW, Group 4 received metformin suspension at a dose of 18 mg, and Group 5 received the SNEDDS formula.



**Figure-2:** The mean insulin level in the serum diabetic rat model. Group 1 received the self-nanoemulsifying drug delivery system (SNEDDS) of *Petiveria alliacea* leaf extract at a dose of 50 mg/kg body weight (BW), Group 2 received it at a dose of 100 mg/kg BW, Group 3 received it at a dose of 200 mg/kg BW, Group 4 received metformin suspension at a dose of 18 mg, and Group 5 received the SNEDDS formula.

**Table-1:** The mean tumor necrosis factor- $\alpha$  levels in the diabetic rat models.

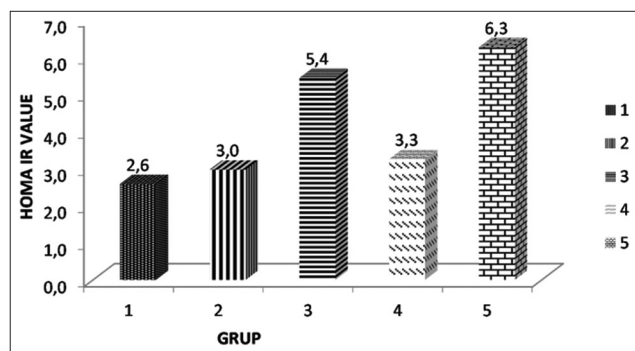
Group	Mean	Minimum	Maximum	Standard deviation
1	29.4	22.04	38.30	5.58561
2	26.9	8.72	44.96	11.97433
3	24.2	15.78	47.29	11.85768
4	26.1	14.36	44.96	10.16328
5	6268.3	35.66	13,619.88	7250.39135

Group 1 received the self-nanoemulsifying drug delivery system (SNEDDS) of *Petiveria alliacea* leaf extract at a dose of 50 mg/kg BW, Group 2 received it at a dose of 100 mg/kg BW, Group 3 received it at a dose of 200 mg/kg BW, Group 4 received metformin suspension at a dose of 18 mg, and Group 5 received the SNEDDS formula

at doses of 50 and 100 mg/kg BW, the HOMA-IR values were 2.6 and 3, indicating normal and borderline levels, respectively.

There was no significant difference in the insulin level in all treatment and control groups; however, a significant difference in the HOMA-IR value was noted between the groups that received the SNEDDS





**Figure-3:** The mean HOMA-IR value in the diabetic rat model. Group 1 received the self-nanoemulsifying drug delivery system (SNEDDS) of *Petiveria alliacea* leaf extract at a dose of 50 mg/kg body weight (BW), Group 2 received it at a dose of 100 mg/kg BW, Group 3 received it at a dose of 200 mg/kg BW, Group 4 received metformin suspension at a dose of 18 mg, and Group 5 received the SNEDDS formula.

of *P. alliacea* leaf extract and metformin and the group that did not receive treatment ( $p < 0.05$ ).

## Discussion

This study shows that the induction of STZ at the dose of 50 mg/kg BW by intraperitoneal injection can cause hyperglycemic conditions, IR (a HOMA-IR value of 6.8) and an increase in the pro-inflammatory mediators, namely, TNF- $\alpha$  and IL-6. These data support the concept of metaflammation, which is a chronic low-level inflammatory condition. This process is highly relevant for the development of diabetes and its complications. This condition contributes to the development of T2DM by increasing IR in peripheral tissues, especially in the liver, muscle, and adipose tissue, and causes damage to the pancreas, which will interfere with insulin secretion [32].

Hyperglycemic conditions can lead to inflammation. Cytokines or markers of inflammation that increase during such conditions include C-reactive protein, TNF $\alpha$ , IL-1, and IL-6 [2,10]. The inflammatory factor can decrease insulin signaling by phosphorylating IRS-1 on some serine residues, and consequently, it decreases the ability of IRS-1 to perform insulin signal transduction. IR is a major defect underlying the development of T2DM and has been defined in the last decade as a metaflammation disease [4]. TNF- $\alpha$  interferes with insulin signal transduction and affects carbohydrate metabolism in cells and tissues. The mechanisms of TNF- $\alpha$  to interfere with insulin signal transduction may be the downregulation of the IR expression and IRS-1 substrate [9,10]. The cytokine IL-6 decreases hepatic insulin sensitivity by activating the pro-inflammatory pathway and concurrently inhibiting IRS signaling [33]. This study is also in line with the research by Bayat *et al* [9]. and Sharma *et al* [31]. exhibiting that induction with STZ in *R. norvegicus* showed the same characteristics of an increased cytokine level and IR [33]. Haidari *et al.* [33] showed that the increased levels of TNF and IL-6 are closely related

to the occurrence of IR. This study shows a relationship between hyperglycemia, inflammation, and IR. Hyperglycemia can cause inflammation, inflammatory conditions will cause IR, and IR conditions will increase blood glucose levels. This circle will worsen the condition of DM and cause complications.

This study demonstrated that the SNEDDS of *P. alliacea* leaf extracts at a dose between 50 and 200 mg/kg BW can significantly reduce the IL-6 and TNF- $\alpha$  levels in the diabetic rat models. This study also showed that the insulin levels in the group of rats that received SNEDDS of *P. alliacea* leaf extracts are higher than those in the control group. This result is in accordance with the HOMA-IR value, where the group that received the SNEDDS of *P. alliacea* leaf extracts has a normal HOMA-IR value. The data showed that the SNEDDS of *P. alliacea* leaf extracts can increase insulin sensitivity in a diabetic rat model.

Based on the results of this study, the mechanism of action of the SNEDDS of *P. alliacea* leaf extract in increasing insulin sensitivity is presumably because the extract can reduce the IL-6 and TNF- $\alpha$  levels. TNF- $\alpha$  inhibits insulin signaling by increasing serine/threonine phosphorylation of IRS-1. IL-6 inhibits insulin signaling by decreasing tyrosine phosphorylation [8]. In addition, the decreased level of inflammatory cytokines causes a decrease in the activation of c-Jun N-terminal kinases (JNKs) so that there is no inhibition of IRS-1. JNKs are members of the mitogen-activated protein kinase superfamily and one of the most studied signal transducers in IR. The JNK signaling pathway can be activated by various factors, one of which is cytokines. JNK activation negatively affects insulin signaling pathways leading to IR. A decreased JNK activity also reduces the activation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B) signaling pathway, causing the decrease in the transcription of inflammatory cytokines [34-36].

The mechanisms of the SNEDDS of *P. alliacea* leaf extracts to reduce the IL-6 and TNF- $\alpha$  levels require further study. However, our previous research on *P. alliacea* leaf extracts proved that the extract could increase the AMPK- $\alpha$ 1 expression in the liver of the diabetic rat models [18]. Therefore, the possible mechanism of action of the SNEDDS of *P. alliacea* leaf extracts is the reduction in the IL-6 and TNF- $\alpha$  levels through AMPK activation. AMPK is a promising target for DM treatment. When AMPK is activated, it will affect various metabolic processes, such as glucose metabolism, lipid metabolism, and mitochondrial metabolism, and it plays a significant role in inflammation, autophagy, and apoptosis [32,37].

The activation of the AMPK pathway decreases the function of the NF- $\kappa$ B system, whereas the NF- $\kappa$ B signaling pathway is a major pathway involved in the activation of both the innate and adaptive immune systems [32,35,38]. The process of inhibiting the NF- $\kappa$ B signaling pathway will inhibit the transcription of inflammatory genes so that the synthesis of IL-6 and

TNF- $\alpha$  cytokines decreases [25,32,34]. Moreover, the AMPK activation inhibits the monocyte chemoattractant protein 1 (MCP-1) secretion [39].

The NF- $\kappa$ B signaling pathway also regulates IL-8, chemokines, and adhesion molecules, such as MCP-1, intercellular adhesion molecule 1 (ICAM-1), and vascular cell adhesion protein 1 (VCAM-1) [40,41]. Several studies have shown that an increase in the serum MCP-1 level is correlated with IR. MCP-1 also initiates the migration and proliferation of ICAM-1 and VCAM-1 [39,40]. These mediators play a pivotal role in the development of the vascular complication of diabetes [41]. Exploration of other pro-inflammatory cytokines and chemokines will be intriguing, but this step could not be performed in this study due to our limitations. Therefore, further research is needed to determine the role and mechanism of *P. alliacea* as therapy in DM.

A drug with a pleiotropic effect is the current approach to diabetes treatment. *P. alliacea* is a medicinal plant with various properties, such as lowering blood glucose levels, increasing AMPK expression, and reducing IR and pro-inflammatory cytokine levels. These data indicate that *P. alliacea* has potential as a therapy in DM because it has a pleiotropic effect.

## Conclusion

Our study showed that the SNEDDS of *P. alliacea* leaf extract reduced the HOMA-IR value and suppressed the TNF- $\alpha$  and IL-6 levels in the STZ-induced diabetic rat model.

## Authors' Contributions

AM: Conceptualized the study, conducted the experiment and laboratory examination, analyzed the data, and wrote and revised the manuscript. NF: Prepared the SNEDDS of *P. alliacea* leaf extract, analyzed the data, and wrote the manuscript. GMS: Analyzed the data and edited the manuscript. All authors read and approved the final manuscript.

## Acknowledgments

The authors are thankful to the Directorate of Research and Community Services, the Ministry of Research Technology and Higher Education in Indonesia for the financial support (Grant no. 4/E1/KP.PTNBH/2019).

## Competing Interests

The authors declare that they have no competing interests.

## Publisher's Note

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