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## Table of contents

## Volume 441

2020

Previous issue
 Next issue

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Accepted papers received: 16 January 2020 Published online: 24 February 2020

View all abstracts

Preface			
OPEN ACCESS			011001
Preface			
➡ View abstract	View article	PDF	
OPEN ACCESS			011002
Peer review state	ment		
	View article	🔁 PDF	

## **Papers**

## **Earth and Environmental Science**

OPEN ACCESS			012001	
The grow-out of abalone ( <i>Haliotis squamata</i> ) at different shelter shape on growth and survival and its marine environmental influences at Lembongan Bay coastal waters				
I Ardi, E Setiadi, Ra	sidi and W S Pranowo			
	Tiew article	🔁 PDF		
OPEN ACCESS			012002	
Stocking density of	of Rotifera, <i>Hexarth</i>	<i>a mira</i> on water quality and product	ion	

020	IOP Conference S	Series: Earth and Environmental Science, Volume 441, 2020 - IOPscience	
I Ardi, S Cahyanings	sih and E Setiadi		
	View article	PDF	
OPEN ACCESS			012
	••••	enan flour to the level of gel strength and dfin bream fish ( <i>Nemipterus nematophorus</i> ) surimi	
D M Astutik, L Sulm	artiwi, E Saputra and	d D Y Pujiastuti	
	View article	PDF	
OPEN ACCESS			012
•	ction of a male offs ce bran suspensio	spring in <i>Moina macrocopa</i> culture through n concentration	
A S Mubarak, D Jus	adi, M Zairin and M	A Suprayudi	
	View article	PDF	
OPEN ACCESS			012
Autonomy Era: Ex		nent of Coastal Communities in the Regional munity Assistance to Obtain the Right to Manage Sea Ibok.	
A Wahyono and M I	lliyani		
	View article	🔁 PDF	
OPEN ACCESS			012
Selective breeding		u and Kunti tilapia ( <i>Oreochromis niloticus</i> ) ti, Klaten-Central Java	012
S H Samara, A W Fa	athurrozi and Sutarn	0	
	View article	🔁 PDF	
OPEN ACCESS			012
Potential of Caule	erpa racemosa extra	acts as sunscreen creams	
E B Ersalina, A A Ab	dillah and L Sulmart	iwi	
	Tiew article	🔁 PDF	
OPEN ACCESS			012
Histopathology of protozoan ectopa	•	Shrimp ( <i>Litopenaeus vannamei</i> ) infested by	_
S R Pribadi, P D W S	Sari and S Subekti		
	View article	PDF	
OPEN ACCESS			012
The effect of iron provide the value	oowder as oxygen a	bsorber active packaging on fish oil total oxidation	
	riastuti and A A Abdi	llah	

	View article	🔁 PDF	
OPEN ACCESS			012
	•	inth on Threadfin Bream ( <i>Nemipterus japonicus</i> ) n, Probolinggo, East Java	
D S Octatriana, P D	W Sari and G Mahas	sri	
	Tiew article	PDF	
OPEN ACCESS			012
•	•	shness based on the handling method in Puncak nung traditional market, Surabaya	
M Sari, J Triastuti, H	I Pramono and Suda	rno	
	View article	🔁 PDF	
OPEN ACCESS			012
	-	y and minimum bactericidal concentration of es extract against <i>Vibrio harveyi</i>	
A Kharisma, W Tjah	janingsih and Setiav	vati Sigit	
	View article	🔁 PDF	
OPEN ACCESS			012
The larasati tilapia	a ( <i>Oreochromis nild</i>	<i>pticus</i> ) fingerling rearing activity in PBIATJanti,	
	•	e through survival rate	
RV Prasetya, Sutarr	no and M B Santanur	nurti	
	View article	🔁 PDF	
OPEN ACCESS			012
Use of hydrogen p	eroxide to improve	potential redox land preparation of land towards	
	tion of traditional s	shrimp vanname ( <i>Litopeaneus vanname</i> ) in Wringin P	utih,
D D Nindarwi, L A S	ari, P D Wulansari, S	H Samara and M B Santanumurti	
	Tiew article	🔁 PDF	
OPEN ACCESS			012
The effect of feed ( <i>Carassius auratu</i>		sp. and high pellet protein to seedling goldfish	
K H Dwiardani, L A	Sari, P. D. W. Sari, D.	D. Nindarwi and S. Arsad	
	Tiew article	PDF	
OPEN ACCESS			012
	-	cy ( <i>Hermetia illucens</i> ) in artificial feed formulation In catfish ( <i>Clarias gariepinus</i> )	
M S Islam, Aguston			

25/2/2020

	View article	PDF	
OPEN ACCESS			012017
retentions of sang	gkuriang catfish ( <i>Cl</i>		
F Azizah, M Arief ar	nd W P Lokapirnasari	_	
✤ View abstract	View article	PDF	
OPEN ACCESS			012018
•	in content, and fatt ent lysine doses in c	y acid of catfish meat ( <i>pangasius</i> sp.) With the commercial feed	
E Aristasari, R A Nu	ır 'Aini, W Nopita, Agu	istono, M Lamid and M A Al-Arif	
✤ View abstract	View article	🔁 PDF	
OPEN ACCESS			012019
(Epinephelus fuso	coguttatus × lanced	performance, and survival rate of cantang grouper platus) in concrete pond at Situbondo, East Java, Indor	nesia
N S Anita and N N I			
➡ View abstract	View article	PDF	
OPEN ACCESS			012020
The effect of temp Aeromonas hydro	· •	d antimicrobial agent on growth and viability of	
M F Ulkhaq, D S Bu	idi and N N Rahayu		
	View article	PDF	
OPEN ACCESS			012021
Utilization of agar L R Dita, Sudarno a	•	natural thickener on liquid bath soap formulation	
➡ View abstract	View article	🔁 PDF	
OPEN ACCESS			012022
		and Protozoan Ectoparasite Infestation Level Of <i>ivelis</i> ) Nursery In UPBL Situbondo, East Java	
G Mahasri, I N D Yo	odharta, D Novalisa a	nd A T Mukti	
	Tiew article	🔁 PDF	
OPEN ACCESS			012023
		um (Cd) in various sizes of blood shells n Bangkalan, Madura	
E S Ulfah, B S Raha	ardja and K T Pursety	0	
	Tiew article	🔁 PDF	

OPEN ACCESS	012024
Identification and prevalence of ectoparasites on the fry of Asian sea bass (Lates	
calcalifer), white shrimp (Litopenaeus vannamei), and blue swimming crab (Portunus pela	agicus)
Budianto, H Suprastyani, Q A'yunin and Z Nadlifah	
OPEN ACCESS	012025
Effect of dense stocking of <i>Gracilaria sp</i> on growth and survival of milkfish ( <i>Chanos chanos forskal</i> ) on polyculture culture systems	
W Isroni, A S Bahri and A A Amin	
OPEN ACCESS	012026
The effect of <i>Dunaliella salina</i> extract on NFkB expression in Cantang Grouper	
(Epinephelus fuscoguttatus x E. lanceolatus) exposed by Viral Nervous Necrosis	
Rani Yuwanita, A Yuniarti, SSP Rahardjo, Q Ayu'nin and AM Madyaratri	
+ View abstract 🔄 View article р PDF	
OPEN ACCESS	012027
The Effectiveness of Vaccines in Gurame ( <i>Osphronemus goramy</i> ) and Challenged <i>Aeromonas hydrophila</i>	
S U Setyaningsih, R Kusdarwati, Rozi and D Handijatno	
OPEN ACCESS	012028
Utilization of By Product <i>Kappaphycus alvarezii</i> as Earthquake Resistant Material Lightweight Concrete	
A A Musthofa, M Z A Bahtiar, F M Ibrahim and A A Abdillah	
+ View abstract 🔲 View article р PDF	
OPEN ACCESS	012029
The effect of density as <i>Skeletonema costatum</i> bioremediation agent of copper (Cu) heavy metal concentration	
N A Pratama, B S Rahardja and L A Sari	
+ View abstract 🔄 View article 🏷 PDF	
OPEN ACCESS	012030
Effectiveness of Heat Shock (40°C) With Different Duration for Tetraploid Formed in Mutiara Catfish ( <i>Clarias sp</i> .) Juvenile	
S Nuswantoro, MS Widodo, F Fariedah and E Artarini	
+ View abstract 🔄 View article 🏷 PDF	
OPEN ACCESS	012031

The use of bromelain enzyme on artificial hatching media as an effort to hatch Nile         tilapia ( <i>Oreochromis niloticus</i> ) eggs outside the mother's mouth         F Fariedah, M S Widodo, S Nuswantoro and Sholikhin         + View abstract       IView article         OPEN ACCESS       01203         Effects of different feed doses of Majapahit leaves ( <i>Crescentia cujete</i> L.) on the growth of Nile tilapia ( <i>Oreochromis niloticus</i> )       01203         S Rahmaningsih, Jumiati and S Awwaliyah       + View abstract       IView article       PDF         OPEN ACCESS       01203         Vaname shrimp ( <i>Litopenaeus vannamel</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       01203         M S A Ningsih, Prayogo and A M Sahidu       + View abstract       Iview article       PDF         OPEN ACCESS       01203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       01203         Effect of probiotic duration and M Lamid       + View abstract       Iview article       PDF	)20	IOP Conference S	eries: Earth and Environmental Science, Volume 441, 2020 - IOPscienc	e
+ View abstract       Image: View article       PDF         OPEN ACCESS       01203         The use of bromelain enzyme on artificial hatching media as an effort to hatch Nile       11203         Itapia ( <i>Oreochromis niloticus</i> ) eggs outside the mother's mouth       F Fariedah, M S Widodo, S Nuswantoro and Sholikhin         + View abstract       Image: View article       PDF         OPEN ACCESS       01203         Effects of different feed doses of Majapahit leaves ( <i>Crescentia cujete</i> L.) on the growth of Nile tilapia ( <i>Oreochromis niloticus</i> )       01203         S Rahmaningsih, Jumiati and S Awwaliyah       + View abstract       Image: View article       01203         PDFN ACCESS       01203         Vaname shrimp ( <i>Litopenaeus vanname</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       01203         Vaname shrimp ( <i>Litopenaeus vanname</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       01203         M S A Ningsih, Prayogo and A M Sahidu       + View abstract       Image: View article       01203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       01203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       01203		analysis of <i>Pangas</i>	<i>sius</i> sp. infected by <i>Edwardsiella tarda</i> causes Edward	dsiellosis
OPEN ACCESS       01203         The use of bromelain enzyme on artificial hatching media as an effort to hatch Nile tilapia ( <i>Oreochromis niloticus</i> ) eggs outside the mother's mouth       1         F Fariedah, M S Widodo, S Nuswantoro and Sholikhin       +         + View abstract       Image: View article       PDF         OPEN ACCESS       01203         Effects of different feed doses of Majapahit leaves ( <i>Crescentia cujete</i> L.) on the growth of Nile tilapia ( <i>Oreochromis niloticus</i> )       01203         S Rahmaningsih, Jumiati and S Awwaliyah       +       View atricle       PDF         OPEN ACCESS       01203         Vaname shrimp ( <i>Litopenaeus vannamel</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       01203         Vaname shrimp ( <i>Litopenaeus vannamel</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       01203         Va N S A Ningsih, Prayogo and A M Sahidu       +       View abstract       Image: View article       PDF         OPEN ACCESS       01203       Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       01203         N Fatmawati, Agustono and M Lamid       +       View abstract       Image: View article       PDF         OPEN ACCESS       01203       The effect of depuration on	Q A'yunin, Budianto,	S Andayani and R Y	uwanita	
OTENS       OTENS         OPEN ACCESS       O1203         S A Ningsih, Prayogo and A M Sahidu       * View article       * PDF         OPEN ACCESS       O1203         Effects of different feed doses of Majapahit leaves ( <i>Crescentia cujete</i> L.) on the growth of Nile tilapia ( <i>Oreochromis niloticus</i> )       S Rahmaningsih, Jumiati and S Awwaliyah         + View abstract       Image: View article       Image: PDF         OPEN ACCESS       O1203         S Rahmaningsih, Jumiati and S Awwaliyah       * View abstract       Image: PDF         OPEN ACCESS       O1203         Vaname shrimp ( <i>Litopenaeus vannamel</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       O1203         M S A Ningsih, Prayogo and A M Sahidu       * View abstract       Image: PDF         OPEN ACCESS       O1203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       O1203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       O1203         Defen Access       O1203       O1203         The effect of depuration on lead levels of the cockles Anadara sp. by using activated carbon as a filter       O1203         The effect of depuration on lead leve		Uiew article	🔁 PDF	
tilapia ( <i>Oreochromis niloticus</i> ) eggs outside the mother's mouth F Fariedah, M S Widodo, S Nuswantoro and Sholikhin + View abstract  View article  PDF OPEN ACCESS O1203 Effects of different feed doses of Majapahit leaves ( <i>Crescentia cujete</i> L.) on the growth of Nile tilapia ( <i>Oreochromis niloticus</i> ) S Rahmaningsih, Jumiati and S Awwaliyah + View abstract  View article  PDF OPEN ACCESS O1203 Vaname shrimp ( <i>Litopenaeus vannamel</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia M S A Ningsih, Prayogo and A M Sahidu + View abstract  View article  PDF OPEN ACCESS O1203 Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient N Fatmawati, Agustono and M Lamid + View abstract  View article  PDF OPEN ACCESS O1203 The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter A R Firdaus, A S Mubarak and W Tjahjaningsih	OPEN ACCESS			012032
<ul> <li>View abstract View article PDF</li> <li>OPEN ACCESS 01203</li> <li>Effects of different feed doses of Majapahit leaves (<i>Crescentia cujete</i> L.) on the growth of Nile tilapia (<i>Oreochromis niloticus</i>)</li> <li>S Rahmaningsih, Jumiati and S Awwaliyah</li> <li>View abstract View article PDF</li> <li>OPEN ACCESS 01203</li> <li>Vaname shrimp (<i>Litopenaeus vannamel</i>) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia</li> <li>M S A Ningsih, Prayogo and A M Sahidu</li> <li>View abstract View article PDF</li> <li>OPEN ACCESS 01203</li> <li>Effect of probiotic duration and dose of coffee peel fermentation (<i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient</li> <li>N Fatmawati, Agustono and M Lamid</li> <li>View abstract View article PDF</li> <li>OPEN ACCESS 01203</li> <li>The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter</li> <li>A R Firdaus, A S Mubarak and W Tjahjaningsih</li> </ul>		•		
OPEN ACCESS       01203         Effects of different feed doses of Majapahit leaves ( <i>Crescentia cujete</i> L.) on the growth of Nile tilapia ( <i>Oreochromis niloticus</i> )       01203         S Rahmaningsih, Jumiati and S Awwaliyah       +         + View abstract       Image: View article       PDF         OPEN ACCESS       01203         Vaname shrimp ( <i>Litopenaeus vannamel</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       01203         M S A Ningsih, Prayogo and A M Sahidu       +       View article       PDF         OPEN ACCESS       01203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       N Fatmawati, Agustono and M Lamid         + View abstract       Image: View article       PDF         OPEN ACCESS       01203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       N Fatmawati, Agustono and M Lamid         + View abstract       Image: View article       PDF       01203         OPEN ACCESS       01203       01203         The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter       01203         A R Firdaus, A S Mubarak and W Tjahjaningsih       01203	F Fariedah, M S Wido	odo, S Nuswantoro a	and Sholikhin	
OTEOS         OTEOS <t< td=""><td>➡ View abstract</td><td>View article</td><td>PDF</td><td></td></t<>	➡ View abstract	View article	PDF	
of Nile tilapia ( <i>Oreochromis niloticus</i> ) S Rahmaningsih, Jumiati and S Awwaliyah + View abstract IView article PDF OPEN ACCESS 01203 Vaname shrimp ( <i>Litopenaeus vannamel</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia M S A Ningsih, Prayogo and A M Sahidu + View abstract IVIEW article PDF OPEN ACCESS 01203 Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient N Fatmawati, Agustono and M Lamid + View abstract IVIEW article PDF OPEN ACCESS 01203 Effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter A R Firdaus, A S Mubarak and W Tjahjaningsih	OPEN ACCESS			012033
<ul> <li>+ View abstract  View article  PDF</li> <li>OPEN ACCESS 01203</li> <li>Vaname shrimp (<i>Litopenaeus vannamel</i>) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia</li> <li>M S A Ningsih, Prayogo and A M Sahidu</li> <li>+ View abstract  View article  PDF</li> <li>OPEN ACCESS 01203</li> <li>Effect of probiotic duration and dose of coffee peel fermentation (<i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient</li> <li>N Fatmawati, Agustono and M Lamid</li> <li>+ View abstract  View article  PDF</li> <li>OPEN ACCESS 01203</li> <li>Effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter</li> <li>A R Firdaus, A S Mubarak and W Tjahjaningsih</li> </ul>		-		
OPEN ACCESS       01203         Vaname shrimp ( <i>Litopenaeus vannamei</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia       M S A Ningsih, Prayogo and A M Sahidu         + View abstract       Image: View article       PDF         OPEN ACCESS       01203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       01203         N Fatmawati, Agustono and M Lamid       Image: View article       PDF         OPEN ACCESS       01203         Diffect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter       01203         A R Firdaus, A S Mubarak and W Tjahjaningsih       Image: View and V	S Rahmaningsih, Jun	niati and S Awwaliya	ah	
Vaname shrimp ( <i>Litopenaeus vannamei</i> ) post-harvest marketing analysis in traditional pond systems at Turi District, Lamongan, East Java, Indonesia M S A Ningsih, Prayogo and A M Sahidu + View abstract IVI View article PDF 0PEN ACCESS 01203 Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient N Fatmawati, Agustono and M Lamid + View abstract IVI View article PDF 0PEN ACCESS 01203 The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter A R Firdaus, A S Mubarak and W Tjahjaningsih	➡ View abstract	View article	🔁 PDF	
pond systems at Turi District, Lamongan, East Java, Indonesia   M S A Ningsih, Prayogo and A M Sahidu   + View abstract   Image: Note of the the text of the text of the text of tex	OPEN ACCESS			012034
<ul> <li>+ View abstract  View article  PDF</li> <li>OPEN ACCESS 01203</li> <li>Effect of probiotic duration and dose of coffee peel fermentation (<i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient</li> <li>N Fatmawati, Agustono and M Lamid</li> <li>+ View abstract  View article  PDF</li> <li>OPEN ACCESS 01203</li> <li>The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter</li> <li>A R Firdaus, A S Mubarak and W Tjahjaningsih</li> </ul>		-		
OPEN ACCESS       01203         Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient       01203         N Fatmawati, Agustono and M Lamid       •         • View abstract       •         • View abstract       •         • OPEN ACCESS       01203         The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter       01203         A R Firdaus, A S Mubarak and W Tjahjaningsih       •	M S A Ningsih, Prayo	go and A M Sahidu		
Effect of probiotic duration and dose of coffee peel fermentation ( <i>Coffea</i> sp.) on crude protein and crude fiber as an alternative fish feed ingredient N Fatmawati, Agustono and M Lamid + View abstract IVIEW article PDF OPEN ACCESS 01203 The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter A R Firdaus, A S Mubarak and W Tjahjaningsih		View article	🔁 PDF	
protein and crude fiber as an alternative fish feed ingredient          N Fatmawati, Agustono and M Lamid         + View abstract       Image: View article         OPEN ACCESS       01203         The effect of depuration on lead levels of the cockles Anadara sp. by using activated carbon as a filter       01203         A R Firdaus, A S Mubarak and W Tjahjaningsih       Image: Market depuice de	OPEN ACCESS			012035
<ul> <li>View abstract  View article  PDF</li> <li>OPEN ACCESS 01203</li> <li>The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter</li> <li>A R Firdaus, A S Mubarak and W Tjahjaningsih</li> </ul>	•			
OPEN ACCESS The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter A R Firdaus, A S Mubarak and W Tjahjaningsih	N Fatmawati, Agusto	no and M Lamid		
The effect of depuration on lead levels of the cockles <i>Anadara</i> sp. by using activated carbon as a filter A R Firdaus, A S Mubarak and W Tjahjaningsih	➡ View abstract	View article	🔁 PDF	
carbon as a filter A R Firdaus, A S Mubarak and W Tjahjaningsih	OPEN ACCESS			012036
	•	ation on lead level	s of the cockles <i>Anadara</i> sp. by using activated	
+ View abstract 💿 View article 🏷 PDF	A R Firdaus, A S Mub	arak and W Tjahjan	ingsih	
		View article	🔁 PDF	
OPEN ACCESS 01203	OPEN ACCESS			012037
Growth performance and survival rate of Boeseman's rainbowfish ( <i>Melanotaenia boesemani</i> ) in natural spawning technique at Depok, West Java, Indonesia	•		-	
W M Akhsan, B Nur and N N Dewi			ווויקטט מו טפאטא, איפטו שמים, וווטטוופטמ	
		_	🔁 PDF	
OPEN ACCESS 01203	OPEN ACCESS			012038

20		Series: Earth and Environmental Science, Volume 441, 2020 - IOPscience	
	guppies ( <i>Poecilia</i> d of pregnant fema	<i>reticulata</i> ) using water of coconut hybrid variety with t le	he
G Meiliana, L Sulma	rtiwi and L Lutfiyah		
➡ View abstract	View article	PDF	
OPEN ACCESS			012039
The effect of comn on laboratory and		increase the population of Skeletonema costatum	
K A Azmi, S Arsad aı	nd L A Sari		
➡ View abstract	View article	PDF	
OPEN ACCESS			012040
center for brackish	n water aquaculture	<i>nos</i> modular system and semi intensive at the e (BBPBAP) Jepara, Central of Java	
M Nisa and K Kismi			
View abstract	View article	PDF	
OPEN ACCESS			012041
Effect of <i>Spirulina</i> rasbora ( <i>Rasbora</i> )		entation in the diet to sperm performance of silver	
M W D Putri, Prayog	o and D S Budi		
➡ View abstract	View article	PDF	
OPEN ACCESS			012042
The variances of he and challenged by	0. 0	mi ( <i>Osphronemus gouramy</i> ) which is vaccinated ophila	
D Alfaniah, R Kusda	rwati, Rozi and D Ha	andijatno	
➡ View abstract	View article	PDF	
OPEN ACCESS			012043
	ity test of mahkota <i>as hydrophilla</i> by ii	dewa leaf extract ( <i>Phaleri amacrocarpa</i> ) against n vitro	
R A Sarendah, Suda	rno and R Kusdarwa	ati	
➡ View abstract	View article	PDF	
OPEN ACCESS			012044
The effect of kerse salmonicida smith		bura L) leaf extract on bacteria Aeromonas	
N Kartika, Sudarno	and D Handijatno		
	View article	PDF	

		Series: Earth and Environmental Science, Volume 441, 2020 - IOPscience	
	al rate of silver bark addition in fish feed	b, <i>Rasbora argyrotaenia</i> under different concentrations d	of
S A Dewi, A S Muba	arak and A T Mukti		
	View article	PDF	
OPEN ACCESS			01204
	rd specific growth ra	uice dregs on catfish ( <i>Pangasius pangasius</i> ) feed ate, efficiency of feed, feed conversion ratio, digestibilit	y of
Z N Arifiina, A P Anj	arwati, M Lamid and	Agustono	
	View article	PDF	
OPEN ACCESS			01204
Addition of lemur <i>Rasbora argyrota</i>		the gonadal maturity level of female silver barb,	
L Agustin, L Sulmar	rtiwi and A S Mubarak	ς	
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The use of mangre		<i>cenia rumphiana</i> as antioxidant feed additive in I survival rate of Nile tilapia fry <i>Oreochromis niloticus</i>	01204
The use of mangro commercial feed		I survival rate of Nile tilapia fry <i>Oreochromis niloticus</i>	01204
The use of mangro commercial feed D Wulansari, L Sulr	towards growth and	I survival rate of Nile tilapia fry <i>Oreochromis niloticus</i>	01204
The use of mangro commercial feed D Wulansari, L Sulr	towards growth and nartiwi and M A Alam	I survival rate of Nile tilapia fry <i>Oreochromis niloticus</i> sjah	
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D D Nindarwi, W P Lokapirnasari, M A Al-Arif, M A Alamsjah and M Lamid

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Nile tilapia ( <i>Oreochromis niloticus</i> ) fish hatchery technique: the survival rate evaluat in IBAT Pandaan, Pasuruan, East Java	tion
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The effect of concrete tanks for the breeding technique of the sand sea cucumber ( <i>Holothuria scabra</i> )	
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<i>Neobenedenia girellae</i> infestation on cobia ( <i>Rachycentron canadum</i> ) in Hurun Bay Lampung, Indonesia	
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OPEN ACCESS	012059
Ecotourism development through legality of mangrove processed products dan river tracing in Cemara Beach, Banyuwangi, East Java, Indonesia	r

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The effect of various concentration of quail egg yolk on s fish ( <i>Tor soro</i> Valenciennes, 1842) post cryopreservation	
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OPEN ACCESS	012061
The fertilization of <i>Tor soro</i> fish (Valenciennes, 1842) usi the effect of skim milk as a cryoprotectant	ng post cryopreservation sperm:
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OPEN ACCESS	012062
Honey effect on sperm motility of kancra fish ( <i>Tor soro</i> Vahours freezing	alenciennes, 1842) after 48
B S D Putri, Abinawanto, O Z Arifin and A H Kristanto	
OPEN ACCESS	012063
The spermatozoa viability of kancra fish ( <i>Tor soro</i> , Valence freezing: effect of brown sugar as natural cryoprotectant	iennes 1842) 48-hour after
M A B Pamungkas, Abinawanto, O Z Arifin and A H Kristanto	
OPEN ACCESS	012064
Feed additive of curcuma flour ( <i>Curcuma xanthorhiza</i> ) in and feed efficiency of tambaqui ( <i>Colossoma macropom</i>	_
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The spermatozoa motility of kancra fish ( <i>Tor soro</i> Valenci process: the application of egg yolk as a cryoprotectant	ennes, 1842) after the frozen
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OPEN ACCESS	012066
Sperm motility of kancra fish ( <i>Tor soro</i> , Valenciennes 18 soybean milk as a natural cryoprotectant	42) after frozen: the effect of
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OPEN ACCESS			012069
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	View article	PDF	
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Degree of nemato from Banyuwangi		nfection in asian swamp eel ( <i>Monopterus albus</i> )	
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	View article	PDF	
OPEN ACCESS			012072
Effect of different ( <i>Oreochromis nild</i>	-	probiotics on the growth performance of Nile Tilapia	
T D Sholihuddin, M	Arief and H Kenconc	ojati	
	View article	PDF	
OPEN ACCESS			012073
-	of <i>Rhadinorhynchi</i> ating net cage in S	<i>us bicircumspinis</i> in barramundi ( <i>Lates calcarifer</i> ) itubondo waters.	
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OPEN ACCESS			012074
Population growth beaches, Indones	•	of the <i>Lorjuk</i> shellfish ( <i>Solen</i> sp.) on Pamekasan	
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OPEN ACCESS			012075
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Morphology, morp ( <i>Rasbora argyrota</i>		me qualitative parameters of silver rasbora	
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OPEN ACCESS			012077
Inventory of ectop cultivated with hig	•	white shrimp ( <i>Litopenaeus vannamei</i> ) that	
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	View article	PDF	
OPEN ACCESS			012078
<i>lanceolatus</i> ) With	Different Feeding F	uvenile ( <i>Epinephelus fuscoguttatus</i> x <i>Epinephelus</i> Frequency	
D Nuraini, Agustono	and L Lutfiyah		
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OPEN ACCESS			012079
The effect of fucox	anthin as coloring	agent on the quality of Shrimp Paste	
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The effect of fucox	anthin as coloring	agent on the quality of catfish sausage	
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OPEN ACCESS			012082
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	Tiew article	PDF	
OPEN ACCESS			012083
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A F Syah and M Sho	oleh		
	View article	🔁 PDF	
OPEN ACCESS			012084
Evaluation of aqu	eous extract of rob	usta coffee ( <i>Coffea canephora</i> ) leaves for	
controlling Argulu	<i>s japonicus</i> infesta	tion on common carp seed	
N Afifah, Kismiyati a	and H Kenconojati		
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OPEN ACCESS			012085
		uration and initial period on hatching rate,	lonioo
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OPEN ACCESS			012086
Addition different Koi fish ( <i>Cyprinus</i>	0 ()	our to artificial feed on color quality and growth of	
D A Sudirman, M A	rief and A H Fasya		
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OPEN ACCESS			012087
Tuna fisheries in f	isheries managem	ent area Republic of Indonesia 572	
N Suyasa, P Raharo	djo, D R Putri and A V	Vidagdo	
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OPEN ACCESS			012088
Optimization of sp	permatozoa cryopre	eservation of Albino pangasius catfish:	
cryoprotectants w	rith various concen	trations and different equilibration times	
U Hasanah, Abinaw	anto, A Alimuddin ar	nd A Boediono	

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OPEN ACCESS			012089
density, case stud	y; Mangrove Cente	ediment (nitrate, phosphate) and <i>Avicennia sp</i> r Bengkak, Banyuwangi Regency, East Java Kenconojati and M F Ulkhaq	
	View article	PDF	
OPEN ACCESS			012090
The optimal n/p ra <i>vulgaris</i>	atio of shrimp cultu	re waste liquid fertilizer on growth of <i>Chlorella</i>	
P A Hidayati, A S Mu	ubarak and Sudarno		
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OPEN ACCESS			012091
Profile of Keranda Kalimantan	ng Fish ( <i>Channa pl</i>	<i>europhthalmus</i> Blkr) Proteins from Central	
Aryani, E Suprayitno	, B B Sasmito and H	ardoko	
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Screening of antiv as probiotic candi		tic acid bacteria originated from aquatic animals	
M Amin, M A Liliyan	ti, N H Nufus and M	Ali	
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OPEN ACCESS			012093
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	View article	PDF	
OPEN ACCESS			012094
•	eolatus x Ephelus i	doparasite helminth infection of cantang grouper <i>fuscoguttatus</i> ) from traditional ponds in the Kampung	{Kerapu of
B Angwarmas, L Sul	martiwi, G Mahasri,	N D Rahayu and G D Pamenang	
	View article	PDF	
OPEN ACCESS			012095
		n the Hybrid Grouper ( <i>Epinephelus fuscoguttatus x</i> ional ponds in the Kampung Kerapu Lamongan East .	lava

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OPEN ACCESS			012096
Different Concentra of Each <i>Moina mac</i>		Suspension on Fecundity and Offspring Production	012000
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	View article	PDF	
OPEN ACCESS			012097
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Biology, Ecology ar East Belitung, Indo		ential of <i>Osteochilus spilurus</i> (Bleeker 1851) in	
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Antiproliferation effects of <i>Glycine max Linn</i> ethanolic extract on induced mammary gland carcinoma in albino rats	
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OPEN ACCESS	012104
Structure communities of macrozoobenthos in mangrove tourism area, Wongsorejo sub- district, Banyuwangi regency, East Java	
D Fatmawati, B S Rahardja, Suciyono, L Lutfiah and M F Ulkhaq	
OPEN ACCESS	012105
Probiotic enriched Daphnia sp: the nutritional profile and enzymatic activities	
E Riyani, A Yuniarti and A M Hariati	
OPEN ACCESS	012106
Effect of different carbon doses of tapioca ( <i>Manihot esculenta</i> ) flour on vegetative cells and spore production of <i>Bacillus megaterium</i>	
I M D Mahariawan, N B Ariffin, W E Kusuma, A Yuniarti, M A G Beltran and A M Hariati	
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Enrichment of feed for growth of cantang grouper ( <i>E. fuscoguttatus x E. lanceolatus</i> ) in floating cages	
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OPEN ACCESS	012108
The efficacy of probiotic with different storage to decrease the total organic matter, ammonia, and total <i>Vibrio</i> on shrimp pond water	
H S Farizky, W H Satyantini and D D Nindarwi	
OPEN ACCESS	012109
Analysis degradation of mangrove vegetation in Tangerang District, Banten Province	
R Haryanti, A Fahrudin and H A Susanto	
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Determination spesies flying fishes (exocoetidae) in makassar strait	
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Potential stock of s	stony corals in Indo	onesia	012111
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Social network and dwellers in Wakato		f cooperation between <i>Sama</i> Bajo and land- I park	
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OPEN ACCESS			012114
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Haematological pa Cr <sup>+3</sup> - Yeast ( <i>Sacca</i>		h ( <i>Clarias</i> sp.) fed by immunostimulant added with <i>e</i> ) and Garlic	
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	Tiew article	🔁 PDF	
OPEN ACCESS			012116
Addition of lemuru barb ( <i>Rasbora arg</i> )		on the fat retention and fatty acid profile of silver	
A Marini, A S Mubara	ak and A T Mukti		
	Tiew article	🔁 PDF	
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Addition of Lemuru barb <i>Rasbora argy</i>		Retention and Feed Utilization Efficiency of silver	
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OPEN ACCESS			012119
The immune response supplementation	onses of <i>Oreochron</i>	<i>nis niloticus</i> under different form of <i>Bacillus</i>	
A Yuniarti, N B Arifir	n, N Muawiyah, M Fa	khri and A M Hariati	
	View article	PDF	
OPEN ACCESS			012120
Response of kutul infected by <i>Myxol</i>	•	rotein adhesion in koi fish ( <i>Cyprinus carpio</i> )	
U Yanuhar, N S Jun	irahma, N R Caesar,	M Musa and G Mahasri	
	View article	PDF	
OPEN ACCESS			012121
		aquaponics system on ammonium (NH <sub>4</sub> <sup>+</sup> ), nitrite ns of catfish ( <i>Clarias</i> sp.) aquaculture	
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	View article	PDF	
OPEN ACCESS			012122
	of molasses on fee ng with biofloc-aqua	ed conversion ratio and water quality in catfish aponic system	
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012124

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Different concentr against <i>Argulus ja</i>		<i>Moringa oleifera</i> leaf aqueous extract immersion ge	
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The effect of using	the initial weight c	of seedlings by the floating method on the	
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Nursery method of	f Jatimbulan Tilapia	a ( <i>Oreochromis niloticus</i> ) in Pasuruan, East Java	
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OPEN ACCESS	012142
Characterization of nano calcium powder from blood cockle ( <i>Anadara</i> sp.) shell produced by using different hydrochloric acid concentration	
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OPEN ACCESS	012143
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cucumber ( <i>Holothuria leucospilot</i> ) that are multi-trophic sea farming	
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The effect of addition of fish bone meal on the concentration of nitrogen (N),	
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Activity test of anti-stress from extract of Datura metel seeds with ethanol solvent	
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OPEN ACCESS	012147
Potential concentration of heavy metal copper (cu) and microalgae growth <i>Spirulina plantesis</i> in culture media	
R M S Budi, B S Rahardja and E D Masithah	

OPEN ACCESS	012148
Isolation and identification of bacteria in gastrointestinal of eel ( <i>Anguilla bicolor</i> ) that has potential as probiotic	
P Lestari, D Suprapto and G Mahasri	
OPEN ACCESS	012149
Effect of fucoidan concentration from <i>Sargassum</i> sp. on skin lotion antioxidant activities	
R C D Putra, M A Alamsjah and L Sulmartiwi	
OPEN ACCESS	012150
Correlation between water quality and prevalence on Koi ( <i>Cyprinus carpio</i> ) which infested by <i>Argulus</i> in Mungkid Subdistrict and Muntilan Subdistrict, Magelang Regency, C Java	entral
R D Yunikasari, Kismiyati and G Mahasri	
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The effectiveness of activated carbon as adsorbent in the oil purification process fish by- product of the fish canning industry	
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The effect of hydrochloric acid concentration and temperature demineralization on characteristics of chitin from penshell ( <i>Atrina pectinata</i> )	
I L Nugroho, E D Masithah and K T Pursetyo	
OPEN ACCESS	012153
Effect of Mengkudu's ( <i>Morinda citrifolia</i> ) distillation with differential fruit ripeness to control <i>Argulus</i> on <i>Carassius auratus auratus</i> .	
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Correlation between Wall Motion Score Index (WMSI) and Anatomical M-mode (AAM) Systolic Thickening with Functional Capacity in Heart Failure among Post-myocardial Infarction Patients

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OPEN ACCESS	012178
Correlation between Global Longitudinal Strain (GLS)-Left Ventricle and TEI Index (TI) with Seattle Heart Failure Model (SHFM) Score in Chronic Heart Failure Patients with Systol Dysfunction	ic
I Kartikasari, A Lefi, D Soemantri and R R Juwita	
OPEN ACCESS	012179
Atria Score for Antithrombotic Agent Utilization in Ischemic Stroke Patients with Atrial Fibrillation	
I P Dewi, K P Dewi and R T Pinzon	
OPEN ACCESS	012180
Renal Artery Stenosis: The Challenging in Diagnosis and Therapy	
I P Dewi and N Mardiana	
OPEN ACCESS	012181
Plasma Histamine Level as a Distinguishing Factor between Stable Coronary Artery Disease and Acute Coronary Syndrome	
I S Prihatiningsih, J N E Putranto, D Soemantri and A S Hadi	
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OPEN ACCESS	012182
Correlation between Left Ventricular Function with Functional Capacity in Post Infarct Myocard Patients with Heart Failure	
I Sudirman, B S Pikir and L G P Rinjani	
OPEN ACCESS	012183
Fibrinogen and Low-Density Lipoprotein (LDL) Cholesterol Levels with the Occurrence of Acute Myocardial Infarction: Is it Correlated?	
I S Warno, J N E Putranto and B Novitalia	
OPEN ACCESS	012184

	View article	🔁 PDF	
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OPEN ACCESS			012185
		Levels and Vascular Age was Measured by Carotid	
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J R S Tengor, D Soema		_	
View abstract	🗐 View article	🔁 PDF	
OPEN ACCESS			012186
Cluster of differentia	ation 36 (CD 36),	ENac, and AQP 2 effects on heart	
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OPEN ACCESS			012188
Correlation betweer	n Serum Galectin-	-3 Level with Wall Motion Score Index in Patients	
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<ul> <li>with ST-Segment Ele</li> <li>K Halid, M Aminuddin</li> <li>View abstract</li> </ul> OPEN ACCESS The Correlation betw <ul> <li>with Acute Coronary</li> <li>L D Yuni, D Soemantri</li> <li>View abstract</li> </ul> OPEN ACCESS Levofloxacin and Dr Prevent It <ul> <li>L G Catelya, C T Tjahjo</li> <li>View abstract</li> </ul> OPEN ACCESS The Effect of Atorvas	evation Myocardi and I P Dewi View article veen Serum Trypt Syndrome After i and M I Ilman View article ug-Induced Long ono and A Hanafi View article	al Infarction PDF Case Level and Ventricular Remodeling in Patient Month of Onset PDF OT Syndrome (diLQTS): The Incidence and How to	012189
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Correlation between Arterial Stiffness with Left Ventricular Mass Index and Diastolic	
Function in Patients with Stage 3 and 4 of Non-Diabetic Chronic Kidney Disease	
L Pramushinta, B S Pikir, Pranawa and D E Wardhani	
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OPEN ACCESS	012193
Thrombolytic Therapy in Octogenarian Patient with STEMI at Buleleng General Hospital	
M A Pasha and I K Susila	
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OPEN ACCESS	012194
The Correlation Between Strain and Strain Rate Left Ventricle with Functional Capacity in	
Post Myocardial Infarction Patients with Heart Failure	
M Muqsith, B S Pikir and M Rifqi	
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OPEN ACCESS	012195
Predicting the Likelihood for Severe CAD and CABG Indication on Elective Patients:	
Comparison of Novel CHA2DS2-VASc-HSF with CHA2DS2 and CHA2DS2-VASc Score	
M J Al-Farabi, I G P G Semita, K A Shonafi, R Ramadhiani, B Jovie and A Andrianto	
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OPEN ACCESS	012196
Early and Late Initiation Time of Statin Administration Effects on Early and Long-Term	
Outcome in Acute Coronary Syndrome Patients: A Literature Review	
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OPEN ACCESS	012197
Predictors of Arteriovenous Fistula Early Failure in End-Stage Renal Disease Patients: Real-World Data in Surabaya	
M R Amadis, J N E Putranto, I Maghfirah and N Mardiana	
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OPEN ACCESS	012198
Different Grades of Body Mass Index are Correlated with Left Atrium and Ventricle Structures in Patients with Hypertensive Heart Disease	
M S Bhisma, M J Al-Farabi and B S Pikir	
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# The different effects of heat shock duration and embryo age on embryonic development and hatching lengths of spotted barb (*Puntius binotatus*) fish

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## The different effects of heat shock duration and embryo age on embryonic development and hatching lengths of spotted barb (Puntius binotatus) fish

#### I Rizal<sup>1</sup>, A T Mukti<sup>2,5</sup>, A S Mubarak<sup>3</sup>, W H Satyantini<sup>2</sup> and Widjiati<sup>4</sup>

<sup>1</sup> Study Program of Aquaculture, Faculty of Fisheries and Marine, Universitas Airlangga, Surabaya 60115, Indonesia

<sup>2</sup> Department of Fish Health Management and Aquaculture, Faculty of Fisheries and Marine, Universitas Airlangga, Surabaya 60115, Indonesia

<sup>3</sup> Department of Marine, Faculty of Fisheries and Marine, Universitas Airlangga, Surabaya 60115, Indonesia

<sup>4</sup> Department of Veterinary Anatomy, Faculty of Veterinary Medicine, Universitas Airlangga, Surabaya 60115, Indonesia

<sup>5</sup> Corresponding author: akhmad-t-m@fpk.unair.ac.id

Abstract. The study aimed to determine the different effects of heat shock duration and embryo age on embryonic development and hatching lengths of spotted barb fish. The method was used the experimental method using a factorial completely randomized design with two factors of treatment, namely heat shock duration (1, 1.5, and 2 min) and embryo age (3, 4, and 5 min after fertilization). The fish embryos were shocked at temperature of 40°C. Each combination was repeated three times. One treatment was conducted without heat shock as control. The results showed that treatments of heat shock duration and embryo age have affected embryonic development and hatching lengths of spotted barb embryo.

#### 1. Introduction

Temperature is one of several determinant environmental variables that affect the development of fish embryos. Temperature causes embryonic mortality at extreme levels and affects metabolic speed, including the maintenance and growth of fish embryos and larvae [1]. Temperature is one of the critical factors and controls during the development of the embryonic and larval phases in fish [2]. Several studies have been conducted related to the effect of temperature on hatching [3, 4]. The water temperature of the media during fish embryogenesis can affect the time and age of the embryo when it hatches [5].

Many studies about the effect of incubation temperature on embryonic development in fish have been conducted. However, the influence of temperature shock, especially heat shock in the polyploidization process on fish embryonic development has not been much studied. Untl now, only few studies on embryonic death due to polyploidization have been reported include successfull ploidy, such as [6, 7, 8, 9, 10, 11]. Although, the effect of cold shock triploidy induction on cell cleavage of spotted sand bass embryo has been carried out [12], however, the influence of heat shock on embryonic development has not been reported. Therefore, this study aimed to determine the effect of heat shock duration and embryo age on embryonic development and hatching lengths of spotted barb (Puntius binotatus) embryo.

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#### 2. Materials and methods

This study was conducted to Technical Unit of Freshwater Aquaculture Development, Umbulan, Pasuruan, East Java, Indonesia and Wet Laboratory, Faculty of Fisheries and Marine, Universitas Airlangga, Surabaya, East Java, Indonesia.

#### 2.1. Artificial fertilization

Eggs and sperm of spotted barb were collected at a clean and dry container separately, especially sperm was added physiological NaCl of 0.9% with ratio 1:9. Then, eggs were added sperm solution of 2-3 drops, stirred gently for 1 min, and added 1-2 mL freshwater at temperature of 27°C and stirred slowly for 1 min. On the other hand, fertilization rate was measured to determine egg quality of spotted barb fish before the treatment of heat shock.

#### 2.2. Heat shock treatment

A number of 50 eggs were distributed in different filter according to each treatment and the number of 50 eggs were spread in control filter. The embryo age of 3, 4, and 5 min after fertilization (maf) were shocked at a water bath of 40°C temperature [13] for 1.0, 1.5, and 2.0 min, respectively.

#### 2.3. Artificial incubation of embryo

The spotted barb embryos were incubated artificially at aquaria of  $60 \times 40 \times 40$  cm size. The water temperature was set around 27-28°C using thermostat. When 30-60 min after fertilization (maf), fish embryos were observed to initial embryonic development. Then, embryo sample was collected to observe the embryonic stage. The first hatched embryos were also observed.

#### 2.4. Data analysis

Data were analyzed statistically and descriptively. Statistically, analysis was used analysis of variance (ANOVA). The treatment differences were determined using Duncan's multiple range test. The statistics are analyzed using SPPS 10 (statistical software). The significant level was determined at p<0.05.

#### 3. Results and discussion

The results showed that the fish eggs used were of good quality with an egg diameter of 0.1 to 0.25 mm and a fertilization rate of 76.92 to 89.38% and there were no significant differences between treatments and control (p>0.05). This proves that the eggs used in this study are relatively homogeneous in terms of quality, so synchronization of egg fertility can be controlled properly. Fertilization observations were made at 4 h after the eggs were fertilized by sperm from the parent spotted barb fish. Determination of the success rate of egg fertilization is seen in the color change, ie the fertilized egg is transparent in color while the unfertilized egg is turbid white [14]. Genetic factors, feed, stress, and poor water quality are the causes of damage to egg quality [15] and can have an impact on survival during the early phases of life of several fish species [1].

Heat shock results in temperature fluctuations from low temperature to high temperature. Temperatures that are too high or too low can inhibit the hatching process, even those that are too extreme can cause embryonic death and hatching failure [16]. The results of microscopic observations note that the development of spotted barb fish consists of several stages, namely: cleavage, morula, blastula, gastrula, organogenesis, and hatching stages. The division stage starts from the first mitosis division to the fifth division. Nelsen [17] states that division is a process of development in embryonic cells, which are cells increasingly becoming smaller or smaller units called blastomers.

The results showed significant differences (p<0.05) between treatments of heat shock duration and embryo age, and their interaction on embryonic development and hatching lengths of spotted barb embryo (Table 1).

IOP Conf. Series: Earth and Environmental Science 441 (2020) 012123 doi:10.1088/1755-1315/441/1/012123

Table 1. The embryonic development and hatching lengths of spotted barb e	embryo after treated heat
shock.	

HSD+EA	Cleavage	Morula	Blastula	Gastrula	Organogenesis	Hatching
(min+maf)	(h)	(h)	(h)	(h)	(h)	(h)
1.0 - 3	$0.88 {\pm} 0.05^{cde}$	2.56±0.08 <sup>cd</sup>	$5.82{\pm}0.06^{ab}$	$6.97 \pm 0.24^{a}$	18.10±0.46 <sup>a</sup>	21.63±0.25 <sup>a</sup>
1.0 - 4	$0.87 \pm 0.04^{cde}$	$2.53 \pm 0.07^{bcd}$	6.85±0.11°	7.25±0.13 <sup>b</sup>	19.48±0.17 <sup>bc</sup>	$22.00{\pm}0.97^{ab}$
1.0 - 5	$1.00{\pm}0.07^{f}$	$2.65 \pm 0.10^{d}$	$6.00 \pm 0.26^{b}$	7.17±0.09 <sup>ab</sup>	19.30±0.28 <sup>b</sup>	$21.81 \pm 0.81^{ab}$
1.5 - 3	$0.75 \pm 0.04^{b}$	$2.40{\pm}0.11^{ab}$	$5.85{\pm}0.87^{ab}$	6.95±0.11 <sup>a</sup>	20.63±0.65 <sup>de</sup>	22.92±1.03 <sup>ab</sup>
1.5 - 4	$0.81{\pm}0.08^{bcd}$	$2.48 \pm 0.72^{abc}$	5.75±0.24 <sup>ab</sup>	$7.00{\pm}0.13^{a}$	18.50±0.43ª	$22.00 \pm 1.00^{ab}$
1.5 - 5	$0.90{\pm}0.05^{de}$	2.56±0.06 <sup>cd</sup>	5.92±0.15 <sup>ab</sup>	7.13±0.12 <sup>ab</sup>	$18.20\pm0.16^{a}$	$20.73 \pm 1.18^{a}$
2.0 - 3	0.66±0.61ª	$2.38{\pm}0.04^{a}$	$5.58 \pm 0.33^{a}$	$6.97 \pm 0.08^{a}$	20.83±0.14e	$22.59 \pm 0.89^{ab}$
2.0 - 4	$0.80 \pm 0.02^{bc}$	$2.50\pm0.09^{abc}$	5.83±0.16 <sup>ab</sup>	$7.00{\pm}0.05^{a}$	$20.00 \pm 0.70^{cd}$	$22.50 \pm 0.90^{ab}$
2.0 - 5	$0.83{\pm}0.04^{bcde}$	$2.46 \pm 0.05^{abc}$	$5.80{\pm}0.14^{ab}$	6.97±0.11 <sup>a</sup>	19.28±0.12 <sup>b</sup>	$21.77 \pm 0.84^{ab}$
Control	$0.92{\pm}0.03^{ef}$	2.58±0.04 <sup>cd</sup>	$5.83{\pm}0.09^{ab}$	$7.08 \pm 0.03^{ab}$	19.25±0.15 <sup>b</sup>	$22.42 \pm 0.28^{b}$

Note: HSD = heat shock duration, EA = embryo age, maf = min after fertilization, h = hours. Different superscripts in the same column show no significant differences (p<0.05).

The treatment of heat shock duration and embryo age have a significant effect (p<0.05) on the initial division of two cells (cleavage). The treatment of heat shock duration of 2.0 min and embryo age of 3 maf being the fastest treatment in reaching the division stage of two cells (cleavage), while the treatment of heat shock duration of 1.0 min and embryo age of 5 maf became the longest treatment in reaching stage of two cells (cleavage). Iswahyudi [18] reports that the first hour after fertilization in spotted barb fishs, while the first mitotic division in common carp (*Cyprinus carpio*) occurred within 20 maf [19].

The results of the ANOVA test for the treatment of heat shock duration and embryo age showed no significant difference (p>0.05), with the time of hatching embryo of 20.73-22.92 h after fertilization. Iswahyudi [18] states that the hatching stage of spotted barb was achieved 24 h after fertilization. Meanwhile according to Haniffa *et al.* [19] showed that eggs of common carp hatch in 71 h 20 min after fertilization at 28°C temperature of incubation. The survival of the 4-day-old caked spotted barb fish larvae resulted from lower temperature shock treatment compared to controls. This is because the haploid embryo will die during hatching and only a small portion can survive [20]. Rustidja [21] states that the death of larvae is caused not only by the presence of defective larvae, but can also be caused by other factors such as water quality media, fungal attack, and inappropriate feeding. The difference in survival between controls and treatments may be due to differences in density in each treatment. Arai [22] states that differences in survival in fish given heat shock treatment, descriptively is not due to heat shock treatment given but rather is determined by the maintenance process carried out for example differences in larval density in each treatment, the nature of cannibalism or larval abnormalities.

#### 4. Conclusion

Treatment of heat shock duration and embryo age influenced embryonic development and hatching lengths of spotted barb fish.

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