

Global systematic review and meta-analysis on prevalence and concentration of aflatoxins in peanuts oil and probabilistic risk assessment

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Review

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Global systematic review and meta-analysis on prevalence and concentration of aflatoxins in peanuts oil and probabilistic risk assessment

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Abstract: Exposure to mycotoxins in food is largely unavoidable, and concerns about their health effects are growing. Consumption of vegetable oils such as peanuts oil has increased, hence several studies have been conducted on concentration of aflatoxins (AFs) in peanuts oil. Search was performed in Scopus and PubMed databases on prevalence and concentration of AFs in peanuts oil from 1 January 2005 to 15 April 29, 2022. Prevalence and concentration of AFs in peanuts oil was meta-analyzed based on country and type of AFs subgroups. In addition, health risk was calculated using monte carlo simulation method. Pooled prevalence of AFB1 in peanuts oil was 47.9%; AFB2, 46.45%; AFG1, 46.92% and AFG2, 54.01%. The Overall prevalence of AFTs was 49.30%, 95%CI (35.80–62.24%). Pooled concentration of AFB1 in peanuts oil was 2.30 $\mu\text{g}/\text{kg}$; AFB2, 0.77 $\mu\text{g}/\text{kg}$; AFG1, 0.07 $\mu\text{g}/\text{kg}$; AFG2, 0.28 $\mu\text{g}/\text{kg}$. The sort of country based on mean of MOEs in the adults consumers was Japan (47,059) > China (17,670) > Ethiopia (7,398) > Sudan (6,974) > USA (1,012) and sort of country based on mean of MOEs in the children was Japan (120,994) > China (46,991) > Ethiopia (19,251) > Sudan (18,200) > USA (2,620). Therefore, adults consumers were

in considerable health risk in Ethiopia, Sudan and USA and for children in USA (MOE < 10,000).

Keywords: aflatoxins; meta-analysis; mycotoxins; peanuts oil; probabilistic risk assessment; vegetable oil.

Introduction

Environmental contamination [1–7] and following food contamination and food security is a severe global health problem [5, 8–15]. Among mycotoxins, aflatoxins (AFs) are one of the secondary and highly toxic metabolites produced by *Aspergillus flavus* and *Aspergillus parasiticus* through a polyketide mechanism [16–19]. The four naturally occurring aflatoxins are aflatoxin B1 (AFB1), aflatoxin B2 (AFB2), aflatoxin G1 (AFG1), and aflatoxin G2 (AFG2), all of which contain the difuran ring and coumarin in their original structure [20–22].

International Agency for Research on Cancer (IARC) classifies AFB1 as one of the most abundant AFs and group I carcinogens [23] and also European Commission in 2002 set AFB1 levels below 20–50 μg in natural feed [24]. These toxins are known to be carcinogenic, teratogenic, mutagenic and immunosuppressive and pose a threat to animal and human health [25–30].

The growth of aflatoxin-producing fungi and level production of toxins in food is related to fungal species, composition of the food, the percentage of active water, temperature and relative humidity [31].

The lethal dose of aflatoxin (LD₅₀) is 1–50 mg/kg for most animal species and highly toxic effects (LD₅₀ less than 1 mg/kg) for other species such as cats, pigs, and dogs [32].

The exposure to AFs, especially aflatoxin B1, can lead to aflatoxicosis. While constant exposure to AFs can lead to further suppression of cancers, immune responses, and other health effects. The lipophilic structure of AFs facilitates their entry into the bloodstream through the gastrointestinal tract and respiratory tract [27, 33]. About 70 to 90

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percent of liver cancers and also third leading cause of death is related to AFs [27]. It is attributed to liver carcinoma (HCC) with cancer [34]. In this regard, the incidence of liver cancer is higher in developing countries [27].

Peanut oil is the fifth most widely used oil in confectionery, candy and pastry. Peanut seeds have high nutritional and commercial value due to their protein, fatty acids, carbohydrates and fiber, in addition to vitamins, calcium and phosphorus [35]. Over the past 20 years, the demand for 13 vegetable oils has increased from 72 million tone in 1995 to 17.5 million in 2015 [36]. Global production of peanut oil was 6.48 million tons in 2021 [37]. China and India are the world's largest markets for peanuts and their derivatives (especially peanut oil) in terms of production and consumption [38].

Experimental studies have shown that aflatoxins present in oily substances can be transferred to the final oil product [39]. The European Commission recommended a maximum level of AFB1 and total aflatoxins of 2 µg/kg and 4 µg/kg, respectively [29, 40]. The Food and Drug Administration (FDA) has recommended a maximum of 20 µg/kg total AFs in peanuts. U.S. regulations also set a maximum acceptable limit of total contamination of AFs in peanuts of 20 µg/kg.

Crude grains that are used for edible oil are often stored in inappropriate conditions for a long time, thus providing the basis for the growth of fungi and then production mycotoxins and following mycotoxins is eventually transferred to the extracted oil [41]. Therefore, the possible presence of AFs in oilseeds is inevitable, and AFs contamination remains an acute problem, especially in areas with high humidity and rainfall levels [27, 29, 42]. It is difficult to prevent or control peanut contamination by poisonous fungi because the danger of fungi occurs naturally [27, 29, 42].

Many investigation have been conducted on prevalence and concentration of AFs in peanut oil [20, 43–47] but not conducted meta-analysis and probabilistic health risk assessment study. Therefore, the current study was performed with the aim of meta-analysis of the prevalence and concentration of AFs in peanut oil and also probabilistic health risk assessment in consumers of peanut oil content AFs.

Material and method

Search strategy

The retrieve of articles was based on Preferred Reporting Items for Systematic Reviews (Figure 1) [48, 49]. The search was conducted in Scopus and PubMed on Prevalence and concentration of AFs in peanuts oil from 1 January 2005 to 15 April 29, 2022. Keywords were consisting of “mycotoxin” OR “Aflatoxins” OR “Ochratoxin A” OR “patulin” OR “fumonisins” OR “zearalenone” OR “nivalenol/deoxynivalenol” AND “Cereal” OR “peanuts oil” OR “goober-pea” OR “*Arachis hypogaea*” OR “puny” OR “seed”. Disagreement between the authors in selecting an paper was resolved by the corresponding author.

Inclusion, exclusion criteria and data extraction

Our criteria for inclusion paper were present positive sample size or mean with range concentration of AFs; measured of concentration of AFs in peanuts oil and descriptive study. In addition, letter to editors; review articles, chapter; books, conferences, and studies measured concentration of AFs in other vegetables oil were excluded. Country, type of vegetable oil, total sample size, positive sample, type of aflatoxins, mean, SD, Range concentration of AFs, LOD and Method of analysis were extracted (Table 1).

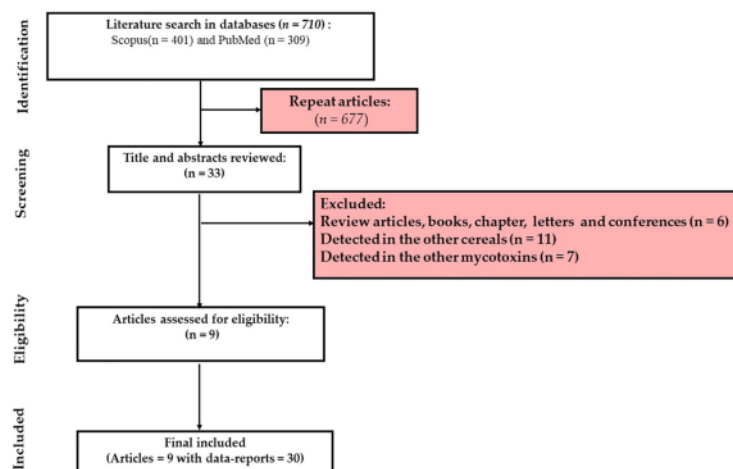


Figure 1: Process selection of study based on PRISMA.

Table 1: Main characteristic included in our study.

Country	Total sample size	Positive sample	Type of mycotoxin	Mean, $\mu\text{g}/\text{kg} \pm \text{SD}$	Range, $\mu\text{g}/\text{kg}$	LOD, $\mu\text{g}/\text{kg}$	Method	References
China	8	5	B1	0.50 ± 0.20	0.03–2.23	0.05	HPLC-MS/MS	[43]
China	8	3	B2	0.50 ± 0.20	0.03–2.23	0.02	HPLC-MS/MS	[43]
Ethiopia	5	5	B1	27.28 ± 10.30	0.07–145.59	0.05	HPLC-MS/MS	[43]
Ethiopia	5	5	B2	27.28 ± 10.30	0.07–145.59	0.02	HPLC-MS/MS	[43]
Ethiopia	5	5	G1	27.28 ± 10.40	0.07–145.59	0.09	HPLC-MS/MS	[43]
Ethiopia	5	5	G2	27.28 ± 11.00	0.07–145.59	0.04	HPLC-MS/MS	[43]
Sudan	21	11	B1	16.30 ± 12.00	2.9–36.1	1.5	HPLC-FLD	[20]
Sudan	21	14	B2	1.00 ± 0.50	0.02–3.6	0.09	HPLC-FLD	[20]
Sudan	21	20	G1	12.90 ± 0.50	3.1–64.1	0.8	HPLC-FLD	[20]
Sudan	21	19	G2	11.60 ± 0.50	0.5–60.7	0.4	HPLC-FLD	[20]
China	52	43	B1	0.93 ± 0.50	0.5–69.4		UPLC-MS/MS	[44]
Japan	8	5	B1	0.62 ± 0.50	0.52–0.72			[50]
Japan	8	5	B2	0.16 ± 0.50	0.09–0.22			[50]
Japan	8	5	G1	0.08 ± 0.50	0.07–0.08			[50]
Japan	8	5	G2					[50]
China	427	47	B1	29.40 ± 44.40	15.4–49.9	1.08	ELISA	[45]
China	427	33	B1	17.10 ± 26.00	8.8–22.2	1.08	ELISA	[45]
China	31	15	B1	1.29 ± 0.32	0.15–2.72	0.05	LC-MS/MS	[46]
China	31	6	B2	0.11 ± 0.03	0.15–0.36	0.02	LC-MS/MS	[46]
China	31	3	G1	0.02 ± 0.00	0.01–0.02	0.09	LC-MS/MS	[46]
USA	3	1	B1	17.00 ± 8.30		0.03	LC-MS/MS	[47]
USA	3	1	B2	3.00 ± 0.75		0.03	LC-MS/MS	[47]
USA	3	0	G1	0.02 ± 0.01		0.03	LC-MS/MS	[47]
USA	3	0	G2	0.04 ± 0.01		0.06	LC-MS/MS	[47]
China	96	22	B1	38.74 ± 47.75	0.26–283.0	0.1	HPLC-FLD	[51]
China	15		B1	0.70 ± 0.40		0.02	HPLC-FLD	[52]
China	120	84	B1	10.94 ± 5.46	0.02–21.85			[52]
China	120	48	B2	1.75 ± 0.87	0.01–3.49		UHPLC-TMS	[52]
China	120	17	G1	0.17 ± 0.08	0.01–0.32		UHPLC-TMS	[52]
China	120	13	G2	0.26 ± 0.12	0.01–0.5		UHPLC-TMS	[52]

Meta-analysis of prevalence and concentration

Meta analysis of prevalence was conducted using “metaprop” command with weighing model of Dersimonian–Laird [53] and also concentration of AFs in peanuts oil was meta-analyzed using average and standard error (SE) [54]. The I-square test used to heterogeneity analysis [55]. Stata 14 (Stata Corporation, College Station, USA) was used to meta-analysis of prevalence and concentration of AFs in peanuts oil.

Probabilistic risk assessment

The health risk in consumers due to the consumption of peanuts oil content of AFs was estimated.

Estimated daily intake was calculated using below equation:

$$EDI = \frac{C \times IR \times EF \times ED}{BW \times AT} \quad (1)$$

where, EDI is estimated daily intake ($\mu\text{g}/\text{kg}\cdot\text{d}$), C is concentration of aflatoxins; IR, ingestion rate of peanuts oil (Table 2) [56]; ED, exposure duration (30 years); EF, exposure frequency (350 days/y), BW, body

weight (children: 15 kg and adults: 70 kg) [57] and AT, average life time (for the both adults and children: 25,550 days).

The margin of exposures in consumers due to AFs was calculated by Equation (2) [60]:

$$MOEs = \frac{BMDL_{10}}{EDI} \quad (2)$$

In this equation, MOEs is margin of exposure; $BMDL_{10}$, value benchmark dose limit ($\mu\text{g}/\text{kg}\cdot\text{d}$). The $BMDL_{10}$ for AFs is equal to

Table 2: Consumption rate of vegetable oil and peanuts oil (kg/n-d).

Country	Vegetable oil [58]	Peanuts oil ^a	
		Mean	SD
USA	30.600	0.612	0.077
Japan	15.600	0.312	0.039
Ethiopia	3.100	0.062	0.008
Sudan	5.500	0.110	0.014
China	8.100	0.162	0.020

^aConsumption rate of Peanuts oil was almost 2% of vegetable oil [59].

0.4 µg/kg-bw [61]. If, MOEs < 10,000, health risk is considered acceptable [60–62]. In order to decreased uncertainties in risk assessment, monte carlo simulation (MCS) method was used in Oracle Crystal Ball software (ver. 11.1.2.4). Mean of MOE was considered as benchmark of risk with 50,000 repetitions. Lognormal distribution for both concentration and ingestion rate and normal distribution for body weight were considered in MCS method [63, 64].

Results and discussion

Results of meta-analysis

The meta-analysis of aflatoxins in peanuts oil (9 articles with 30 data-reports) (Figure 1) revealed pooled prevalence of AFB1 in peanuts oil was 47.9%, 95%CI (26.96–69.19%); AFB2, 46.45%, 95%CI (30.06–68.91%); AFG1, 46.92%, 95%CI (10.08–85.52%); and AFG2, 54.01%, 95%CI (5.38–98.38%) (Appendix 1). Overall prevalence of total AF was 50.30%, 95%CI (35.80–62.84%) (Appendix 1). Pooled concentration of AFB1 in peanuts oil was 2.30 µg/kg, 95%CI (1.94–2.65 µg/kg); AFB2, 0.77 µg/kg, 95%CI (0.58–0.96 µg/kg); AFG1, 0.07 µg/kg, 95%CI (0.03–0.11 µg/kg); AFG2, 0.28 µg/kg, 95%CI (0.03–0.52 µg/kg). As can be seen from the results, the prevalence and concentration of aflatoxins in peanuts was different, though not significant. In the current study, the pooled concentration of AFB1 in peanuts oil was greatly lower than maximum levels (MLs) of 2–8 µg/kg for AFB1 and 15 µg/kg for total aflatoxins that was established by the European Commission [65]. As mentioned from studies, the differences observed in AFs concentration in peanuts oil can be correlated with several factors such as oilseeds species, weather situations, management of seeds and chemical properties of each of produces (moisture, pH, temperature). Also, the AFs can be entered into oil crops during the managing condition post- and preharvest of the product, processing conditions and storage [66, 67]. Various studies have surveyed the concentration of aflatoxins in peanut oil. For example, Fang et al. (2022) reported from 120 peanut oil sample, the prevalence of AFB1, AFB2, AFG1, and AFG2 was 70.0, 40.0, 14.2 and 10.8%, respectively [65]. Among different edibles oil studied by these authors, peanut oil had the most contamination. They indicated type of packaging and also sampling strategies were effectively affecting the concentration of AFs [65]. In another study, Chen et al. reported that the average AFB1 and AFs contamination in peanut oil of Ethiopian was 76.98 and 27.28 µg/kg, respectively [68]. Deng et al. showed from among 52 peanut oil samples, contamination of AFB1 were detected in 43 samples with the content in the ranges of 0.5–69.4 µg/kg [44]. Furthermore, in similar study Qi et al. found that the average and

range of AFB1 in peanut oil was 29.4 µg/kg [69]. Moreover, Elzupire et al. indicated the mean level of AFT in peanut oil samples was 32.0 µg/kg [20]. In conducted by Yang et al. the levels of AFB1, AFB2 and AFG1 in peanut oils ranged 0.15–2.72, 0.15–0.36 and 0.01–0.02 µg/kg, respectively [46]. According to these investigations, excessive discrepancy between AFT level in peanut oils in our study and those of previous studies may be associated to the species and quality of seeds and also time harvest of crops. The grain quality can be related with different factors such as physical-chemical properties (oxygen, moisture and temperature), the quantity of fungal contamination and duration of storage [70]. Elzupir et al. indicated the main source of AFs contamination in peanut oils is often utilize the oil seeds with low quality that stored over prolonged periods at high temperature and humidity conditions [20]. Contamination of Peanut with AFT at the pre-harvest during may happen because of the collision of the crops with the contaminated soil with fungal. Even the harvest season of the crops can effect on concentration of peanuts to AFs contamination [71, 72]. Consistent with these studies, Qi et al. showed during the harvest stage of peanut seeds, they may they may be stored by rural households and oil processors in an open pile and/or in nylon bags, can easy result to fungal infection [73]. Storage conditions of peanuts are another important factor effective in the observed changes on AFs contamination in peanut oils. According to the investigation conducted by Idris et al. AFs contamination in the peanut oils can be as a result of the unprincipled storage of the seeds in suitable conditions for fungal production and growth [74]. Among other significant issues, several process techniques have key influence on the AFs concentration in oilseed. According to a results obtained by Bordin et al. processes methods such as scaling, heat treatment, wet and dry milling, neutralization, bleaching and deodorization have a many effect on the content of AFs in the peanut oils [70].

Prevalence and concentration of aflatoxins in peanut oil

Based on the obtained data, the sort of country based on prevalence of AFB1 in peanuts oil was Ethiopia (100%) > Japan (62.5%) > Sudan (52.38%) > China (40.48%) > USA (33.33%) (Appendix 2), for AFB2 was Ethiopia (100%) > Sudan (66.67%) > Japan (62.5%) > USA (33.33%) > China (31.76%) > (Appendix 3), in regarding AFG1 was Ethiopia (100%) > Sudan (95.24%) > Japan (62.5%) > China (12.96%) > USA (0.00%) (Appendix 4), and also AFG2 was Ethiopia (100%) > Sudan (90.48%) > Japan

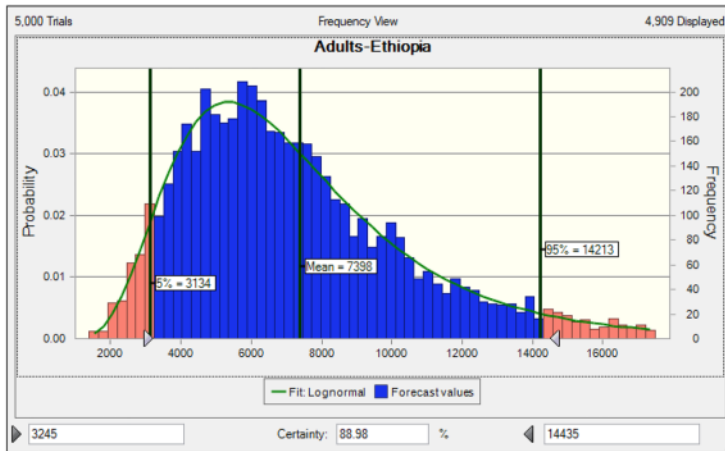
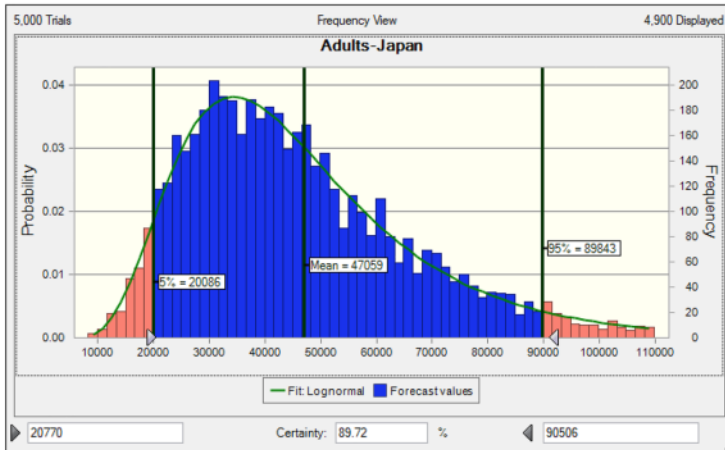
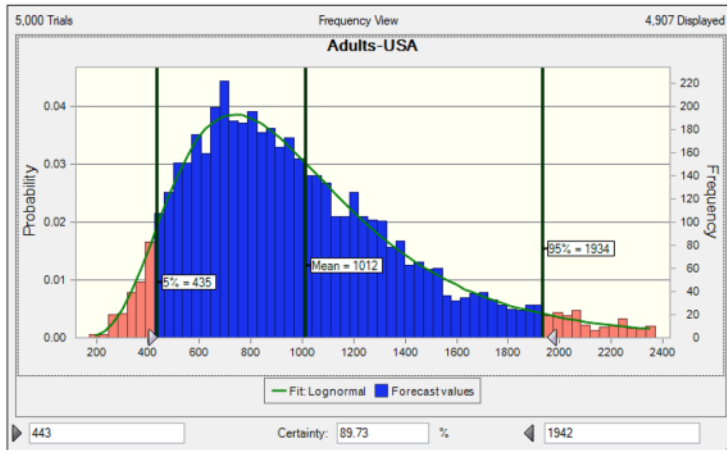


Figure 2: MOE in adults due to consumption peanuts oil content of aflatoxins.

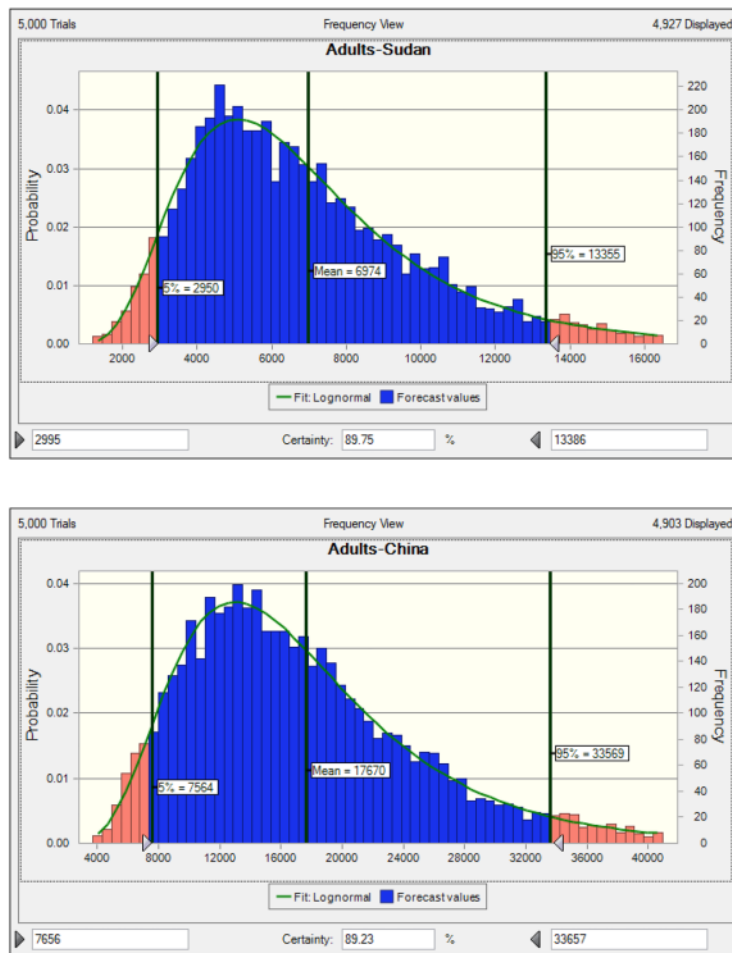


Figure 2: Continued.

(62.5%) > China (10.83%) > USA (0.00%) (Appendix 5). The sort of country based on concentration of AFB1 in peanuts oil was Ethiopia (27.28 µg/kg) > USA (17.00 µg/kg) > Sudan (16.30 µg/kg) > China (4.29 µg/kg) (Appendix 6). The sort of country based on concentration of AFB1 in peanuts oil was Ethiopia (27.28 µg/kg) > USA (3.00 µg/kg) > Sudan (1.00 µg/kg) > China (0.78 µg/kg) > Japan (0.16 µg/kg) (Appendix 7). The sort of country based on concentration of AFG1 in peanuts oil was Ethiopia (27.28 µg/kg) > Sudan (12.90 µg/kg) > China (0.09 µg/kg) > Japan (0.08 µg/kg) > USA (0.02 µg/kg) (Appendix 8). The sort of country based on concentration of AFG2 in peanuts oil was Ethiopia (27.28 µg/kg) > Sudan (11.60 µg/kg) > China (0.25 µg/kg) > USA (0.04 µg/kg) (Appendix 9). Based on the results, there is a significant difference between various countries

in terms of contamination to aflatoxins. Comparisons between several countries showed that the level of AFs contamination varied in different countries and regions. Oyedele et al. and Kabak reported, in Europe (26.7%) of samples were contamination with AFs in range from 0.6 to 49.6 µg/kg. In Canada, only 8.3% of samples had AFs ranging from 0.1 to 28 µg/kg. On the contrary, it was seen that the high AFs level in samples were related to other Asiatic countries [75, 76]. According to these studies, AFs content is significantly dependent on geographical position of peanut producers. As mentioned in previous studies in the poorest regions and developing countries in the world with humid and warm weather conditions usually AFs concentration increase because of post-harvest fungal growth [77]. Similarly, Kumar et al. showed in warm and

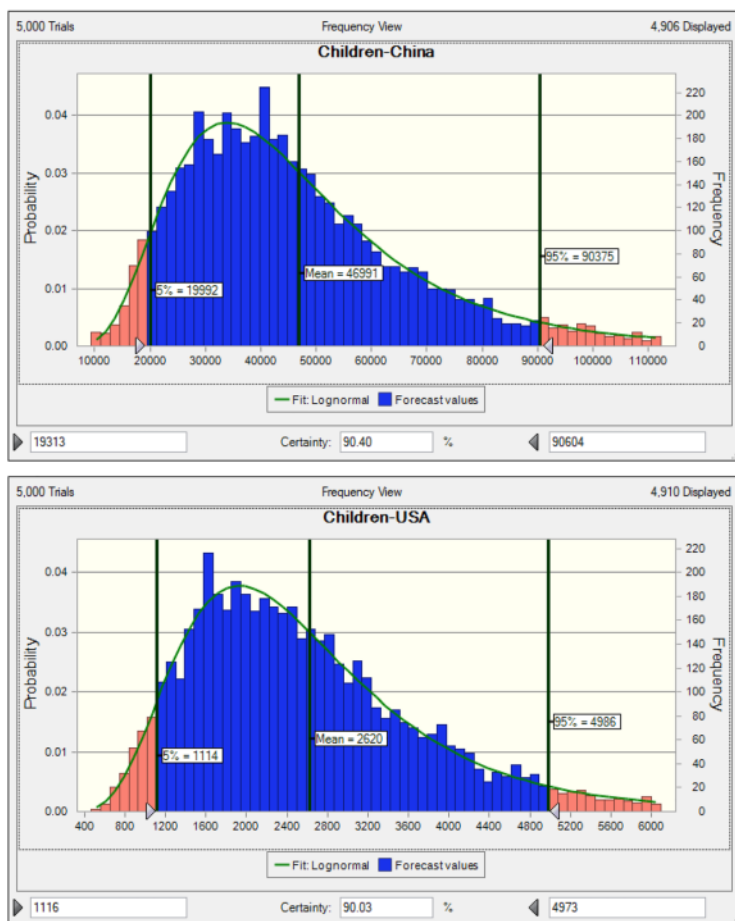


Figure 3: MOE in children due to consumption peanuts oil content of aflatoxins.

wet climate condition, was more favorable growth of the fungal. In this status to decrease the AFB1 content, recommended for farmers to harvest the raw peanut in good weather condition such as (i.e. sunshine, less rain and fog) and crops should process and store under appropriate temperature and humidity [78]. Besides, hot temperatures and high humidity are the main reasons of fungal growth in subtropical and tropical areas, [71]. Other studies have shown that the species of fungus that infects peanuts and as well as agricultural methods has an important role in the variable content of aflatoxins in peanut oil. Ismail et al. indicated aflatoxigenic fungi, particularly *A. Parasiticus* and *A. flavus* have strong affinity [27] contaminate peanut [79]. Nabizadeh et al. stated that the occurrence of AFs in edible oils in Iran country may be related to different agricultural methods and techniques and also environmental conditions [80]. Similar to these findings, in studies conducted by Chen et al. on samples of peanut oils from

Ethiopia and China, they reported which oils samples from Ethiopia were more highly contaminated than the Chinese samples possibly due to traditional agricultural, lower awareness among farmers, climatic conditions, late harvesting, and inadequate drying of oil seeds and peanuts. Moreover, authors stated low level of contamination in peanut oils from China can be related to modern technologies, strong agricultural policies and chemical analyses [43]. In line to these studies, Qiu and Fu (2012) reported all positive samples of peanut oil were lower than the regulatory limit, which was attributed to technologies used by alkali-refining processing for AFB1 removal [81].

Risk assessment

The sort of country based on mean of MOEs in the adults consumers was Japan (47,059) > China (17,670) > Ethiopia

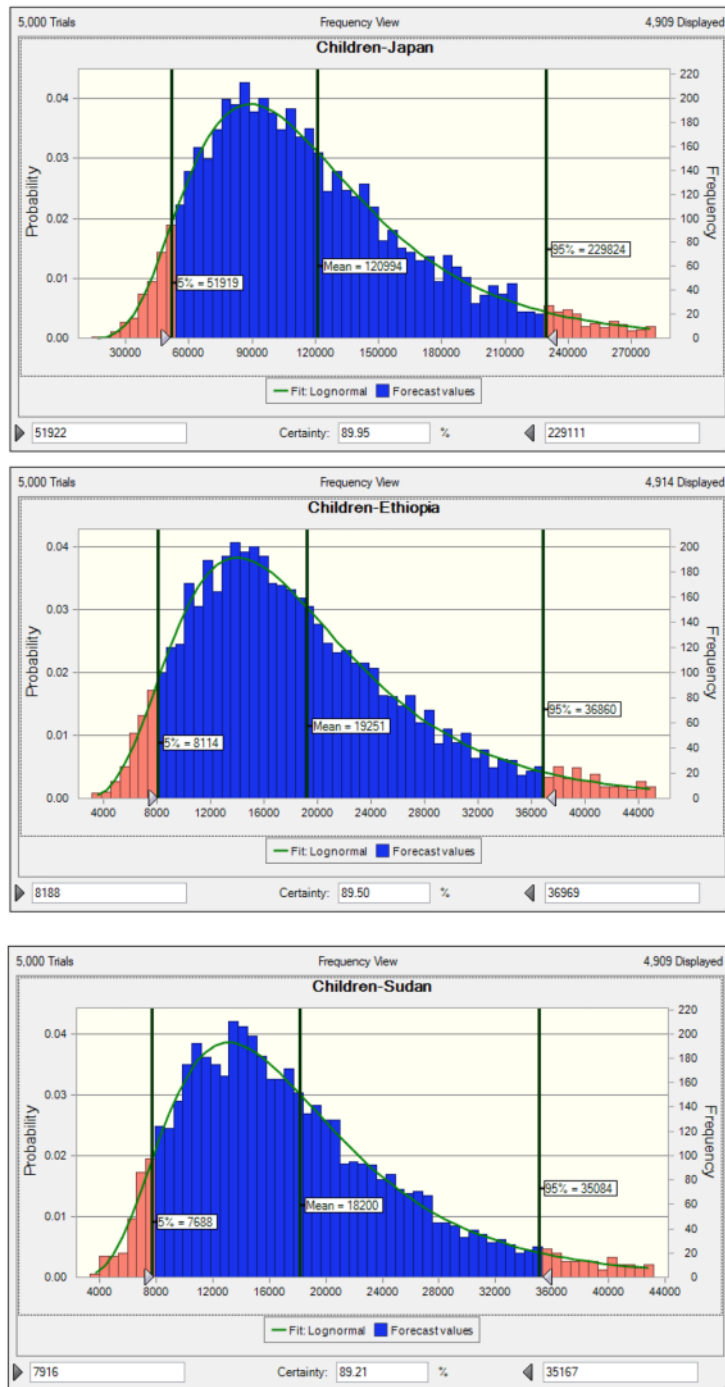


Figure 3: Continued.

(7,398) > Sudan (6,974) > USA (1,012) and in children Japan (120,994) > China (46,991) > Ethiopia (19,251) > Sudan (18,200) > USA (2,620) (Figures 2 and 3). When MOEs value less than 10,000, health risk is considerable [60–62], hence MOE was less than 10,000 for adults in Ethiopia, Sudan and USA and for children in USA.

Conclusions

This study was designed with the aim of meta-analysis of prevalence and concentration of Afs in peanut oil and also the health risk in the adults and children consumers were estimated using MCS method. The sort of Afs in peanuts oil based on pooled concentration was AFB1 > AFB2 > AFG2 > AFG1. Afs in approximately 50% of the peanut oil

samples was observed, hence control plans are recommended to reduce the concentration of Afs in peanut oil. The probabilistic health risk assessment revealed that adults consumers in Ethiopia, Sudan and USA and also children in USA were in considerable health risk.

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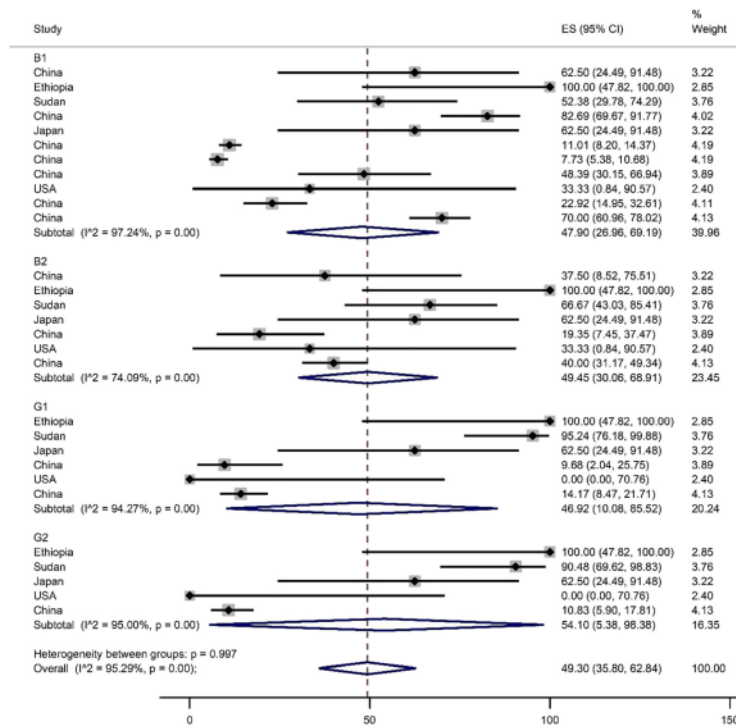
Author contributions: All authors have accepted responsibility for the entire content of this manuscript and approved its submission.

Competing interests: Authors state no conflict of interest.

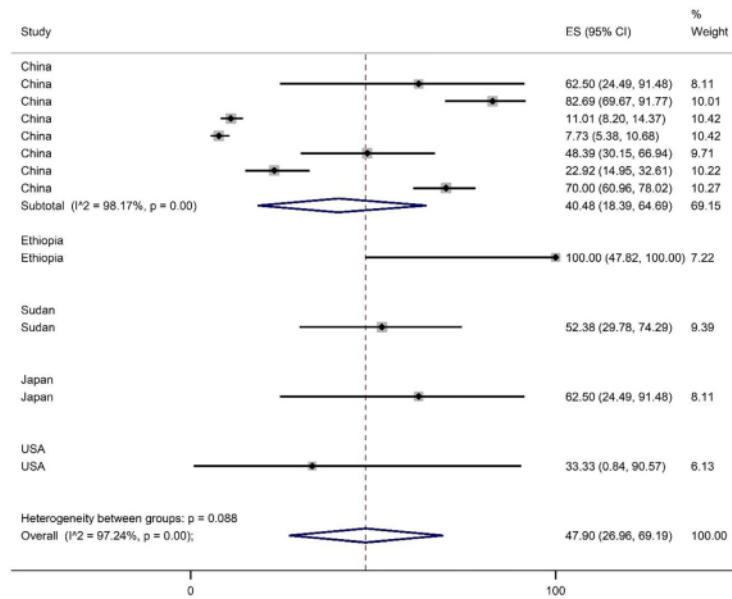
Informed consent: Not applicable.

Ethical approval: The conducted research is not related to either human or animal use.

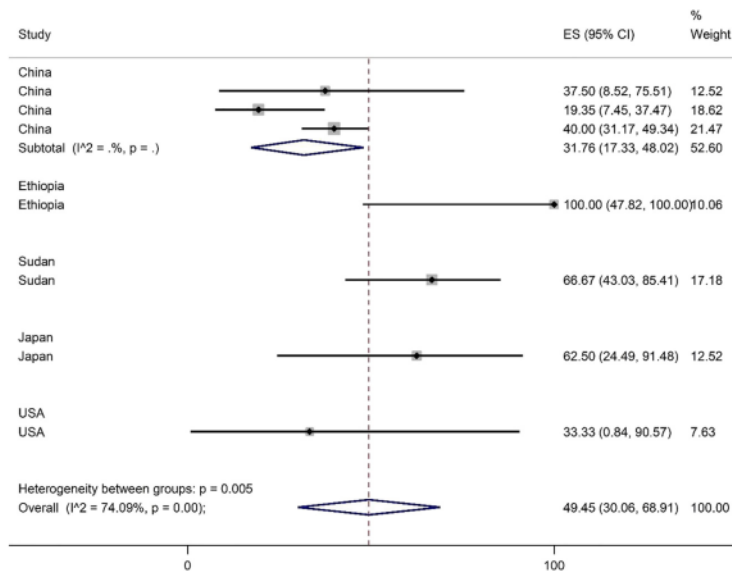
Appendix figures



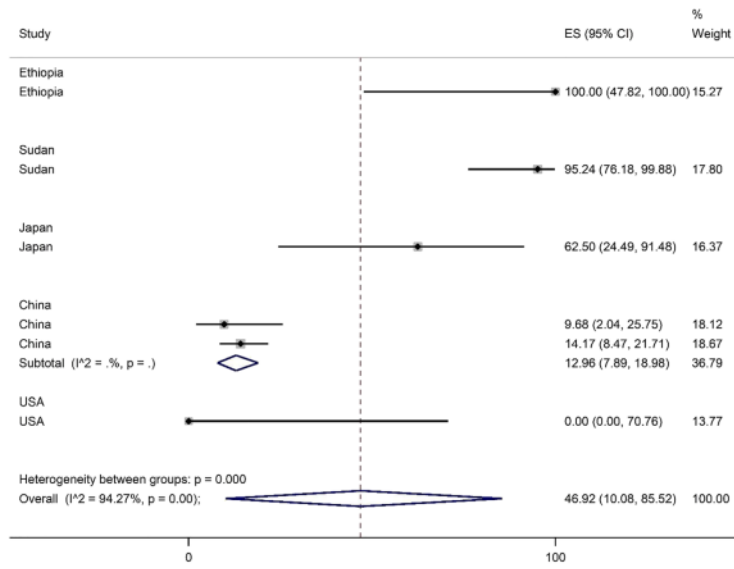
Appendix 1: Meta-analysis of prevalence of aflatoxins in peanuts oil based on county subgroups.



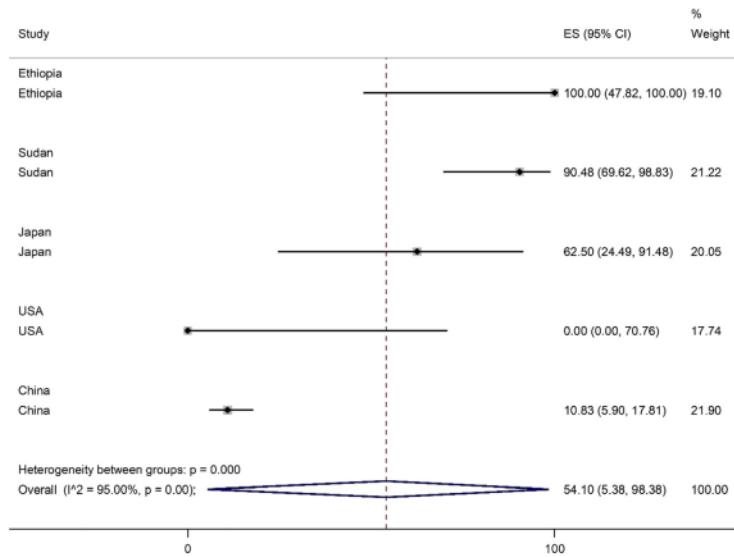
Appendix 2: Meta-analysis of prevalence of AFB1 in peanuts oil.



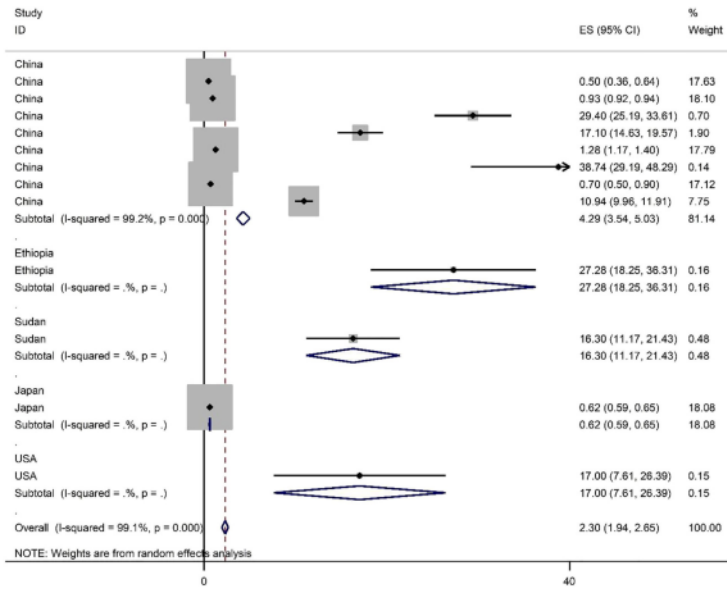
Appendix 3: Meta-analysis of prevalence of AFB2 in peanuts oil.



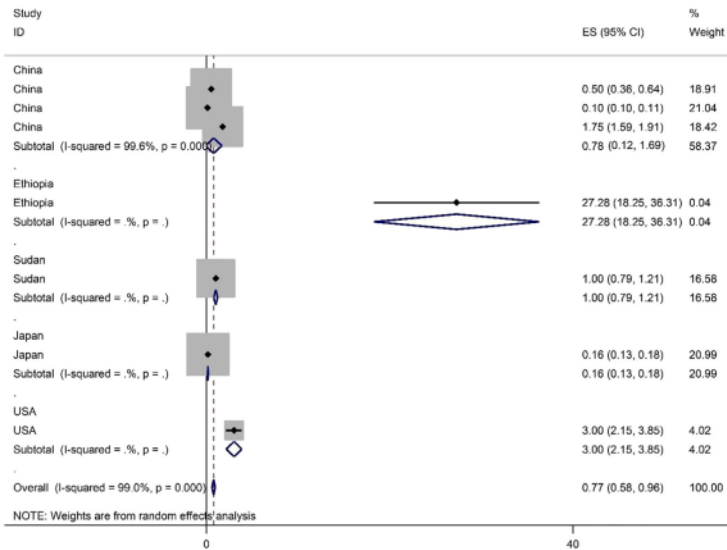
Appendix 4: Meta-analysis of prevalence of AFG1 in peanuts oil.



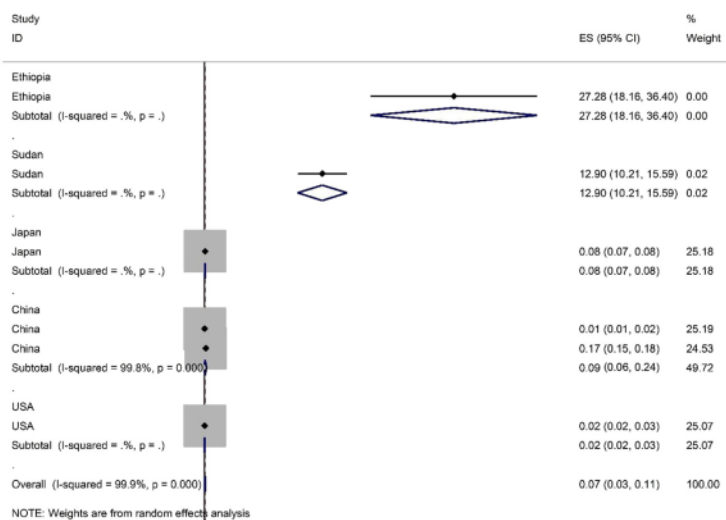
Appendix 5: Meta-analysis of prevalence of AFG2 in peanuts oil.



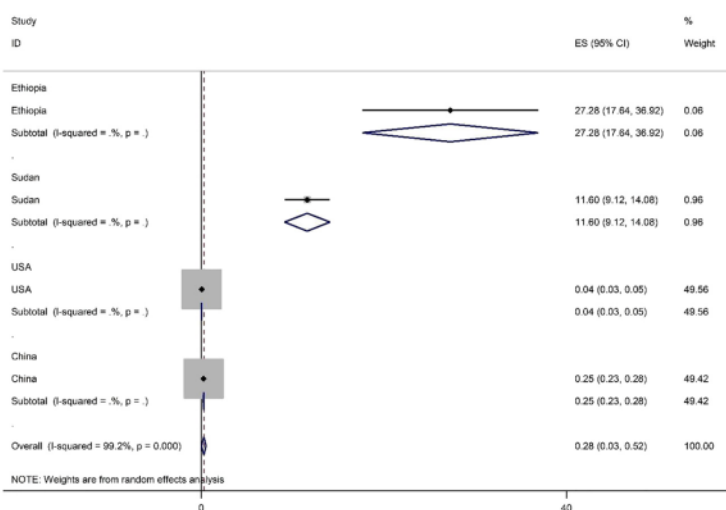
Appendix 6: Meta-analysis of concentration of AFB1 in peanuts oil (µg/kg).



Appendix 7: Meta-analysis of concentration of AFB2 in peanuts oil (µg/kg).



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Appendix 8: Meta-analysis of concentration of AFG1 in peanuts oil ($\mu\text{g}/\text{kg}$).



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Appendix 9: Meta-analysis of concentration of AFG2 in peanuts oil ($\mu\text{g}/\text{kg}$).

References

1. Azari A, Babaie A-A, Rezaei-Kalantary R, Esrafil A, Moazzen M, Kakavandi B. Nitrate removal from aqueous solution by carbon nanotubes magnetized with nano zero-valent iron. *J Mazandaran Univ Med Sci* 2014;23:15–27.
2. Moghaddam MH, Nabizadeh R, Dehghani MH, Akbarpour B, Azari A, Yousefi M. Performance investigation of Zeolitic Imidazolate framework-8 (ZIF-8) in the removal of trichloroethylene from aqueous solutions. *Microchem J* 2019;150: 104185.
3. Badi MY, Esrafil A, Pasalari H, Kalantary RR, Ahmadi E, Gholami M, et al. Degradation of dimethyl phthalate using persulfate activated by UV and ferrous ions: optimizing operational parameters mechanism and pathway. *J Environ Health Sci Eng* 2019;17:685–700.
4. Lai W-F. Non-conjugated polymers with intrinsic luminescence for drug delivery. *J Drug Deliv Sci Technol* 2020;59:101916.
5. Ding W, Meng Q, Dong G, Qi N, Zhao H, Shi S. Metabolic engineering of threonine catabolism enables *Saccharomyces cerevisiae* to produce propionate under aerobic conditions. *Biotechnol J* 2022;17:2100579.
6. Obireddy SR, Lai W-F. Preparation and characterization of 2-hydroxyethyl starch microparticles for co-delivery of multiple bioactive agents. *Drug Deliv* 2021;28:1562–8.
7. Noor NM, Suberi IAM, Susanti D, Mukail Y, Adam A, Saad S, et al. Potential of dried and fresh extracts of *sansevieria trifasciata* to mitigate alexdrium tamiyavanichii, A toxic dinoflagellate. *Sci Heritage J* 2018;2:18–20.

8. Imran M, Cao S, Wan S, Chen Z, Saleemi MK, Wang N, et al. Mycotoxins—a global one health concern: a review. *Agrobiol Records* 2020;2:1–16.
9. Roudbari A, Rafiei Nazari R, Shariatifar N, Moazzen M, Abdolshahi A, Mirzamohammadi S, et al. Concentration and health risk assessment of polycyclic aromatic hydrocarbons in commercial tea and coffee samples marketed in Iran. *Environ Sci Pollut Control Ser* 2021;28:4827–39.
10. Homayonpour P, Jalali H, Shariatifar N, Amanlou M. Effects of nano-chitosan coatings incorporating with free/nano-encapsulated cumin (*Cuminum cyminum* L.) essential oil on quality characteristics of sardine fillet. *Int J Food Microbiol* 2021;341:109047.
11. Karami H, Shariatifar N, Nazmara S, Moazzen M, Mahmoodi B, Mousavi Khaneghah A. The concentration and probabilistic health risk of potentially toxic elements (PTEs) in edible mushrooms (wild and cultivated) samples collected from different cities of Iran. *Biol Trace Elem Res* 2021;199:389–400.
12. Samiee S, Fakhri Y, Sadighara P, Arabameri M, Rezaei M, Nabizadeh R, et al. The concentration of polycyclic aromatic hydrocarbons (PAHs) in the processed meat samples collected from Iran's market: a probabilistic health risk assessment study. *Environ Sci Pollut Control Ser* 2020;27:21126–39.
13. Ke Z, Bai Y, Bai Y, Chu Y, Gu S, Xiang X, et al. Cold plasma treated air improves the characteristic flavor of Dry-cured black carp through facilitating lipid oxidation. *Food Chem* 2022;377:131932.
14. Liu G, Nie R, Liu Y, Mehmood A. Combined antimicrobial effect of bacteriocins with other hurdles of physicochemic and microbiome to prolong shelf life of food: a review. *Science of the Total Environment* 2022;825:154058.
15. Mahato S, Bhuju S, Shrestha J. Effect of *Trichoderma viride* as biofertilizer on growth and yield of wheat. *Malays J Sustain Agric* 2018;2:1–5.
16. Huffman J, Gerber R, Du L. Recent advancements in the biosynthetic mechanisms for polyketide-derived mycotoxins. *Biopolymers* 2010;93:764–76.
17. Sadighara P, Ghanati K. The aflatoxin B1 content of peanut-based foods in Iran: a systematic review. *Rev Environ Health* 2022;37:29–33.
18. Behfar M, Heshmati A, Mehri F, Khaneghah AM. Removal of ochratoxin A from grape juice by clarification: a response surface methodology study. *Foods* 2022;11:1432.
19. Heshmati A, Mozaffari Nejad AS, Mehri F. Occurrence, dietary exposure, and risk assessment of aflatoxins in wheat flour from Iran. *Int J Environ Anal Chem* 2021:1–14. <https://doi.org/10.1080/03067319.2021.2011254>.
20. Elzupir AO, Suliman MA, Ibrahim IA, Fadul MH, Elhussein AM. Aflatoxins levels in vegetable oils in Khartoum State, Sudan. *Mycotoxin Res* 2010;26:69–73.
21. Ji N, Diao E, Li X, Zhang Z, Dong H. Detoxification and safety evaluation of aflatoxin B1 in peanut oil using alkali refining. *J Sci Food Agric* 2016;96:4009–14.
22. Einolghozati M, Heshmati A, Mehri F. The behavior of aflatoxin M1 during lactic cheese production and storage. *Toxin Rev* 2021:1–9. <https://doi.org/10.1080/15569543.2021.1979044>.
23. Miliță NM, Mihăescu G, Chifiriuc C. Aflatoxins—health risk factors. *Bacteriologia Virusologia Parazitologia Epidemiologia*. 2010;55:19–24.
24. Hathout AS, Abel-Fattah SM, Abou-Sree YH, Fouzy AS. Incidence and exposure assessment of aflatoxins and ochratoxin A in Egyptian wheat. *Toxicol Rep* 2020;7:867–73.
25. Aranega JPRB, Oliveria CAF. Occurrence of mycotoxins in pastures: A systematic review. *Qual. Assur. Saf. Crop* 2022;14:135–44.
26. Chen J, Liu F, Li Z, Tan L, Zhang M, Xu D. Solid phase extraction based microfluidic chip coupled with mass spectrometry for rapid determination of aflatoxins in peanut oil. *Microchem J* 2021;167:106298.
27. Nourbakhsh F, Tajbakhsh E. Neurotoxicity mechanism of Ochratoxin A. *Qual. Assur. Saf. Crop* 2021;13:34–45.
28. Ji J, Xie W. Removal of aflatoxin B1 from contaminated peanut oils using magnetic attapulgite. *Food Chem* 2021;339:128072.
29. Magzoub R, Yassin A, Abdel-Rahim A, Gubartallah E, Miskam M, Saad B, et al. Photocatalytic detoxification of aflatoxins in Sudanese peanut oil using immobilized titanium dioxide. *Food Control* 2019;95:206–14.
30. Xia X, Wang H, Yang H, Deng S, Deng R, Dong Y, et al. Dual-terminal stemmed aptamer beacon for label-free detection of aflatoxin B1 in broad bean paste and peanut oil via aggregation-induced emission. *J Agric Food Chem* 2018;66:12431–8.
31. Şengül Ü, Yalçın E, Şengül B, Çavuşoğlu K. Investigation of aflatoxin contamination in maize flour consumed in Giresun, Turkey. *Qual. Assur. Saf. Crop* 2016;8:385–91.
32. Balina A, Kebede A, Tamiru Y. Review on aflatoxin and its impacts on livestock. *J Dairy Sci* 2018;6:e555685.
33. Finotti E, Parroni A, Zaccaria M, Domin M, Momeni B, Fanelli C, et al. Aflatoxins are natural scavengers of reactive oxygen species. *Sci Rep* 2021;11:1–9.
34. Cai P, Zheng H, She J, Feng N, Zou H, Gu J, et al. Molecular mechanism of aflatoxin-induced hepatocellular carcinoma derived from a bioinformatics analysis. *Toxins* 2020;12:203.
35. Tosun H, Ayyıldız T. Occurrence of aflatoxin M1 in organic dairy products. *Qual. Assur. Saf. Crop* 2013;5:215–9.
36. Mielke T. World markets for vegetable oils and animal fats. *Biokerosene: Springer*; 2018:147–88 pp.
37. Shahbandeh M. Peanut-oil-production-volume-worldwide. Available from: <https://www.statista.com/statistics/613483/peanut-oil-production-volume-worldwide/2021>.
38. Dean L. Peanut protein-processes and applications. A review. *J Nutr Food Sci* 2021;44:031.
39. Bordin K, Sawada MM, Rodrigues CEC, da Fonseca CR, Oliveira CAF. Incidence of aflatoxins in oil seeds and possible transfer to oil: a review. *Food Eng Rev* 2014;6:20–8.
40. Ma X, Wang W, Chen X, Xia Y, Wu S, Duan N, et al. Selection, identification, and application of Aflatoxin B1 aptamer. *Eur Food Res Technol* 2014;238:919–25.
41. Abdolmaleki K, Khedri S, Alizadeh L, Javanmardi F, Oliveira CA, Khaneghah AM. The mycotoxins in edible oils: an overview of prevalence, concentration, toxicity, detection and decontamination techniques. *Trends Food Sci Technol* 2021;115:500–11.
42. Mao J, He B, Zhang L, Li P, Zhang Q, Ding X, et al. A structure identification and toxicity assessment of the degradation products of aflatoxin B1 in peanut oil under UV irradiation. *Toxins* 2016;8:332.
43. Chen L, Molla AE, Getu KM, Ma A, Wan C. Determination of aflatoxins in edible oils from China and Ethiopia using immunoaffinity column and HPLC-MS/MS. *J AOAC Int* 2019;102:149–55.
44. Deng H, Su X, Wang H. Simultaneous determination of aflatoxin B1, bisphenol A, and 4-nonylphenol in peanut oils by liquid-liquid extraction combined with solid-phase extraction and ultra-high

- performance liquid chromatography-tandem mass spectrometry. *Food Anal Methods* 2018;11:1303–11.
45. Qi N, Yu H, Yang C, Gong X, Liu Y, Zhu Y. Aflatoxin B₁ in peanut oil from western guangdong, China, during 2016–2017. *Food Addit Contam B* 2019;12:45–51.
 46. Yang L-X, Liu Y-P, Miao H, Dong B, Yang N-J, Chang F-Q, et al. Determination of aflatoxins in edible oil from markets in Hebei Province of China by liquid chromatography–tandem mass spectrometry. *Food Addit Contam B* 2011;4:244–7.
 47. Zhang K, Xu D. Application of stable isotope dilution and liquid chromatography tandem mass spectrometry for multi-mycotoxin analysis in edible oils. Oxford University Press; 2019:1651–6 pp.
 48. Higgins JP, Green S. *Cochrane handbook for systematic reviews of interventions*. John Wiley & Sons; 2011.
 49. Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gøtzsche PC, Ioannidis JP, et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. *Plos Med* 2009;6: 15–25.
 50. Kamimura H, Nishijima M, Tabata S, Yasuda K, Ushiyama H, Nishima T. Survey of mycotoxins contamination in edible oil and fate of mycotoxins during oil-refining processes. *Food hygiene saf Sci* 1986;27:59–63_1.
 51. Zhang H-X, Zhang P, Fu X-F, Zhou Y-X, Peng X-T. Rapid and sensitive detection of aflatoxin B₁, B₂, G₁ and G₂ in vegetable oils using bare Fe₃O₄ as magnetic sorbents coupled with high-performance liquid chromatography with fluorescence detection. *J Chromatogr Sci* 2020;58:678–85.
 52. Yu L, Ma F, Zhang L, Li P. Determination of aflatoxin B₁ and B₂ in vegetable oils using Fe₃O₄/rGO magnetic solid phase extraction coupled with high-performance liquid chromatography fluorescence with post-column photochemical derivatization. *Toxins* 2019;11:621.
 53. Nyaga VN, Arbyn M, Aerts M. Metaprop: a stata command to perform meta-analysis of binomial data. *Arch Publ Health* 2014; 72:1–10.
 54. Peck R, Olsen C, Devore JL. *Introduction to statistics and data analysis*. Cengage Learning; 2015.
 55. Higgins J, White IR, Anzueto-Cabrera J. Meta-analysis of skewed data: combining results reported on log-transformed or raw scales. *Stat Med* 2008;27:6072–92.
 56. Helgilibrary. Egg consumption per capita in Jordan. Available from: <https://www.helgilibrary.com/indicators/egg-consumption-per-capita/jordan/2021>.
 57. EPA. Recommended use of body weight; 2015:15–23pp. Available from: <https://www.epa.gov/sites/production/files/2013-09/documents/recommended-use-of-bw34.pdf>.
 58. Helgilibrary. Vegetable oil consumption per capita. 2017. Available from: <https://www.helgilibrary.com/indicators/vegetable-oil-consumption-per-capita/>.
 59. Colombo CA, Berton LHC, Diaz BG, Ferrari RA. Macauba: a promising tropical palm for the production of vegetable oil. *OCL* 2018;25:D108.
 60. Authority EFS. Outcome of a public consultation on the draft risk assessment of aflatoxins in food. Wiley Online Library; 2020. Report No 2397–8325.
 61. EFSA. Scientific Opinion adopted. *EFSA J* 2020;18:28–36. <https://efsa.onlinelibrary.wiley.com/doi/epdf/10.2903/j.efsa.2020.6040>.
 62. Adetunji MC, Alike OP, Awa NP, Atanda OO, Mwanza M. Microbiological quality and risk assessment for aflatoxins in groundnuts and roasted cashew nuts meant for human consumption. *J Toxicol* 2018;2018. <https://doi.org/10.1155/2018/1308748>.
 63. Fakhri Y, Rahmani J, Oliveira CAF, Franco LT, Corassin CH, Saba S, et al. Aflatoxin M₁ in human breast milk: a global systematic review, meta-analysis, and risk assessment study (Monte Carlo simulation). *Trends Food Sci Technol* 2019;88:333–42.
 64. Pirsaeheb M, Fakhri Y, Karami M, Akbarzadeh R, Safaei Z, Fatahi N, et al. Measurement of permethrin, deltamethrin and malathion pesticide residues in the wheat flour and breads and probabilistic health risk assessment: a case study in Kermanshah, Iran. *Int J Environ Anal Chem* 2019;99:1353–64.
 65. Fang L, Zhao B, Zhang R, Wu P, Zhao D, Chen J, et al. Occurrence and exposure assessment of aflatoxins in Zhejiang province, China. *Environ Toxicol Pharmacol* 2022;103847. <https://doi.org/10.1016/j.etap.2022.103847>.
 66. Fakhri Y, Sarafraz M, Nematollahi A, Ranaei V, Soleimani-Ahmadi M, Thai VN, et al. A global systematic review and meta-analysis of concentration and prevalence of mycotoxins in birds' egg. *Environ Sci Pollut Control Ser* 2021;28:59542–50.
 67. Einolghozati M, Talebi-Ghane E, Ranjbar A, Mehri F. Concentration of aflatoxins in edible vegetable oils: a systematic meta-analysis review. *Eur Food Res Technol* 2021;247:2887–97.
 68. Chen LY, Molla AE, Getu KM, Ma AD, Wan CS. Determination of aflatoxins in edible oils from China and Ethiopia using immunoaffinity column and HPLC-MS/MS. *J AOAC Int* 2019;102:149–55.
 69. Qi N, Yu H, Yang C, Gong X, Liu Y, Zhu Y. Aflatoxin B₁ in peanut oil from western guangdong, China, during 2016–2017. *Food Addit Contam Part B Surveillance* 2019;12:45–51.
 70. Bordin K, Sawada MM, da Costa Rodrigues CE, da Fonseca CR, Oliveira CAF. Incidence of aflatoxins in oil seeds and possible transfer to oil: a review. *Food Eng Rev* 2014;6:20–8.
 71. Bhat R, Reddy KRN. Challenges and issues concerning mycotoxins contamination in oil seeds and their edible oils: updates from last decade. *Food Chem* 2017;215:425–37.
 72. Wu L, Ding X, Li P, Du X, Zhou H, Bai YZ, et al. Aflatoxin contamination of peanuts at harvest in China from 2010 to 2013 and its relationship with climatic conditions. *Food Control* 2016; 60:117–23.
 73. Filbert ME, Brown DL. Aflatoxin contamination in Haitian and Kenyan peanut butter and two solutions for reducing such contamination. *J Hunger Environ Nutr* 2012;7:321–32.
 74. Idris YM, Hassan SA, Mariod AA. Physicochemical characteristics and aflatoxin levels in two types of Sudanese sesame oil. *J Am Oil Chem Soc* 2013;90:989–98.
 75. Oyedele OA, Ezekiel CN, Sulyok M, Adetunji MC, Warth B, Atanda OO, et al. Mycotoxin risk assessment for consumers of groundnut in domestic markets in Nigeria. *Int J Food Microbiol* 2017;251:24–32.
 76. Kabak B. Aflatoxins in hazelnuts and dried figs: occurrence and exposure assessment. *Food Chem* 2016;211:8–16.
 77. Williams JH, Phillips TD, Jolly PE, Stiles JK, Jolly CM, Aggarwal D. Human aflatoxicosis in developing countries: a review of toxicology, exposure, potential health consequences, and interventions. *Am J Clin Nutr* 2004;80:1106–22.
 78. Kumar P, Mahato DK, Kamle M, Mohanta TK, Kang SG. Aflatoxins: a global concern for food safety, human health and their management. *Front Microbiol* 2017;7:2170.
 79. Ismail A, Gonçalves BL, de Neeff DV, Ponzilacqua B, Coppa CF, Hintzsche H, et al. Aflatoxin in foodstuffs: occurrence and

- recent advances in decontamination. *Food Res Int* 2018;113:74–85.
80. Nabizadeh S, Shariatifar N, Shokoohi E, Shoeibi S, Gavahian M, Fakhri Y, et al. Prevalence and probabilistic health risk assessment of aflatoxins B₁, B₂, G₁, and G₂ in Iranian edible oils. *Environ Sci Pollut Control Ser* 2018;25:35562–70.
81. Qiu W-Q, Fu W-S. Contamination of aflatoxins in peanuts and peanut products from Fujian. *Chin J Health Lab Technol* 2012;22:2446–8.

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