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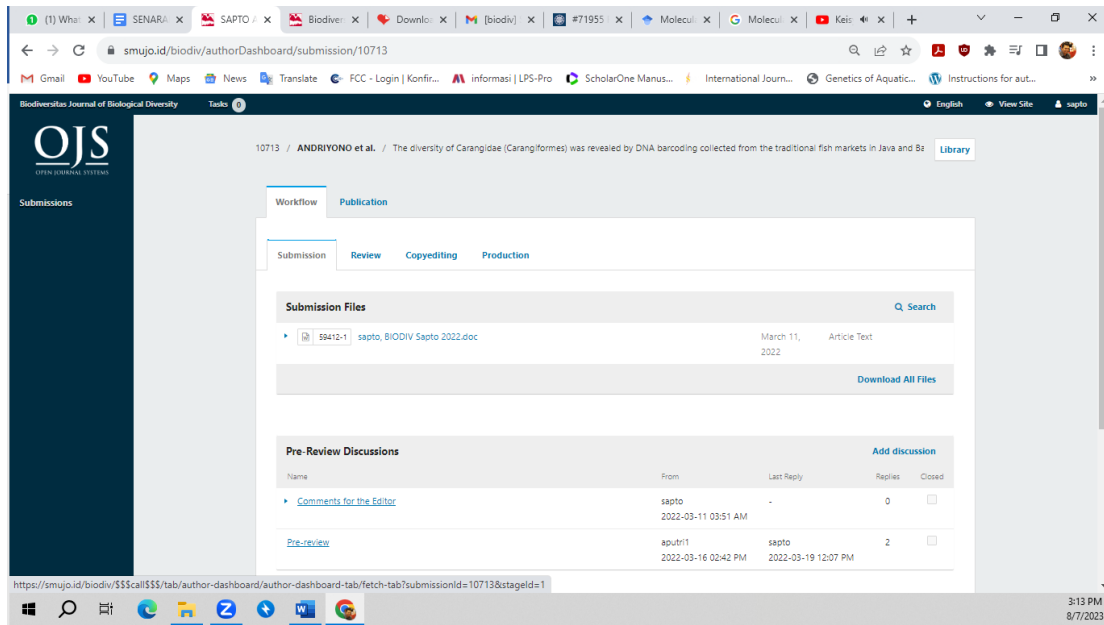
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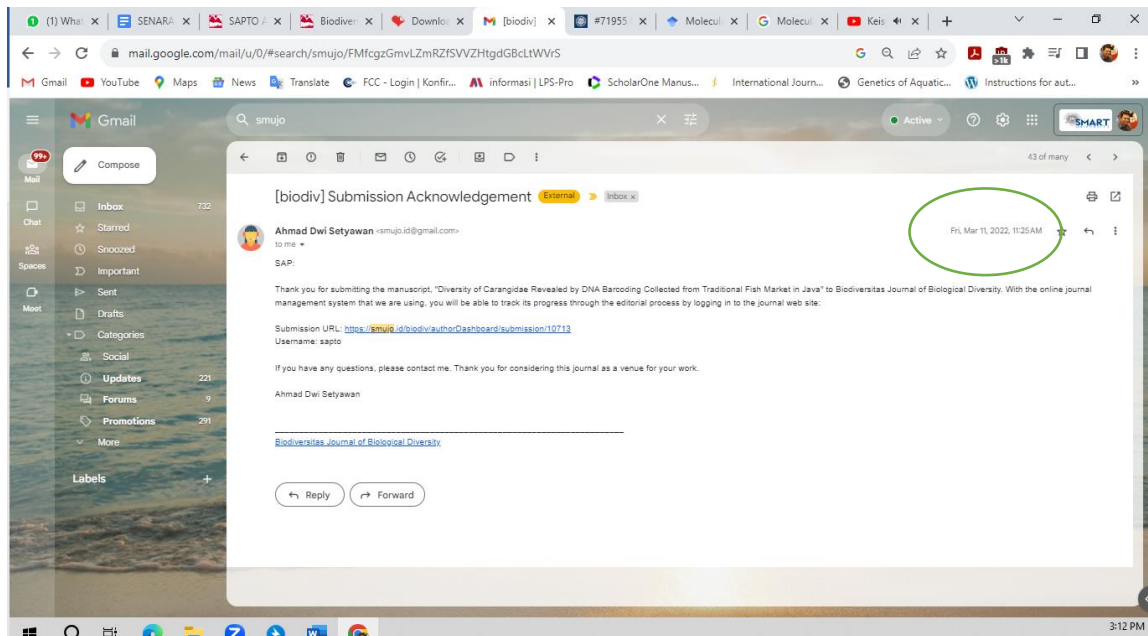
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1	Bukti Submit dan Artikel yang di submit	11 Maret 2022	Page 2,
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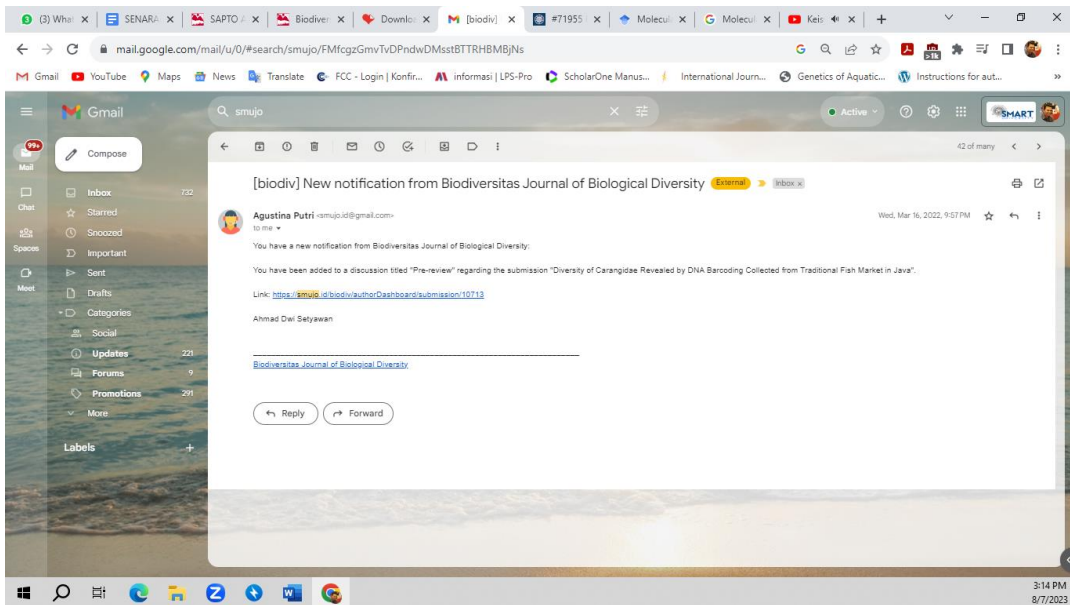
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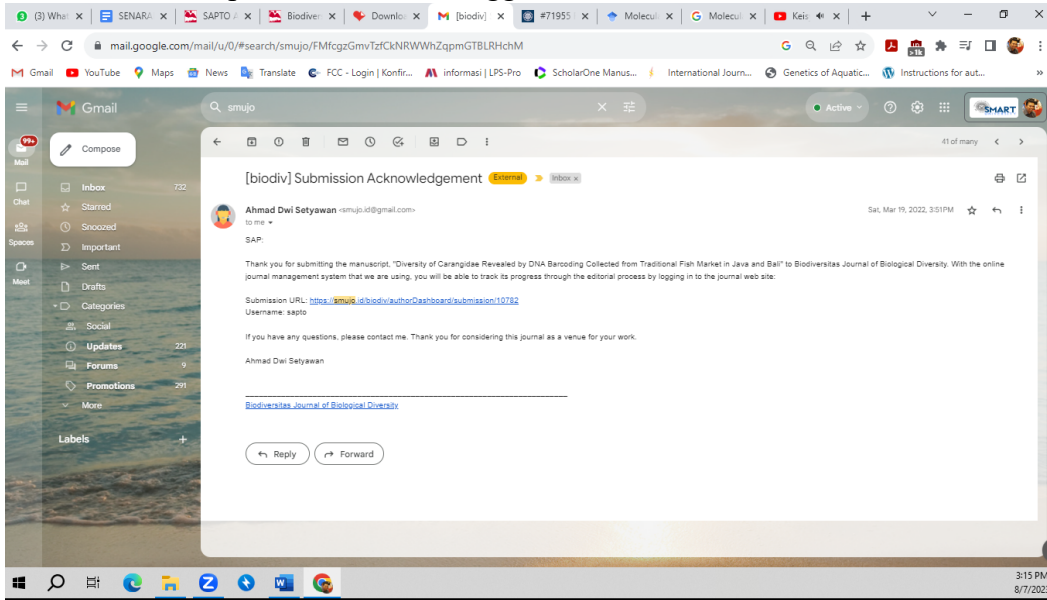
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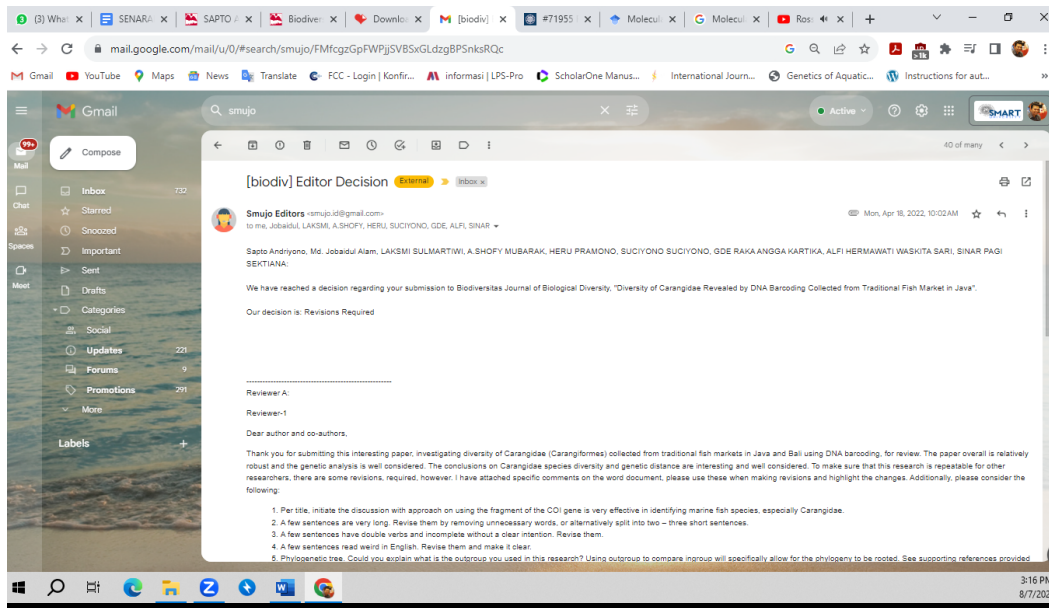
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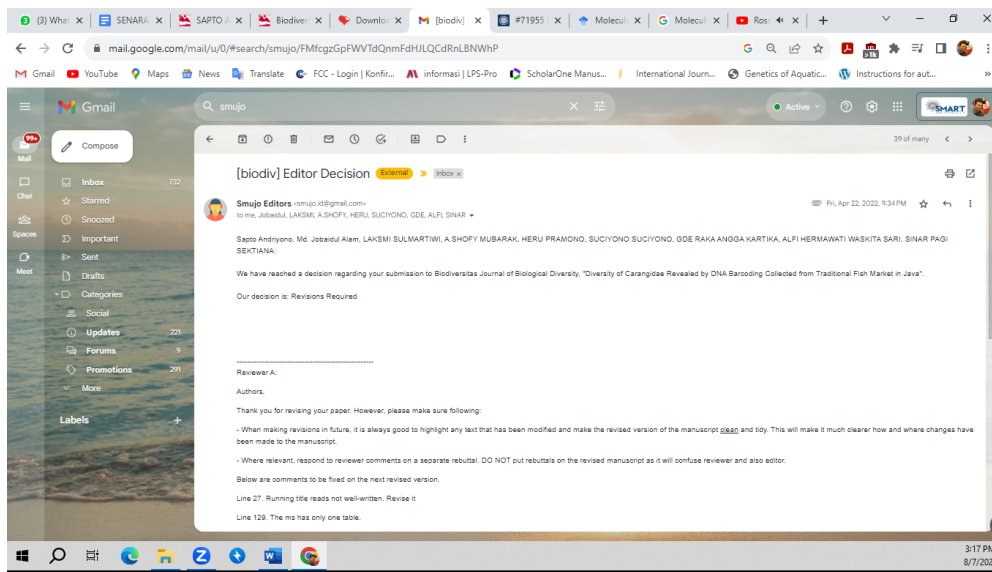


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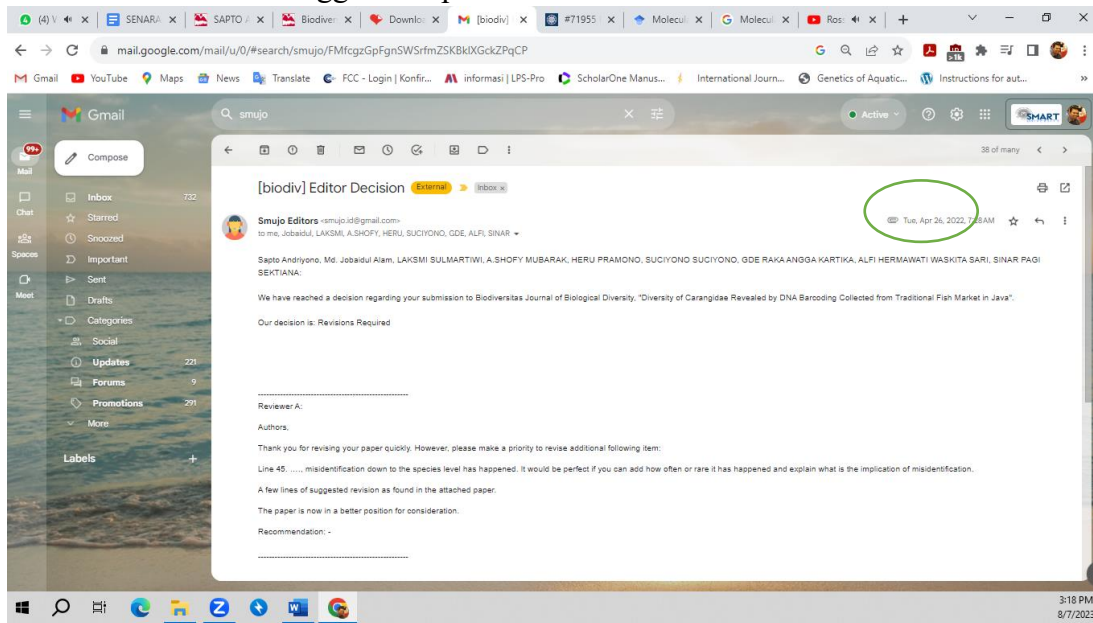


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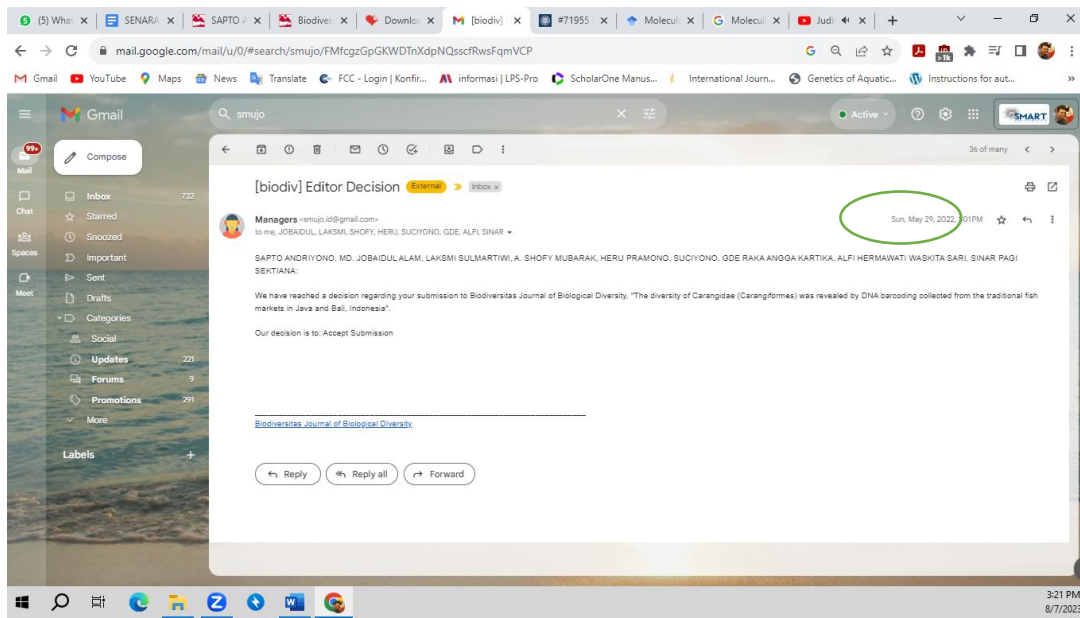
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SAPTO ANDRIYONO

Diversity of Carangidae Revealed by DNA Barcoding Collected from Traditional Fish Market in Java

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Abstract. Biodiversity has been utilised in various ways, including in fulfilling the protein needs of fish for coastal communities. For the island of Java with the largest population, the intensive fisheries in the Java Sea are sufficient to support domestic food needs. This Carangid fish diversity study in Java is the beginning to identify commercial fish in Indonesia, which have been exploited for a long time. In this study, identification was carried out molecularly in the Cytochrome c oxidase subunit I (COI) gene region with the universal primary set and found a great variety of Carangids fish species. A total of 33 fish specimens have been identified, indicating two suborder groups, Caranginae (31) and Scomberoidenae (2). The Caranginae suborder group is more dominated with the most types of which are *Megalaspis cordyla* (3), *Atule mate* (3), *Decapterus macarellus* (4). Meanwhile, the Scomberoidinae suborder is *Scomberoides commersonianus* and *Scomberoides tala*. This study also found two species that have the potential to be ciguatera poisoning agents that need to be watched out for (*Decapterus macarellus* and *Selar crumenophthalmus*).

Keywords: biodiversity, commercial, conservation, fisheries, genetic study, Java

Running title: Andriyono et al. Diversity of Carangidae

INTRODUCTION

In Indonesia, an archipelago distributed in tropical areas called the Indo-Malay Archipelago (IMA), Incredible biodiversity in marine life. This region has become recognised as one of the centres of marine biodiversity in various taxon fish, molluscs, and various other taxa (Lohman et al. 2011), located between the Pacific Ocean in the North and the Indian Ocean in the South. The island of Java, located in the southern part of Indonesia, has the potential for diversity, strongly influenced by the Indian Ocean compared to the Pacific Ocean region. However, the Java Sea's existence in the middle of the Indonesian archipelago allows the mixing of Pacific waters through the Makassar Strait gap between Kalimantan and Sulawesi Islands. The existence of the South China Sea also enables fish resources to enter the Java Sea through the Riau Islands and into the Java Sea. This considerable fishery potential causes capture fisheries in the Java Sea to be very potential, dominated by the Carangid fish.

Carangidae includes fish with varying body sizes ranging from 16 cm-250 cm, with irregular body shapes (elongated, fusiform and compressed) (Randall 1995). Fish from this family are commonly found in fisheries commodities such as scads, jacks, trevallies, pompanos, and rainbow runners. As a source of food and animal protein, Carangids have become essential raw material in Southeast Asia's processing industry (Mohsin and Ambak 1996). Even though intensive exploitation continues, taxonomy and scientific naming is still confused (Laroche 1984) because of the similarities in morphology, size and colour pattern (Lakra et al. 2009), which causes ambiguity in this family taxonomy (Froese 2009). In the life cycle, the juvenile stage to adulthood sometimes undergoes significant morphological changes and pigmentation (Bohlke and Chaplin 1993), which results in misidentification down to the species level. It is found in juvenile *Alectis ciliary* long filament but will shorten and eventually disappear as it grows to maturity (Randall et al. 1997).

This DNA barcode method uses a reasonably short but standardised area of mitochondrial DNA (Hebert et al. 2003b) with an identification accuracy up to the species level (Aquilino et al. 2011), which is high enough to approach 100% and has been widely accepted to date. Thus, the identification based on morphological needs to be complemented by other identification methods to ensure identification accuracy. The method of identification that is currently being developed

49 molecularly allows to reduce errors in identification. This identification is referred to as barcoding DNA. This DNA
50 barcode method uses a reasonably short but standardised area of mitochondrial DNA (Hebert et al. 2003b) with an
51 identification accuracy up to the species level (Aquilino et al. 2011) high enough to be close to 100%, and to date has
52 been widely accepted (Hebert et al. 2003a). Open-access databases are continually being developed in connection with
53 the emergence of misidentification through the molecular approach (Ratnasingham and Hebert 2007). Simplifying the
54 identification process through molecular methods is relatively easy and allows finding new taxa (Hajibabaei et al. 2007).

55 In this study, we carried out tests on Carangidae fish from Java's northern and southern regions and analysed the COI
56 sequence's genetic diversity. This analysis is expected to be important information for sustainable management of
57 Carangidae fish resources because most of them are not reported and are not well identified.

58

MATERIALS AND METHODS

59 Sampling site

60 Thirty-three fish samples (**Table 1**) were collected from the five traditional fish markets around Jawa Island during
61 July 2019. In the northern part of Java, samples were obtained from the Banten (6°0'50.00'S-106°10'21.00" E), Gresik
62 (6°52'56.65'S-112°12'15.87" E), and Pekalongan (6°51'32,10"S 109°41'09,52"E). While Southern Java was represented
63 samples from Malang (8°26'06.65'S-112°40'55.31"), the Banyuwangi (8°12'07.52'S-114°23'07.18" E), and Denpasar
64 (8°45'23.00'S-115°10'05.68"E). Morphologically identification was conducted according to the guideline from FAO
65 (Heemstra 1993), and species confirmation has been carried out with molecular identification carried out in this study
66 using the COI gene region. No specific permit was required for this study because collection from the local traditional
67 fish market was dead upon purchasing. All samples have been photographed was applied to every single specimen using
68 a digital camera.

69

70 DNA extraction and PCR Condition

71 All samples were preserved in 90% ethanol for further laboratory experiments. In preparation for the genomic
72 DNA extraction process, cutting the tissue in each specimen was done aseptically (around 0.5 cm) and immediately
73 washed to remove ethanol under running water. After the washing process is carried out, the sample is put in a microtube
74 containing 6X lysis buffer and mixed with the TissueLyser II (Qiagen). The use of Accuprep® Genomic DNA extraction
75 Kit (Bioneer) was carried out according to the Kit protocol and followed by measuring the concentration of the extracted
76 with nanoDrop (ThermoFisher Scientific D1000). The extract is used in the application process by the PCR or stored at -
77 70oC for further analysis.

78 The amplification process of genomic DNA from samples was carried out using a universal primer set, namely FISH-
79 BCL (5'-TCA ACY AAT CAY AAA GAT ATY GGC AC-3 ') and FISH-BCH (5'-TAA ACT TCA GGG TGA CCA
80 AAA AAT CA -3 ') (Baldwin et al. 2009, Handy et al. 2011). The target DNA amplified is the cytochrome c oxidase I
81 (COI) region, about 600 bp in length. The volume of PCR mixture is 20µL consisting of 11.2 µL ultra-pure water, 1 µL
82 forward and reverse primer (0.5 µM), 0.2 µL Ex Taq DNA polymerase (TaKaRa, Japan), 2 µL 10X ExTag Buffer, 2 µL
83 dNTPs (1 µM, TaKaRa, Japan), and 2 µL genomic DNA as a template. The PCR condition was carried out under the
84 following setting: 95oC for 5 min in initial denaturation, followed by denaturation at 95oC for 30 s in 40 cycles, 50oC for
85 30 s in annealing, and 72oC for 45 s in extension step, and a final extension at 72oC for 5 min. After obtaining the
86 desired band on the electrophoresis gel (1.5% agarose), the PCR products were purified with the AccuPrep® Gel
87 purification kit (Bioneer, Korea).

88

89 Data Analysis and Phylogenetic analysis

90 All sequences are then aligned with the help of Mega7 software, including the sequences obtained from the GenBank
91 database for comparison. The pairwise evolutionary distance among the species is determined by the Kimura 2-Parameter
92 method. The Neighbor-joining (NJ) tree was constructed, and 1000 bootstrap analysis was carried out by Mega7 (Kumar et
93 al. 2016).

94

95

RESULTS AND DISCUSSION

96 Results

97 A total of 33 successful identification samples consisted of 2 sub-families within Carangidae, Caranginae (31) and
98 Scomberoidenae (2). In this study, the Caranginae more dominated the general catches of fisherman, including
99 *Megalaspis cordyla* (3), *Atule mate* (3), *Decapterus macarelus* (4). Meanwhile, local fishers' common fish caught in Java
100 and Bali and commercial fish, which benefit domestic consumers. In general, the types of fish collected in the Carangids
101 group have low prices and become food sources of protein for the community.

102

103 **Genetic distance**

104 Genetic distance analysis was carried out using Mega7 compared it to the GenBank database. Several sequences have
105 unique, potentially as a haplotype that Indonesia has founded. Within subfamily Scomberoidinae, *Scomberoides tala* is
106 similar to the Indo-Malay (JX261091) than *Scomberoides commersonianus* has a very close genetic distance with the
107 Indo-Malay sequence (JX261017) is 0.0019. Beside, within sub-family of Caranginae, we also found potentially haplotype
108 Indonesia in *Decapterus muruadsi* (0.0079), *Megalaspis cordyla* (0.0019), *Carangoides armatus* (0.0019), *Atropos*
109 *Atropos* (0.0019), *Alepes vari* (0.0019), *Atule mate* (0.0059), Selar boops (0.019), *Selar crumenophthalmus* (0.0039),
110 *Carangoides malabaricus* (0.0019), *Carangoides chrysophrys* (0.0019), and *Selaroides leptolepis* (0.0019).
111

112 **Phylogenetic reconstruction**

113 In the phylogenetic tree produced, two clades have been formed: the subfamily Cranginae and Scomberoidinae
114 (**Figure 1**). Morphologically, Scomberoidinae is closer to the Family of Scombridae with fusiform in body shape (**Figure**
115 **2**), but its body was flattened compared to the torpedo-like Scombridae. Scomberoidinae, in etymology, have a meaning is
116 similar to tunny or mackerel. In this study, *Scomberoides tala* and *Scomberoides commersonianus* were successfully
117 identified from Banyuwangi and Gresik, respectively.
118

119 **Carangidae status in IUCN and CITES**

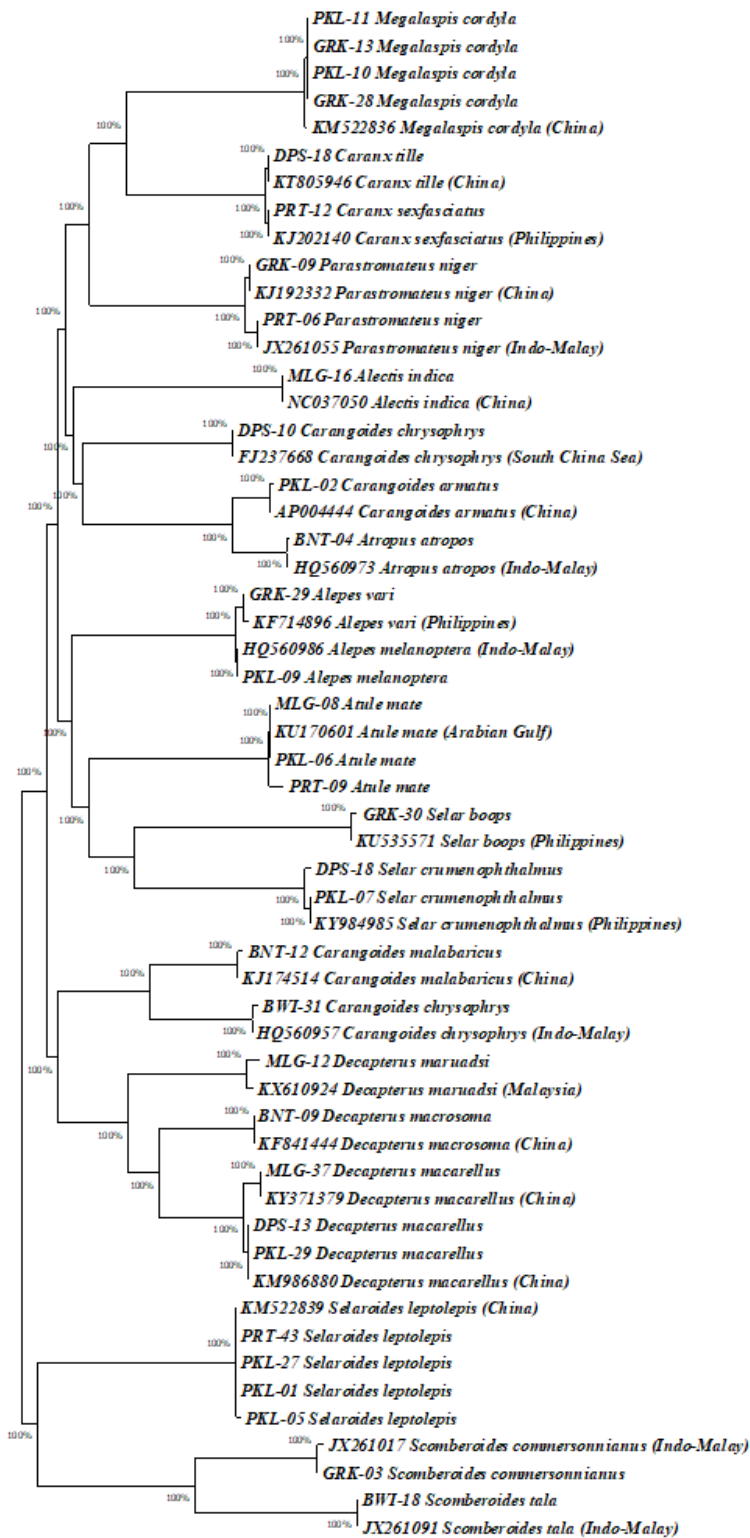
120 Almost all Carangidae species in this study have the status of Least Concern (LC), and the international trade
121 (CITES) status is not evaluated (**Table 2**). From several reports, the fishing of Carangids species is carried out
122 intensively, and most of them turn it into fish for processed fishery products such as surimi or only as raw material for
123 fish meal. Research on fish catches in the Madura Strait found that Carangidae fish catch was 8% (Purwangka and
124 Mubarak 2018), while other studies found Carangidae species of 38.51% of the catch (Khatami et al. 2018). Apart from
125 Java, the Carangidae species that traditional fishers catch in Kalimantan also entirely dominate around 16.67% (Alfian et
126 al. 2020).
127

128 **Discussion**

129 Shallow marine fishery resources, especially in the Java Sea, have provided benefits for meeting animal protein needs
130 in the Java Island region, the most populous island area in Indonesia. Indonesian capture fisheries production in Indonesia
131 until 2017 reached 6,603,630.58 tonnes / year with a value of 185,798,801 US \$ (Nainggolan et al. 2019). From several
132 ports on Java, the Java sea fishery shows quite intensive fishery activity with a fairly large catch value. There are potential
133 ports with very diverse catches along the north coast of the island of Java. However, studies conducted by Java sea capture
134 fisheries in the Central Java region show that the Carangids group such as *Decapterus* spp. is a caught fish that dominates,
135 especially during the peak season in September-November each year (Chodrijah and Hariati 2017).
136

137 Carangidae fishery is a fishery potential that is the mainstay of fulfilling domestic consumption compared to tuna
138 fisheries which are exported products (Yusuf et al. 2018). Carangidae commodity, processed products are an effort to
139 increase the product's added value besides being sold in fresh fish. One of the most famous processed products with
140 Carangids fish raw materials is the scavenging with the main fish species in steam fish (pindang) from the genus
141 *Decapterus* spp and *Atule* spp (Lubis et al. 2019).
142

143 The results of surveys in some fish landing areas in the north coast of Java and the south coast illustrate that the
144 Javanese marine fishery makes Carangids a relatively high catch (Table 1). On the other hand, in the southern region of
145 Java, it is the result of large pelagic fisheries such as tuna and skipjack (Firdaus 2019). Previous research has also
146 confirmed that the Java sea fishery with the Carangids commodity is becoming quite popular (Chodrijah and Hariati 2017).
147 Meanwhile, the Indian sea fishery or the southern coast of Java, tuna fisheries are the mainstay and are the mainstay in
148 Malang (Wiadnya et al. 2018) and Pelabuhan ratu (Mertha et al. 2017), which is well known as a centre for tuna fish
149 catches in Southern Java (Nurani et al. 2017). Besides, Bali Island, directly facing the Indian Ocean, also has a potential
tuna landing centre in Benoa (Ahmad et al. 2019).



Caranginae

Scomberoidinae

0.020

Figure 1. Phylogenetic tree of Carangidae using the neighbour-joining algorithm by Mega7

150
 151
 152
 153



154
155 **Figure 2.** Scomberoidinae: *Scomberoides commersonnianus* from Gresik and *Scomberoides tala* from Banyuwangi
156 (black line: 1 cm).

157
158 **CONCLUSIONS**

159
160 This research successfully identified the diversity of Carangid fish landed in several fishery centres in Java. Of the 33
161 carangids fish, the most extensive composition obtained from these locations was *Megalaspis cordyla* (3), *Atule mate* (3),
162 *Decapterus macarellus* (4). Generally, these fish are consumed locally to meet people's protein needs in coastal areas of
163 Java. Phylogenetic analysis shows that the suborder Caranginae and Scomberoidinae are separated. Fish in the Caranginae
164 suborder dominate compared to other sub-orders. In this study, we identified that *Decapterus macarellus* and *Selar*
165 *crumenophthalmus* had been reported to be the causes of ciguatera poisoning that need to be watched out for and get
166 attention in the processing post-harvest processes.
167
168

169 **ACKNOWLEDGEMENTS**

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172 supported the collection of samples throughout northern and southern Java, including Bali samples.
173

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234

Tabel 2. IUCN and CITES status of all Carangids species

No.	Species	Sub-Family	Habitat distribution	Common Name	IUCN Red List Status	CITES	Threat to humans
1	<i>Alectis indica</i>	Caranginae	Indo-Pacific	Indian threadfish	LC	NE	Harmless
2	<i>A. melanoptera</i>	Caranginae	Indo-Pacific	Blackfin scad	LC	NE	Harmless
3	<i>A. vari</i>	Caranginae	Indo-West Pacific	Herring scad	LC	NE	Harmless
4	<i>Atropus atropus</i>	Caranginae	Indo-West Pacific	Cleftbelly trevally	LC	NE	Harmless
5	<i>Atule mate</i>	Caranginae	Indo-Pacific	Yellowtail scad	LC	NE	Harmless
6	<i>C. armatus</i>	Caranginae	Indo-West Pacific	Longfin trevally	LC	NE	Harmless
7	<i>C. chrysophrys</i>	Caranginae	Indo-Pacific	Longnose trevally	LC	NE	Harmless
8	<i>C. malabaricus</i>	Caranginae	Indo-West Pacific	Malabar trevally	LC	NE	Harmless
9	<i>Caranx sexfasciatus</i>	Caranginae	Indo-Pacific	Bigeye trevally	LC	NE	Harmless
10	<i>Caranx tille</i>	Caranginae	Indo-West Pacific	Tille trevally	LC	NE	Harmless
11	<i>Decapterus macarellus</i>	Caranginae	Circumglobal.	Mackerel scad	LC	NE	Ciguatera poisoning
12	<i>D. macrosoma</i>	Caranginae	Indo-Pacific and Southeast Atlantic	Shortfin scad	LC	NE	Harmless
13	<i>D. maruadsi</i>	Caranginae	Indo-West Pacific	Japanese scad	LC	NE	Harmless
14	<i>Megalaspis cordyla</i>	Caranginae	Indo-West Pacific	Torpedo scad	LC	NE	Harmless
15	<i>Parastromateus niger</i>	Caranginae	Indo-West Pacific	Black pomfret	LC	NE	Harmless
16	<i>Selar boops</i>	Caranginae	Pacific Ocean	Oxeye scad	LC	NE	Harmless
17	<i>S. crumenophthalmus</i>	Caranginae	Circumtropical	Bigeye scad	LC	NE	Ciguatera poisoning
18	<i>Selaroides leptolepis</i>	Caranginae	Indo-West Pacific	Yellow stripe scad	LC	NE	Harmless
19	<i>Scomberoides tala</i>	Scomberoidinae	Indo-West Pacific	Barred queenfish	LC	NE	Harmless
20	<i>S. commersonianus</i>	Scomberoidinae	Indo-West Pacific	Talang queenfish	LC	NE	Harmless

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1 **The Diversity of Carangidae (Carangiformes) was revealed by DNA**
 2 **barcoding collected from the traditional fish markets in Java and Bali,**
 3 **Indonesia.**

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15 **Abstract.** Biodiversity has been utilised in various ways, including in fulfilling the protein needs of fish for coastal communities. For the island of Java, with the largest population in Indonesia, the intensive fisheries in the Java Sea are sufficient to support domestic food needs. This Carangid fish diversity study in Java is the beginning to identify commercial fish in Indonesia, which have been exploited for a long time. In this study, identification was carried out molecularly in the Cytochrome c oxidase subunit I (COI) gene region with the universal primary set and found a great variety of Carangids fish species. Thirty-three fish specimens have been identified, indicating two suborder groups, Caranginae (31) and Scomberoidenae (2). The Caranginae suborder group is more dominated with the most types of which are *Megalaspis cordyla* (3), *Atule mate* (3), and *Decapterus macarellus* (4). Meanwhile, the Scomberoidinae suborder is *Scomberoides commersonnianus* and *Scomberoides tala*. This study also found two species that have the potential to be ciguatera poisoning agents that need to be watched out for (*Decapterus macarellus* and *Selar crumenophthalmus*). Food safety in the fisheries sector has received considerable attention for a long time. Fishery commodities in highly polluted habitats are among the chains in increasing heavy metals concentration and the other residual chemical compounds that may impact human health.

26 **Keywords:** biodiversity, commercial, conservation, fisheries, genetic study

27 **Running title:** The Diversity of Carangidae (Carangiformes)

28 **INTRODUCTION**

29 In Indonesia, an archipelago is distributed in tropical areas called the Indo-Malay Archipelago (IMA), with incredible biodiversity in marine life. This region has been recognised as one of the centres of marine biodiversity in various taxon fish, molluscs, and various other taxa (Lohman et al. 2011), located between the Pacific Ocean in the North and the Indian Ocean in the South. The island of Java, located in the southern part of Indonesia, has the potential for diversity, strongly influenced by the Indian Ocean compared to the Pacific Ocean region. However, the Java Sea's existence in the middle of the Indonesian archipelago allows the mixing of Pacific waters through the Makassar Strait gap between Kalimantan and Sulawesi Islands. The presence of the South China Sea also enables fish resources to enter the Java Sea through the Riau Islands and into the Java Sea. This considerable fishery potential causes capture fisheries in the Java Sea to be very potential, dominated by the Carangid fish.

38 The Carangidae family (order Carangiformes) includes fish with varying body sizes ranging from 16 cm-250 cm, with irregular body shapes (elongated, fusiform, and compressed) (Randall 1995). Fish from this family are commonly found in fisheries commodities such as scads, jacks, trevallies, pompanos, and rainbow runners. In addition, Carangids have become essential raw materials in Southeast Asia's processing industry as a food and animal protein source. Even though intensive exploitation continues, taxonomy and scientific naming is still confused because of the similarities in morphology, size, and colour pattern, which causes ambiguity in this family taxonomy (FROESE 2009). In addition, the juvenile stage to adulthood sometimes undergoes significant morphological changes and pigmentation in the life cycle. As a result, misidentification down to the species level has happened. For example, it is found in juvenile *Alectis ciliaris* long filament but will shorten and eventually disappear as it grows to maturity (Randall et al. 1997).

47 This DNA barcode method uses a reasonably short but standardised area of mitochondrial DNA with an identification accuracy up to the species level (Aquilino et al. 2011), which is high enough to approach 100% and has been widely

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49 accepted to date. Thus, the identification based on morphological needs to be complemented by other identification
50 methods to ensure identification accuracy. The form of identification currently being developed molecularly reduces
51 errors in identification. This identification is referred to as barcoding DNA. This DNA barcode method uses a reasonably
52 short but standardised area of mitochondrial DNA. That region is Cytochrome c Oxidase subunit I (COI). This gene
53 region has been widely used for a molecular approach-based identification with accuracy up to the species level high
54 enough to be close to 100%. DNA barcoding has been globally accepted (Hebert et al. 2003). Open-access databases are
55 continually being developed in connection with the emergence of misidentification through the molecular approach
56 (Ratnasingham and Hebert 2007). Simplifying the identification process through molecular methods is relatively easy
57 and allows for finding new taxa (Hajibabaei et al. 2007).
58 This study carried out tests on Carangidae fish from Java's northern and southern regions and analysed the COI
59 sequence's genetic diversity. This analysis is essential for the sustainable management of Carangidae fish resources
60 because most of them are not reported and are not well identified.

61 MATERIALS AND METHODS

62 Sampling site

63 Thirty-three fish samples (Table 1) were collected from the five traditional fish markets around Jawa Island in July
64 2019. In the northern part of Java, samples were obtained from the Banten-BNT (6°0'50.00"S-106°10'21.00"-E), Gresik-
65 GRK (6°52'56.65"S-112°12'15.87" E), and Pekalongan-PKL (6°51'32.10"S 109°41'09.52"E). While Southern Java was
66 represented samples from Pelabuhanratu-PRT (6°59'20.92"S 106°32'29.91"E), Malang-MLG (8°26'06.65"S-
67 112°40'55.31"E), the Banyuwangi-BWI (8°12'07.52"S-114°23'07.18"E), and Denpasar-DPS (8°45'23.00"S-
68 115°10'05.68"E). Morphological identification was conducted according to the guideline from FAO (Heemstra 1993),
69 and species confirmation was carried out with molecular identification carried out in this study using the COI gene
70 region. No specific permit was required for this study because collection from the local traditional fish market was dead
71 upon purchasing. All samples were photographed for every single specimen using a digital camera.

72 DNA extraction and PCR Condition

73 All samples were preserved in 90% ethanol for further laboratory experiments. In preparation for the genomic
74 DNA extraction process, cutting the tissue in each specimen was done aseptically (around 0.5 cm) and immediately
75 washed to remove ethanol under running water. After washing process was carried out, the sample was put in a
76 microtube containing 6X lysis buffer and mixed with the TissueLyser II (Qiagen). The use of Accuprep® Genomic DNA
77 extraction Kit (Bioneer) was carried out according to the Kit protocol and followed by measuring the concentration of the
78 extracted with nanoDrop (ThermoFisher Scientific D1000). The extract was used in the application process by the PCR or
79 stored at -70oC for further analysis.

80 The amplification process of genomic DNA from samples was carried out using a universal primer set, namely FISH-
81 BCL (5'-TCA ACY AAT CAY AAA GAT ATY GGC AC-3 ') and FISH-BCH (5'-TAA ACT TCA GGG TGA CCA
82 AAA AAT CA -3 ') (Baldwin et al. 2009, Handy et al. 2011). The target DNA amplified is the cytochrome c oxidase I
83 (COI) region, about 600 bp in length. The volume of the PCR mixture is 20µL consisting of 11.2 µL ultra-pure water, 1
84 µL forward and reverse primer (0.5 µM), 0.2 µL Ex Taq DNA polymerase (TaKaRa, Japan), 2 µL 10X ExTag Buffer, 2
85 µL dNTPs (1 µM, TaKaRa, Japan), and 2 µL genomic DNA as a template. The PCR condition was carried out under the
86 following setting: 95°C for 5 min in initial denaturation, followed by denaturation at 95°C for 30 s in 40 cycles, 50°C for
87 30 s in annealing, and 72°C for 45 s in extension step, and a final extension at 72°C for 5 min. After obtaining the desired
88 band on the electrophoresis gel (1.5% agarose), the PCR products were purified with the AccuPrep® Gel purification kit
89 (Bioneer, Korea).

90 Data Analysis and Phylogenetic analysis

91 All sequences are then aligned with the help of Mega7 software, including the sequences obtained from the GenBank
92 database for comparison. The pairwise evolutionary distance among the species is determined by the Kimura 2-Parameter
93 method. The Neighbor-joining (NJ) tree was constructed, and 1000 bootstrap analysis was carried out by Mega7 (Kumar et
94 al. 2016).

97 RESULTS AND DISCUSSION

98 Results

99 Thirty-three successful identification samples consisted of 2 sub-families within Carangidae, Caranginae (31) and
100 Scomberoidenae (2). In this study, the Caranginae dominated the general catches of fisherman, including *Megalaspis*
101 *cordyla* (3), *Atule mate* (3), and *Decapterus macarellus* (4). Meanwhile, local fishers' common fish caught in Java and

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Bali and commercial fish, which benefit domestic consumers. In general, the types of fish collected in the Carangids group have low prices and become food sources of protein for the community.

Genetic distance

Genetic distance analysis was carried out using Mega7 compared to the GenBank database. Several sequences have unique, potentially as a haplotype that Indonesia has founded. Within subfamily Scomberoidinae, *Scomberoides tala* is similar to the Indo-Malay (JX261091) than *Scomberoides commersonianus* has a very close genetic distance with the Indo-Malay sequence (JX261017) is 0.0019. The smaller the genetic distance, the closer the similarity and may indicate that a particular species may have similar origins (Dwifajri et al, 2022; Tapilatu et al, 2021). In addition, within sub-family of Caranginae, we also found potentially haplotype Indonesia in *Decapterus muruadsi* (0.0079), *Megalaspis cordyla* (0.0019), *Carangoides armatus* (0.0019), *Atropos Atropos* (0.0019), *Alepes vari* (0.0019), *Atule mate* (0.0059), *Selar boops* (0.019), *Selar crumenophthalmus* (0.0039), *Carangoides malabaricus* (0.0019), *Carangoides chrysophrys* (0.0019), and *Selaroides leptolepis* (0.0019).

Phylogenetic reconstruction

In the phylogenetic tree produced, two clades have been formed: the subfamily Cranginae and Scomberoidinae (Figure 1). Morphologically, Scomberoidinae is closer to the Family of Scombridae with fusiform in body shape (Figure 2), but its body was flattened compared to the torpedo-like Scombridae. Scomberoidinae, in etymology, has a meaning similar to tunny or mackerel. In this study, *Scomberoides tala* and *Scomberoides commersonianus* were successfully identified from Banyuwangi and Gresik, respectively.

Carangidae status in IUCN and CITES

Almost all Carangidae species in this study have the status of Least Concern (LC), and the international trade (CITES) status is not evaluated (Table 2). From several reports, the fishing of Carangids species is carried out intensively, and most of them turn it into fish for processed fishery products such as surimi or only as raw material for fish meal. For example, research on fish catches in the Madura Strait found that Carangidae fish catch was 8% (Purwangka and Mubarak 2018), while other studies found Carangidae species of 38.51% of the catch (Khatami et al. 2018). Apart from Java, the Carangidae species that traditional fishers catch in Kalimantan also entirely dominate around 16.67% (Alfian et al. 2020).

Discussion

Shallow marine fishery resources, especially in the Java Sea, have provided benefits for meeting animal protein needs in the Java Island region, the most populous island area in Indonesia. Indonesian capture fisheries production in Indonesia until 2017 reached 6,603,630.58 tonnes/year with a value of USD 185,798,801-US\$ (Nainggolan et al. 2019). From several ports on Java, the Java sea fishery shows quite intensive fishery activity with a fairly large catch value. There are potential ports with very diverse catches along the north coast of the island of Java. However, studies conducted by Java sea capture fisheries in the Central Java region show that the Carangids group, such as *Decapterus* spp., is a caught fish species that dominates, especially during the peak season in September-November each year (Chodrijah and Hariati 2017). Another study stated that as a migratory pelagic fish, *Decapterus* spp. into the waters of the Java Sea from the Indian Ocean. The migration occurred in the western season (January-March) and increased during the southeast monsoon (June-July), including two species, *Decapterus macrosoma* and *Decapterus ruselli* (Prasetyo et al. 2018).

Carangidae fishery is a fishery potential that is the mainstay of fulfilling domestic consumption compared to tuna fisheries which are exported products (Yusuf et al. 2018). Carangidae commodity, processed products are an effort to increase the product's added value besides being sold in fresh fish. One of the most famous processed products with Carangids fish raw materials is the scavenging with the main fish species in steam fish (pindang) from the genus *Decapterus* spp and *Atule* spp (Lubis et al. 2019).

The surveys in some fish landing areas on the north coast of Java and the south coast illustrate that the Javanese marine fishery makes Carangids a relatively high catch (Table 1). On the other hand, in the southern region of Java, it is the result of large pelagic fisheries such as tuna and skipjack (Firdaus 2019). Previous research has also confirmed that the Java sea fishery with the Carangids commodity is becoming quite popular (Chodrijah and Hariati 2017). Meanwhile, in the Indian sea fishery or the southern coast of Java, tuna fisheries are the mainstay and are the mainstay in Malang (Wiadnya et al. 2018). Pelabuhan ratu (Mertha et al. 2017) is well known as a centre for tuna fish catches in Southern Java (Nurani et al. 2017). Besides, Bali Island, directly facing the Indian Ocean, also has a potential tuna landing centre in Benoa (Ahmad et al. 2019).

However, Carangids fish's diversity has provided benefits even though, economically, it is only developed into a domestic product with traditional processing technologies such as dried salted fish and pindang. Phylogenetic tree analysis shows that the carangid diversity caught in the waters around Java is mainly in the Carangidae suborder group. However, a small proportion is included in the Scomberoidinae suborder group. In the Caranginae group, *Megalaspis cordyla* and *Selaroides leptolepis* dominate Java's northern region (Gresik and Pekalongan). In this phylogenetics, two distinct clade sub-orders between the Caranginae and Scomberoidinae sub-orders (Figure 1).

162 All carangid fish status in the IUCN Red list status and CITES classify all species in this order into the Least Concern
163 (LC) category and Not Evaluated (NE). Attention and regulation are needed in the management of capture fisheries in
164 particular. For instance, the northern part of Java reported having experienced overfishing (Sadhotomo and Atmadja
165 2016). Another interesting point is that apart from being an essential source of animal protein for coastal communities,
166 two types of fish have been reported to be the causes of Ciguatera poisoning that need to be watched out for in
167 *Decapterus macarellus* (Lange et al. 1992), and *Selar crumenophthalmus* (Rongo and van Woesik 2012). The type of
168 *Decapterus macarellus*, suspected of containing poison is smoked fish imported from the Philippines. The possibility of
169 smoked fish meat has palytoxin, which causes ciguatera poisoning (Kodama et al. 1989). Ciguatera poisoning is a form
170 of ichthyo-sarcotomism, which means the content of some toxin in that body. This condition occurs when marine fishes
171 inadvertently ingest particular dinoflagellates that produce ciguatoxins (Rongo and van Woesik 2012). Until now, no
172 identified reports about ciguatera poisoning in Indonesia. Food safety in the fisheries sector has received considerable
173 attention for a long time. Fishery commodities that harvested from highly polluted habitats are among the chains in
174 increasing heavy metals concentration that may impact humans (REF?). Furthermore, fish that live in waters with a
175 reasonably complex food chain, allowing food to enter the fish's body, would trigger the cause of ciguatera that has been
176 reported in several other countries outside Indonesia (Chan 2016). Although there have been no reports of ciguatera cases
177 in Indonesia (including Brunei, Cambodia, Myanmar, and North Korea) (Chan 2015b), there needs to be serious attention
178 to Indonesian fishery products, which are also exported commodities. A report states that Indonesian fishery products
179 are reported to cause ciguatera from snapper fish on the commercial market in Germany (Friedemann 2019). However, only
180 China (Chan 2015a) and Hongkong (Chan 2014a) received reports of ciguatera due to fishery products on hump-head
181 wrasse (Chan 2013) and tiger grouper (Chan 2014b), which caused some people to receive medical care.

182 183 **Sub-Family Caranginae**

184 In this study, the sub-family Caranginae dominates with 31 species identified from 31 species (94%), while the remaining
185 groups are Scomberoidinae (4%). In the Caranginae group, it can be found both in the Java Sea and the Indian Ocean. In
186 the Java Sea area, this study found the types of *Megalaspis cordyla* (**GRK** and **PKL**), *Alapes vari* (GRK), *Alepes*
187 *melanoptera* (**PKL**), *Selar boops* (**GRK**), *Carangoides malabaricus* (**BNT**), and *Selaroides leptolepis* (**PKL** and **PRT**).
188 Based on the phylogenetic tree (Figure 1), the species *Megalaspis cordyla* from Gresik and Pekalongan has close related
189 and probably in the same population and share a habitat in Jawa Sea (Maskur et al. 2020). However, the Indian Ocean and
190 Java Sea regions also share other types of Caranginae, such as *Decapterus macarellus* (**DPS**, **PKL** and **MLG**), and
191 *Decapterus macrosoma* (**BNT**), *Decapterus muruadi* (**MLG**), *Carangoides chrysophrys* (**BWI**) and *Carangoides*
192 *malabaricus* (**BNT**). In brief, the two species of *Decapterus* (*D. macarellus* and *D. Macrosoma*) are pretty difficult to
193 distinguish morphologically. The primary and key characteristic in the scattered scales on the lineal lateralis part. Research
194 on the morphological and molecular characteristics of these two species of *Decapterus* has been carried out clearly (Zhang
195 et al. 2020). These types of fish have become one of the targets of artisanal fisheries in Java, both in the southern and
196 northern regions including Bali. Research in the Java Sea waters states that several fish species, such as small pelagic,
197 dominate the catch, including the type of *Selaroides leptolepis*, *Selar crumenophthalmus*, *Dussumieria acuta*,
198 *Rastrellinger branchysoma*, and *Atule mate* (Khatami et al. 2019).

199 Many studies on artisanal fisheries in Java have been carried out, and *Selaroides* (Wijayanto et al. 2019a, Wijayanto et
200 al. 2019b) is one of the catch targets for medium-sized nets such as the use of Danish seine fishing gear, which is
201 considered an environmentally friendly fishing gear compared to trawling (Adhawati et al., 2017a, Adhawati et al., 2017b).
202 This artisanal fishery has driven the economy and even continues today. For example, capture fisheries in Pekalongan with
203 mini purse seine fishing gear that get catches belonging to the Caranginae group, such as the type of *Decapterus* sp
204 (Maulana et al. 2017). In addition, artisanal fisheries in the northern waters of Java are also highly developed, such as in
205 Banten Barlian et al., 2020, Barlian et al., 2021), Tegal (Wijayanto et al. 2019a), Lamongan (Syamsuddin et al. 2020), dan
206 Madura (Yonvitner et al. 2021).

207 208 **Sub-Family Scomberoidinae**

209 Although this group of fish is only 4% of this study, economically, it is included in the fish group with a fairly
210 reasonable price. This fish of the Scomberoidinae group has morphological characteristics similar to the Family
211 Scombridae, which includes tuna and mackerel with a streamlined body shape to become a group of fast swimming fish. In
212 this study, *Scomberoides commersonianus* is a group of fish that is quite attractive and becomes potential raw material
213 for fishery products. The method of catching this type of fish varies considerably; the use of blue LEDs is beneficial and
214 increases the catch of this species (Mawardi and Riyanto 2017). Processed products from the *Scomberoides* fish species are
215 Talang-Talang salted fish in Aceh (Melawati et al. 2019, Riski 2017), fillets (Farsanipour et al. 2020), Surimi, and fish
216 nugget (Moosavi-Nasab et al. 2019, Oujifard and Morammazi 2020). However, the types of processed fishery products
217 from these fish groups in Indonesia are limited.

218

CONCLUSIONS

219 This research successfully identified the diversity of Carangid fish landed in several fishery centres in Java. Of the 33
 220 carangids fish, the most extensive composition obtained from these locations was *Megalaspis cordyla* (4), *Selaroides*
 221 *leptolepis* (4), *Atule mate* (3), and *Decapterus macarellus* (3). Generally, these fish are consumed locally to meet people's
 222 protein needs in coastal areas of Java. Phylogenetic analysis shows that the suborder Caranginae and Scomberoidinae are
 223 separated. Fish in the Caranginae suborder dominate compared to other sub-orders. Scomberoidinae in this study has
 224 unique morphological characteristics that are sometimes confused with the Scombridae group. This reinforces the use of
 225 molecular identification to complement morphological identification.

226

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380 **Table 1.** Carangids species list from Jawa, Indonesia

No.	Species	Sampling Site						
		GRK	PKL	BNT	PRT	MLG	BWI	DPS
1	<i>Alectis indica</i>					1		
2	<i>Alepes melanoptera</i>		1					
3	<i>Alepes vari</i>	1						
4	<i>Atropus atropos</i>			1				
5	<i>Atule mate</i>		1		1	1		
6	<i>Carangoides armatus</i>		1					
7	<i>Carangoides chrysophrys</i>						1	
8	<i>Carangoides malabaricus</i>			1				
9	<i>Caranx sexfasciatus</i>				1			
10	<i>Caranx tille</i>							
11	<i>Decapterus macarellus</i>		1			1		
12	<i>Decapterus macrosoma</i>			1				
13	<i>Decapterus mariuadi</i>					1		
14	<i>Megalaspis cordyla</i>	2	2					
15	<i>Parastromateus niger</i>	1			1			
16	<i>Selar boops</i>	1						
17	<i>Selar crumenophthalmus</i>		1					
18	<i>Selaroides leptolepis</i>		3		1			
19	<i>Scomberoides commersonianus</i>	1						
20	<i>Scomberoides tala</i>						1	
	TOTAL	6	10	3	4	4	2	

413
 414 **Remarks:** GRK = Gresik; PKL= Pekalongan; BNT= Banten; PRT= Pelabuhan Ratu; MLG= Malang; BWI= Banyuwangi;
 415 and DPS= Denpasar
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Tabel 2. IUCN and CITES status of all Carangids species

No.	Species	Sub-Family	Habitat distribution	Common Name	IUCN Red List Status	CITES	Threat to humans
1	<i>Alectis indica</i>	Caranginae	Indo-Pacific	Indian threadfish	LC	NE	Harmless
2	<i>A. melanoptera</i>	Caranginae	Indo-Pacific	Blackfin scad	LC	NE	Harmless
3	<i>A. vari</i>	Caranginae	Indo-West Pacific	Herring scad	LC	NE	Harmless
4	<i>Atropus atropus</i>	Caranginae	Indo-West Pacific	Cleftbelly trevally	LC	NE	Harmless
5	<i>Atule mate</i>	Caranginae	Indo-Pacific	Yellowtail scad	LC	NE	Harmless
6	<i>C. armatus</i>	Caranginae	Indo-West Pacific	Longfin trevally	LC	NE	Harmless
7	<i>C. chrysophrys</i>	Caranginae	Indo-Pacific	Longnose trevally	LC	NE	Harmless
8	<i>C. malabaricus</i>	Caranginae	Indo-West Pacific	Malabar trevally	LC	NE	Harmless
9	<i>Caranx sexfasciatus</i>	Caranginae	Indo-Pacific	Bigeye trevally	LC	NE	Harmless
10	<i>Caranx tille</i>	Caranginae	Indo-West Pacific	Tille trevally	LC	NE	Harmless
11	<i>Decapterus macarellus</i>	Caranginae	Circumglobal.	Mackerel scad	LC	NE	Ciguatera poisoning
12	<i>D. macrosoma</i>	Caranginae	Indo-Pacific and Southeast Atlantic	Shortfin scad	LC	NE	Harmless
13	<i>D. maruadsi</i>	Caranginae	Indo-West Pacific	Japanese scad	LC	NE	Harmless
14	<i>Megalaspis cordyla</i>	Caranginae	Indo-West Pacific	Torpedo scad	LC	NE	Harmless
15	<i>Parastromateus niger</i>	Caranginae	Indo-West Pacific	Black pomfret	LC	NE	Harmless
16	<i>Selar boops</i>	Caranginae	Pacific Ocean	Oxeye scad	LC	NE	Harmless
17	<i>S. crumenophthalmus</i>	Caranginae	Circumtropical	Bigeye scad	LC	NE	Ciguatera poisoning
18	<i>Selaroides leptolepis</i>	Caranginae	Indo-West Pacific	Yellow stripe scad	LC	NE	Harmless
19	<i>Scomberoides tala</i>	Scomberoidinae	Indo-West Pacific	Barred queenfish	LC	NE	Harmless
20	<i>S. commersonianus</i>	Scomberoidinae	Indo-West Pacific	Talang queenfish	LC	NE	Harmless

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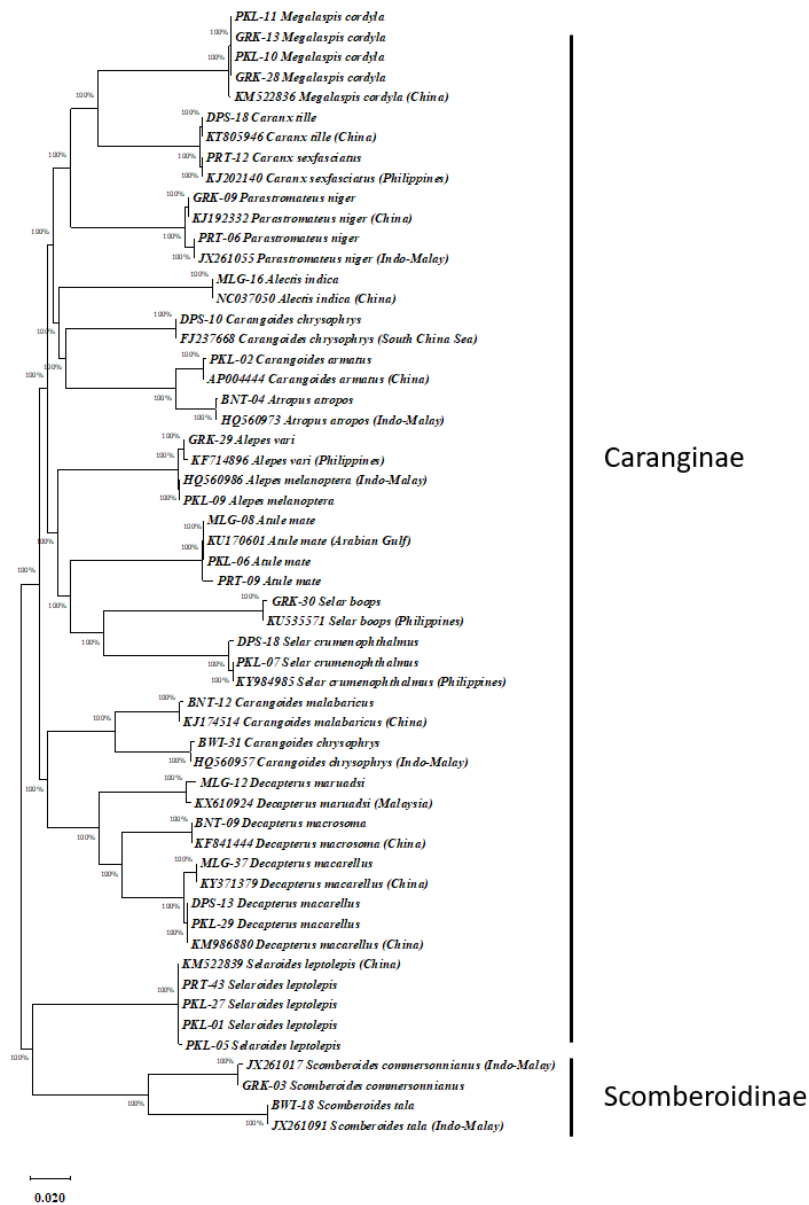


Figure 1. Phylogenetic tree of Carangidae using the neighbour-joining algorithm by Mega7.

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Figure 2. Scomberoidinae: *Scomberoides commersonianus* from Gresik and *Scomberoides tala* from Banyuwangi (black line: 1 cm).

Diversity of Carangidae (Carangiformes) revealed by DNA barcoding collected from traditional fish market in Java and Bali, Indonesia

Abstract. Biodiversity has been utilised in various ways, including in fulfilling the protein needs of fish for coastal communities. For the island of Java with the largest population in Indonesia, the intensive fisheries in the Java Sea are sufficient to support domestic food needs. This Carangid fish diversity study in Java is the beginning to identify commercial fish in Indonesia, which have been exploited for a long time. In this study, identification was carried out molecularly in the Cytochrome c oxidase subunit I (COI) gene region with the universal primary set and found a great variety of Carangids fish species. A total of 33 fish specimens have been identified, indicating two suborder groups, Caranginae (31) and Scomberoidenae (2). The Caranginae suborder group is more dominated with the most types of which are *Megalaspis cordyla* (3), *Atule mate* (3), *Decapterus macarellus* (4). Meanwhile, the Scomberoidenae suborder is dominated by *Scomberoides commersonianus* and *Scomberoides tala*. This study also found two species that have the potential to be ciguatera poisoning agents that need to be watched out for (*Decapterus macarellus* and *Selar crumenophthalmus*). Food safety in the fisheries sector has received considerable attention for a long time. Fishery commodities that live in highly polluted habitats are among the chains in increasing heavy metals concentration and the other residual chemical compounds that may impact human health.

Keywords: biodiversity, commercial, conservation, fisheries, genetic study

Running title: Andriyono et al. Diversity of Carangidae

INTRODUCTION

In Indonesia, an archipelago is distributed in tropical areas called the Indo-Malay Archipelago (IMA), with incredible biodiversity in marine life. This region has become recognised as one of the centres of marine biodiversity in various taxon fish, molluscs, and various other taxa (Lohman et al. 2011), located between the Pacific Ocean in the North and the Indian Ocean in the South. The island of Java, located in the southern part of Indonesia, has the potential for diversity, strongly influenced by the Indian Ocean compared to the Pacific Ocean region. However, the Java Sea's existence in the middle of the Indonesian archipelago allows the mixing of Pacific waters through the Makassar Strait gap between Kalimantan and Sulawesi Islands. The presence of the South China Sea also enables fish resources to enter the Java Sea through the Riau Islands and into the Java Sea. This considerable fishery potential causes capture fisheries in the Java Sea to be very potential, dominated by the Carangid fish.

Carangidae family (order Carangiformes) includes fish with varying body sizes ranging from 16 cm-250 cm, with irregular body shapes (elongated, fusiform and compressed) (Randall 1995). Fish from this family are commonly found in fisheries commodities such as scads, jacks, trevallies, pompanos, and rainbow runners. As a food and animal protein source, Carangids have become essential raw materials in Southeast Asia's processing industry. Even though intensive exploitation continues, taxonomy and scientific naming is still confused because of the similarities in morphology, size and colour pattern, which causes ambiguity in this family taxonomy (Froese 2009). In the life cycle, the juvenile stage to adulthood sometimes undergoes significant morphological changes and pigmentation which results in misidentification down to the species level. It is found in juvenile *Alectis ciliaris* long filament but will shorten and eventually disappear as it grows to maturity (Randall et al. 1997).

This DNA barcode method uses a reasonably short but standardised area of mitochondrial DNA with an identification accuracy up to the species level (Aquilino et al. 2011), which is high enough to approach 100% and has been widely accepted to date. Thus, the identification based on morphological needs to be complemented by other identification

49 methods to ensure identification accuracy. The method of identification currently being developed molecularly allows
50 reducing errors in identification. This identification is referred to as barcoding DNA. [This DNA barcode method uses a
51 reasonably short but standardised area of mitochondrial DNA with an identification accuracy up to the species level high
52 enough to be close to 100%, and to date has been widely accepted (Hebert et al. 2003)]. Open-access databases are
53 continually being developed in connection with the emergence of misidentification through the molecular approach
54 (Ratnasingham and Hebert 2007). Simplifying the identification process through molecular methods is relatively easy
55 and allows finding new taxa (Hajibabaei et al. 2007).

56 This study carried out tests on Carangidae fish from Java's northern and southern regions and analysed the COI
57 sequence's genetic diversity. This analysis is expected to be important for sustainable management of Carangidae fish
58 resources because most of them are not reported and are not well identified.

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59 MATERIALS AND METHODS

60 Sampling site

61 Thirty-three fish samples (Table 1) were collected from the five traditional fish markets around Jawa Island during
62 July 2019. In the northern part of Java, samples were obtained from the Banten-BNT (6°0'50.00'S-106°10'21.00" E),
63 Gresik-GRK (6°52'56.65"S-112°12'15.87" E), and Pekalongan-PKL (6°51'32.10"S 109°41'09.52"E). While Southern Java
64 was represented samples from Pelabuhanratu-PRT (6°59'20.92"S 106°32'29.91"E), Malang-MLG (8°26'06.65"S-
65 112°40'55.31"E), the Banyuwangi-BWI (8°12'07.52"S-114°23'07.18"E), and Denpasar-DPS (8°45'23.00'S-
66 115°10'05.68"E). Morphologically identification was conducted according to the guideline from FAO (Heemstra 1993),
67 and species confirmation has been carried out with molecular identification carried out in this study using the COI gene
68 region. No specific permit was required for this study because collection from the local traditional fish market was dead
69 upon purchasing. All samples have been photographed was applied to every single specimen using a digital camera.

70 DNA extraction and PCR Condition

71 All samples were preserved in 90% ethanol for further laboratory experiments. In preparation for the genomic
72 DNA extraction process, cutting the tissue in each specimen was done aseptically (around 0.5 cm) and immediately
73 washed to remove ethanol under running water. After the washing process is carried out, the sample is put in a microtube
74 containing 6X lysis buffer and mixed with the TissueLyser II (Qiagen). The use of Accuprep® Genomic DNA extraction
75 Kit (Bioneer) was carried out according to the Kit protocol and followed by measuring the concentration of the extracted
76 with nanoDrop (ThermoFisher Scientific D1000). The extract is used in the application process by the PCR or stored at -
77 70oC for further analysis.

78 The amplification process of genomic DNA from samples was carried out using a universal primer set, namely FISH-
79 BCL (5'-TCA ACY AAT CAY AAA GAT ATY GGC AC-3 ') and FISH-BCH (5'-TAA ACT TCA GGG TGA CCA
80 AAA AAT CA -3 ') (Baldwin et al. 2009, Handy et al. 2011). The target DNA amplified is the cytochrome c oxidase I
81 (COI) region, about 600 bp in length. The volume of PCR mixture is 20µL consisting of 11.2 µL ultra-pure water, 1 µL
82 forward and reverse primer (0.5 µM), 0.2 µL Ex Taq DNA polymerase (TaKaRa, Japan), 2 µL 10X ExTag Buffer, 2 µL
83 dNTPs (1 µM, TaKaRa, Japan), and 2 µL genomic DNA as a template. The PCR condition was carried out under the
84 following setting: 95oC for 5 min in initial denaturation, followed by denaturation at 95oC for 30 s in 40 cycles, 50oC for
85 30 s in annealing, and 72oC for 45 s in extension step, and a final extension at 72oC for 5 min. After obtaining the
86 desired band on the electrophoresis gel (1.5% agarose), the PCR products were purified with the AccuPrep® Gel
87 purification kit (Bioneer, Korea).

88 Data Analysis and Phylogenetic analysis

89 All sequences are then aligned with the help of Mega7 software, including the sequences obtained from the GenBank
90 database for comparison. The pairwise evolutionary distance among the species is determined by the Kimura 2-Parameter
91 method. The Neighbor-joining (NJ) tree was constructed, and 1000 bootstrap analysis was carried out by Mega7 (Kumar et
92 al. 2016).

93 RESULTS AND DISCUSSION

94 Results

95 A total of 33 successful identification samples consisted of 2 sub-families within Carangidae, Caranginae (31) and
96 Scomberoidenae (2). In this study, the Caranginae dominated the general catches of fisherman, including *Megalaspis*
97 *cordyla* (3), *Atule* mate (3), *Decapterus macarellus* (4). Meanwhile, local fishers' common fish caught in Java and Bali
98 and commercial fish, which benefit domestic consumers. In general, the types of fish collected in the Carangids group
99 have low prices and become food sources of protein for the community.

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Genetic distance

Genetic distance analysis was carried out using Mega7 compared to the GenBank database. Several sequences have unique, potentially as a haplotype that Indonesia has founded. Within subfamily Scomberoidinae, *Scomberoides tala* is similar to the Indo-Malay (JX261091) than *Scomberoides commersonnianus* has a very close genetic distance with the Indo-Malay sequence (JX261017) is 0.0019. Beside, within sub-family of Caranginae, we also found potentially haplotype Indonesia in *Decapterus muruadi* (0.0079), *Megalaspis cordyla* (0.0019), *Carangoides armatus* (0.0019), *Atropos atropos* (0.0019), *Alepes vari* (0.0019), *Atule mate* (0.0059), *Selar boops* (0.019), *Selar crumenophthalmus* (0.0039), *Carangoides malabaricus* (0.0019), *Carangoides chrysophrys* (0.0019), and *Selaroides leptolepis* (0.0019).

Phylogenetic reconstruction

In the phylogenetic tree produced, two clades have been formed: the subfamily Cranginae and Scomberoidinae (Figure 1). Morphologically, Scomberoidinae is closer to the Family of Scombridae with fusiform in body shape (Figure 2), but its body was flattened compared to the torpedo-like Scombridae. Scomberoidinae, in etymology, have a meaning is similar to tunny or mackerel. In this study, *Scomberoides tala* and *Scomberoides commersonnianus* were successfully identified from Banyuwangi and Gresik, respectively.

Carangidae status in IUCN and CITES

Almost all Carangidae species in this study have the status of Least Concern (LC), and the international trade (CITES) status is not evaluated (Table 2). From several reports, the fishing of Carangids species is carried out intensively, and most of them turn it into fish for processed fishery products such as surimi or only as raw material for fish meal. Research on fish catches in the Madura Strait found that Carangidae fish catch was 8% (Purwangka and Mubarak 2018), while other studies found Carangidae species of 38.51% of the catch (Khatami et al. 2018). Apart from Java, the Carangidae species that traditional fishers catch in Kalimantan also entirely dominate around 16.67% (Alfian et al. 2020).

Discussion

Shallow marine fishery resources, especially in the Java Sea, have provided benefits for meeting animal protein needs in the Java Island region, the most populous island area in Indonesia. Indonesian capture fisheries production in Indonesia until 2017 reached 6,603,630.58 tonnes / year with a value of 185,798,801 US \$ (Nainggolan et al. 2019). From several ports on Java, the Java sea fishery shows quite intensive fishery activity with a fairly large catch value. There are potential ports with very diverse catches along the north coast of the island of Java. However, studies conducted by Java sea capture fisheries in the Central Java region show that the Carangids group such as *Decapterus* spp] is a caught fish that dominates, especially during the peak season in September-November each year (Chodrijah and Hariati 2017).

Carangidae fishery is a fishery potential that is the mainstay of fulfilling domestic consumption compared to tuna fisheries which are exported products (Yusuf et al. 2018). Carangidae commodity, processed products are an effort to increase the product's added value besides being sold in fresh fish. One of the most famous processed products with Carangids fish raw materials is the scavenging with the main fish species in steam fish (pindang) from the genus *Decapterus* spp and *Atule* spp (Lubis et al. 2019).

The surveys in some fish landing areas on the north coast of Java and the south coast illustrate that the Javanese marine fishery makes Carangids a relatively high catch (Table 1). On the other hand, in the southern region of Java, it is the result of large pelagic fisheries such as tuna and skipjack (Firdaus 2019). Previous research has also confirmed that the Java sea fishery with the Carangids commodity is becoming quite popular (Chodrijah and Hariati 2017). Meanwhile, the Indian sea fishery or the southern coast of Java, tuna fisheries are the mainstay and are the mainstay in Malang (Wiadnya et al. 2018). Pelabuhan ratu (Mertha et al. 2017), which is well known as a centre for tuna fish catches in Southern Java (Nurani et al. 2017). Besides, Bali Island, directly facing the Indian Ocean, also has a potential tuna landing centre in Benoa (Ahmad et al. 2019).

However, Carangids fish's diversity has provided benefits even though economically, it is only developed into a domestic product with traditional processing technologies such as dried salted fish and pindang. Phylogenetic tree analysis shows that the carangid diversity caught in the waters around Java is mainly in the Carangidae suborder group. However, a small proportion is included in the Scomberoidinae suborder group. In the Caranginae group, the species *Megalaspis cordyla* and *Selaroides leptolepis* dominate Java's northern region (Gresik and Pekalongan). In this phylogenetics, two distinct clade sub-orders between the Caranginae and Scomberoidinae sub-orders (Figure 1).

All carangid fish status in the IUCN Red list status and CITES classify all species in this order into the Least Concern (LC) category and Not Evaluated (NE). Attention and regulation are needed in the management of capture fisheries in particular. For instance, the northern part of Java reported having experienced overfishing (Sadhotomo and Atmadja 2016). Another interesting point is that apart from being an essential source of animal protein for coastal communities, two types of fish have been reported to be the causes of Ciguatera poisoning that need to be watched out for in *Decapterus macarellus* (Lange et al. 1992) and *Selar crumenophthalmus* (Rongo and van Woessik 2012). Food safety in the fisheries sector has received considerable attention for a long time. Fishery commodities that live in highly polluted habitats are among the chains in increasing heavy metals concentration that may impact humans. Besides, fish that live in

Commented [W12]: Need more detailed explanation why *Decapterus* spp. Dominates in the Central Java?

Commented [W13]: A more detailed explanation of how these two species caused Ciguatera. Ciguatera comes from Dinoflagellates attached to algae or seagrass. It needs to be related to the habitat conditions of the two species and their eating habits.

163 waters with a reasonably complex food chain, allowing food to enter the fish's body, can trigger the cause of ciguatera
164 that has been reported in several other countries outside Indonesia (Chan 2016). Although there have been no reports of
165 ciguatera cases in Indonesia (including Brunei, Cambodia, Myanmar and North Korea) (Chan 2015a), there needs to be
166 serious attention to Indonesian fishery products, which are also exported commodities. A recent report states that
167 Indonesian fishery products are reported to cause ciguatera from snapper fish on the commercial market in Germany
168 (Friedemann 2019). However, only China (Chan 2015b) and Hongkong (Chan 2014a) received reports of ciguatera due
169 to fishery products on hump-head wrasse (Chan 2013) and tiger grouper (Chan 2014b), which caused some people to
170 receive medical care.

171 **Sub-Family Caranginae**

173 In this study, the sub-family Caranginae dominates with 31 species identified from 31 species (94%) while the remaining
174 groups are Scomberoidinae (4%). In the Caranginae group, it can be found both in the Java Sea and the Indian Ocean. In
175 the Java Sea area, this study found the types of *Megalaspis cordyla* (GRK and PKL), *Alapes vari* (GRK), *Alepes*
176 *melanoptera* (PKL and GRK: Gresik), *Selar boops* (GRK), *Carangoides malabaricus* (BNT), and *Selaroides leptolepis*
177 (PKL and PRT). However, the Indian Ocean and Java Sea regions also share other types of Caranginae, such as
178 *Decapterus macarellus* (DPS, and PKL and MLG), *Decapterus macrosoma* (BNT), *Decapterus muruadsi* (MLG),
179 *Carangoides chrysophrys* (BWI) and *Carangoides malabaricus* (BNT). These types of fish become one of the targets of
180 artisanal fisheries in Java, both in the southern and northern regions.

181 Many studies on artisanal fisheries in Java have been carried out, and *Selaroides* (Wijayanto et al. 2019a, Wijayanto et
182 al. 2019b) is one of the catch targets for medium-sized nets such as the use of Danish seine fishing gear, which is
183 considered an environmentally friendly fishing gear compared to trawling (Adhawati Sri Suro et al. 2017a, Adhawati Sri S
184 et al. 2017b). This artisanal fishery has driven the economy and even continues today. For example, capture fisheries in
185 Pekalongan with mini purse seine fishing gear that get catches belonging to the Caranginae group, such as the type of
186 *Decapterus* sp (Maulana et al. 2017). In addition, artisanal fisheries in the northern waters of Java are also highly
187 developed, such as in Banten (Barlian Erland et al. 2020, Barlian E et al. 2021), Tegal (Wijayanto et al. 2019a), Lamongan
188 (Syamsuddin et al. 2020), dan Madura (Yonvitner et al. 2021).

190 **Sub-Family Scomberoidinae**

191 Although this group of fish is only 4% of this study, economically, it is included in the fish group that has a fairly
192 reasonable price. This fish of the Scomberoidinae group has morphological characteristics similar to the Family
193 Scombridae, which includes tuna and mackerel with a streamlined body shape to become a group of fast swimming fish. In
194 this study, *Scomberoides commersonianus* is a group of fish that is quite attractive and becomes potential raw material
195 for fishery products. The method of catching this type of fish varies greatly; the use of blue LEDs is beneficial and
196 increases the catch of this species (Mawardi and Riyanto 2017). Processed products from the Scomberoides fish species are
197 Talang-Talang salted fish in Aceh (Melawati et al. 2019, Riski 2017), fillets (Farsanipour et al. 2020), Surimi and fish
198 nugget (Moosavi-Nasab et al. 2019, Oujifard and Morammazi 2020). However, the types of processed fishery products
199 from these fish groups in Indonesia are limited.

200 **CONCLUSIONS**

201 This research successfully identified the diversity of Carangid fish landed in several fishery centres in Java. Of the 33
202 carangids fish, the most extensive composition obtained from these locations was *Megalaspis cordyla* (3), *Atule mate* (3),
203 *Decapterus macarellus* (4). Generally, these fish are consumed locally to meet people's protein needs in coastal areas of
204 Java. Phylogenetic analysis shows that the suborder Caranginae and Scomberoidinae are separated. Fish in the Caranginae
205 suborder dominate compared to other sub-orders. In this study, we identified that *Decapterus macarellus* and *Selar*
206 *crumenophthalmus* had been reported to be the causes of ciguatera poisoning that need to be watched out for and get
207 attention in the processing post-harvest processes.

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211 supported the collection of samples throughout northern and southern Java, including Bali samples.
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Commented [W14]: In Figure 1, the similarity is 100% the same between samples from GRK and PKL. Are there habitat similarities between GRK and PKL? Or is it just because of the large population of the species?

Commented [W15]: Morphologically the *D. macarellus* and *D. macrosoma* are similar and it is difficult to distinguish them. There needs to be an explanation of the differences between these two species. Because from Figure 1 there is a similarity between *D. macarellus* from PKL and DPS, but there is a slight difference from MLG

Commented [W16]: How about *Selaroides leptolepis*? The number is quite large, there are 3 from PKL and 1 from PRT (Table 1). Are the locations between PKL and PRT similar, so the diversity is the same?

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Table 1. Carangids species list from Jawa, Indonesia

No.	Species	Sampling Site						
		GRK	PKL	BNT	PRT	MLG	BWI	DPS
1	<i>Alectis indica</i>					1		
2	<i>Alepes melanoptera</i>		1					
3	<i>Alepes vari</i>	1						
4	<i>Atropus atropus</i>			1				
5	<i>Atule mate</i>		1		1	1		
6	<i>Carangoides armatus</i>		1					
7	<i>Carangoides chrysophrys</i>						1	1
8	<i>Carangoides malabaricus</i>			1				
9	<i>Caranx sexfasciatus</i>				1			
10	<i>Caranx tille</i>							1
11	<i>Decapterus macarellus</i>		1			1		1
12	<i>Decapterus macrosoma</i>			1				
13	<i>Decapterus maruadsi</i>					1		
14	<i>Megalaspis cordyla</i>	2	2					
15	<i>Parastromateus niger</i>	1			1			
16	<i>Selar boops</i>	1						
17	<i>Selar crumenophthalmus</i>		1					1
18	<i>Selaroides leptolepis</i>		3		1			
19	<i>Scomberoides commersonianus</i>	1						
20	<i>Scomberoides tala</i>						1	
	TOTAL	7	10	3	4	4	2	4

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GRK = Gresik; PKL= Pekalongan; BNT= Banten; PRT= Pelabuhan Ratu; MLG= Malang; BWI= Banyuwangi; and DPS= Denpasar

Tabel 2. IUCN and CITES status of all Carangids species

No.	Species	Sub-Family	Habitat distribution	Common Name	IUCN Red List Status	CITES	Threat to humans
1	<i>Alectis indica</i>	Caranginae	Indo-Pacific	Indian threadfish	LC	NE	Harmless
2	<i>A. melanoptera</i>	Caranginae	Indo-Pacific	Blackfin scad	LC	NE	Harmless
3	<i>A. vari</i>	Caranginae	Indo-West Pacific	Herring scad	LC	NE	Harmless
4	<i>Atropus atropus</i>	Caranginae	Indo-West Pacific	Cleftbelly trevally	LC	NE	Harmless
5	<i>Atule mate</i>	Caranginae	Indo-Pacific	Yellowtail scad	LC	NE	Harmless
6	<i>C. armatus</i>	Caranginae	Indo-West Pacific	Longfin trevally	LC	NE	Harmless
7	<i>C. chrysophrys</i>	Caranginae	Indo-Pacific	Longnose trevally	LC	NE	Harmless
8	<i>C. malabaricus</i>	Caranginae	Indo-West Pacific	Malabar trevally	LC	NE	Harmless
9	<i>Caranx sexfasciatus</i>	Caranginae	Indo-Pacific	Bigeye trevally	LC	NE	Harmless
10	<i>Caranx tille</i>	Caranginae	Indo-West Pacific	Tille trevally	LC	NE	Harmless
11	<i>Decapterus macarellus</i>	Caranginae	Circumglobal.	Mackerel scad	LC	NE	Ciguatera poisoning
12	<i>D. macrosoma</i>	Caranginae	Indo-Pacific and Southeast Atlantic	Shortfin scad	LC	NE	Harmless
13	<i>D. maruadsi</i>	Caranginae	Indo-West Pacific	Japanese scad	LC	NE	Harmless
14	<i>Megalaspis cordyla</i>	Caranginae	Indo-West Pacific	Torpedo scad	LC	NE	Harmless
15	<i>Parastromateus niger</i>	Caranginae	Indo-West Pacific	Black pomfret	LC	NE	Harmless
16	<i>Selar boops</i>	Caranginae	Pacific Ocean	Oxeye scad	LC	NE	Harmless
17	<i>S. crumenophthalmus</i>	Caranginae	Circumtropical	Bigeye scad	LC	NE	Ciguatera poisoning
18	<i>Selaroides leptolepis</i>	Caranginae	Indo-West Pacific	Yellow stripe scad	LC	NE	Harmless
19	<i>Scomberoides tala</i>	Scomberoidinae	Indo-West Pacific	Barred queenfish	LC	NE	Harmless
20	<i>S. commersonianus</i>	Scomberoidinae	Indo-West Pacific	Talang queenfish	LC	NE	Harmless

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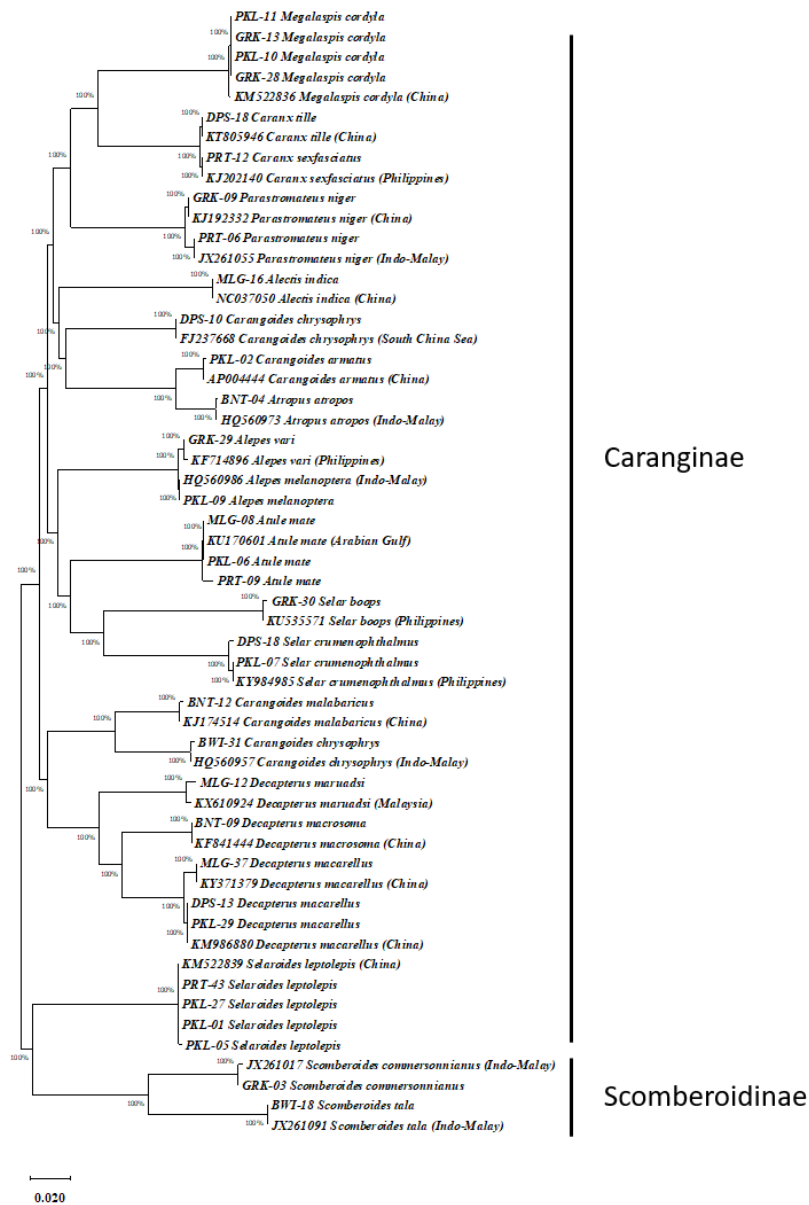


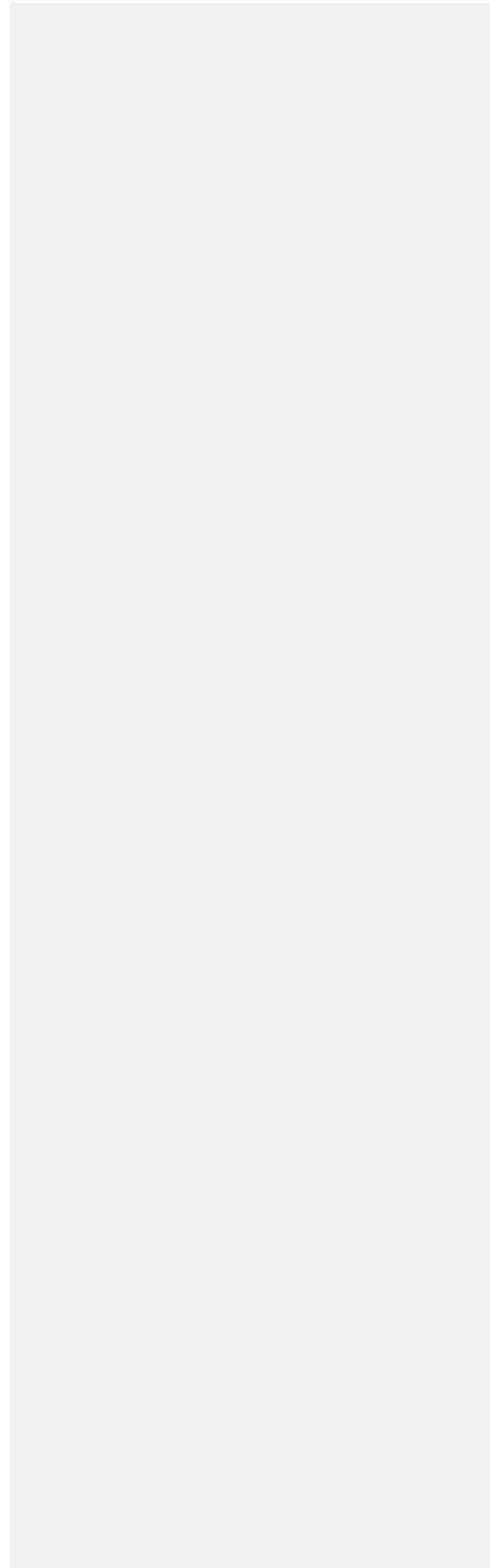
Figure 1. Phylogenetic tree of Carangidae using the neighbour-joining algorithm by Mega7.

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Figure 2. Scomberoidinae: *Scomberoides commersonianus* from Gresik and *Scomberoides tala* from Banyuwangi (black line: 1 cm).



The diversity of Carangidae (Carangiformes) was revealed by DNA barcoding collected from the traditional fish markets in Java and Bali, Indonesia

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Abstract. Andriyono S, Alam MJ, Sumartiw L, Mubarak AS, Pramono H, Suciyono, Kartika GRA, Sari AHW, Sektiana SP. 2022. The diversity of Carangidae (Carangiformes) was revealed by DNA barcoding collected from the traditional fish markets in Java and Bali, Indonesia. *Biodiversitas* 23: 2799-2806. Biodiversity has been utilized in various ways, including in fulfilling the protein needs of fish for coastal communities. For the island of Java, with the largest population in Indonesia, the intensive fisheries in the Java Sea are sufficient to support domestic food needs. This Carangid fish diversity study in Java is the beginning to identify commercial fish in Indonesia, which have been exploited for a long time. In this study, identification was carried out molecularly in the Cytochrome c oxidase subunit I (COI) gene region with the universal primary set and found a great variety of Carangids fish species. Thirty-three fish specimens have been identified, indicating two suborder groups, Caranginae (31) and Scomberoidenae (2). The Caranginae suborder group is more dominated with the most types of which are *Megalaspis cordyla* (3), *Atule mate* (3), and *Decapterus macarellus* (4). Meanwhile, the Scomberoidenae suborder is *Scomberoides commersonnianus* and *Scomberoides tala*. This study also found two species that have the potential to be ciguatera poisoning agents that need to be watched out for (*Decapterus macarellus* and *Selar crumenophthalmus*). Food safety in the fisheries sector has received considerable attention for a long time. Fishery commodities in highly polluted habitats are among the chains in increasing heavy metals concentration and the other residual chemical compounds that may impact human health.

Keywords: Biodiversity, commercial, conservation, fisheries, genetic study

INTRODUCTION

In Indonesia, an archipelago is distributed in tropical areas called the Indo-Malay Archipelago (IMA), with Incredible biodiversity in marine life. This region has been recognized as one of the centers of marine biodiversity in various taxon fish, molluscs, and various other taxa (Lohman et al. 2011), located between the Pacific Ocean in the North and the Indian Ocean in the South. The island of Java, located in the southern part of Indonesia, has the potential for diversity, strongly influenced by the Indian Ocean compared to the Pacific Ocean region. However, the Java Sea's existence in the middle of the Indonesian archipelago allows the mixing of Pacific waters through the Makassar Strait gap between Kalimantan and Sulawesi Islands. The presence of the South China Sea also enables fish resources to enter the Java Sea through the Riau Islands and into the Java Sea. This considerable fishery potential causes capture fisheries in the Java Sea to be very potential, dominated by the Carangid fish.

The Carangidae family (order Carangiformes) includes fish with varying body sizes ranging from 16 cm-250 cm, with irregular body shapes (elongated, fusiform, and compressed) (Randall 1995). Fish from this family are commonly found in fisheries commodities such as scads, jacks, trevallies, pompanos, and rainbow runners. In addition, Carangids have become essential raw materials in Southeast Asia's processing industry as a food and animal protein source. Even though intensive exploitation continues, taxonomy and scientific naming is still confused because of the similarities in morphology, size, and color pattern, which causes ambiguity in this family taxonomy (Froese 2009). In addition, the juvenile stage to adulthood sometimes undergoes significant morphological changes and pigmentation in the life cycle. As a result, misidentification down to the species level has often happened. The color difference in the *Cichla* species occurs in the *Cichla kelberi* species which has a color similar to that of *Cichla temensis* (Sastraprawira et al. 2020). The implication of misidentification is mislabeling of fisheries

products (Guardone et al. 2017). For instance, the type of *Cichla ocellaris* circulating in Indonesia has a higher price than other types of this fish. The *Cichla* fish in the ornamental fish market is known as peacock bass or peacock cichlids with attractive color patterns. Misidentification cases based on morphology also occur in juvenile *Alectis ciliary* long filament but will shorten and eventually disappear as it grows to maturity (Randall et al. 1997).

This DNA barcode method uses a reasonably short but standardized area of mitochondrial DNA with an identification accuracy up to the species level (Aquilino et al. 2011), which is high enough to approach 100% and has been widely accepted to date. Thus, the identification based on morphological needs to be complemented by other identification methods to ensure identification accuracy. The form of identification currently being developed molecularly reduces errors in identification. This identification is referred to as barcoding DNA. This DNA barcode method uses a reasonably short but standardized area of mitochondrial DNA. That region is Cytochrome c Oxidase subunit I (COI). This gene region has been widely used for a molecular approach-based identification with accuracy up to the species level high enough to be close to 100%. DNA barcoding has been globally accepted (Hebert et al. 2003). Open-access databases are continually being developed in connection with the emergence of misidentification through the molecular approach (Ratnasingham and Hebert 2007). Simplifying the identification process through molecular methods is relatively easy and allows for finding new taxa (Hajibabaei et al. 2007).

This study carried out tests on Carangidae fish from Java's northern and southern regions and analyzed the COI sequence's genetic diversity. This analysis is essential for the sustainable management of Carangidae fish resources because most of them are not reported and are not well identified.

MATERIALS AND METHODS

Sampling site

Thirty-three fish samples (Table 1) were collected from the five traditional fish markets around Jawa Island in July 2019. In the northern part of Java, samples were obtained from the Banten-BNT (6°0'50.00"S-106°10'21.00"E), Gresik-GRK (6°52'56.65"S-112°12'15.87" E), and Pekalongan-PKL (6°51'32.10"S 109°41'09.52"E). While Southern Java was represented samples from Pelabuhan Ratu-PRT (6°59'20.92"S 106°32'29.91"E), Malang-MLG (8° 26' 06.65' S - 112° 40' 55.31" E), the Banyuwangi-BWI (8° 12' 07.52' S - 114°23'07.18" E), and Denpasar-DPS (8° 45' 23.00' S - 115° 10' 05.68" E). Morphological identification was conducted according to the guideline from FAO (Heemstra 1993), and species confirmation was carried out with molecular identification carried out in this study using the COI gene region. No specific permit was required for this study because collection from the local traditional fish market was dead upon purchasing. All samples were photographed for every single specimen using a digital camera.

Table 1. Carangids species list from Jawa, Indonesia

Species	Sampling location						
	GRK	PKL	BNT	PRT	MLG	BWI	DPS
<i>Alectis indica</i>					1		
<i>Alepes melanoptera</i>		1					
<i>Alepes vari</i>	1						
<i>Atropus atropos</i>			1				
<i>Atule mate</i>		1		1	1		
<i>Carangoides armatus</i>		1					
<i>Carangoides chrysophrys</i>						1	1
<i>Carangoides malabaricus</i>			1				
<i>Caranx sexfasciatus</i>				1			
<i>Caranx tille</i>							1
<i>Decapterus macarellus</i>		1			1		1
<i>Decapterus macrosoma</i>			1				
<i>Decapterus maruadsi</i>					1		
<i>Megalaspis cordyla</i>	2	2					
<i>Parastromateus niger</i>	1			1			
<i>Selar boops</i>	1						
<i>Selar crumenophthalmus</i>		1					1
<i>Selaroides leptolepis</i>		3		1			
<i>Scomberoides commersonianus</i>	1						
<i>Scomberoides tala</i>						1	
TOTAL	6	10	3	4	4	2	4

Note: GRK = Gresik; PKL= Pekalongan; BNT= Banten; PRT= Pelabuhan Ratu; MLG= Malang; BWI= Banyuwangi; DPS= Denpasar

DNA extraction and PCR condition

All samples were preserved in 90% ethanol for further laboratory experiments. In preparation for the genomic DNA extraction process, cutting the tissue in each specimen was done aseptically (around 0.5 cm) and immediately washed to remove ethanol under running water. After the washing process was carried out, the sample was put in a microtube containing 6X lysis buffer and mixed with the TissueLyser II (Qiagen). The use of Accuprep® Genomic DNA extraction Kit (Bioneer) was carried out according to the Kit protocol and followed by measuring the concentration of the extracted with nanoDrop (ThermoFisher Scientific D1000). The extract was used in the application process by the PCR or stored at -70°C for further analysis.

The amplification process of genomic DNA from samples was carried out using a universal primer set, namely FISH-BCL (5'-TCA ACY AAT CAY AAA GAT ATY GGC AC-3 ') and FISH-BCH (5'-TAA ACT TCA GGG TGA CCA AAA AAT CA -3 ') (Baldwin et al. 2009, Handy et al. 2011). The target DNA amplified is the cytochrome c oxidase I (COI) region, about 600 bp in length. The volume of the PCR mixture is 20 µL consisting of 11.2 µL ultra-pure water, 1 µL forward and reverse primer (0.5 µM), 0.2 µL Ex Taq DNA polymerase (TaKaRa, Japan), 2 µL 10X ExTag Buffer, 2 µL dNTPs (1 µM, TaKaRa, Japan), and 2 µL genomic DNA as a template. The PCR condition was carried out under the following setting: 95°C for 5 min in initial denaturation, followed by denaturation at 95°C for 30 s in 40 cycles, 50°C for 30 s in annealing, and 72°C for 45 s in extension step, and a final extension at 72°C for 5 min. After obtaining the desired band on the electrophoresis gel (1.5% agarose), the PCR products were purified with the AccuPrep® Gel purification kit (Bioneer, Korea).

Data analysis and phylogenetic analysis

All sequences are then aligned with the help of Mega7 software, including the sequences obtained from the GenBank database for comparison. The pairwise evolutionary distance among the species is determined by the Kimura 2-Parameter method. The Neighbor-joining (NJ) tree was constructed, and 1000 bootstrap analysis was carried out by Mega7 (Kumar et al. 2016).

RESULTS AND DISCUSSION

Results

Thirty-three successful identification samples consisted of 2 sub-families within Carangidae, Caranginae (31) and Scomberoidenae (2). In this study, the Caranginae dominated the general catches of fisherman, including *Megalaspis cordyla* (3), *Atule mate* (3), and *Decapterus macarellus* (4). Meanwhile, local

fishers' common fish caught in Java and Bali and commercial fish, benefit domestic consumers. In general, the types of fish collected in the Carangids group have low prices and become food sources of protein for the community.

Genetic distance

Genetic distance analysis was carried out using Mega7 compared to the GenBank database. Several sequences have unique, potentially as a haplotype that Indonesia has founded. Within subfamily Scomberoidinae, *Scomberoides tala* is similar to the Indo-Malay (JX261091) than *Scomberoides commersonianus* has a very close genetic distance with the Indo-Malay sequence (JX261017) is 0.0019. The smaller the genetic distance, the closer the similarity and may indicate that a particular species may have similar origins (Tapilatu et al. 2021; Dwifajri et al. 2022). In addition, within sub-family of Caranginae, we also found potentially haplotype Indonesia in *Decapterus maruadsi* (0.0079), *Megalaspis cordyla* (0.0019), *Carangoides armatus* (0.0019), *Atropos Atropos* (0.0019), *Alepes vari* (0.0019), *Atule mate* (0.0059), *Selar boops* (0.019), *Selar crumenophthalmus* (0.0039), *Carangoides malabaricus* (0.0019), *Carangoides chrysophrys* (0.0019), and *Selaroides leptolepis* (0.0019).

Phylogenetic reconstruction

In the phylogenetic tree produced, two clades have been formed: the subfamily Cranginae and Scomberoidinae (Figure 1). Morphologically, Scomberoidinae is closer to the Family of Scombridae with fusiform in body shape (Figure 2), but its body was flattened compared to the torpedo-like Scombridae. Scomberoidinae, in etymology, has a meaning similar to tunny or mackerel. In this study, *Scomberoides tala* and *Scomberoides commersonianus* were successfully identified from Banyuwangi and Gresik, respectively.

Carangidae status in IUCN and CITES

Almost all Carangidae species in this study have the status of Least Concern (LC), and the international trade (CITES) status is not evaluated (Table 2). From several reports, the fishing of Carangids species is carried out intensively, and most of them turn it into fish for processed fishery products such as surimi or only as raw material for fish meal. For example, research on fish catches in the Madura Strait found that Carangidae fish catch was 8% (Purwangka and Mubarak 2018), while other studies found Carangidae species of 38.51% of the catch (Khatami et al. 2018). Apart from Java, the Carangidae species that traditional fishers catch in Kalimantan also entirely dominate around 16.67% (Alfian et al. 2020).

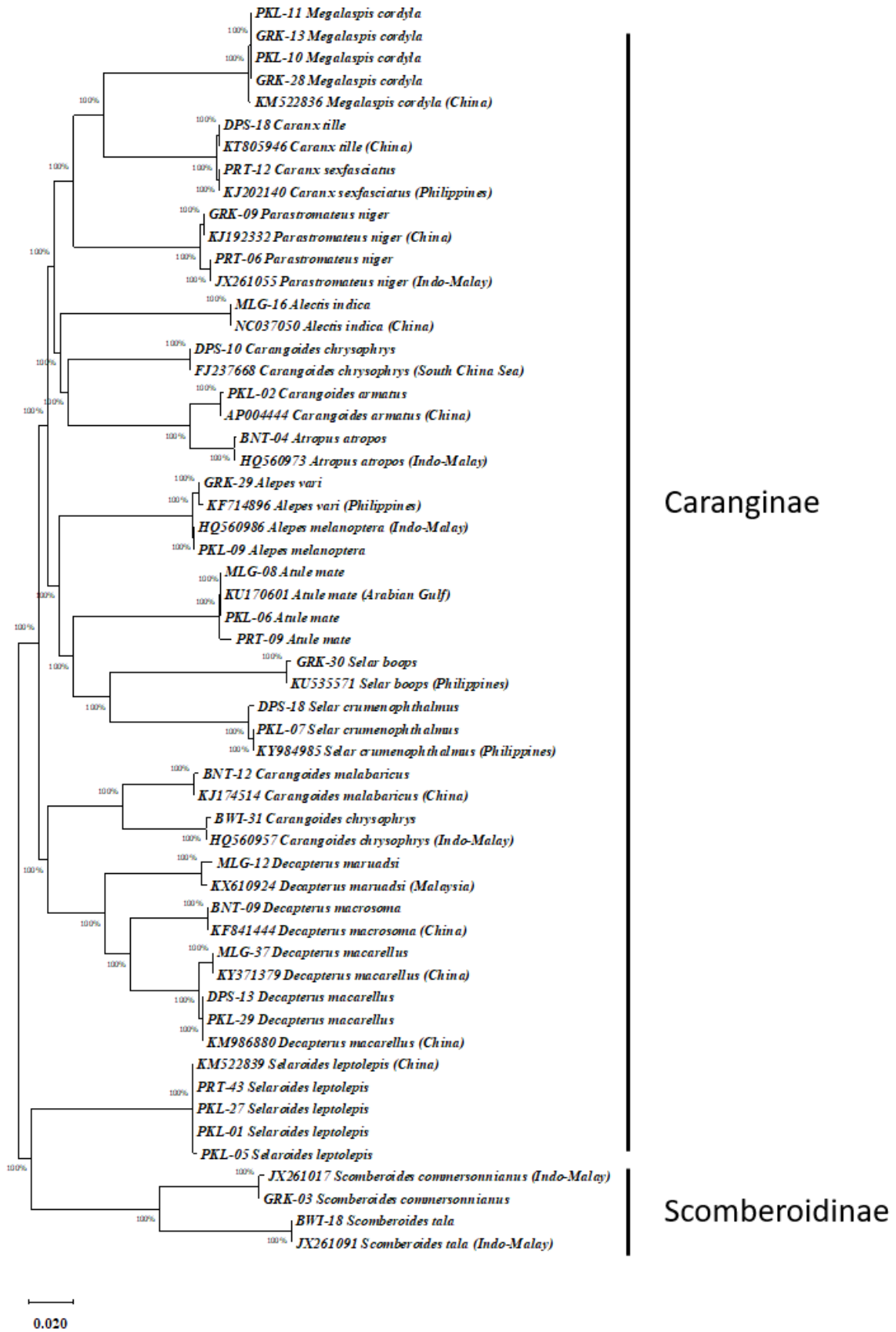


Figure 1. Phylogenetic tree of Carangidae using the neighbor-joining algorithm by Mega7



Figure 2. Scomberoidinae: *Scomberoides commersonianus* from Gresik and *Scomberoides tala* from Banyuwangi (Bar = 1 cm)

Tabel 2. IUCN and CITES status of all Carangids species

Species	Sub-family	Habitat distribution	Common name	IUCN Red List Status	CITES	Threat to humans
<i>Alectis indica</i>	Caranginae	Indo-Pacific	Indian threadfish	LC	NE	Harmless
<i>A. melanoptera</i>	Caranginae	Indo-Pacific	Blackfin scad	LC	NE	Harmless
<i>A. vari</i>	Caranginae	Indo-West Pacific	Herring scad	LC	NE	Harmless
<i>Atropus atropus</i>	Caranginae	Indo-West Pacific	Cleftbelly trevally	LC	NE	Harmless
<i>Atule mate</i>	Caranginae	Indo-Pacific	Yellowtail scad	LC	NE	Harmless
<i>C. armatus</i>	Caranginae	Indo-West Pacific	Longfin trevally	LC	NE	Harmless
<i>C. chrysophrys</i>	Caranginae	Indo-Pacific	Longnose trevally	LC	NE	Harmless
<i>C. malabaricus</i>	Caranginae	Indo-West Pacific	Malabar trevally	LC	NE	Harmless
<i>Caranx sexfasciatus</i>	Caranginae	Indo-Pacific	Bigeye trevally	LC	NE	Harmless
<i>Caranx tille</i>	Caranginae	Indo-West Pacific	Tille trevally	LC	NE	Harmless
<i>Decapterus macarellus</i>	Caranginae	Circumglobal.	Mackerel scad	LC	NE	Ciguatera poisoning
<i>D. macrosoma</i>	Caranginae	Indo-Pacific and Southeast Atlantic	Shortfin scad	LC	NE	Harmless
<i>D. maruadsi</i>	Caranginae	Indo-West Pacific	Japanese scad	LC	NE	Harmless
<i>Megalaspis cordyla</i>	Caranginae	Indo-West Pacific	Torpedo scad	LC	NE	Harmless
<i>Parastromateus niger</i>	Caranginae	Indo-West Pacific	Black pomfret	LC	NE	Harmless
<i>Selar boops</i>	Caranginae	Pacific Ocean	Oxeye scad	LC	NE	Harmless
<i>S. crumenophthalmus</i>	Caranginae	Circumtropical	Bigeye scad	LC	NE	Ciguatera poisoning
<i>Selaroides leptolepis</i>	Caranginae	Indo-West Pacific	Yellow stripe scad	LC	NE	Harmless
<i>Scomberoides tala</i>	Scomberoidinae	Indo-West Pacific	Barred queenfish	LC	NE	Harmless
<i>S. commersonianus</i>	Scomberoidinae	Indo-West Pacific	Talang queenfish	LC	NE	Harmless

Discussion

Shallow marine fishery resources, especially in the Java Sea, have provided benefits for meeting animal protein needs in the Java Island region, the most populous island area in Indonesia. Indonesian capture fisheries production in Indonesia until 2017 reached 6,603,630.58 tonnes/year with a value of USD 185,798,801 (Nainggolan et al. 2019). From several ports on Java, the Java Sea fishery shows quite intensive fishery activity with a fairly large catch value. There are potential ports with very diverse catches along the north coast of the island of Java. However, studies conducted by Java sea capture fisheries in the Central Java region show that the Carangids group, such as *Decapterus* spp., is a caught fish species that dominates, especially during the peak season in September-November each year (Chodrijah and Hariati 2017). Another study stated that as a migratory pelagic fish, *Decapterus* spp. into the waters of the Java Sea from the Indian Ocean. The migration occurred in the western season (January-March) and increased during the southeast monsoon (June-July), including two species, *Decapterus macrosoma* and *Decapterus russelli* (Prasetyo et al. 2018).

Carangidae fishery is a fishery potential that is the mainstay of fulfilling domestic consumption compared to tuna fisheries which are exported products (Yusuf et al. 2018). Carangidae commodity, processed products are an effort to increase the product's added value besides being sold in fresh fish. One of the most famous processed products with Carangids fish raw materials is the scavenging with the main fish species in steam fish (pindang) from the genus *Decapterus* spp and *Atule* spp (Lubis et al. 2019).

The surveys in some fish landing areas on the north coast of Java and the south coast illustrate that the Javanese marine fishery makes Carangids a relatively high catch (Table 1). On the other hand, in the southern region of Java, it is the result of large pelagic fisheries such as tuna and skipjack (Firdaus 2019). Previous research has also confirmed that the Java sea fishery with the Carangids commodity is becoming quite popular (Chodrijah and Hariati 2017). Meanwhile, in the Indian sea fishery or the southern coast of Java, tuna fisheries are the mainstay and are the mainstay in Malang (Wiadnya et al. 2018). Pelabuhan Ratu (Mertha et al. 2017) is well known as a center for tuna fish catches in Southern Java (Nurani et al.

2017). Besides, Bali Island, directly facing the Indian Ocean, also has a potential tuna landing center in Bena (Ahmad et al. 2019).

However, Carangids fish's diversity has provided benefits even though, economically, it is only developed into a domestic product with traditional processing technologies such as dried salted fish and pindang. Phylogenetic tree analysis shows that the carangid diversity caught in the waters around Java is mainly in the Carangidae suborder group. However, a small proportion is included in the Scomberoidinae suborder group. In the Caranginae group, *Megalaspis cordyla* and *Selaroides leptolepis* dominate Java's northern region (Gresik and Pekalongan). In this phylogenetics, two distinct clade suborders between the Caranginae and Scomberoidinae suborders (Figure 1).

All carangid fish status in the IUCN Red list status and CITES classify all species in this order into the Least Concern (LC) category and Not Evaluated (NE). Attention and regulation are needed in the management of capture fisheries in particular. For instance, the northern part of Java reported having experienced overfishing (Sadhotomo and Atmadja 2016). Another interesting point is that apart from being an essential source of animal protein for coastal communities, two types of fish have been reported to be the causes of Ciguatera poisoning that need to be watched out for in *Decapterus macarellus* (Lange et al. 1992), and *Selar crumenophthalmus* (Rongo and van Woesik 2012). The type of *Decapterus macarellus*, suspected of containing poison is smoked fish imported from the Philippines. The possibility of smoked fish meat has palytoxin, which causes ciguatera poisoning (Kodama et al. 1989). Ciguatera poisoning is a form of ichthyo-sarcotoxism, which means the content of some toxin in that body. This condition occurs when marine fishes inadvertently ingest particular dinoflagellates that produce ciguatoxins (Rongo and van Woesik 2012). Until now, no identified reports about ciguatera poisoning in Indonesia. Food safety in the fisheries sector has received considerable attention for a long time. Fishery commodities that harvested from highly polluted habitats are among the chains in increasing heavy metals concentration that may impact humans (REF?). Furthermore, fish that live in waters with a reasonably complex food chain, allowing food to enter the fish's body, would trigger the cause of ciguatera that has been reported in several other countries outside Indonesia (Chan 2016). Although there have been no reports of ciguatera cases in Indonesia (including Brunei, Cambodia, Myanmar, and North Korea) (Chan 2015a), there needs to be serious attention to Indonesian fishery products, which are also exported commodities. A report states that Indonesian fishery products are reported to cause ciguatera from snapper fish on the commercial market in Germany (Friedemann 2019). However, only China (Chan 2015b) and Hongkong (Chan 2014b) received reports of ciguatera due to fishery products on hump-head wrasse (Chan 2013) and tiger grouper (Chan 2014a), which caused some people to receive medical care.

Sub-Family Caranginae

In this study, the sub-family Caranginae dominates with 31 species identified from 31 species (94%), while the remaining groups are Scomberoidinae (4%). In the Caranginae group, it can be found both in the Java Sea and the Indian Ocean. In the Java Sea area, this study found the types of *Megalaspis cordyla* (GRK and PKL), *Alepes vari* (GRK), *Alepes melanoptera* (PKL), *Selar boops* (GRK), *Carangoides malabaricus* (BNT), and *Selaroides leptolepis* (PKL and PRT). Based on the phylogenetic tree (Figure 1), the species *Megalaspis cordyla* from Gresik and Pekalongan has close related and probably in the same population and share a habitat in Jawa Sea (Maskur et al. 2020). However, the Indian Ocean and Java Sea regions also share other types of Caranginae, such as *Decapterus macarellus* (DPS, PKL and MLG), and *Decapterus macrosoma* (BNT), *Decapterus maruadsi* (MLG), *Carangoides chrysophrys* (BWI) and *Carangoides malabaricus* (BNT). In brief, the two species of *Decapterus* (*D. macarellus* and *D. macrosoma*) are pretty difficult to distinguish morphologically. The primary and key characteristic in the scattered scales on the lineal lateralis part. Research on the morphological and molecular characteristics of these two species of *Decapterus* has been carried out clearly (Zhang et al. 2020). These types of fish have become one of the targets of artisanal fisheries in Java, both in the southern and northern regions including Bali. Research in the Java Sea waters states that several fish species, such as small pelagic, dominate the catch, including the type of *Selaroides leptolepis*, *Selar crumenophthalmus*, *Dussumieria acuta*, *Rastrelliger brachysoma*, and *Atule mate* (Khatami et al. 2019).

Many studies on artisanal fisheries in Java have been carried out, and *Selaroides* (Wijayanto et al. 2019a, Wijayanto et al. 2019b) is one of the catch targets for medium-sized nets such as the use of Danish seine fishing gear, which is considered an environmentally friendly fishing gear compared to trawling (Adhawati et al. 2017a, Adhawati et al. 2017b). This artisanal fishery has driven the economy and even continues today. For example, capture fisheries in Pekalongan with mini purse seine fishing gear that get catches belonging to the Caranginae group, such as the type of *Decapterus* sp (Maulana et al. 2017). In addition, artisanal fisheries in the northern waters of Java are also highly developed, such as in Banten (Barlian et al. 2020; Barlian et al. 2021), Tegal (Wijayanto et al. 2019a), Lamongan (Syamsuddin et al. 2020), and Madura (Yonvitner et al. 2021).

Sub-Family Scomberoidinae

Although this group of fish is only 4% of this study, economically, it is included in the fish group with a fairly reasonable price. This fish of the Scomberoidinae group has morphological characteristics similar to the Family Scombridae, which includes tuna and mackerel with a streamlined body shape to become a group of fast swimming fish. In this study, *Scomberoides commersonianus* is a group of fish that is quite attractive and becomes potential raw material for fishery products. The method of catching this type of fish varies

considerably; the use of blue LEDs is beneficial and increases the catch of this species (Mawardi and Riyanto 2017). Processed products from the Scomberoides fish species are Talang-Talang salted fish in Aceh (Melawati et al. 2019; Riski 2017), fillets (Farsanipour et al. 2020), Surimi, and fish nugget (Moosavi-Nasab et al. 2019, Oujifard and Morammazi 2020). However, the types of processed fishery products from these fish groups in Indonesia are limited.

In conclusion, this research successfully identified the diversity of Carangid fish landed in several fishery centers in Java. Of the 33 carangids fish, the most extensive composition obtained from these locations was *Megalaspis cordyla* (4), *Selaroides leptolepis* (4), *Atule mate* (3), and *Decapterus macarellus* (3). Generally, these fish are consumed locally to meet people's protein needs in coastal areas of Java. Phylogenetic analysis shows that the suborder Caranginae and Scomberoidinae are separated. Fish in the Caranginae suborder dominate compared to other sub-orders. Scomberoidinae in this study has unique morphological characteristics that are sometimes confused with the Scombridae group. This reinforces the use of molecular identification to complement morphological identification.

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