Snapper by Sapto Andriyono

Submission date: 09-Jan-2023 05:05PM (UTC+0800)

Submission ID: 1990126672

File name: 48702-161358-2-PB.pdf (339.27K)

Word count: 4858 Character count: 25292

Molecular Identification of Snapper (Perciformes: Lutjanidae) Landed at Pondokdadap Fishing Port of Sendang Biru, Malang, Indonesia

Sapto Andriyono^{1*}, Novian Aji Pradana², Laksmi Sulmartiwi¹, Andi Aliah Hidayani³, Md. Jobaidul Alam⁴, Adrian Damora⁵, Ahasan Habib⁶

¹Department of Marine, Fisheries and Marine Faculty, Universitas Airlangga
 ²Aquaculture Study Program, Faculty of Fisheries and Marine, Universitas Airlangga
 JI. Wijaya Kusuma No. 13 Banyuwangi, Jawa Timur 68411 Indonesia
 ³Fisheries Department, Faculty of Marine Science and Fisheries, Hasanuddin University
 JI. Perintis Kemerdekaan No.Km. 10 Makassar, Sulawesi Selatan 90245 Indonesia
 ⁴Department of Fisheries, Ministry of Fisheries and Livestock, Dhaka, Bangladesh
 Dhaka 1205, Bangladesh

5 Department of Aquaculture, Faculty of Marine and Fisheries, Universitas Syiah Kuala

Jl. Teuku Nyak Arief No.441, Kopelma Darussalam, Syiah Kuala, Kota Banda Aceh, Aceh 23111 Indonesia

6 Faculty of Fisheries and Food Science, Universiti Malaysia Terengganu
21030 Kuala Nerus, Terengganu, Malaysia

Email: sapto.andriyono@fpk.unair.ac.id

Abstract

Snapper is a type of demersal marine fish from the Lutjanidae family. The Lutjanidae family spread throughout the world and currently has 123 species in 21 genera, one of which is the Lutjanus genus (Miller and Thomas, 2007). To this day, the records of capture fisheries production data for snapper in Malang are still very limited to certain types. Morphological identification that has been carried out so far is still challenging to obtain accurate results because of the many similarities between the observed species or the loss of characteristics. Therefore, molecular identification is necessary to determine the types of snappers in this area and their conservation status. This study aims to determine the types of snappers using a molecular approach by Cytochrome Oxidase subunit I (COI) gene marker. Phylogenetic tree reconstruction and genetic distance calculations were performed using Mega X software through the neighbour-joining (NJ) algorithm. The results of the identification snapper based on a molecular approach with DNA barcoding revealed that the four snapper samples were L. gibbus, L. rufolineatus, L. bengalensis, and L. erythropterus. Based on the results of the compilation of the phylogenetic tree, it can be seen that the L. bengalensis sample is closely related to L. rufolineatus while L. gibbus, and L. erythropterus each form a separate clade from the two previous Lutjanus species. Based on their conservation status at the IUCN, the four species of snapper found are in the Least Concern category.

Keywords: diversity, gene, identification, phylogenetic, snapper

Introduction

Sendang Biru is one of the coastal areas that prioritise managing marine fisheries resources in Malang Regency, East Java (Andriyono et al., 2019). The development makes Sendang Biru a centre for the capture industry (Aliviyanti et al., 2021). Snapper is one of the fish catches obtained on this region (Luthfi et al., 2016). Snapper is a type of demersal fish of the family Lutjanidae. The family Lutjanidae, spread throughout the world, currently has 123 species in 21 genera, one of which is the genus Lutjanus (Miller and Cribb, 2007). Based on morphology and habitat characteristics, 30 species of snapper from the genus Lutjanus found in Indonesian waters (Allen et al., 2013, Halim et al., 2020).

Snapper in nature plays the role as one of the large-sized apex predatory fishes that inhabit tropical coastal ecosystems around the world. Ecologically, the existence of this fish is important because it acts as a peak predator with extensive food habits. This fish can eat other small fish, cephalopods, crabs, shrimps, and other benthic crustaceans to control the stability of the aquatic ecosystem in which it lives (Simonsen et al., 2015). Snapper is also one of the captured fishery commodities that is usually used as consumption fish sold in the form of fresh fish, fillets, and processed products (Oktaviyani, 2018). The production of this fish has increased every year. This follows data from the Central Statistics Agency (BPS) of Malang Regency (2020), where the total production of this fish reached 57.05 tons in 2018 and 2019 to 108.24 tons. Based on data from BPS

Received: 02-09-2022

Accepted: 19-11-2022

Malang Regency (2020), recording data on the production of capture fisheries for snapper in Malang is still very limited to certain types. This is due to the difficulty of identifying species in the field and at the time of simultaneous landing with other types of fish at the fish auction site. Identification of a species can be made morphologically as well as molecularly. Morphological identification that has been carried out so far is still challenging to obtain accurate results because of the many similarities between the observed specifications. In addition, the loss of distinctive features in observed species due to adaptation to the environment is also an obstacle in identifying a species morphologically (Prehadi et al., 2015).

One alternative to identify that can be done in addition to morphological is molecular identification by DNA barcoding. DNA barcoding is a globally agreed method for identifying plant and animal species based on DNA sequence variations (Coissac et al., 2016) from nitrogenous base pair regions in the Cytochrome Oxidase subunit I (COI) gene (Powers et al., 2018). Since its introduction in 2003, the DNA barcoding technique has become the golden standard or the main standard for molecular taxonomy (Fadli et al., 2020). DNA-based identification barcoding has been well received globally for its various advantages, such as being very simple and using a universal tool applicable to all organisms, both in fresh samples and processed products (Kress et al., 2015). Some examples of research that utilizes the DNA barcoding technique include the use of DNA barcoding to identify fish larvae at different stages of development (Wibowo et al., 2018), identification of the discovery of new and cryptic fish species (Farhana et al., 2018), identification of fish species that 5 have similar morphological characters (Bingpeng et al., 2018).

DNA barcoding is effective for identifying a species with fast and accurate results based on the Cytochrome Oxidase subunit I (COI), even if specimen are larvae (Li et al., 2016). The COI gene is one of the protein-encoding genes found in mitochondria that has a distinctive character in each species so it becomes a standard gene as a marker gene when identifying an animal species (Pentinsaari et al., 2016). Therefore, the COI genes in DNA barcoding has two advantages, not only for species identification and for metabarcoding (Andújar et al., 2018), (Tan et al., 2019) as well. Therefore, research on the identification of snappers based on DNA markings of the COI gene needs to be carried out to provide genetic information. It is expected to be supporting data in the management of conservation areas and fishing zones in the waters of South Malang, especially from Sendang Biru.

Materials and Methods

Sampling of fish

A total of 4 samples were collected from traditional fish market of Pondokdadap fishing port at Sendang Biru, Malang in the middle of March 2020. All samples collected from the local traditional fisherman were dead upon purchase. The digital camera was used to take the individual photograph before further treatments. Morphologically, identification and species confirmation were carried out with molecular identification in this study. No specific permit was required for this study.

DNA extraction and PCR amplification

Each specimen has been collected based on the morphological characters and directly preserved in 90% ethanol for further experimental purposes. Genomic DNA extracted using an Accuprep® Genomic DNA Extraction Kit (Bioneer) according to the product guidelines. The pereiopod fin, around 1 cm tissues, was dissected and mixed with 6X lysis buffer, which was further homogenized by the TissueLyser II (Qiagen). Quantification of purified genomic DNA performed by nanoDrop (Thermofisher Scientific D1000), aliquoted and stored at the -70°C for further analysis.

One set of universal fish primer targeting cytochrome c oxidase I (COI) region, BCL-BCH were used to obtain the partial sequences of each gene (Madduppa et al., 2016). The PCR mixture (20µL) included 11.2 µL ultra-pure water, 1 µL primer forward and reverse (0.5 µM), 0.2 µL Ex Taq DNA polymerase (TaKaRa, Japan), 2 µL 10X ExTag Buffer, 2 µL dNTPs (1 µM, TaKaRa, Japan), and 2 µL genomic DNA as template. The PCR condition carried out under the following setting: 95°C for 5 min in initial denaturation, followed by denaturation at 95°C for 30 s in 40 cycles, 50°C for 30 s in annealing, and 72°C for 45 s in extension step, and a final extension at 72°C for 5 min. The PCR products purified with the AccuPrep®Gel purification kit (Bioneer, Korea).

Data analysis

Forward and reverse sequence were edited and aligned using MEGAX (Kumar et al., 2018). All sequences were then aligned to the reference on GenBank database by BLASTN (https://blast.ncbi.nlm.nih.gov/Blast.cgi). The pairwise evolutionary distance among the family determined by the Kimura 2-Parameter method. The Neighbor-joining (NJ) tree constructed, and 1000 bootstrap analysis was carried by Mega X and genetic distance used a nucleotide substitution model by comparing a DNA sequence of one nucleotide with another nucleotide (Kumar et al., 2018).

Result and Discussion

Morphological identification

The samples obtained from the Sendang Biru traditional fish market were 4 snappers with different species consisting of *Lutjanus bengalengsis*, *L. rufolineatus*, *L. gibbus*, and *L. erythropterus*. The most striking difference between each species is each snapper sample's color and body pattern (Figure 1.). In addition to body color and pattern, morphological identification also observed morphometric and meristic characters in snapper samples (Table 1.).

Molecular identification

Molecular identification of snapper samples was carried out using the DNA barcoding method. The sequence data obtained were then analyzed and matched with the sequences found in GenBank at NCBI (National Center for Biotechnology Information) using BLASTN (Basic Local Alignment Search Tool Nucleotide) based on the degree of similarity (Table 2). Based on the results of the BLASTN analysis, sample MLKK1 was identified as having 96.72% similarity to the species L. gibbus (Humpback red snapper) access number MF409615, sample MLKK2 was identified to have 99.19% similarity to the species L. rufolineatus (Yellow lined snapper) access number MN870411, sample MLKK3 was identified as having 99.54% similarity to L. bengalensis (Bengal snapper) access number EU600137, while MLKK4 sample was identified to have 100% similarity to L. erythropterus (Crimson snapper) species access number GU673841.

Phylogentic tree reconstruction

Based on the results of the phylogenetic tree reconstruction (Figure 2.), samples of snapper landed at the Pondokdadap Sendang Biru Fishing Port, obtained 4 clades formed in the family Lutjanidae with the genus Lutjanus. The clade L. bengalensis is phylogenetically close to the clade L. rufolineatus, while the clade L. gibbus is closely related to the clade L. erythropterus. Both L. bengalensis (MLKK3) and L. rufolineatus (MLKK2) has low genetic distance with close species from other region (Table 3.), but L erythropterus from Malang has sifnificant genetic distance with sam species from Australia (0.18) and Malaysia (0.19). As a fish associated with coral reef ecosystems, Lutjanus fish species make coral reefs a habitat for rearing ground and feeding ground (Halim et al., 2020, Tony et al., 2020). The coral reef habitat will experience different speciation in each region. The Indian Ocean area has different characteristics from the Malaysian waters (South China Sea) and the Australian area which is influenced by the Pacific Ocean which is the main barrier in the distribution of shallow marine fish species. This pattern is known as allopatric speciation (Rocha and Bowen, 2008).

The diversity of potential marine fish in Indonesia needs serious attention (Suman et al., 2017). Not only in sustainable management (Atmaja and Nugroho, 2017), accurate species determination is also a must in providing a valid database at the species level. Many morphological identifications have been carried out. However, in marine fish species there are morphological similarities in both shape and color which causes confusion and inconsistency in naming fish species. In this study,

Table 1. Morphometric and Meristic Measurements of Snapper Samples

		Sar	nple ID	
Parameters	MLKK1	MLKK2	MLKK3	MLKK4
Total length	26,3 cm	26,8 cm	27,5 cm	26,4 cm
Standard length	21, <mark>5</mark> cm	21,5 cm	22,5 cm	20,5 cm
Head length	8 cm	8,3 cm	8,6 cm	8,2 cm
Height	8,5 cm	8,6 cm	10,8 cm	11,2 cm
Head height	6,5 cm	6,8 cm	9,2 cm	8,6 cm
Tail base height	2,5 cm	2,8 cm	3,0 cm	2,8 cm
Dorsal fin	D.X, 14	D.X, 14	D.XI, 14	D.XI, 14
Pectoral fin	P.17	P.16	P.16	P.17
ventral fin	V.I,6	V.I,6	V.I,6	V.I,5
anal fin	A.III.8	A.III, 8	A.III, 8	A.III, 9
caudal fin	C.18	C.18	C.20	C.18





A. Sampel MLKK1 L. gibbus

B. Sampel MLKK2 L. rufolineatus





C. Sampel MLKK3 L. bengalensis

D. Sampel MLKK4 L. erythropterus

Figure 1. Four species of Snappers Landed at Pondokdadap Fishing Port in traditional fish market of Sendang Biru, South Malang.

Table 2. BLASTN Results of Snapper Samples with NCBI GenBank Database

No.	Sample Code	21 Species Name/ Common Name	No. Access GenBank	Identity (%)
1.	MLKK1	Lutjanus gibbus / Humpback red snapper	MF409615	96,72%
2.	MLKK2	Lutjanus rufolineatus / Yellow lined snapper	MN870411	99,19%
3.	MLKK3	Lutjanus bengalensis / Bengal snapper	EU600137	99,54%
4.	MLKK4	Lutjanus erythropterus / Crimson snapper	GU673841	100%

apart from observing specimens based on morphological characteristics, a molecular approach was used to improve data accuracy in identification at the species level. Of the 4 specimens collected, the morphological characteristics showed that the specimens were able to be identified based on their morphometric characteristics, so that all samples were identified as *L. bengalensis*, *L. rufolineatus*, *L. gibbus*, and *L. erythropterus*. To increase accuracy in identification, we also carried out molecular identification of the COI gene section (Andriyono and Suciyono, 2020) which has been agreed as a universal area for identification at the species level.

Morphological observations on the four samples showed that the MLKK1 sample had similarities with Lutjanus gibbus/ humpback red snapper, also known as jinaha snapper. The distinctive features or key morphological identification of this fish are having a compressed body shape with a grayish red body color, the caudal fin is clearly branched with dark red rounded lobes, and on the dorsal fin there are 10 hard spines and 13-14 soft spines (Thi et al., 2015). The MLKK2 sample has similarities with Lutjanus rufolineatus/ yellow lined snapper, also known as badur snapper. The key to morphological identification of the L. rufolineatus species is that there are 6 yellow stripes on each side of its body, it has a pale red compressed body shape, the tail is brownish yellow, and on the dorsal fin there are 10 hard spines and 12-13 soft spines (Allen et al., 2013).

The MLKK3 sample is similar to the Lutjanus bengalensis/bengal snapper species, also known as yellow snapper with a key Identification. The body of this fish is compressed, the body color is bright yellow with 4 grayish white stripes on each side of the body, there is a deep groove on the front operculum, the caudal fin is broad with a straight tip, has 11 spines and 12-14 soft rays on the dorsal fin (Iwatsuki et al., 2016). The MLKK4 sample has similarities to the L. erythropterus/ crimson snapper or often referred to as the red snapper. The identification key in common in the form of having a pink to dark red compressed body shape from the tip of the head to the tail. Another characteristic are the tip of the snout is slightly pointed and relatively small. Besides, the preoperculum notch is not very pronounced and on the dorsal fin there are 11 hard spines and 16-17 soft spines (Sarkar et al., 2021).

Reconstruction of the phylogenetic tree of snapper samples landed at the Pondokdadap Fishing Port, Sendang Biru, obtained 4 clades formed in the family Lutjanidae with the genus Lutjanus. The clade *L. bengalensis* is phylogenetically close to the clade *L. rufolineatus*, while the clade *L. gibbus* is closely related to the clade *L. erythropterus*. In the phylogenetic tree reconstruction, there is also a clade

of the Nemipterus virgatus species as a comparison or outgroup. Reconstruction of phylogenetic trees is supported by the results of genetic distance analysis in a species (Akbar and Labenua 2018). The results of the genetic distance analysis showed that the MLKK3 sample was close to the L. bengalensis EU600137 (China) and LC075762 (Indian Ocean) samples, with a genetic distance of 0.00 (zero). The MLKK2 specimen was closely related to the L. rufolineatus specimen MN870411 (Indonesia) and had a genetic distance of 0.01. The MLKK1 specimen was closely related to L. gibbus MN870581 (Ambon, Indonesia), MK566973 (France) and MF409615 (Reunion) with a genetic distance of 0.01 each. The MLKK4 specimen was closely related to L. erythropterus specimens GU673841 (Indonesia) and GU673202 (Australia) with a genetic distance of 0.00 and 0.01, respectively. Research on the Lutjanidae species in peninsular Malaysia (Malacca Strait and South China Sea) also shows that there is a variation in genetic distance (Halim et al., 2022).

Based on the conservation status that refers to the IUCN (International Union for the Conservation of Nature and Natural Resources), L. gibbus, L. rufolineatus, L. bengalensis, and L. erythropterus species are included in the Least Concern or low risk category (IUCN Red List, 2021). Least Concern is a species that has been evaluated but its status is still under the status of almost endangered or it can be said that it does not fall into any category. The IUCN conservation status categories include the category of extinction (EX), category of extinction in the wild (EW), category of critically (CR), category of threatened or critical (EN), category of vulnerable (VU), category of near threatened (NT), the category of low risk (LC) and the category of lack of information (DD) (https://www.iucnredlist.org/). Then based on their trading status according to CITES, these four snapper species are included in the Not Evaluated category, so that they are still classified as safe for international trade.

Table 3. Genetic Distance of Snapper COI gene Sequences from Sendang Biru with Snapper COI gene Sequences on NCBI GenBank

N.o.	Name of Cassin							7			10		10	12
No.	Name of Spesies MLKK1 Lutianus	1	2	3	4	5	6	7	8	9	10	11	12	13
	gibbus													
2.	MN870581 <i>L. gibbu</i> s (Ambon)	0,01			_									
3.	MK566973 <i>L. gibbus</i> (Perancis)	0,01	0,00											
4.	MF409615 <i>L. gibbus</i> (Reunion)	0,01	0,00	0,00	┛									
5.	MLKK2 Lutjanus rufolineatus	0,17	0,16	0,16	0,16									
6.	MN870411 L.rufolineatus (Ambon)	0,16	0,15	0,15	0,15	0,01								
7.	MLKK3 Lutjanus bengalensis	0,17	0,16	0,16	0,16	0,07	0,06							
8.	EU600137 L.bengalensis (China)	0,17	0,16	0,16	0,16	0,06	0,05	0,00						
9.	LC075762 <i>L.</i> bengalensis (Indian Ocean)	0,17	0,16	0,16	0,16	0,07	0,06	0,00	0,00					
10.	MLKK4 Lutjanus erythropterus	0,18	0,17	0,17	0,17	0,17	0,16	0,18	0,17	0,18	J			
11.	GU673841 L. erythropterus (Australia)	0,18	0,17	0,17	0,17	0,17	0,16	0,18	0,17	0,18	0,18	0,00		
12.	GU67202 L. erythropterus (Malaysia)	0,19	0,18	0,18	0,18	0,18	0,17	0,19	0,18	0,19	0,19	0,01	0,01	
13.	KP112336 Nemipterus virgatus (China)	0,24	0,23	0,23	0,23	0,21	0,20	0,21	0,21	0,21	0,21	0,21	0,21	0,20

Conclusion

Based on morphological and molecular identification, the types of snappers that landed from Sendang Biru, South Malang waters were *Lutjanus bengalensis*, *L. rufolineatus*, *L. gibbus*, and *L. erythropterus*. Based on the results of the compilation of the phylogenetic tree, it can be seen that the *L. bengalensis* sample is closely related to *L. rufolineatus* while *L. gibbus*, and *L. erythropterus* each form a separate clade from the two previous Lutjanus species. Based on their conservation status at the IUCN, the four species of snapper found are in the Least Concern category, while based on their trading status on CITES, these four species are in the Not Evaluated category.

Acknowledgement

We would like to deliver our gratitude to the PUF Research Grant 2020 internal fund research program from Faculty of Fisheries and Marine Affairs, Universitas Airlangga has been providing support in this research. We also thank the research team colleagues who have helped in the sample collection in South Malang, East Java

References

- Akbar, N.N. & Labenua, R. 2018. Keragaman genetik ikan cakalang (*Katsuwonus pelamis*) di perairan laut Maluku Utara. *Depik* 7: 164-176. https://doi.org/10.13170/depik.7.2.11156
- Aliviyanti, D., Semedi, B., Yona, D., Asadi, M.A., Kasitowati, R.D., Dewi, C.S.U., Lutfi, O.M. & Isdianto, A., 2021. Upaya Penguatan Manajemen Pemasaran Hasil Perikanan Berbasis Media Online di TPI Sendangbiru, Kabupaten Malang, Indonesia. Abdi Geomedisains, 1(2): 59-67. https://doi.org/10. 23917/abdigeomedisains.v1i2.199
- Allen, G.R., Erdmann, M.V., Randall, J.E., Ching, P., Rauzon, M.J., Hayashi, L.A., Thomas, M.D., Robertson, D.R., Taylor, L. & Coste, M., 2013. Reef fishes of the East Indies. *Philosophy East and West*, 63(2): p. 1292
- Andriyono, S. & Suciyono, S. 2020. Molecular Identification and Phylogenetic Tree Reconstruction of Marine Fish from the Essential Wetland Area of Banyuwangi, Indonesia. Egypt. J. Aquat. Biol. Fish, 24: 427-439. https://doi.org/ 10.21608/ejabf.2020.87032
- Andriyono, S., Alam, M.J. & Kim, H.W. 2019. Environmental DNA (eDNA) metabarcoding:

- Diversity study around the Pondok Dadap fish landing station, Malang, Indonesia. *Biodivers. J.* 20(12): 3772-3781. https://doi.org/10.13057/biodiv/d201241
- Andújar, C., Arribas, P., Yu, D.W., Vogler, A.P. & Emerson, B.C. 2018. Why the COI barcode should be the community DNA metabarcode for the metazoa. *Mol. Ecol.*, 27: 3968–3975 https://doi.org/10.1111/mec.14844
- Atmaja, S.B. & Nugroho, D. 2017. Upaya-upaya pengelolaan sumber daya ikan yang berkelanjutan di Indonesia. *J. Kebijakan Perikanan Indonesia*, 3: 101-113. https://doi.org/ 10.15578/jkpi.3.2.2011.101-113
- Bingpeng, X., Heshan, L., Zhilan, Z., Chunguang, W., Yanguo, W. & Jianjun, W. 2018. DNA barcoding for identification of fish species in the Taiwan Strait. PloS One, 13: e0198109. https://doi.org/ 10.1371/journal.pone.0198109
- Coissac, E., Hollingsworth, P.M., Lavergne, S. & Taberlet, P. 2016. From barcodes to genomes: extending the concept of DNA barcoding. *Mol. Ecol.*, 25: 1423–1428 https://doi.org/10.11 11/mec.13549
- Fadli, N., Nor, S.A.M., Othman, A.S., Sofyan, H. & Muchlisin, 2020. DNA barcoding of commercially important reef fishes in Weh Island, Aceh, Indonesia. Peer J. 8: e9641. https://doi.org/ 10.7717/peerj.9641
- Farhana, N.S., Muchlisin, Z.A., Duong, T.Y., Tanyaros, S., Page, L.M., Zhao, Y., Adamson, E.A., Khaironizam, M., de Bruyn, M. & Siti A.M.N., 2018. Exploring hidden diversity in Southeast Asia's Dermogenys spp. (Beloniformes: Zenarchopteridae) through DNA barcoding. Sci. Rep., 8: 1-11. https://doi.org/10.1038/s415 9 8-018-29049-7
- Halim, L.J., Rahim, I., Mahboob, S., Al-Ghanim, K.A., Asmiaty, A.M.A.T. & Naim, D.M. 2022. Phylogenetic relationships of the commercial red snapper (*Lutjanidae* sp.) from three marine regions. *J. King Saud. Univ. Sci.*, 34: p.101756. https://doi.org/10.1016/j.jksus.2021.101756
- Halim, A., Loneragan, N.R., Wiryawan, B., Hordyk, A.R., Sondita, M.F.A. & Yulianto, I., 2020. Evaluating data-limited fisheries for grouper (Serranidae) and snapper (Lutjanidae) in the Coral Triangle, eastern Indonesia. Reg. Stud. Mar. Sci., 38: p.101388. https://doi.org/10.10 16/j.rsma.2020.101388

- Iwatsuki, Y.U.K.I.O., Al-Mamry, J.M. & Heemstra, P.C. 2016. Validity of a blue stripe snapper, Lutjanus octolineatus (Cuvier 1828) and a related species, L. bengalensis (Bloch 1790) with a new species (Pisces; Lutjanidae) from the Arabian Sea. Zootaxa, 4098: 511-528. https://doi.org/ 10.11646/zootaxa.4098.3.5
- Kress, W.J., García-Robledo, C., Uriarte, M. & Erickson, D.L., 2015. DNA barcodes for ecology, evolution, and conservation. *Trends Ecol. Evol.*, 30: 25-35. https://doi.org/10.1016/j.tree.2014. 10.008
- Kumar, S., Stecher, G., Li, M., Knyaz, C. & Tamura, K.. 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol. Biol. Evol.*, 35(6): p.1547. https://doi.org/10.1093/molbev/msy096
- Li, Q., Li, M., Li, D., Shi, W., Li, J., Ning, T. & Duan, Y. 2016. Identification of the Larvae of Helicoverpa assulta and H. Armigera (Lepidoptera: Noctuidae) Through Morphological Difference and DNA Barcoding. 3rd Int. Conf. Smart Mat. Nanotechnol. Eng. pp.306-309.
- Luthfi, O.M., Pujarahayu, P., Wahyudiarto, A., Fakri, S.R., Sofyan, M., Ramadhan, F., Murian, S., Tovani, I., Mahmud, M., Adi, D. & Abdi, F. 2016. Biodiversitas dan populasi ikan karang di perairan selat sempu sendang biru kabupaten malang jawa timur. J. Kelautan: Indonesian J. Mar. Sci. Technol. 9: 43-49. https://doi.org/10.21107/jk.v9i1.1019
- Madduppa, H., Ayuningtyas, R.U., Subhan, B. & Arafat, D. 2016. Exploited but unevaluated: DNA barcoding reveals skates and stingrays (Chordata, Chondrichthyes) species landed in the Indonesian fish market. Ilmu Kelautan: Indonesian Journal of Marine Science, 21: 77-84. https://doi.org/10.14710/ik.ijms.21.2.77-84
- Miller, T.L. & Cribb, T.H. 2007. Phylogenetic relationships of some common Indo-Pacific snappers (Perciformes: Lutjanidae) based on mitochondrial DNA sequences, with comments on the taxonomic position of the Caesioninae. *Mol. Phylogenet. Evol.*, 44: 450-460. https://doi.org/10.1016/j.ympev.2006.10.029
- Oktaviyani, S. 2018. Mengenal marga Lutjanus, salah satu komoditas unggulan dalam perikanan tangkap. Oseana 43: 29-39. https://doi.org/10.14203/oseana.2018.Vol.43No.3.61

- Pentinsaari, M., Salmela, H., Mutanen, M. & Roslin, T. 2016. Molecular evolution of a widely-adopted taxonomic marker (COI) across the animal tree of life. Sci. Rep. 6: 1-12. https://doi.org/10.1038/srep35275
- Powers, T., Harris, T., Higgins, R., Mullin, P. & Powers, K. 2018. Discovery and Identification of Species Using COI DNA Barcoding. J. Nematol. 50: 399-412. https://doi.org/10.21307/jofnem-2018-029
- Prehadi, P., Sembiring, A., Kurniasih, E.M., Rahmad, R., Arafat, D., Subhan, B. & Madduppa, H.H. 2015. DNA barcoding and phylogenetic reconstruction of shark species landed in Muncar fisheries landing site in comparison with Southern Java fishing port. *Biodivers. J.*, 16(1): 55-61. https://doi.org/10.13057/biodiv/d160107
- Rocha, L. & Bowen, B. 2008. Speciation in coral-reef fishes. J. Fish Biol. 72: 1101-1121. https://doi.org/ 10.1111/j.1095-8649.2007.01770.x
- Sarkar, P., Islam, M.J., Habib, A.H.M., Neogi, A.K. & Habib, K.A. 2021. Two New Records of Snapper (Perciformes, Lutjanidae) from Saint Martin's Island, Bangladesh. J. Ocean Univ. China, 20: 439-444. https://doi.org/10.1007/s11802-02 1-4566-x
- Simonsen, K.A., Cowan, J.H. & Boswell, K.M. 2015. Habitat differences in the feeding ecology of red snapper (*Lutjanus campechanus*, Poey 1860): a comparison between artificial and natural reefs in the northern Gulf of Mexico. *Environ. Biol. Fishes*, 98: 811-824. https://doi.org/10.1007/ s10641-014-0317-9
- Suman, A., Irianto, H.E., Satria, F. & Amri, K. 2017. Potensi dan tingkat pemanfaatan sumber daya ikan di wilayah pengelolaan perikanan Negara Republik Indonesia (WPP NRI) Tahun 2015 serta Opsi Pengelolaannya. J. Kebijakan Perikanan Indonesia, 8: 97-100. https://doi.org/10.155 78/jkpi.8.2.2016.97-100
- Thi, O.T., Ha, Q.V.D. & Thuy, B.D. 2015. Phylogenetic relationships of emperors (Lethrinidae) and Snappers (Lutjanidae) in Vietnam based on mitochondrial DNA sequences. Pages 15-16. Int. Conf. Biol. Environ. Food Eng. Singapore May. 15-16 pages.
- Tony, F., Soemarno, S., Wiadnya, D.G.R. & Hakim, L. 2020. Diversity of reef fish in Halang Melingkau Island, South Kalimantan, Indonesia. *Biodiv. J.* 21 (10): 4804-4812. https://doi.org/10.13057/biodiv/d211046.

ILMU KELAUTAN: Indonesian Journal of Marine Sciences December 2	022 Vol 27(4):307-314
Wibowo, A., Panggabean, A.S., Zamroni, A., Priatna, A. & Yusuf, H.N. 2018. Using DNA barcode to improve the identification of marine fish larvae,	case study coastal water near Jakarta and Banda Sea, Indonesia. <i>Indones. Fish. Res. J.</i> , 24: 23-30. https://doi.org/10.15578/ifrj.24.1.2018.37-44

Snapper

ORIGINALITY REPORT

6% SIMILARITY INDEX

1%
INTERNET SOURCES

6%
PUBLICATIONS

)%

STUDENT PAPERS

PRIMARY SOURCES

1 Bhi MY

Bhuiyan, M.S.A.. "Identification of SNPs in MYOD gene family and their associations with carcass traits in cattle", Livestock Science, 200912

<1%

Publication

Agus Alim Hakim, Yessi Natalina Sidabutar, Ali Mashar, Zairion, Zulhamsyah Imran, Yusli Wardiatno. "Application of cytochrome oxidase subunit 1 partial gene for species validation of Macrobrachium sintangense from Lake Lido, West Java", IOP Conference

Series: Earth and Environmental Science, 2022

<1%

Publication

Dian Adi Syahputra, Munthadar M. Isa, Muhammad Jailani. "Catastrophic appendicitis operation, successful repair in tertiary limited resources hospital: A case report", Annals of Medicine and Surgery, 2021

<1%

Publication

4

Estefany Chavarria-Quicaño, Victor Contreras-Jácquez, Armando Carrillo-Fasio, Francisco De

<1%

la Torre-González et al. "Bacteria from the Sonoran Desert with nematicidal potential against phytopathogenic nematodes", Research Square Platform LLC, 2022

Publication

H S Farizky, W H Satyantini, D D Nindarwi. "
The efficacy of probiotic with different storage to decrease the total organic matter, ammonia, and total on shrimp pond water ", IOP Conference Series: Earth and Environmental Science, 2020

<1%

Publication

P. P. Glenning. "The Short Primigravida", The Australian and New Zealand Journal of Obstetrics and Gynaecology, 1965

<1%

Publication

Syazwani Aqilah Alias, Norizah Mhd Sarbon.
"Rheological, physical, and mechanical properties of chicken skin gelatin films incorporated with potato starch", npj Science of Food, 2019

<1%

Publication

M Lukman, N Fitriya, H Prayitno, A Afdal, A Nasir, T Kaisupi, P Avianto, U E Hernawan. "LIPI COREMAP-CTI Nusa Manggala oceanographic survey, I: oceanic mesozooplankton community from epipelagic zones of North-Eastern part of the Indonesian

<1%

waters -adjacent to the Southwest Pacific Ocean", IOP Conference Series: Earth and Environmental Science, 2020

Publication

"Reef Fish Spawning Aggregations: Biology, Research and Management", Springer Nature, 2012

<1%

Publication

Jae Yong Jung, Ah Reum Choi, Yoo Kyung Lee, Hong Kum Lee, Kwang-Hwan Jung. "Spectroscopic and photochemical analysis of proteorhodopsin variants from the surface of the Arctic Ocean", FEBS Letters, 2008

<1%

Publication

Cecilia Chu, Mohammed Rizman-Idid, Chong Ving Ching. "Phylogenetic relationships of selected genera of Lutjanidae inferred from mitochondrial regions, with a note on the taxonomic status of Pinjalo pinjalo", Ciencias Marinas, 2013

<1%

Publication

Lia Juliana Halim, Iradhatullah Rahim, Shahid Mahboob, K.A. Al-Ghanim, Darlina Md Naim. "Phylogenetic relationships of the commercial Red Snapper (Lutjanidae sp.) from three marine regions", Journal of King Saud University - Science, 2021

<1%

Publication

Young-Hwa Kim, Gyung Hye Huh. "Members of the ran family of stress-inducible small GTP-binding proteins are differentially regulated in sweetpotato plants", Journal of Plant Biotechnology, 2013

<1%

Publication

A Rahmah, A M Makhfirah, A Damora, E Miswar, R M Aprilla, A Sembiring. "Vulnerability status of longtail tuna in Southwest Aceh waters, Indonesia ", IOP Conference Series: Earth and Environmental Science, 2019

<1%

Publication

Giorgi Epitashvili, Matthias Geiger, Jonas Astrin, Fabian Herder, Bella Japoshvili, Levan Mumladze. "Towards retrieving the Promethean treasure: a first molecular assessment of the freshwater fish diversity of Georgia", Biodiversity Data Journal, 2020

<1%

<1%

I. Stolarek, L. Handschuh, A. Juras, W. Nowaczewska, H. Kóčka-Krenz, A. Michalowski, J. Piontek, P. Kozlowski, M. Figlerowicz. "Goth migration induced changes in the matrilineal genetic structure of the central-east European population", Scientific

Publication

Reports, 2019

17	Ultraconserved Elements Alongside COI to Strengthen Metabarcoding Studies", Frontiers in Ecology and Evolution, 2019 Publication	<1%
18	Xiao-Xuan Zhang, Qi-Dong Tan, Dong-Hui Zhou, Xiao-Ting Ni, Guang-Xue Liu, Yan-Chuan Yang, Xing-Quan Zhu. "Prevalence and molecular characterization of Cryptosporidium spp. in dairy cattle, northwest China", Parasitology Research, 2015 Publication	<1%
19	Patricia Mariana Domingues. "Sub-Saharan Africa descendents in Rio de Janeiro (Brazil): population and mutational data for 12 Y-STR loci", International Journal of Legal Medicine, 04/13/2007 Publication	<1%
20	G. Waelchli. "Mikrospektroskopische Untersuchungen der gefärbten Kugeln in der Retina von Vögeln", Albrecht von Græfe's Archiv für Ophthalmologie, 1881	<1%
21	Richard N. Muallil, Ahalnida M. Tambihasan,	_1

Marylyn J. Enojario, Yunadzmal N. Ong, Cleto

L. Nañola. "Inventory of commercially

important coral reef fishes in Tawi-Tawi

Islands, Southern Philippines: The Heart of the Coral Triangle", Fisheries Research, 2020

Off

Publication

Exclude quotes On Exclude matches

Exclude bibliography On

Snapper

GRADEMARK REPORT

FINAL GRADE

/100

GENERAL COMMENTS

Instructor

PAGE 1
PAGE 2
PAGE 3
PAGE 4
PAGE 5
PAGE 6
PAGE 7
PAGE 8