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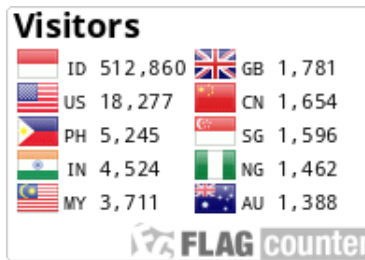
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Pengaruh Lama Perendaman Asam Sitrat terhadap Kadar Logam Berat Timbal (Pb) pada Daging Lorjuk (*Solen* sp.)

The Effect of Citric Acid Soaking Time on The Levels of Lead (Pb) in Lorjuk Meat (*Solen* sp.)

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Abstrak

Timbal (Pb) merupakan salah satu bahan pencemar pada lingkungan perairan yang sering dipermasalahkan karena bersifat toksik dan berbahaya terhadap biota perairan, serta berdampak tidak langsung terhadap manusia yang mengonsumsinya. Lorjuk (*Solen* sp.) merupakan salah satu biota perairan yang dapat mengakumulasi timbal dalam tubuhnya karena merupakan *filter feeder*. Kadar logam berat timbal yang terakumulasi dalam tubuh lorjuk dapat diturunkan dengan cara melakukan perendaman menggunakan *chelating agent*, seperti asam sitrat karena dapat mengikat logam sehingga membebaskan bahan pangan dari cemaran logam seperti timbal. Tujuan penelitian ini yaitu untuk mengetahui pengaruh perbedaan lama perendaman dengan asam sitrat terhadap kadar logam berat timbal (Pb) serta lama perendaman optimal untuk menurunkan kadar logam berat timbal (Pb) pada daging lorjuk. Penelitian ini menggunakan Rancangan Acak Lengkap (RAL) yang terdiri dari tiga perlakuan perbedaan lama perendaman daging lorjuk (1,5 jam; 3 jam dan 4,5 jam). Hasil penelitian menunjukkan bahwa lama perendaman daging lorjuk dengan asam sitrat selama 4,5 jam merupakan perlakuan terbaik untuk menurunkan kadar logam berat dalam daging lorjuk sebesar 14,38%. Nilai organoleptik daging lorjuk setelah perendaman 4,5 jam adalah kenampakan 6,07 (utuh, warna daging spesifik jenis, cerah dan bersih), bau 7,40 (sangat segar) dan tekstur 7,07 (elastis, padat dan kompak); kadar air 78,25%; kadar protein 8,67%; kadar lemak 0,65% dan kadar abu 0,61%.

Kata kunci: asam sitrat, perendaman, *Solen* sp., timbal

Abstract

Lead (Pb) is one of the pollutants in the aquatic environment that is often questioned because it has toxic and dangerous properties for aquatic biota and indirect impacts on humans who consume it. Lorjuk (*Solen* sp.) is one of the aquatic biotas that can accumulate lead in its body because it is a filter feeder. The levels of lead accumulated in the body of lorjuk can be reduced by immersion using a chelating agent, such as citric acid because can bind metals thereby freeing food from metal contamination such as lead. This study aimed to determine the effect of different soaking times with citric acid on the levels of lead (Pb) and determine the optimal soaking time to reduce the levels of lead (Pb) in lorjuk meat. This study used a Completely Randomized Design (CRD) which consisted of three treatments for soaking lorjuk meat (1.5 hours; 3 hours and 4.5 hours). The results showed that the duration of immersion in citric acid affected the levels of lead (Pb) in lorjuk meat. The best treatment is soaking for 4,5 hours because it could reduce the levels of lead with a percentage decrease of 14,38%, organoleptic values on appearance parameter 6,07 (whole, specific meat color, bright and clean); odor 7,40 (very fresh) and texture 7,07 (elastic, solid and compact), moisture content 78,25%; protein content 8,67%; lipid content 0,65% and ash content 0,61%.

Keywords: citric acid, lead, soaking, *Solen* sp.

1. Pendahuluan

Lorjuk (*Solen* sp.) merupakan salah satu jenis bivalvia yang memiliki nilai ekonomis tinggi (Nurjanah *et al.*, 2021). Daging lorjuk mengandung 78,59% air; 1,53% abu; 14,48% protein dan 1,72% lemak (Nurjanah *et al.*, 2013). Lorjuk dapat ditemukan di pantai yang landai dan datar, seperti Pantai Kenjeran Surabaya. Pantai Kenjeran merupakan salah satu tempat wisata yang lokasinya dekat dengan pemukiman penduduk dan tempat berakhirnya aliran sungai di Surabaya sehingga mudah tercemar oleh limbah, baik limbah rumah tangga maupun limbah industri yang mengandung logam berat timbal (Pb). Kadar logam berat timbal (Pb) yang terkandung dalam sedimen di Pantai Kenjeran sebesar 1,646 mg/kg (Tyas and Kuntjoro, 2018). Angka tersebut sudah melebihi baku mutu yang ditetapkan dalam Peraturan Pemerintah Lingkungan Hidup Nomor 51 Tahun 2004 yaitu sebesar 0,005 mg/kg. Tingginya konsentrasi timbal (Pb) pada sedimen Pantai Kenjeran tersebut dapat berpengaruh terhadap lorjuk yang hidup dalam sedimen dan merupakan *filter feeder* sehingga dapat mengakumulasi timbal (Pb) (Hassan and Kanakaraju, 2016). Pada beberapa penelitian ditemukan kadar timbal (Pb) pada daging lorjuk sebesar 0,019–0,28 mg/kg (Mulyati and Pujiono, 2020; Nurjanah *et al.*, 2013). Batas maksimum cemaran logam berat timbal (Pb) pada kerang menurut BPOM (2018), yaitu 0,2 mg/kg.

Timbal (Pb) merupakan logam yang sangat beracun dan memengaruhi hampir setiap organ dalam tubuh, utamanya pada sistem saraf (Wani *et al.*, 2015). Pada biota perairan, timbal dapat mengakibatkan gangguan pada beberapa faktor genetik, pola pemijahan, tingkah laku, penurunan kemampuan untuk berorientasi, menghindari musuh, bermigrasi dan bersaing (Amnan, 1994 *dalam* Sari *et al.*, 2014). Seseorang yang mengonsumsi biota perairan yang terkontaminasi Pb dapat menyebabkan keracunan hingga berdampak buruk terhadap sistem organ, seperti sistem kardiovaskular, ginjal, sistem kekebalan tubuh dan sistem saraf (Wani *et al.*, 2015).

Solusi untuk mengurangi kadar timbal yang terakumulasi pada tubuh lorjuk yaitu dengan melakukan perendaman daging lorjuk menggunakan *chelating agent* seperti asam sitrat (Meidianasari, 2010 *dalam* Galih *et al.*, 2016). Asam sitrat dipilih sebagai *chelating agent* karena bernilai ekonomis, ramah lingkungan serta mudah diperoleh di toko kimia (Izza *et al.*, 2015). Menurut Alpatih *et al.* (2010) yang dikutip oleh Izza *et al.* (2015) asam sitrat memiliki tiga gugus COOH sehingga apabila asam sitrat dilarutkan dalam air maka dapat membentuk suatu ion yang disebut ion sitrat yang dapat berikatan dengan ion logam. Penelitian ini bertujuan mengetahui pengaruh lama perendaman asam sitrat terhadap kadar logam berat Pb pada daging lorjuk serta lama perendaman yang

paling baik untuk memperoleh daging lorjuk dengan kadar logam berat timbal (Pb) seminimal mungkin sehingga aman untuk dikonsumsi.

P2 : larutan asam sitrat 2% dengan lama perendaman 3 jam
P3 : larutan asam sitrat 2% dengan lama perendaman 4,5 jam

2. Material dan Metode

Material

Bahan utama penelitian adalah lorjuk yang diperoleh dari pengepul yang mendapatkannya dari daerah Pantai Kenjeran, Surabaya. Asam sitrat diperoleh dari Bohr Chemical. Bahan-bahan yang digunakan pada pengujian kadar timbal (Pb) menggunakan metode AAS, yaitu larutan standar Pb, HNO₃ 65%, H₂O₂, larutan matrik *modifier*, air deionisasi. Bahan yang digunakan untuk pengujian proksimat (kadar air, kadar protein, kadar lemak, kadar abu), antara lain H₂SO₄, HCl, NaOH 40%, tablet kjeldahl, H₃BO₃ 5%, metil merah, Bromocresol Green, Heksana.

Metode

Rancangan Penelitian

Penelitian eksperimental ini menggunakan Rancangan Acak Lengkap (RAL) terdiri dari tiga perlakuan dan lima ulangan. Berikut merupakan perlakuan penelitian :

P1 : larutan asam sitrat 2% dengan lama perendaman 1,5 jam

$$\% \text{ penurunan kadar logam berat timbal (Pb)} = \frac{C_{\text{sebelum}} - C_{\text{sesudah}}}{C_{\text{sebelum}}} \times 100\%$$

Keterangan:

C_{sebelum} = kadar logam berat timbal (Pb) sebelum perendaman dengan asam sitrat
C_{sesudah} = kadar logam berat timbal (Pb) sesudah perendaman dengan asam sitrat

Preparasi sampel daging lorjuk mengacu SNI (2011). Sampel dihaluskan dengan *blender* atau *homogenizer* hingga homogen dan tempatkan sampel dalam wadah polystyrene tertutup. Sebanyak 2 g sampel halus dimasukkan ke dalam tabung sampel (*vesse*l). Untuk kontrol positif, ditambahkan 0,2 ml larutan standar Pb pada sampel, kemudian divortex. Selanjutnya secara berurutan, menambahkan 5-10 ml HNO₃ 65% dan 2

Pengujian Organoleptik

Pengujian organoleptik menggunakan *scoresheet* SNI 3460.1:2009 dengan responden sebanyak 30 orang mahasiswa Fakultas Perikanan dan Kelautan Universitas Airlangga. Parameter yang diuji yaitu kenampakan, bau dan tekstur. Pengujian organoleptik terhadap daging lorjuk dilakukan dua kali, yaitu sebelum perendaman dan sesudah perendaman dengan asam sitrat.

Pengujian Kadar Logam Berat Timbal (Pb)

Pengujian kadar logam berat timbal (Pb) pada daging lorjuk dilakukan dua kali. Pengujian pertama dilakukan setelah pengujian organoleptik untuk mengetahui kadar Pb awal pada daging lorjuk. Pengujian kedua dilakukan setelah daging lorjuk direndam dengan asam sitrat 2% dengan lama perendaman yang berbeda (1,5 jam, 3 jam, dan 4,5 jam) untuk mengetahui kadar Pb pada daging lorjuk setelah perendaman dengan asam sitrat. Persentase penurunan kadar Pb dapat dihitung menggunakan rumus menurut Prihatini and Mulyati (2013).

ml H₂O₂, kemudian didestruksi menggunakan *microwave*. Hasil destruksi dipindahkan ke labu ukur 50 ml, ditambahkan larutan matrik *modifier*, kemudian air deionisasi hingga tanda batas pada labu ukur. Larutan standar kerja Pb disiapkan minimal 5 titik konsentrasi. Selanjutnya, melakukan pembacaan pada larutan standar kerja Pb, sampel dan kontrol positif pada alat *Atomic Absorption Spectrophotometer* (AAS) pada panjang gelombang 283,3 nm.

Perendaman Daging Lorjuk dalam Larutan Asam Sitrat

Proses perendaman daging lorjuk

diawali dengan menyiapkan larutan asam sitrat 2% dengan melarutkan 2 g asam sitrat dengan akuades hingga 100 ml. Selanjutnya daging lorjuk sebanyak 100 g dimasukkan ke dalam Beaker glass kemudian ditambahkan larutan asam sitrat 2% sebanyak 100 ml kemudian direndam dengan lama waktu berbeda, yaitu 1,5 jam, 3 jam, dan 4,5 jam. Setelah proses perendaman, daging lorjuk dicuci menggunakan aquades sebanyak 3 kali kemudian dilakukan pengujian kadar logam berat timbal (Pb) dan analisis proksimat.

Pengujian Kadar Air

Tahap awal pengujian kadar air berdasarkan *Association of Official Analytical Chemist* (2007) yaitu mengeringkan cawan porselen bersih dalam oven dengan suhu 105°C selama 1 jam, kemudian cawan didinginkan dalam desikator selama 30 menit dan ditimbang. Selanjutnya sebanyak 5 g sampel ditimbang dan diletakkan pada cawan dan dikeringkan dengan oven bersuhu 105°C selama 5-6 jam hingga beratnya konstan. Selanjutnya cawan didinginkan dalam desikator selama 30 menit dan ditimbang beberapa kali hingga beratnya tetap. Persentase kadar air dihitung dengan menggunakan rumus berikut:

$$\% \text{ Kadar Air} = \frac{(B - C)}{(B - A)} \times 100\%$$

Keterangan:

A = berat cawan kosong (g)

B = berat cawan dan sampel sebelum dikeringkan (g)

C = berat cawan dan sampel sesudah dikeringkan (g)

Pengujian Kadar Protein

Pengujian kadar protein berdasarkan *Association of Official Analytical Chemist* (2007) dapat dilakukan dengan tiga tahap, yaitu destruksi, destilasi dan titrasi. Pada tahap destruksi, sampel sebanyak 0,1 g dimasukkan ke dalam labu Kjeldahl, ditambah ¼ tablet Kjeldahl dan 2,5 ml H₂SO₄. Proses destruksi dilakukan hingga larutan menjadi jernih.

Larutan hasil destruksi dipindahkan ke dalam alat destilasi, selanjutnya dilakukan pencucian labu Kjeldahl sebanyak 5-6 kali menggunakan akuades dan air bilasan diletakkan dalam alat destilasi. Larutan NaOH 40% sebanyak 5 ml ditambahkan ke dalam alat destilasi. Cairan pada ujung kondensor ditampung dengan Erlenmeyer yang berisi larutan H₃BO₃ sebanyak 15 ml dan 3 tetes indikator (cairan metilen merah dan metilen biru). Proses destilasi berlangsung hingga mendapatkan larutan berwarna biru kehijauan.

Larutan hasil destilasi kemudian dititrasi menggunakan larutan HCl 0,01 N hingga larutan berwarna merah muda. Volume titran dibaca dan dicatat. Persentase kadar protein dihitung dengan menggunakan rumus berikut:

$$\% \text{ Nitrogen} = \frac{(\text{volume HCl sampel} - \text{volume HCl blanko}) \times N \text{ HCl} \times 14,007}{\text{bobot sampel}} \times 100\%$$

$$\% \text{ Kadar Protein} = \% N \times fk$$

Keterangan:

N HCl = normalitas HCl

fk = faktor konversi (6,25)

Pengujian Kadar Lemak

Pengujian kadar lemak mengacu pada *Association of Official Analytical Chemist* (2007). Sampel sebanyak 0,5 g pada kertas saring diletakkan pada alat ekstraksi Soxhlet. Selanjutnya sampel disiram dengan pelarut n-Hexane dan dilakukan refluks selama 6 jam. Pelarut n-

Hexane dalam labu lemak didestilasi dan ditampung. Hasil ekstraksi pada labu lemak dikeringkan dalam oven pada suhu 105°C dan didinginkan dalam desikator, kemudian ditimbang. Persentase kadar lemak dihitung dengan menggunakan rumus berikut:

$$\% \text{ Lemak} = \frac{W3 - W2}{W1} \times 100\%$$

Keterangan:

W1 = berat sampel (g)

W2 = berat labu lemak kosong (g)

W3 = berat labu lemak dengan lemak (g)

Pengujian Kadar Abu

Pengujian kadar abu mengacu pada *Association of Official Analytical Chemist* (2007). Cawan porselen dikeringkan dalam oven bersuhu 105°C selama 1 jam, kemudian didinginkan dalam desikator selama 30 menit dan ditimbang. Sebanyak 5 g sampel diletakkan dalam cawan dan dipijarkan di atas nyala api hingga tidak berasap lagi. Selanjutnya sampel diletakkan ke dalam tanur dengan suhu 600°C selama 5 jam dan ditimbang. Persentase kadar abu dapat dihitung dengan menggunakan rumus berikut:

$$\% \text{ Kadar Abu} = \frac{\text{berat abu}}{\text{berat sampel}} \times 100\%$$

Keterangan:

Berat abu = berat sampel dan cawan setelah ditanur – berat cawan kosong

Analisis Data

Data hasil pengujian kadar logam berat timbal (Pb) dan pengujian proksimat daging lorjuk dianalisis menggunakan *Analysis of Variance* (ANOVA), kemudian dilanjutkan *Duncan Multiple Range Test* (DMRT) dengan tingkat kepercayaan 95%. Data hasil pengujian organoleptik pada daging lorjuk dianalisis menggunakan uji *Kruskal-Wallis* dengan tingkat kepercayaan 95%, kemudian dilakukan uji *Mann Whitney* dengan tingkat kepercayaan 95%. Analisis data dilakukan dengan menggunakan perangkat lunak SPSS 23.

3. Hasil dan Pembahasan

Kadar Logam Berat Timbal (Pb)

Kandungan logam berat timbal (Pb) pada lorjuk sebesar 0,211 mg/kg (Tabel 1). Hasil tersebut melebihi batas maksimum pencemaran logam berat pada produk perikanan menurut Peraturan Badan Pengawas Obat dan Makanan Nomor 5 Tahun 2018 tentang Batas Maksimum dalam Pangan Olahan yaitu sebesar 0,20 mg/kg.

Tabel 1. Kadar logam berat timbal (Pb) pada daging lorjuk (*Solen* sp.)

Perlakuan	Kadar Timbal sebelum perlakuan (mg/kg)	Kadar Timbal Akhir (mg/kg) ± SD	Penurunan Kadar Timbal (mg/kg) ± SD	Persentase Penurunan Kadar Timbal (%) ± SD	Batas Maksimum Kadar Timbal Pada Lorjuk* (mg/kg)
P1 (1,5 jam)		0,194 ± 0,003 ^a	0,017 ± 0,003 ^a	8,01 ± 1,42 ^a	
P2 (3 jam)	0,211	0,186 ± 0,002 ^b	0,025 ± 0,002 ^b	11,69 ± 0,72 ^b	0,20
P3 (4,5 jam)		0,181 ± 0,003 ^c	0,030 ± 0,003 ^c	14,38 ± 1,19 ^c	

Keterangan: - Notasi huruf *superscript* yang berbeda berarti terdapat perbedaan nyata pada uji DMRT dengan tingkat kepercayaan 95%.

* Peraturan Badan Pengawas Obat dan Makanan Nomor 5 Tahun 2018 Tentang Batas Maksimum dalam Pangan Olahan.

Perbedaan lama waktu perendaman daging lorjuk dengan menggunakan asam sitrat 2% berpengaruh terhadap penurunan kadar timbal pada daging lorjuk. Penurunan kadar timbal terbesar terdapat pada perendaman selama 4,5 jam yaitu 0,030 mg/kg (14,38%), kemudian penurunan kadar timbal pada perendaman selama 3 jam yaitu 0,025 mg/kg (11,69%), dan yang terendah ditunjukkan oleh perendaman

selama 1,5 jam yaitu 0,017 mg/kg (8,01%) (Tabel 1). Semakin lama waktu perendaman maka semakin kecil pula kadar timbal yang ada pada daging lorjuk. Abadiana and Nurhayati (2013) membuktikan bahwa semakin lama waktu perendaman daging kerang darah dengan asam sitrat, semakin lama pula interaksi antara logam dengan asam sitrat, sehingga asam sitrat tersebut memiliki waktu yang lama untuk mengikat logam.

Logam yang terikat oleh asam sitrat akan dilepaskan dalam air melalui proses pelarutan. Asam sitrat merupakan zat pengkhelet (*chelating agent*) yang memiliki tiga gugus fungsional karboksil (-COOH). Gugus karboksil yang dimiliki oleh asam sitrat dapat mengalami deprotonasi atau melepaskan proton (H+) saat dilarutkan dalam air. Lepasnya proton (H+) tersebut dapat membentuk ion sitrat yang kemudian bereaksi dengan ion logam dan terjadi khelasi (Priyadi *et al.*, 2013), sehingga kadar logam berat timbal yang ada pada daging lorjuk dapat berkurang.

Nilai Organoleptik

Daging lorjuk sebelum perlakuan dan P1 (perendaman 1,5 jam) memiliki

kenampakan yang utuh, warna daging spesifik jenis, cerah dan bersih; P2 (perendaman 3 jam) memiliki kenampakan yang utuh, warna daging spesifik jenis, agak cerah dan bersih (Tabel 2). Nilai tersebut menunjukkan bahwa daging lorjuk sebelum perlakuan, P1 dan P2 memenuhi persyaratan mutu daging lorjuk yang layak untuk dikonsumsi menurut SNI 3460.1 (2009) karena memiliki nilai lebih dari 7 yang merupakan nilai minimal organoleptik. Pada P3 (perendaman 4,5 jam) memiliki kenampakan yang utuh, namun berwarna agak pucat dan kusam sehingga belum memenuhi persyaratan mutu daging lorjuk yang layak konsumsi menurut SNI 3460.1 (2009) karena tidak mencapai nilai minimal organoleptik.

Tabel 2. Nilai organoleptik daging lorjuk (*Solen sp.*)

Parameter	Rata-Rata ± SD			
	Sebelum Perlakuan	P1 (1,5 jam)	P2 (3 jam)	P3 (4,5 jam)
Kenampakan	8,67 ± 0,76 ^a	8,07 ± 1,02 ^b	7,40 ± 1,10 ^c	6,67 ± 0,76 ^d
Bau	8,60 ± 0,81 ^a	8,40 ± 0,93 ^a	7,67 ± 0,96 ^b	7,40 ± 0,81 ^b
Tekstur	8,87 ± 0,51 ^a	8,40 ± 0,93 ^b	7,53 ± 0,90 ^c	7,07 ± 0,83 ^d

Keterangan: Notasi huruf *superscript* yang berbeda berarti terdapat perbedaan nyata pada uji *Mann Whitney* dengan tingkat kepercayaan 95%.

Perubahan warna daging menjadi agak pucat dan kusam disebabkan oleh mioglobin dalam daging terlarut selama perendaman (Jayanti, 2018). Mioglobin adalah bagian dari rantai protein yang ada pada daging dan dapat larut dalam pH < 6 (asam) (Wodi *et al.*, 2014). Selain itu, mioglobin merupakan faktor yang berpengaruh terhadap warna pada daging sehingga apabila mioglobin terlarut dalam larutan asam sitrat, maka terjadi penurunan warna pada daging (Jayanti, 2018).

Nilai organoleptik parameter bau daging lorjuk sebelum perlakuan dan daging lorjuk P1 (perendaman 1,5 jam) memiliki bau yang sangat segar serta pada daging lorjuk P2 (perendaman 3 jam) dan P3 (perendaman 4,5 jam) memiliki bau yang segar. Nilai tersebut memenuhi persyaratan mutu daging lorjuk yang layak konsumsi menurut SNI 3460.1 (2009) karena memiliki nilai lebih dari 7. Meskipun terjadi penurunan nilai organoleptik parameter bau, namun

daging lorjuk masih layak untuk dikonsumsi karena tidak berbau tengik. Menurut Jayanti (2018), penggunaan asam sitrat dapat mencegah munculnya bau tengik pada daging lorjuk karena merupakan antioksidan.

Nilai organoleptik parameter tekstur daging lorjuk sebelum perlakuan dan daging lorjuk P1 (perendaman 1,5 jam) memiliki tekstur yang elastis, padat dan kompak, sedangkan pada daging lorjuk P2 (perendaman 3 jam) dan P3 (perendaman 4,5 jam) memiliki tekstur elastis, padat, namun kurang kompak. Nilai tersebut memenuhi persyaratan mutu daging lorjuk menurut SNI 3460.1 (2009) karena memiliki nilai lebih dari 7. Kadar air yang meningkat karena pengaruh lama perendaman dapat memengaruhi tekstur daging (Al Chusein and Ibrahim, 2012).

Analisis Proksimat

Analisis proksimat merupakan analisis yang dilakukan untuk mengetahui kandungan gizi pada suatu bahan

(Nurjanah *et al.*, 2021). Analisis proksimat daging lorjuk dilakukan sebelum perlakuan dan sesudah perlakuan. Kadar air daging lorjuk mengalami peningkatan setelah perendaman dengan asam sitrat (Tabel 3). Peningkatan kadar air setelah perendaman dikarenakan air rendaman masuk ke daging lorjuk menggantikan ion logam yang ditarik oleh gugus fungsi milik asam sitrat (Al Chusein and Ibrahim, 2012). Kadar air suatu bahan pangan memiliki peran penting untuk menjaga

kelembaban dan kestabilan bahan pangan tersebut (Henggu *et al.*, 2021) yang dikutip oleh Maharani *et al.*, 2021). Semakin tinggi kadar air yang ada pada bahan pangan, maka semakin besar kemungkinan terjadinya kerusakan bahan pangan secara biokimia maupun mikrobiologi sehingga berpengaruh pula terhadap umur simpan bahan pangan (Maharani *et al.*, 2021; Daud *et al.*, 2019 dalam Dayanti, 2021).

Tabel 3. Nilai proksimat daging lorjuk (*Solen sp.*)

Parameter	Rata-Rata (%) ± SD			
	Sebelum Perlakuan	P1 (1,5 jam)	P2 (3 jam)	P3 (4,5 jam)
Air	66,80 ± 0,083 ^a	67,99 ± 0,093 ^b	69,79 ± 0,045 ^c	78,25 ± 0,114 ^d
Protein	8,92 ± 0,025 ^c	8,83 ± 0,030 ^{bc}	8,79 ± 0,015 ^b	8,67 ± 0,091 ^a
Lemak	0,78 ± 0,021 ^c	0,73 ± 0,015 ^b	0,70 ± 0,010 ^b	0,65 ± 0,026 ^a
Abu	0,85 ± 0,020 ^c	0,76 ± 0,040 ^b	0,66 ± 0,035 ^a	0,61 ± 0,015 ^a

Keterangan: Notasi huruf *superscript* yang berbeda berarti terdapat perbedaan nyata pada uji DMRT dengan tingkat kepercayaan 95%.

Kadar protein daging lorjuk mengalami penurunan setelah perendaman dengan asam sitrat (Tabel 3). Hal tersebut merupakan efek samping dari penurunan kadar logam berat timbal (Pb) pada daging lorjuk. Logam berat timbal (Pb) dapat terakumulasi dalam daging lorjuk karena berikatan dengan gugus sulfihidril pada protein daging lorjuk (Mirawati *et al.*, 2016), sehingga ketika ion logam membentuk senyawa kompleks dengan ion sitrat dan terlarut dalam larutan asam sitrat maka protein pun ikut terlarut (Maulana and Umroh, 2017; Saputri and Rachmadiarti, 2015; Chotimah *et al.*, 2016). Protein merupakan komponen terbesar setelah air dan juga merupakan zat pembangun serta pengatur jaringan tubuh merupakan komponen terbesar setelah air (Winarno, 1994 dalam Maharani *et al.*, 2021), sehingga apabila kadar air pada daging lorjuk mengalami peningkatan, maka kadar protein mengalami penurunan.

Kadar lemak daging lorjuk juga mengalami penurunan setelah perendaman dengan asam sitrat (Tabel 3). Setiawan *et al.* (2012) membuktikan bahwa lemak membentuk emulsi yang halus pada saat perendaman dengan asam sitrat, kemudian lemak tersebut larut

dalam larutan asam sitrat, sehingga kadar lemak dalam daging turun. Secara tidak langsung larutnya lemak dalam larutan asam sitrat dapat berpengaruh pada turunnya kadar logam berat timbal (Pb) pada daging lorjuk. Hal tersebut dikarenakan timbal merupakan jenis logam berat yang dapat larut dalam lemak (Kristanto, 2002 dalam Setiawan *et al.*, 2012).

Kadar abu daging lorjuk juga mengalami penurunan (Tabel 3). Penurunan kadar abu dapat terjadi karena kandungan mineral yang ada pada daging lorjuk terlarut dalam larutan asam sitrat (Al Chusein and Ibrahim, 2012). Menurut Sudarmadji (2003) yang dikutip oleh Trisyani (2019), jumlah kadar abu pada suatu bahan pangan dapat menentukan kualitas bahan pangan tersebut. Suatu bahan pangan yang memiliki kadar abu dengan jumlah yang terlalu tinggi menunjukkan bahwa kualitas bahan pangan tersebut kurang baik, meskipun zat abu dalam bahan pangan tetap dibutuhkan sesuai dengan kadar yang disarankan.

4. Kesimpulan

Asam sitrat dapat digunakan untuk

menurunkan kadar logam berat timbal (Pb) dalam daging lorjuk. Lama waktu perendaman asam sitrat 4,5 jam paling baik untuk menurunkan kadar logam berat timbal (Pb) sebesar 14,38% dengan memiliki nilai organoleptik pada parameter kenampakan 6,07 (utuh, warna daging spesifik jenis, cerah dan bersih), bau 7,40 (sangat segar) dan tekstur 7,07 (elastis, padat dan kompak); kadar air 78,25%; kadar protein 8,67%; kadar lemak 0,65% dan kadar abu 0,61%.

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SURAT KETERANGAN

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Telah melakukan penelitian yang dipublikasi pada bulan Februari tahun 2023 dengan judul sebagai berikut:

The Effect of Citric Acid Soaking Time on The Levels of Lead (Pb) in Lorjuk Meat (Solen sp.)

Adapun penelitian ini sudah mengacu pada prosedur pertimbangan etik dari:

1. *American Fisheries Society* (AFS, 2014) yang berjudul *Guideline for the Use of Fishes in Research* yang menyebutkan bahwa: penelitian dalam kondisi laboratorium baru mengatur tentang hewan percobaan berupa ikan hidup (hal 43 ; terlampir), dan
2. *Canadian Council on Animal Care* (CCAC, 2005) yang berjudul *Guideline on the Care and Use of Fish in Research, Teaching and Testing* yang menyebutkan bahwa: pedoman tersebut hanya digunakan untuk hewan uji berupa ikan hidup (Kelas: Chondrichthyes, Agnatha, dan Osteichthyes) dan Avertebrata (Kelas: Cephalopoda) (hal 13,14 ; terlampir).

Sedangkan dalam penelitian tersebut tidak menggunakan hewan hidup sebagai bahan penelitiannya. Sehingga penelitian tersebut tidak perlu dilakukan ***Uji Ethical Clearence***.

Demikian Surat Keterangan ini kami buat untuk dapat dipergunakan sebagai persyaratan pengusulan Jabatan Fungsional **Guru Besar** atas nama Dr. Endang Dewi Masithah, Ir., MP.

Surabaya, 27 April 2023

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Guidelines for the Use of Fishes in Research

Use of Fishes in Research Committee members:

J. A. Jenkins, Chair, H. L. Bart, Jr., J. D. Bowker, P. R. Bowser, J. R. MacMillan, J. G. Nickum, J. D. Rose, P. W. Sorensen, and G. W. Whitley on behalf of the American Fisheries Society; J. W. Rachlin and B. E. Warkentine on behalf of the American Institute of Fishery Research Biologists; and H. L. Bart on behalf of the American Society of Ichthyologists and Herpetologists

American Fisheries Society
Bethesda, Maryland
2014

A suggested citation format for this book follows.

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Cover art: Close-up photograph of Brown Trout, *Salmo trutta*, from the South Fork of the Cache la Poudre River, Colorado, taken by James Rose in 2010.

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7. Laboratory Activities

7.1 General Principles

Working with live fishes under laboratory conditions requires attention to many details concerning the requirements for, and limits of tolerance of, the particular species under study. Acceptable physical facilities and an adequate supply of water with good quality must be provided, even if the fishes are to be held for only short periods of time. Although fish may tolerate marginal facilities and conditions for a few hours or even several days, holding them under less than optimal conditions will affect the results of the research. Standards for humane treatment of animals must also be maintained, regardless of the length of time that the fishes are held.

The reader should note that some content of section 7 is not restricted to laboratory activities, but may be applicable to field situations, as well.

7.2 Confinement, Isolation, and Quarantine

Prior to bringing fishes into a laboratory, facilities and plans should be in place to ensure that the fish cannot escape, especially species not native to the watershed, and that the introduced fishes can be isolated physically from fishes already present. Each holding unit should have its own set of nets and other equipment. Facilities and equipment used for previous studies should be disinfected prior to use in new studies, typically with a chlorinated disinfectant or another disinfectant such as Virkon[®] Aquatic (www.wchemical.com/). If the introduced fishes may carry disease agents, especially pathogens or parasites that are not endemic to the area, quarantine-level facilities should be used. The level of quarantine required will vary with the seriousness of the known or suspected disease agent (see section 2.5 Fish Health Management: Control of Pathogens and Parasites).

Individual fish with suspected ill health should be quarantined from the others so as to negate the potential for spread of potential disease agents. Such fish should be evaluated by an individual with expertise in fish diseases (fish pathologist or veterinarian), and the proper therapeutant should be applied as directed. Providing guidance for the treatment of specific diseases is beyond the scope of this document. The investigator is strongly urged to establish a working relationship with individuals with expertise in fish health with whom they may consult.

Experimentation with nonindigenous fishes, transgenic fishes, or other genetically modified fishes is a special situation that requires additional precautions to preclude their escape. Permitting with site visits by state wildlife agencies may be required for holding nonindigenous species (see section 3.4 Permits and Certificates). The specific barriers may be similar to those used to prevent the escape of disease agents but must be developed to fit the physical characteristics of the laboratory or experimental facility. The USDA has developed

Canadian Council on Animal Care



guidelines on:

***the care and use of
fish in research,
teaching and
testing***

This document, the CCAC *guidelines on: the care and use of fish in research, teaching and testing*, has been developed by the *ad hoc* subcommittee on fish of the Canadian Council on Animal Care (CCAC) Guidelines Committee.

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the care and use of fish in research, teaching and testing



A. PREFACE

The Canadian Council on Animal Care (CCAC) is the national peer review agency responsible for setting and maintaining standards for the care and use of animals used in research, teaching and testing throughout Canada. In addition to the *Guide to the Care and Use of Experimental Animals*, vol. 1, 2nd ed., 1993 and vol. 2, 1984, which provide the general principles for the care and use of animals, the CCAC also publishes detailed guidelines on issues of current and emerging concerns. The CCAC *guidelines on: the care and use of fish in research, teaching and testing* is the seventh of this series. This document supersedes Chapter I - Fish, *Guide to the Care and Use of Experimental Animals*, vol. 2 (CCAC, 1984).

These guidelines aim to provide information for investigators, animal care committees, facility managers and animal care staff that will assist in improving both the care given to fishes and the manner in which experimental procedures are carried out.

The present document has drawn substantially from the work of organizations listed in Appendix A. Their contributions to the development of these guidelines are gratefully acknowledged.

The guidelines have been developed by the CCAC subcommittee on fish and were reviewed by a total of 69 experts. A preliminary first draft was agreed on by the subcommittee and circulated to experts in June 2002 (including representatives of the organizations listed in Appendix A), and a second draft was circulated for widespread comment in June 2003. A final review was carried out in August 2004 involving all individuals who had previously provided significant input to the development process. The development of these guidelines also involved consultation with the Canadian Association for Laboratory Animal Science (CALAS) and the Canadian Society of Zoologists (CSZ) through workshops held at annual meetings in Québec City (June 2003), Acadia University (May 2004), and Hamilton (June 2004). Consultations were also held at the Aquaculture Association of Canada and AquaNet annual meetings in Québec City (October 2004), and at the CCAC Workshop on the Fish Guidelines in Vancouver (April 2005).

The guidelines have been organized in a format that should facilitate easy access to relevant sections. Early sections provide an ethical overview relevant to the use of fishes in research, teaching and testing. This is followed

by a brief overview of regulations and responsibilities relevant to the care and use of fishes in science in Canada. The remainder of the document provides guidelines to assist in caring for fishes in laboratory facilities, followed by guidelines to help in the development and review of experimental protocols. An overview of the CCAC *guidelines on: the care and use of fish in research, teaching and testing* is provided through a summary of the guidelines listed in

this document prior to the beginning of the main text.

The refinement of animal care and use guidelines is a continuous process. These guidelines are intended to provide assistance in the implementation of best practices, and should not be viewed as regulations. Where regulatory requirements are involved or where it is absolutely imperative to adhere to a particular guideline, the term *must* has been used.

B. INTRODUCTION

The greatest challenge in providing *guidelines on: the care and use of fish* is the wide variety of fishes used in Canada and the diversity of their habits, behavior, life history, and environmental and husbandry requirements. In addition, the scientific information required to define the preferred conditions for fish well-being is limited. While considerable research has been conducted on culture strategies and environmental and water quality requirements, such studies have generally been aimed at determining conditions that optimize production in aquaculture systems, rather than improving the welfare of fishes, and have not usually addressed the difference between *tolerance* and *preference* (Fisher, 2000).

An important consideration in these guidelines is the naturally high mortality rates of juveniles in species whose ecological strategies include the generation of large numbers of progeny to ensure adequate survival in the wild. In addition, many experimental populations of species with usually high survival contain individuals that will not thrive to adulthood even under the best environmental conditions. In some situations, a population-based (or a group of study fish) approach to well-being may be appropriate, but individuals that are not likely to thrive should be euthanized as soon as they are identified.

Another consideration for these guidelines is the general acceptance by the public of the current killing methods used in harvesting wild fishes or in recreational angling. In general, the public appears to be willing to accept these killing methods for food production but not when fishes are used for research. These guidelines accept that for research, teaching, and testing use of any animal, including fishes, more emphasis will be placed on individual well-being than is generally accepted for the commercial harvesting or production of animals for food. It is recognized, however, that in some instances investigators may obtain fishes from people involved in commercial or recreational harvesting and have little influence over the capture methods.

These guidelines apply to fishes held in facilities for research, teaching and testing, as well as to fishes that are studied in their natural habitats.

1. Definition of Fish

For the purpose of these guidelines, fishes are defined as all bony and cartilaginous fish genera (classes Chondrichthyes [cartilaginous fishes], Agnatha, and Osteichthyes [bony fishes]). Fish eggs, embryos or larvae that have not developed beyond exclusive reliance on their own yolk nutrients are not covered by these guidelines. Similarly, invertebrates (except cephalopods) are not covered under the CCAC system of surveillance, but institutions are encouraged to foster respect for these animals by ensuring that holding facilities and levels of husbandry meet standards equivalent to those used for fishes.

2. Rationale for Guidelines on the Care and Use of Fish

The use of fishes as experimental subjects has increased substantially over the past two decades. This increase in use is a result of the rapid development of the aquaculture industry, requirements for testing involving fishes as indicators of environmental change, and the use of fishes as a replacement for mammals in biomedical, pharmacological and genetic research (DeTolla *et al.*, 1995; Fabacher & Little, 2000). The trend toward the use of fishes as a replacement for studies that would previously have used mammals as experimental subjects is not discouraged. However, it must also be recognized that fishes have the capacity to perceive noxious stimuli. Noxious stimuli are those stimuli that are damaging or potentially damaging to normal tissue (e.g., mechanical pressure, extremes of temperature and corrosive chemicals). Whether or not fishes have the capacity to experience any of the adverse states usually associated with pain in mammals is subject to a great deal of debate in the scientific literature (FAWC, 1996; FSBI, 2002; Rose, 2002; Braithwaite & Huntingford, 2004). Nonetheless, fishes are capable of behavioral,

physiological and hormonal responses to stressors (including noxious stimuli) which can be detrimental to their well-being. These CCAC guidelines both support the leadership role that Canadians play in fish research, and ensure that the welfare of fishes is carefully considered during the use of fishes for research, teaching and testing, recognizing that better welfare will result in better science.

3. Ethical Overview

Guideline 1:

Fishes used in research, teaching and testing must be treated with the respect accorded to other vertebrate species.

The CCAC's surveillance system for animals used in research, teaching and testing is based on the principles of humane science, i.e. the Three Rs of Russell and Burch (Russell & Burch, 1959) - Reduction, Replacement and Refinement. For the CCAC, these principles are laid out in its *policy statement on: ethics of animal investigation* (CCAC, 1989). The *ethics of animal investigation* applies to all species covered by the CCAC system, i.e. all vertebrates and cephalopods.

In addition, the CCAC system takes a "moral stewardship" approach to the use of animals in science as explained in the CCAC Experimental Animal User Training Core Topics - Module 2, Ethics in Animal Experimentation (http://www.ccac.ca/en/CCAC_Programs/ETCC/Module02/toc.html).

The first guideline statement in the CCAC *guidelines on: institutional animal user training* (CCAC, 1999a) states, "Institutions must strive through their training programs to sustain an institutional culture of respect for animal life".

3.1 Principles of the Three Rs

According to the CCAC *policy statement on: ethics of animal investigation* (CCAC, 1989), it is the responsibility of the local animal care committee (ACC) to ensure that fishes are used only if the investigator's best efforts to find a non-animal model have failed.

As for any other species covered by the CCAC system, investigators using fishes are required to use the most humane methods on the smallest

number of animals necessary to obtain valid information. This requires the use of a sound research strategy, including: identification of key experiments that determine whether a particular line of enquiry is worth pursuing; use of pilot studies; staging of *in vitro* to *in vivo* experiments where possible; and implementation of staged increase in test stimuli where possible (Balls *et al.*, 1995). The numbers and species of animals required depend on the questions to be explored. Field studies, aquaculture studies and laboratory studies require different statistical designs; field studies and aquaculture production typically require the use of larger numbers of animals. The life stage of the fishes used in each study will also affect the numbers of animals needed. Studies of early life stages typically require large numbers of individuals. In all cases, studies should be designed to use the fewest animals necessary. Heffner *et al.* (1996) and Festing *et al.* (2002) provide discussions on the appropriate treatment of samples and experimental units. Investigators are encouraged to consult with a statistician to develop study designs that have the appropriate statistical power to accomplish the research objectives (Nickum *et al.*, 2004).

The CCAC *policy statement on: ethics of animal investigation* (CCAC, 1989) also requires adherence to the following principles:

- animals must be maintained in a manner that provides for their optimal health and well-being, consistent with the demands imposed by the experimental protocol;
- animals must not be subjected to pain and/or distress that is avoidable and that is not required by the nature of the relevant protocol;
- expert opinion must attest to the potential value of studies with all animals, including fishes (e.g., scientific merit for research, see CCAC *policy statement on: the importance of independent scientific merit of animal based research projects* [CCAC, 2000a]; pedagogical value for teaching; and the appropriateness of the method to provide data for testing according to current regulatory requirements);
- if pain or distress is a justified component of