



ISSN: 0975-3575

PHARMACOGNOSY JOURNAL

An Official Journal of Phcog.net



Editorial Board (2020-21)

Editors & Editorial Board Members (2021)

Dr.Djemli Samir

Department of Biology , Applied Neuroendocrinology Laboratory
Badji Mokhtar Annaba University
Algeria

Dr. Raghava Naidu, Ph.D

Department of Human Oncology,
University of Wisconsin,
1111, Highland Ave, Madison,
Wisconsin 53705, USA

Dr.Karim Raafat

Associate Professor of Pharmacognosy and Phytochemistry,
Pharmaceutical Sciences Department,
Faculty of Pharmacy,
Beirut Arab University (BAU),
Beirut 115020, Lebanon

Ourlad Alzeus Tantengco, MD-PhD Molecular Medicine

College of Medicine, University of the Philippines Manila
Pedro Gil Street, Ermita, Manila, Philippines, 1000

Janib Achmad

Lecturer of Faculty of Fisheries and Marine Science,
University of Khairun Ternate
Kampus 2 Jalan Pertamina, Kelurahan Gambesi,
Ternate Selatan

Muammar Fawwaz, Ph.D

Department of Pharmaceutical Chemistry
Faculty of Pharmacy
Universitas Muslim Indonesia
Makassar 90231, South Sulawesi, Indonesia

Hany Ezzat Khalil

Associate Professor,
College of Clinical Pharmacy,
King Faisal University,
KSA

Emad Yousif

Department of Chemistry
College of Science
Al-Nahrain University
Baghdad, Iraq

Sughosh Upasani

R.C Patel Institute of pharmacy,
Shirpur, Dist-Dhule, Maharashtra,
India.

Gurusiddaiah suresh kumar

Scientist
Dept of biochemistry
CSIR-CFTRI
Mysore, Karnataka, INDIA

Arjun Patra

Assistant Professor
School of Pharmaceutical Sciences
Guru Ghasidas Central University
Koni, Bilaspur - 495 009
Chattisgarh, India

Francis O. Atanu, Ph.D

Department of Biochemistry
Faculty of Natural Sciences
Kogi State University
Anyigba, Nigeria.

Vijay Kumar Chattu

Faculty of Medical Sciences
University of the West Indies
St. Augustine, Trinidad & Tobago.

Dr.Kunle Okaiyeto, PhD

Applied and Environmental Microbiology Research Group (AEMREG)
Department of Biochemistry and Microbiology
University of Fort Hare
Alice campus
5700, Alice
South Africa.

Dr. Srisailam Keshetti, Ph.D

Principal, University College of Pharmaceutical Sciences, Satavahana University
Karimnagar 505001
Telangana
INDIA

Dr. Gayathri M Rao

Associate Professor
Department of Biochemistry
Kasturba Medical College, Mangaluru.

Shuge Tian

Experimental Teaching Demonstration Center of TCM in Xinjiang Medical University
Department of traditional medicine, TCM
Xinjiang Medical University
Xinjiang CHINA 830054

Dr. Ramachandra Setty Siddamsetty,

Professor, Govt College of Pharmacy,
Mission Road, Bengaluru, INDIA

Dr. (Mrs.) Sayyada Khatoon

HOD, Pharmacognosy Division
CSIR-National Botanical Research Institute,
Rana Pratap Marg, Post Box 436,
Lucknow-226001 (U.P.) India

Dr. A. Sajeli Begum

Department of Pharmacy
Birla Institute of Technology & Science
Hyderabad, India

Olga Silva

Department of Pharmacological Sciences,
Faculdade de Farmácia,
Universidade de Lisboa, Portugal

Xinwen Wang

Department of Clinical Pharmacy
University of Michigan
USA

Roman Lysiuk

Department of Pharmacognosy and Botany,
Danylo Halytsky Lviv National Medical University,
Pekarska,69., Lviv 79010, Ukraine

Arif Nur Muhammad Ansori

Universitas Airlangga
Indonesia

PharmacognJ Vol 14, Issue 6 (Suppl), Nov-Dec, 2022

[View](#)

[What links here](#)

Submitted by [sys1](#) on Wed, 12/28/2022 - 14:46

Medicinal Plants Adopted to Treat Children's Diseases by Traditional Pediatrics "Women Healers" In The Souss Massa Region (Agadir Idaoutanan, Inzegane Ait Meloul and Chtouka Ait Baha) Morocco

Differences in interleukin-6 and interleukin-17 expression in covid-19 post-mortem lung tissue biopsy compared with noncovid- 19

Kaempferia galanga L. Extract Administration Attenuate Aquaporin-4 Expression in Traumatic Brain Injury: An Experimental Study in Rats

Effect of Pomegranate Extract On N-Terminal Pro Brain Natriuretic Peptide and Asymmetric Dimethylarginine Levels in Children with Pulmonary Artery Hypertension in Acyanotic Congenital Heart Disease

Evaluation of Experimental Cerebral Malaria of Curcumin and Kaempferol in Plasmodium berghei ANKA-Infected Mice

Molecular Docking Estrogen Receptor Alpha Antagonist and P53- MDM2 Inhibitor, ADMET Prediction of Alkaloid Compound from Mitragyna speciosa for Breast Cancer Therapy

Left Atrial Myxoma Presented with an Obstructive Shock, Right Ventricle Dysfunction and Pulmonary Hypertension

Risk Factors for Hepatotoxicity From L-Asparaginase Chemotherapy In Children With Acute Lymphoblastic Leukemia

Phytochemical Approach Including Total Phenolic and Flavonoid Contents and Evaluation of in vitro ABTS Antioxidant Capacity and Lipoxygenase Inhibition of Anisoscadium lanatum

Cichorioside a biocoumarin modulates lipid and glucose storage on 3T3-L1 cell lines: In vitro and in silico approach

Relationship Histopathology Grading of Meningioma with the Use of Medroxyprogesterone Acetate (MPA) as A Hormonal Contraceptive

Correlation Between the Suitability of Empirical and Definitive Therapies According to Culture Results with the Clinical Outcomes of Patients with Bacteremia Due to Carbapenem- Resistant Acinetobacter baumannii (CRAB) at Dr. Soetomo Tertiary Referral Hosp

GC-MS Analysis of Volatiles Present in Papea Capensis Extracts

Phytochemical Analysis and Antioxidant Activity of Water Hyacinth Flowers (*Eichhornia Crassipes*) Extract

Research on External Signs and Chemical Composition of Medicinal Plant Raw Material -Leaves of *Ficus Elastica*

Antiparasitic effect of *Psidium guajava* on promastigotes and axenic amastigotes of *Leishmania*

The Serum Formulation of Hati Tanah Tuber Ethanol Extract from Central Kalimantan

Job fulfilment and its related variables among pharmacy certificate holders in Jordan: A Cross-sectional Study

Chemical Profiling of Nonpolar Compounds of *Onopordum Acanthium* using GCMASS

Antioxidant, Antimicrobial, and Antiplasmodial Activities of *Sonchus arvensis* L. Leaf Ethyl Acetate Fractions

Senna Siamea Hexane Extract: Potent Antifungal Activity Against *Candida albicans*, *Candida Krusei* and Identification of Its Chemicals Content

DFT and Pharmacokinetic Study of Some Heterocyclic Aspirin Derivatives as The Cyclooxygenase Inhibitors: An In-Silico Approach

Fingerprint and Multivariate Analysis of *Apium Graveolens* L. From Different Geographic with Spectroscopic ATR-FTIR

Antibacterial Effect of *Nigella sativa* L. Seed from Indonesia

Successful Intrapericardial Fibrinolysis on Acute Recurrent Purulent Pericarditis with Impending Cardiac Tamponade during Pandemic Situation: A Rare Case Report

Bio-Evaluation, In-Vitro and In-Vivo Anti-Inflammatory Activity, Therapeutic Efficacy, and Genotoxicity of the Potentials of the Green Seaweed *Valoniopsis Pachynema* using Zebra Fish Larvae (*Danio Rerio*) as an Animal Model

Aquaporin 4 Expression on End Feet Astrocyte Before and After Cerebrospinal Fluid Drainage of Hydrocephalus Mice Model

Effects of Pomegranate Extract Supplementation (*Punica granatum* L.) on Clinical Manifestations of Pulmonary Arterial Hypertension in Children with Acyanotic Congenital Heart Disease

The Role of Breast Milk on Reducing the Risk of Neonatal Sepsis in Preterm and Low Birth Weight Infants: A Systematic Review and Meta-Analysis

Histopathological Perspectives of Multiple Organs in a Red- Footed Tortoise (*Chelonoidis carbonaria*) with Suspected Metabolic Bone Disease: A Case Report

Differences in interleukin-6 and interleukin-17 expression in covid-19 post-mortem lung tissue biopsy compared with non-covid-19

Etty Hary Kusumastuti^{1,*}, Priangga Adi Wiratama¹, Grace Ariani¹, Stephanie Natasha Djuanda¹, Alphaenia Rahniayu¹, Nila Kurniasari¹, Dyah Fauziah¹, Anny Setijo Rahaju¹, Isnin Anang Marhana², Alfian Nur Rosyid², Dwi Wahyu², Gilang Muhammad Setyo Nugroho², Adhitri Anggoro², I Komang Rusgi Yandi², Bambang Pujo Semedi³, Jilientasia Godrace Lilihata³, Ummi Maimunah⁴, Supriadi⁴, Achmad Lefi⁵, Lalu Galih Pratama Rinjani⁵, Edi Suyanto⁶, Ricardo Ardian Nugraha⁶

¹Department of Anatomical Pathology, Faculty of Medicine, Universitas Airlangga – Dr. Soetomo General Academic Hospital, Surabaya, INDONESIA.

²Department of Pulmonology and Respiratory Medicine, Faculty of Medicine, Universitas Airlangga – Dr. Soetomo General Academic Hospital, Surabaya, INDONESIA.

³Department of Anesthesiology and Reanimation, Faculty of Medicine, Universitas Airlangga University – Dr. Soetomo General Academic Hospital, Surabaya, INDONESIA.

⁴Department of Internal Medicine, Faculty of Medicine, Universitas Airlangga – Dr. Soetomo General Academic Hospital, Surabaya, INDONESIA.

⁵Department of Cardiology and Vascular Medicine, Faculty of Medicine, Universitas Airlangga – Dr. Soetomo General Academic Hospital, Surabaya, INDONESIA.

⁶Department of Forensics and Medicolegal Medicine, Faculty of Medicine, Universitas Airlangga – Dr. Soetomo General Academic Hospital, Surabaya, INDONESIA.

Correspondence

Etty Hary Kusumastuti

Department of Anatomical Pathology, Faculty of Medicine, Universitas Airlangga – Dr. Soetomo General Academic Hospital, Surabaya, INDONESIA.

Email: ettyhary@fk.unair.ac.id

History

- Submission Date: 08-10-2022;
- Review completed: 28-11-2022;
- Accepted Date: 05-12-2022.

DOI : 10.5530/pj.2022.14.184

Article Available online

<http://www.phcogj.com/v14/i6>

Copyright

© 2022 Phcogj.Com. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.



ABSTRACT

Background: COVID-19 has spread rapidly around the world. It is necessary to study lung tissue of post-mortem COVID-19 patients to determine the molecular alteration particularly the role of IL-6 and IL-17 in causing fatality. **Objective:** This study aims to determine the differences in the expressions of IL-6 and IL-17 in lung tissue of post-mortem COVID-19 patients compared to non-COVID-19 patients. This study also aimed to analyze the correlation between the expressions of IL-6 and IL-17 in lung tissue of post-mortem COVID-19 patients. **Methods:** This research is an observational analytic study with cross-sectional approach. The samples were 15 paraffin blocks of post-mortem lung tissue biopsy of COVID-19 patients, and 15 paraffin blocks of inflammatory lung tissue biopsy or surgery of non-COVID-19 patients. IL-6 and IL-17 expressions were evaluated by immunohistochemical procedure. **Result:** There was a significant difference in the expression of IL-6 in the COVID-19 group and the non-COVID-19 group with a p-value = 0.001 ($p < 0.05$). There was a significant difference in the expression of IL-17 in the COVID-19 group and the non-COVID-19 group with p-value = 0.001 ($p < 0.05$). There was a significant correlation between the expressions of IL-6 and IL-17 in the COVID-19 group, with the Spearman coefficient value (rs) of 0.548 with $p = 0.034$ ($p < 0.05$). **Conclusion:** There are differences in the expression of IL-6 and IL-17 between COVID-19 and non-COVID-19 lung tissue. There is a significant correlation between the expressions of IL-6 and IL-17 in post-mortem lung tissue of COVID-19 patients.

Key words: Biopsy, COVID-19, IL-6, IL-17, Post mortem lung tissue.

INTRODUCTION

Since December 2019, Corona Virus Disease (COVID-19) caused by Severe Acute Respiratory Syndrome Corona Virus 2 (SARS CoV-2) has spread rapidly around the world. SARS-CoV2 is an RNA (ribonucleic acid) virus with a new variant belonging to the Coronaviridae family which was first described in February 2020.¹ The World Health Organization stated that until March 2021, the number of confirmed cases was 125 million with a death toll of 2.8 million. COVID-19 has also had a negative impact on the global economy and health system. Lack of knowledge regarding the characteristics and pathogenesis of the SARS CoV-2 virus causes difficulties in controlling the course of the COVID-19 disease. It is necessary to conduct research on lung tissue of post-mortem COVID-19 patients to determine the molecular alteration that plays a role in causing fatality.

The pathogenesis of COVID-19 can generally be categorized into three successive and overlapping phases, namely the pulmonary phase, the proinflammatory phase and the prothrombotic phase.² In the proinflammatory phase, SARS-CoV-2 causes the release of proinflammatory cytokines in very large quantities that cause interstitial pneumonia and acute respiratory distress syndrome (ARDS). In a more advanced

state will develop sepsis and cytokine storm (cytokine storm) with hypercoagulability and multiorgan dysfunction.^{2,3} Several types of cytokines that play a role in the pathogenesis of COVID-19 are IL-1 β , IL-2, IL-4, IL-6, IL-8, IL12, IL-17, G-CSF, GM-CSF, IFN γ , MCP-1 and TNF α . Cytokines IL-6 and IL-17 are known to play an important role and influence disease fatality, which levels of both are known to be elevated in blood.^{1,4-6}

Interleukin-6 (IL-6) is one of the cytokines that is activated in the presence of infection or injury. IL-6 is secreted by various cell types and plays a role in the regulation of various physiological processes. Elevated IL-6 serum levels are associated with an inflammatory response, respiratory failure and affect mortality in COVID-19 patients.⁷⁻⁹ Interleukin-17 (IL-17) is a cytokine synthesized by Th17 lymphocytes and is increased in inflammatory processes and autoimmune diseases. IL-17 is a proinflammatory cytokine that plays a role in tissue damage, physiological stress, and infection.^{5,10}

A number of studies have been carried out through blood serum analysis to study the important role of IL-6 and IL-17 in the pathogenesis of COVID-19, but lack studies to analyze the role of IL-6 and IL-17 in lung tissue of patients with COVID-19. This study aims to determine the differences in the expression of IL-6 and IL-17 in lung tissue of post-mortem

Cite this article: Kusumastuti EH, Wiratama PA, Ariani G, Djuanda SN, Rahniayu A, Kurniasari N, et al. Differences in interleukin-6 and interleukin-17 expression in covid-19 post-mortem lung tissue biopsy compared with non-covid-19. Pharmacogn J. 2022;14(6)Suppl: 887-892.

COVID-19 patients compared to non-COVID-19 patients and to analyze the correlation between the expression of IL-6 and IL-17 in lung tissue of post-mortem COVID-19 patients.

MATERIAL AND METHODS

The design of this research is an observational analytic with cross-sectional approach. This research is a branch of research titled 'Multi-organ pathological finding in COVID-19 infection through post-mortem core biopsy' and has received ethical approval from the Health Research Ethics Committee of Dr. Soetomo general hospital number 0022/KEPK/VII/2020.

The number of research samples is 30, consisting of 15 paraffin blocks of post-mortem pulmonary tissue biopsy of patients with COVID-19 and 15 paraffin blocks of pulmonary tissue biopsy or surgery of non-COVID-19 pneumonia patients. IL-6 expression was evaluated by an immunohistochemical procedure using IL-6 antibody GeneTex Antibodies Rabbit Polyclonal antibody, unconjugated (GTX17623). Antigen retrieval using pH 6, with a dilution of 1:100. IL-17 expression was also evaluated by an immunohistochemical procedure using anti-GeneTex Antibodies Rabbit Recombinant-Human IL-17 (IL-17A) polyclonal antibody, unconjugated (GTX31174) antibody, with a concentration of 0.25 g/ml. The expression of IL-6 was positive for brown color in the cytoplasm of lymphocytes and macrophages, while the expression of IL-17 was positive for brown color in the cytoplasm of lymphocyte cells. The assessment is carried out in five fields of view with high power fields. The number of positive cells is calculated as a percentage. The assessment was carried out by two pathologists in a double-blind manner and then an average of the two was made. Analysis of expression differences was done with independent T-test. Analysis of correlation was done with Spearman correlation test. Statistic result is significant if $p < 0.05$.

RESULT

The data obtained in this study showed the sex distribution in the COVID-19 group, the number of subjects being male was 8 people (53%) and women were 7 people (47%). While in the non-COVID-19 group, the number of male subjects was 11 people (73%) and 4 women (27%).

Age data showed that in the COVID-19 group, the highest number was from the 41-50-year-old and 51-60-year age group, each of which amounted to 5 people (33.3%). Meanwhile, from the non-COVID-19 group, the highest number was in the age range of 61-70 years, namely 5 people (33.3%).

The results of this study showed the mean expression of IL-6 in the COVID-19 group was $80.67\% \pm 7.03$, while in the non-COVID-19 group it was 21.50 ± 11.60 . The difference between the two was then assessed using the Independent T test. The results of statistical analysis showed a significant difference between the expression of IL-6 in the COVID-19 group and the non-COVID-19 group with p value = 0.001 ($p < 0.05$).

The results of IL-6 expression in the COVID-19 and non-COVID-19 groups are presented in table 3. The average percentage of IL-6 expression in the COVID-19 group is higher than the non-COVID-19 group.

The results of this study showed the mean expression of IL-17 in the COVID-19 group was $75.33\% \pm 14.93$, while in the non-COVID-19 group it was $45.67\% \pm 17.31$. The difference between the two was then assessed using the Independent T test. The results of statistical analysis showed a significant difference between the expression of IL-17 in the COVID-19 group and the non-COVID-19 group with p value = 0.001 ($p < 0.05$).

Table 1: Distribution COVID-19 and non COVID-19 groups by gender.

	COVID-19 (n)	Non COVID-19 (n)
Male	8	11
Female	7	4

Table 2: Distribution COVID-19 and non-COVID-19 groups by age.

	COVID-19		Non COVID-19	
	Number	Percentage	Number	Percentage
< 10 years	0	0%	1	6.67%
11-20 years	0	0%	1	6.67%
21-30 years	1	6.67%	0	0%
31-40 years	3	20%	1	6.67%
41-50 years	5	33.3%	3	20%
51-60 years	5	33.3%	3	20%
61-70 years	1	6.67%	5	33.3%
> 70 years	0	0%	1	6.67%
Total	15	100%	15	100%

Table 3: Table of results of the assessment of IL-6 expression in the COVID-19 and non-COVID-19 groups.

Groups	n	Average (%)	SD	Min	Max	p
COVID-19	15	80.67	7.03	70	90	0.001
Non COVID-19	15	21.50	11.60	7.5	55	

Table 4: Table of results of the assessment of IL-17 expression in the COVID-19 and non-COVID-19 groups.

Groups	n	Average (%)	SD	Min	Max	P
COVID-19	15	75.33	14.93	30	90	0.001
Non COVID-19	15	45.67	17.31	20	75	

Table 5: Correlation of IL-6 and IL-17 expression in all samples.

		IL-17 expression
IL-6 expression	r_s	0.779
	p	0.001
	n	30

Table 6: Correlation of IL-6 and IL-17 expression in the COVID-19 patient group.

		IL-17 expression
IL-6 expression	r_s	0.548
	p	0.034
	n	15

The results of IL-17 expression in the COVID-19 and non-COVID-19 groups are presented in table 4. The average percentage of IL-17 expression in the COVID-19 group is higher than the non-COVID-19 group. Figure 2 shows an overview of IL-17 expression in the COVID-19 and non-COVID-19 groups.

The correlation between the expression of IL-6 and IL-17 in lung tissue of COVID-19 and non-COVID-19 patients was statistically tested using the Spearman correlation test. The results of the analysis showed a significant correlation between the expression of IL-6 and IL-17 in all samples (COVID-19 and non-COVID-19 groups) with a Spearman correlation coefficient (r_s) of 0.779 with $p = 0.001$ ($p < 0.05$), which means that the higher the expression of IL-6, the higher the expression of IL-17.

In the group of COVID-19 patients, a significant correlation was found between the expression of IL-6 and IL-17. The results of statistical analysis using the Spearman correlation test showed the Spearman coefficient (r_s) of 0.548 with $p = 0.034$ ($p < 0.05$) which means that the

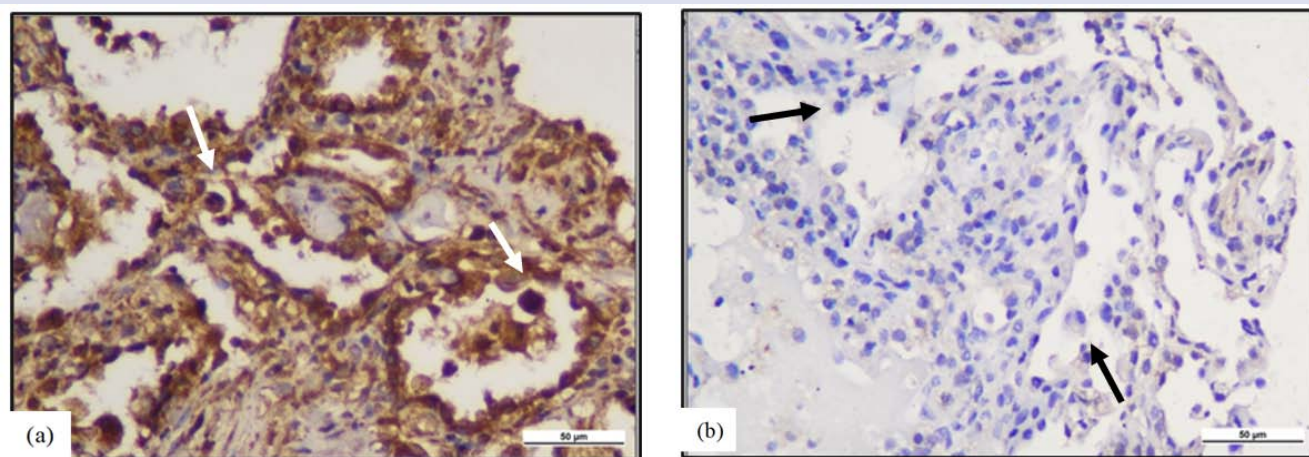


Figure 1: IL-6 expression in the COVID-19 and non-COVID-19 groups, magnification 400x. (a) IL-6 is positively expressed in lung tissue of COVID-19 patients (white arrow), (b) IL-6 is negatively expressed in lung tissue of non-COVID-19 patients (black arrow).

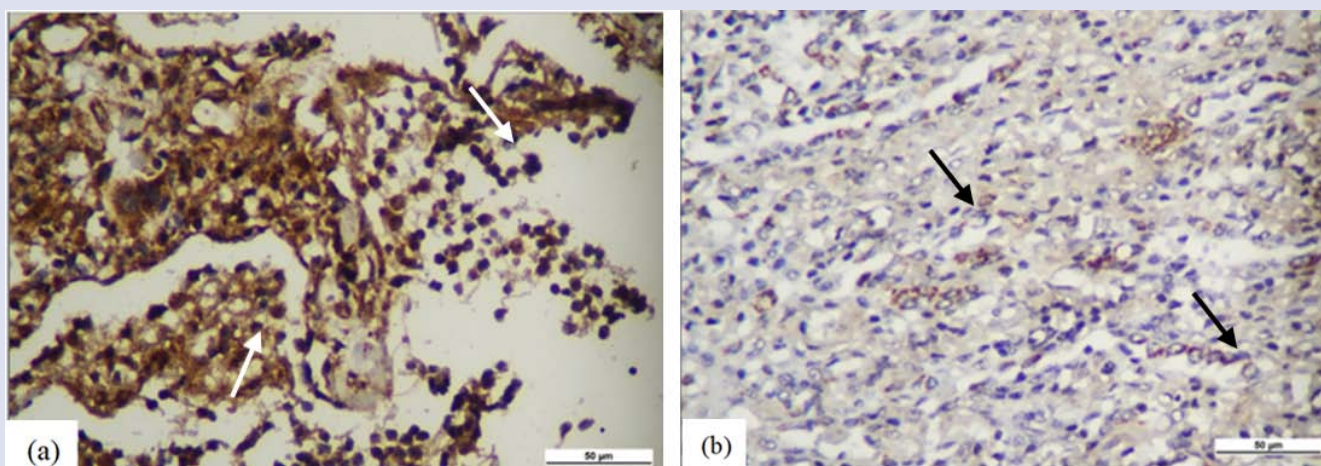


Figure 2: IL-17 expression in the COVID-19 and non-COVID-19 groups, magnification 400x. (a) IL-17 is positively expressed in lung tissue of COVID-19 patients (white arrow), (b) IL-17 is negatively expressed in lung tissue of non-COVID-19 patients (black arrow).

high expression of IL-6 in the tissue will be directly proportional to the expression of IL-17.

DISCUSSION

The data obtained in this study shows the sex distribution of COVID-19 cases, where the incidence of COVID-19 in men (53%) and women (47%) is almost the same. The incidence of disease and fatality in males was found to be slightly higher than that of females. This is in line with WHO data where the percentage of male sufferers is more (58%) in the UK 57% and 51.1% in China.¹⁰⁻¹²

Almost the same figure was also found in the incidence of COVID 19 in Indonesia. Several studies have shown higher morbidity and mortality rates in men, namely 58.94%,¹³ 56.5% with a fatality rate of 63.8%.¹⁴ Several studies have linked this event to several factors including sex hormones, the X chromosome (playing a role in high innate and adaptive immunity) and high expression of ACE 2 receptors, and lifestyle. Men smoke and consume alcohol more often.^{15,16}

The age range of the sample is between 5 and 88 years, with the mean age of the entire sample being 48.93 years. From these data, it was found that in the COVID-19 group, the highest number was from the 41-50-year-old and 51-60-year age group, each of which amounted to

5 people (33.3%). This is slightly different from several other studies where the highest fatality rate was found in the age range > 60 years, followed by the 40-59-year age group. While the smallest mortality rate was obtained from the age group < 10 years.^{11,14} The high mortality rate in the old age group is most likely due to the weak immune system in old age and the number of comorbid diseases that aggravate the course of COVID-19. In this study, the highest number was from the age group of 41-50 years and 51-60 years. This is different from other studies because this study did not assess the proportion of fatalities in each age group.

In addition to old age and male gender, several other risk factors that play a role in increasing mortality rates in COVID patients are the presence of co-morbidities such as diabetes, cardiovascular, cerebrovascular and kidney diseases, and obesity.^{15,17} Several socioeconomic conditions can also play a role, for example lack of socioeconomic conditions, high levels of air pollution and diurnal temperature variations.¹⁷

The data obtained from this study showed that the mean expression of IL-6 in the COVID-19 group was significantly higher than in the non-COVID-19 group. Previous studies have also shown high expression of IL-6 in lung tissue of COVID-19 patients, and is a predictor of fatality and disease progression.^{7,9,18,19}

Interleukin-6 is a pleiotropic cytokine that is activated in the presence of infection or injury, one of which is a viral infection. Interleukin-6 is secreted by various cell types (eg: T lymphocytes, macrophages, and fibroblasts) and plays a role in the regulation of various physiological processes.⁹ The role of IL-6 in the inflammatory process including COVID infection is like a double-edged sword, because it has two effects, namely pro- and anti-inflammatory. There are two pathways in the IL-6 signaling process, namely the classical pathway and the IL-6 signal trans pathway. The classical pathway is considered the anti-inflammatory pathway whereas the signal trans pathway is the pro-inflammatory pathway. Therefore, in various inflammatory conditions and autoimmune diseases, the role of the IL-6 signaling trans pathway is that inhibiting this process is needed to relieve inflammation and damage that occurs.²⁰ One of the roles of IL-6 as part of the pro-inflammatory function is in the activation and differentiation of Th17, and induces the expression of granzyme B and perforin on CD8 T lymphocytes that can eliminate the virus. Activation of these cytotoxic T cells will stimulate the release of TNF- and IFN- which causes neutrophil migration. In addition, IL-6 also plays a role in thermoregulation (as an endogenous pyrogenic cytokine) and amplifies the immune response to viral infections. The IL-6 signal is known to induce the production of IL-27 by monocytes and macrophages in the respiratory tract, thereby stimulating T cell maturation in the lungs.²⁰ Interleukin 6 has also been shown to be associated with inflammatory responses in the acute phase, including CRP and LDH.²¹ On the other hand, the anti-inflammatory function of IL-6 plays a role in the progression of viral infection, thereby causing viral persistence in the host. Interleukin-6 stimulates the Th2 response while inhibiting Th1 proliferation through two mechanisms. First, by stimulating CD4 T cells to secrete IL-4 which will stimulate Th2 and suppress the production of IFN- γ by CD4 T cells and cause Th1 polarization (antiviral response). Interleukin-6 will cause Th17 differentiation which will then produce IL-17 which will increase the expression of anti-apoptotic molecules so that the virus is stronger and can survive.¹⁹ A study proved that increased expression of IL-6 in lung tissue indicates the severity of COVID-19 sufferers.²²

Interleukin-17 plays a role in the pathogenesis of ARDS by increasing the infiltration of neutrophils into the lung. Increased signaling of IL-17A and Th17 or regulatory T cells (Treg) is known to be positively correlated with disease severity in MERS infection, where the cause is a type of virus that belongs to the same family as MERS. The overexpression of Th17 cells and the excessive cytotoxic effect of CD8+ T cells will cause tissue damage in COVID-19 patients with pneumonia. Like IL-6, high expression of IL-17A in lung tissue is also directly proportional to lung inflammation and poor outcomes (ARDS) and multi-organ dysfunction.^{23,24}

Various studies have shown the important role of IL-17A as a regulator of PMN infiltration. IL-17 is known to play a role in neutrophil maturation and differentiation by increasing the release of granulocyte-colony stimulating factor (G-CSF). This situation will cause the differentiation of hematopoietic progenitor cells CD34 into neutrophils. In addition, IL-17 can stimulate granulopoiesis markers and other chemokines, such as Growth-regulated Oncogene- α (GRO- α) which will regulate neutrophil penetration into tissues. Interleukin-17 also plays a role in the release of cytokines and other inflammatory chemokines such as IL-1, IL-6, TNF- α , macrophage inflammatory protein-2 (MIP-2), IL-8, Interferon-inducible protein-10 (IP-10). All these molecules will be used by neutrophils in the process of chemotaxis.²⁵

According to Hou *et al.*, IL-17 can also increase the expression of antiapoptotic molecules that increase the survival of virus-infected cells and block the process of destruction by cytotoxic T cells.²⁶ Therefore, the role of IL-17 in COVID inflammation is important in amplification of the inflammatory process.²⁵

The results of this study showed that the mean expression of IL-17 in the COVID-19 group was significantly higher than the non-COVID-19 group. Several previous studies showed high IL-17 expression in lung tissue of COVID-19 patients and was a predictor of fatality and disease progression.²³⁻²⁷ A similar study conducted by Avezedo *et al.*, which assessed the expression of IL-17 and IL-8 in post-mortem lung tissue of patients with COVID-19 also showed results that were in line with this study.²⁸

The results of the analysis of this study showed that there was a positive and significant correlation between the expression of IL-6 and IL-17 in all samples (COVID-19 and non-COVID-19 groups) as well as when analyzed only in the COVID-19 group. The COVID-19 group showed a higher correlation. This phenomenon can be seen in general that the pathogenesis of COVID-19 may be different from other infectious processes. In COVID-19, there is an exaggerated immune response that is characteristic of this disease. The state of hypercytokinemia or what is called a cytokine storm is a collection of symptoms caused by the activation of T lymphocytes, macrophages and the release of various cytokines which then activate other immune cells. In the laboratory, cytokine storm is characterized by an increase in various proinflammatory cytokines such as IL-1, IL-6, IL-12, IFN- γ . Measurement of these levels, especially IL-6 is considered useful as a predictor of fatality rates and the need for respiratory equipment, and can be used as a potential therapeutic target.^{7,9,19}

The SARS-CoV2 virus which is the etiology of COVID-19 has a surface protein, namely the surface spike (S) protein, which will bind to the Angiotensin Converting Enzyme-2 (ACE-2) receptor found in pulmonary alveolar epithelial cells to enter the body. The ACE-2 receptor is very important in viral virulence, because it has been shown that cells that do not have this receptor are resistant to viral infection. Angiotensin Converting Enzyme-2 is not only expressed in the respiratory tract, but also in the small intestine, pancreas, kidney, heart, esophagus, bladder and brain. This has led to multi-organ involvement in COVID-19. Under normal conditions, ACE-2 functions to cause the conversion of angiotensin II to angiotensin 1-7 in the renin-angiotensin-aldosterone system (RAA system). Various studies have reported increased levels of Angiotensin II in COVID-19 patients compared to healthy individuals, which is associated with viral load and lung damage. This situation is caused by the virus binding to ACE-2, so that Angiotensin II cannot be converted, accumulates and causes an imbalance in the RAA system. Angiotensin II buildup will cause a change in equilibrium towards a proinflammatory state.¹⁹ In the absence of ACE-2, angiotensin II will bind to the angiotensin I receptor which will cause an oxidative stress cascade, resulting in the formation of Reactive Oxygen Species (ROS) and an increase in IL-6 expression. On the other hand, the increased expression of IL-6 will also increase the expression of angiotensin I receptors and attach to endothelial cell walls and cause cellular and vascular inflammation. Interleukin-6 and angiotensin II will cause an increase in the expression of each other continuously so that over time it will aggravate the state of oxidative stress.¹⁹

The role of IL-6 in inflammatory processes including COVID infection is like a double-edged sword, because it has two effects, namely pro- and anti-inflammatory.^{9,19} In pro-inflammatory, one of the functions of IL-6 is in the activation and differentiation of Th17 and the induction of granzyme B and perforin in CD8 T lymphocyte cells. Interleukin-6 causes the production of IL-17 which will further increase the expression of antiapoptosis molecules leading to increasing the survival of virus-infected cells and blocking the process of digestion by cytotoxic T cells.²⁶ Interleukin-17 mainly causes neutrophil migration to the lungs thus causing tissue damage during the inflammatory process.¹⁹ Interleukin-17 also plays a role in the release of cytokines and other inflammatory chemokines such as IL-1, IL-6, TNF- α , macrophage

inflammatory protein-2 (MIP-2), IL-8, Interferon-inducible protein-10 (IP-10).¹⁹

The synergistic interaction between IL-17A and IL-6 plays an important role in the formation of pulmonary fibrosis and imbalances of the respiration system. IL-6 not only plays a role in the differentiation of T helper (Th) cells into Th17 cells through ROR γ t which is the JAK-STAT3 pathway but also in the process of pulmonary fibrosis (causing collagen deposition) due to the function of epithelial cells and fibroblasts that are aberrant (works synergistically with IL-8).^{6,29}

CONCLUSION

There are significant differences between IL-6 and IL-17, which are higher expressed in the lung tissue of covid-19 patients than the lung tissue of the non-COVID-19 group. There was also a strong correlation between IL-6 and IL-17 in the lung tissue of COVID-19 patients, as well as in the entire sample. Although not a single factor, IL-6 and IL-17 play a role in the pathogenesis of COVID-19.

ACKNOWLEDGEMENT

The authors are grateful for support from the director and research and development unit Dr. Soetomo General Academic Hospital.

ETHICS AND CONSENT

This research is a branch of research titled 'Multi-organ pathological finding in COVID-19 infection through post-mortem core biopsy' and has received ethical approval from the Health Research Ethics Committee of Dr. Soetomo general hospital number 0022/KEPK/VII/2020.

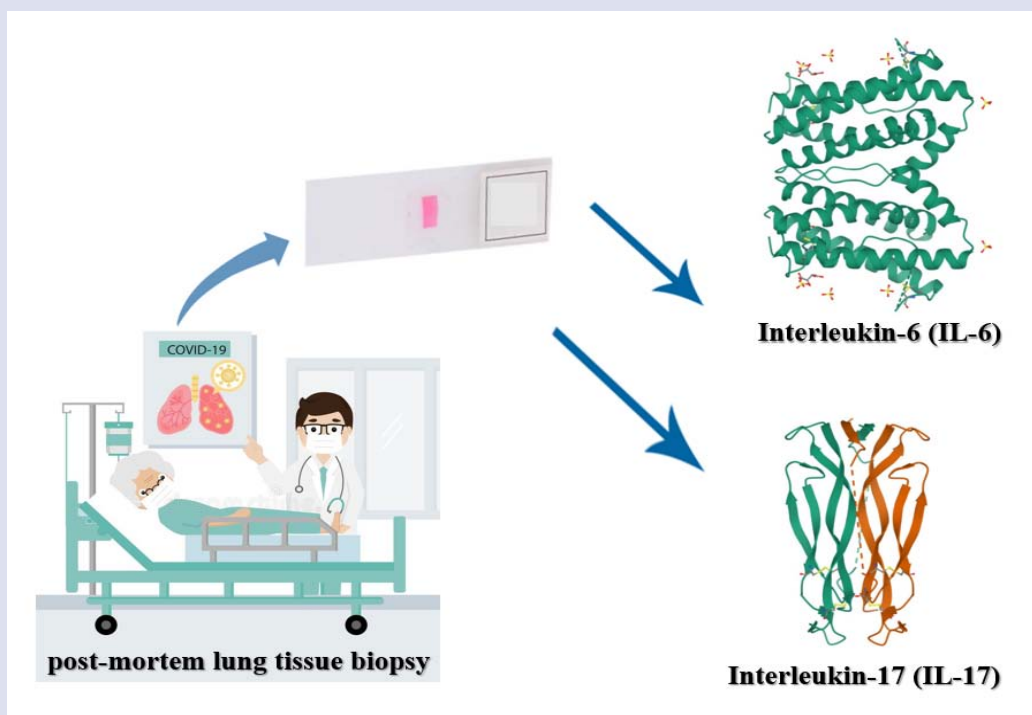
CONFLICTS OF INTEREST/PLAGIARISM REPORT

There are no conflicts of interest in this research.

REFERENCES

- Zhou P, Yang XL, Wang XG. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(7798):270-3.
- Lee C, Choi WJ. Overview of COVID-19 inflammatory pathogenesis from the therapeutic perspective. *Arch Pharm Res*. 2021;44(1):99-116.
- Maiese A, Manetti AC, La Russa R. Autopsy findings in COVID-19-related deaths: a literature review. *Forensic Sci Med Pathol*. 2021;17(2):279-96.
- Mendoza VM. Interleukin-17: A potential therapeutic target in COVID-19. *J Infection*. 2020;81(2):136-8.
- Costela-Ruiz VJ, Illescas-Montes R, Puerta-Puerta JM, Ruiz C, Melguizo-Rodríguez L. SARS-CoV-2 infection: The role of cytokines in COVID-19 disease. *Cytokine Growth Factor Rev*. 2020;54:62-75.
- Shibabaw T. Inflammatory Cytokine: IL-17A Signaling Pathway in Patients Present with COVID-19 and Current Treatment Strategy. *J Inflamm Res*. 2020;13(3):673-80.
- Herold T, Jurinovic V, Arnreich C. Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19. *J Allergy Clin Immunol*. 2020;146(1):128-36.
- Zhang ZL, Hou YL, Li DT, Li FZ. Laboratory findings of COVID-19: a systematic review and meta-analysis. *Scand J Clin Lab Invest*. 2020;80(6):441-7.
- Santa Cruz A, Mendes-Frias A, Oliveira AI. Interleukin-6 is a Biomarker for the Development of Fatal Severe Acute Respiratory Syndrome Coronavirus 2 Pneumonia. *Front Immunol*. 2021;12(1):613422.
- Fadlallah S, Sham Eddin MS, Rahal EA. IL-17A in COVID-19 Cases: a meta-analysis. *J Infect Dev Ctries*. 2021;15(11):1630-9.
- Brown AE, Heinsbroek E, Kall MM. Epidemiology of Confirmed COVID-19 Deaths in Adults, England, March-December 2020. *Emerg Infect Dis*. 2021;27(5):1468-71.
- Park SE. Epidemiology, virology, and clinical features of severe acute respiratory syndrome -coronavirus-2 (SARS-CoV-2; Coronavirus Disease-19). *Clin Exp Pediatr*. 2020;63(4):119-24.
- Sutaryono, Andasari SD, Kasjono HS. Diagnosis and epidemiology of Coronavirus (COVID-19) outbreak in Indonesia. *J Teknologi Laboratorium*. 2020;9(1):49-57.
- Hikmawati I, Setiyabudi R. Epidemiology of COVID-19 in Indonesia: common source and propagated source as a cause for outbreaks. *J Infect Dev Ctries*. 2021;15(5):646-52.
- Noor AU, Maqbool F, Bhatti ZA, Khan AU. Epidemiology of CoViD-19 Pandemic: Recovery and mortality ratio around the globe. *Pak J Med Sci*. 2020;36(COVID19-S4):S79-84.
- Bwire GM. Coronavirus: why men are more vulnerable to Covid-19 than women? *SN Compr Clin Med*. 2020;2(7):874-6.
- Cao Y, Hiyoshi A, Montgomery S. COVID-19 case-fatality rate and demographic and socioeconomic influencers: worldwide spatial regression analysis based on country-level data. *BMJ Open*. 2020;10(11):e043560.
- Smetana K, Brabek J. Role of interleukin-6 in lung complications in patients with COVID-19: therapeutic implications. *In Vivo*. 2020;34(3 Suppl):1589-92.
- Shekhawat J, Gauba K, Gupta S. Interleukin-6 Perpetrator of the COVID-19 Cytokine Storm. *Indian J Clin Biochem*. 2021;36(4):440-50.
- Su H, Lei CT, Zhang C. Interleukin-6 Signaling Pathway and Its Role in Kidney Disease: An Update. *Front Immunol*. 2017;8:405.
- Adiatmaja CO, Nugraha J, Utari A. Correlation between Interleukin-6, CRP, and LDH in COVID-19 patients of Dr. Soetomo Teaching Hospital, Surabaya, Indonesia. *Res J Pharm Technol*. 2022;15(10):4753-7.
- Nugroho GMS, Marhana IA, Kusumastuti EH, Semedi BP, Maimunah U, Lefi A, *et al.* Interleukin-6 (IL-6) expression of lung tissue in COVID-19 patient severity through core biopsy post mortem. *Ann Med Surg*. 2022;82(2022):104648.
- Kang YW, Lee SC, Jeon SM, Jo EK. Roles of interleukin-17 and Th17 responses in COVID-19. *J Bacteriol Virol*. 2021;51(3):89-102.
- Feng CM, Wang XM, Li MD. Serum interleukin-17 predicts severity and prognosis in patients with community acquired pneumonia: a prospective cohort study. *BMC Pulm Med*. 2021;21(1):393.
- Maione F, Casillo GM, Raucci F. Interleukin-17A (IL-17A): A silent amplifier of COVID-19. *Biomed Pharmacother*. 2021;142:111980.
- Hou W, Kang HS, Kim BS. Th17 cells enhance viral persistence and inhibit T cell cytotoxicity in a model of chronic virus infection. *J Exp Med*. 2009;206(2):313-28.
- Pourgholaminejad A, Pahlavanneshan S, Basiri M. COVID-19 immunopathology with emphasis on Th17 response and cell-based immunomodulation therapy: Potential targets and challenges. *Scand J Immunol*. 2022;95(2):e13131.
- Azevedo MLV, Zanchettin AC, Vaz de Paula CB. Lung Neutrophilic Recruitment and IL-8/IL-17A Tissue Expression in COVID-19. *Front Immunol*. 2021;12:656350.
- Darif D, Hammi I, Kihel A, El Idrissi Saik I, Guessous F, Akarid K. The pro-inflammatory cytokines in COVID-19 pathogenesis: What goes wrong? *Microb Pathog*. 2021;153:104799.

GRAPHICAL ABSTRACT



ABOUT AUTHORS



Etty Hary Kusumastuti is a Pathologist at Dr. Soetomo General Academic Hospital and as a lecturer in Department of Anatomical Pathology, Faculty of Medicine, Universitas Airlangga, Indonesia. She completed her residency in Anatomical Pathology at the Faculty of Medicine of Universitas Airlangga, Indonesia. She graduated doctoral degree in clinical medicine at the Faculty of Medicine of Universitas Airlangga, Indonesia. She is one of the most expert in cytopathology of respiratory tract. She has participated in many conferences, workshops, and courses as a speaker and participant. She writes a number of scientific papers on various subjects of pathology.



Isnin Anang Marhana is an interventional pulmonologist at Dr. Soetomo General Academic Hospital, as well as a lecturer in Pulmonology and Respiratory Medicine, Faculty of Medicine, Universitas Airlangga, Indonesia. After graduating as a resident of pulmonology at Faculty of Medicine, Universitas Airlangga, he then underwent interventional bronchoscopy training in St. Marianna University, Kawasaki, Japan. He is a researcher and the author of a number of studies in the field of lungs and especially COVID19.

Cite this article: Kusumastuti EH, Wiratama PA, Ariani G, Djuanda SN, Rahniayu A, Kurniasari N, et al. Differences in interleukin-6 and interleukin-17 expression in covid-19 post-mortem lung tissue biopsy compared with non-covid-19. *Pharmacogn J.* 2022;14(6) Suppl: 887-892.



Source details

Pharmacognosy Journal

Scopus coverage years: from 2009 to 2022

Publisher: Pharmacognosy Network Worldwide

ISSN: 0975-3575

Subject area: Pharmacology, Toxicology and Pharmaceutics: Pharmacology

Pharmacology, Toxicology and Pharmaceutics: Drug Discovery

Source type: Journal

CiteScore 2021

1.9



SJR 2021

0.258



SNIP 2021

0.718



[View all documents >](#)

[Set document alert](#)

[Save to source list](#)

[CiteScore](#) [CiteScore rank & trend](#) [Scopus content coverage](#)

i Improved CiteScore methodology



CiteScore 2021 counts the citations received in 2018-2021 to articles, reviews, conference papers, book chapters and data papers published in 2018-2021, and divides this by the number of publications published in 2018-2021. [Learn more >](#)

CiteScore 2021

$$1.9 = \frac{1,760 \text{ Citations 2018 - 2021}}{946 \text{ Documents 2018 - 2021}}$$

Calculated on 05 May, 2022

CiteScoreTracker 2022

$$1.9 = \frac{1,635 \text{ Citations to date}}{863 \text{ Documents to date}}$$



Last updated on 05 February, 2023 • Updated monthly

CiteScore rank 2021

Category	Rank	Percentile
Pharmacology, Toxicology and Pharmaceutics	#219/303	27th
└ Pharmacology		
Pharmacology, Toxicology and Pharmaceutics	#116/154	25th
└ Drug Discovery		

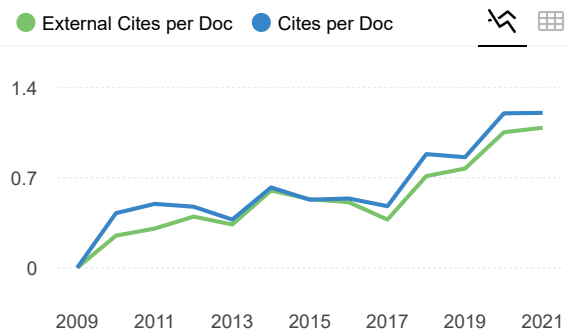
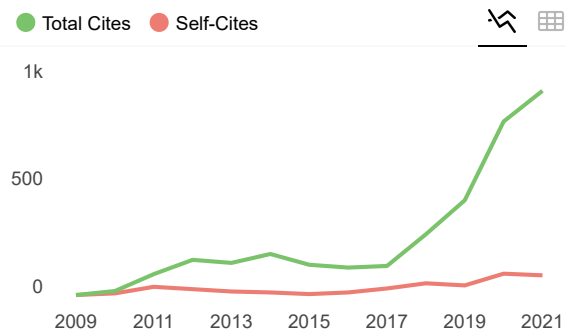
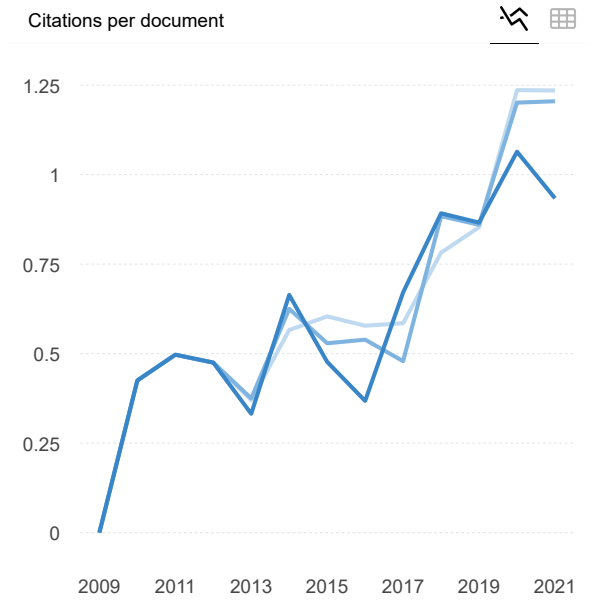
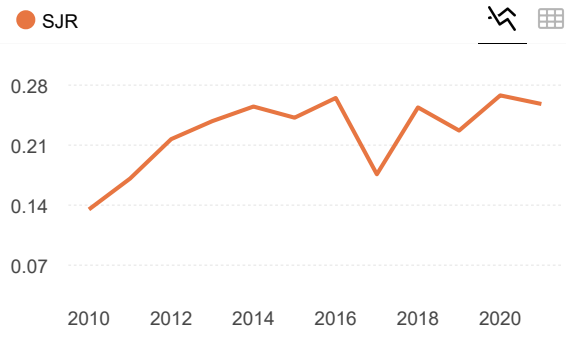
[View CiteScore methodology >](#) [CiteScore FAQ >](#) [Add CiteScore to your site](#)

Pharmacognosy Journal

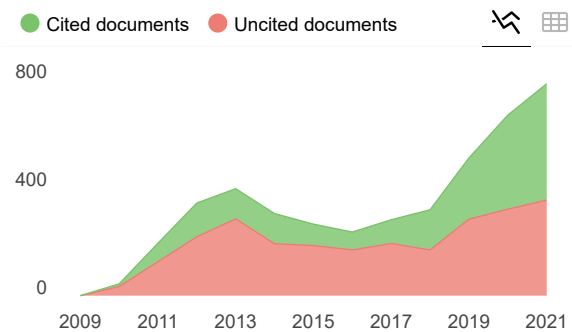
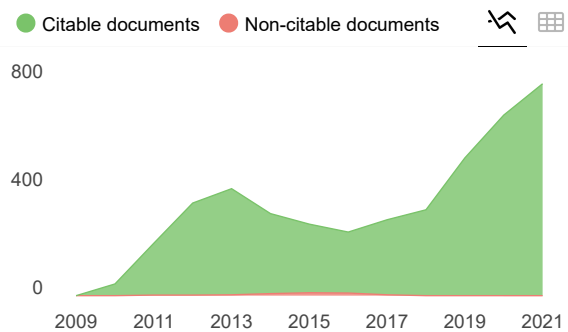
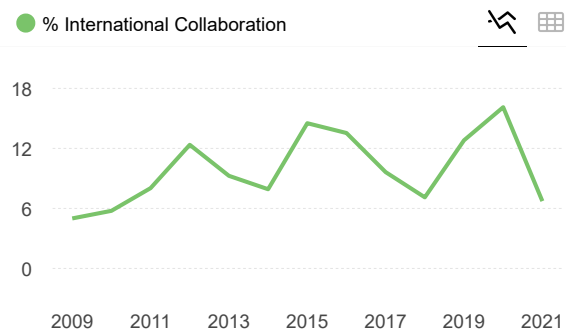
COUNTRY India  Universities and research institutions in India  Media Ranking in India	SUBJECT AREA AND CATEGORY Pharmacology, Toxicology and Pharmaceutics Drug Discovery Pharmacology	PUBLISHER EManuscript Technologies	H-INDEX 25
PUBLICATION TYPE Journals	ISSN 09753575	COVERAGE 2009-2021	INFORMATION Homepage How to publish in this journal editor@phcogj.com

SCOPE

Pharmacognosy Journal (Phcog J.) covers different topics in natural product drug discovery, and also publishes manuscripts that describe pharmacognostic investigations, evaluation reports, methods, techniques and applications of all forms of medicinal plant research



● Cites / Doc. (4 years)
● Cites / Doc. (3 years)
● Cites / Doc. (2 years)



Pharmacognosy Journal

Show this widget in your own website

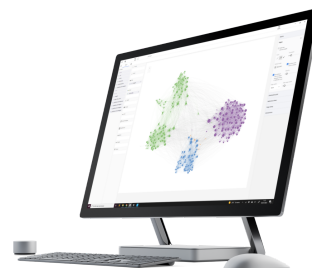
Q3 Pharmacology
best quartile

SJR 2021
0.26

powered by scimagojr.com

Just copy the code below and paste within your html code:

```
<a href="https://www.scimagojr.com" style="color: #0070C0; text-decoration: none; border: 1px solid #0070C0; padding: 2px 5px;">https://www.scimagojr.com
```



SURAT EXEMPTION
(" LETTER OF EXEMPTION ")

Ref. No. : 1196/LOE/301.4.21/2023

Judul Protokol Penelitian : Korelasi antara N-cadherin dan MMP-9 pada berbagai stadium N radiologis pada squamous cell carcinoma laring

Dokumen yang disetujui : 1878/125/4/XII/2022 (versi: 3)

Tanggal terbit : 21 Januari 2023

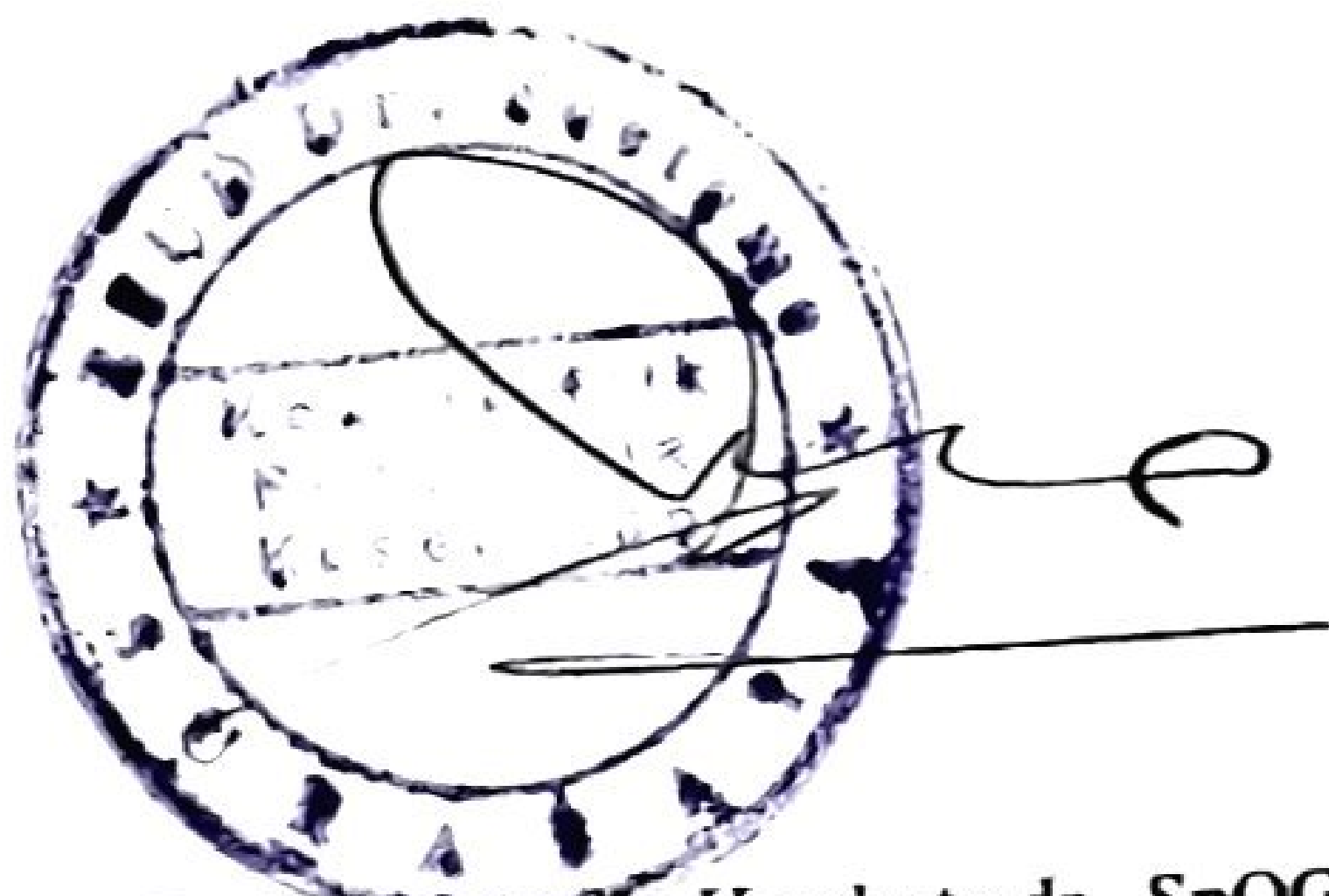
Berlaku sampai : 21 Januari 2024

Peneliti Utama : Dr. Etty Hary Kusumastuti, dr., Sp.PA (K)., FIAC

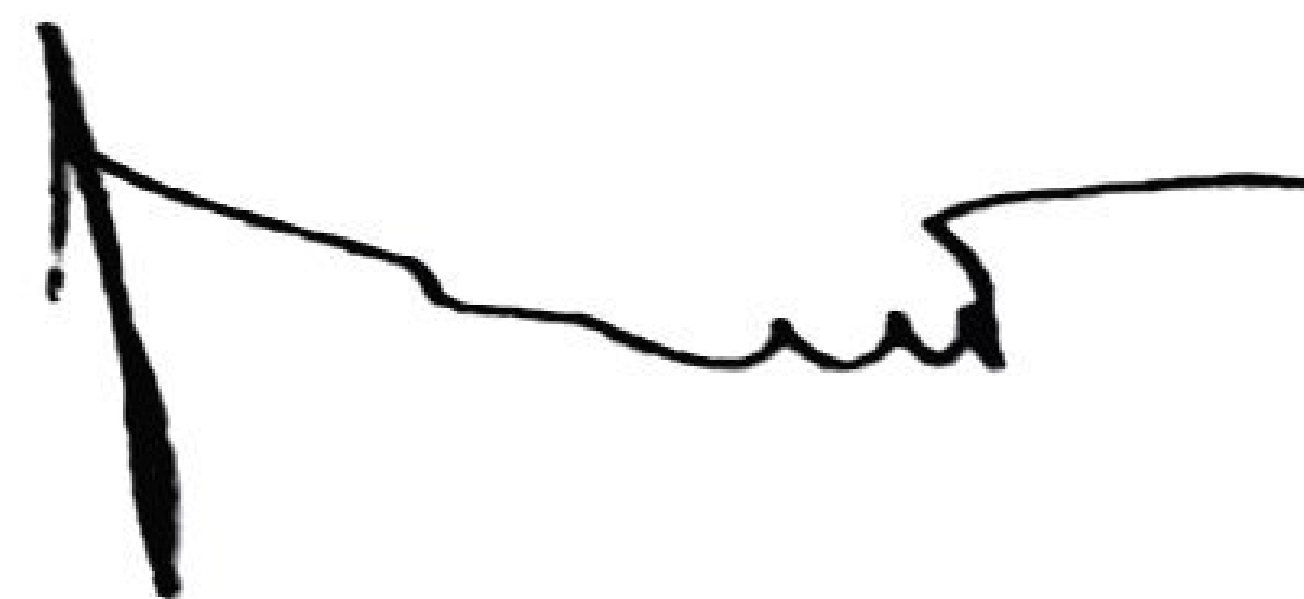
Peneliti Lain : 1. Grace Ariani, dr., SpPA(K)
2. Stephanie Natasha Djuanda, dr

Instalasi/Tempat Penelitian : RSUD Dr. Soetomo

Komite Etik Penelitian Kesehatan RSUD Dr Soetomo menyatakan bahwa dokumen diatas sesuai dengan The Office for Human Research Protections (OHRP) dibawah persyaratan the U.S. Department of Health and Human Services (HHS) Regulasi 45 CFR bagian 46 untuk **exempt review**.



Prof. Dr. Hendy Hendarto dr., SpOG(K)
Ketua Panel 1



Dra. Siti Farida SpFRS. Apt
Sekretaris Panel 1