

[HOME](#) / [ARCHIVES](#) / [Advanced online](#)

## Advanced online

PUBLISHED: 2023-01-01

### EDITORIAL

---



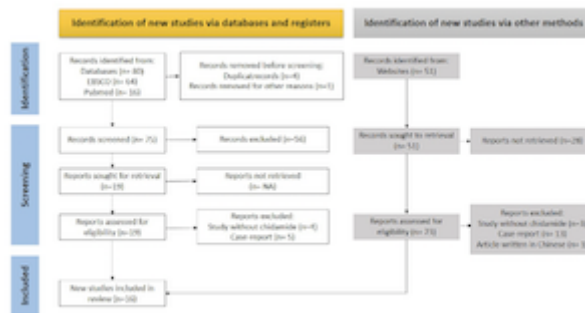
### COP27 Climate Change Conference: Urgent action needed for Africa and the world

Lukoye Atwoli, Gregory E. Erhabor, Aiah A. Gbakima, Abraham Haileamlak, Jean-Marie Kayembe Ntumba, James Kigera, Laurie Laybourn-Langton, Robert Mash, Joy Muhia, Fhumulani Mavis Malaudzi, David Ofori-Adjei, Fricay Okonofua, Arash Rashidian, Maha El Adawy, Siaka Sidibe, Abdelmadjid Snouber, James Tumwine, Mohammad Sahar Yassien, Paul Yonga, Lilia Zakhama, Chris Zielinski

[FULL TEXT](#)[PDF](#)[Citations](#) 0

### REVIEWS

---



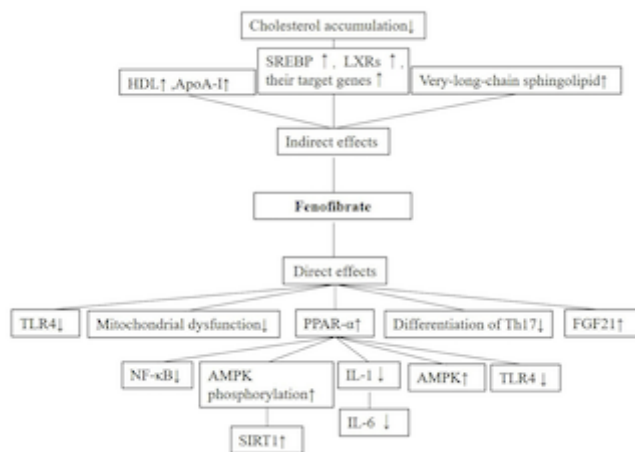
## The role of chidamide in the treatment of B-cell non-Hodgkin lymphoma: An updated systematic review

Hastono Ridwansyah, Indra Wijaya, Muhammad Hasan Bashari, Achmad Hussein Sundawa Kartamihardja, Bethy Suryawathy Hernowo

FULL TEXT

PDF

Citations 0



## Anti-inflammatory role of fenofibrate in treating diseases

Lv Jin, Hu Hua, Yong Ji, Zhanjun Jia, Mingqi Peng, Songming Huang

FULL TEXT

PDF

Citations 1



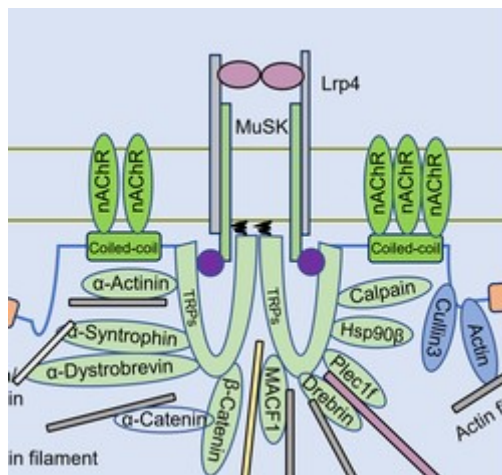
## Myocarditis and coronavirus disease 2019 vaccination: A systematic review and meta-summary of cases

Pandit Bagus Tri Saputra, Roy Bagus Kurniawan, Desy Trilistyoti, Makhyani Jibril Al Farabi, Hendri Susilo, Mochamad Yusuf Alsagaff, Yudi Her Oktaviono, Henry Sutanto, Arief Gusnanto, Citrawati Dyah Kencono Wungu

FULL TEXT

PDF

Citations 0



### The role of Rapsyn in neuromuscular junction and congenital myasthenic syndrome

Xufeng Liao, Yingxing Wang, Xinsheng Lai, Shunqi Wang

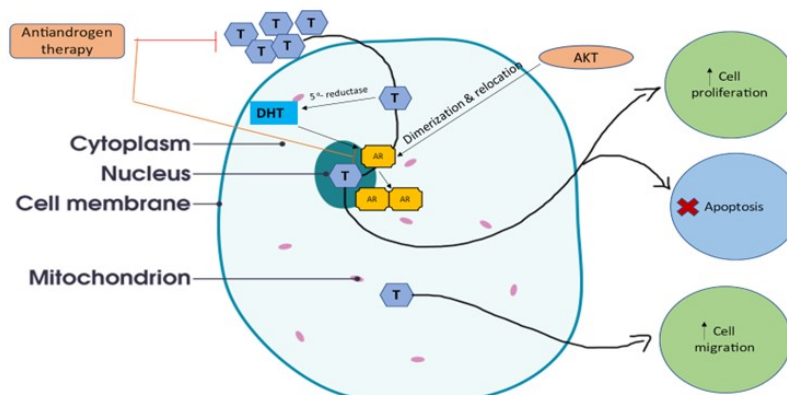
📄 FULL TEXT
📄 PDF
📄 Citations
0

Type of videolaryngoscope	Description	Examples
Macintosh-style videolaryngoscope (Macintosh-style VL)	<ul style="list-style-type: none"> <li>Macintosh-style blades and video technology</li> <li>Similar insertion procedure to the Macintosh laryngoscope</li> <li>A tube introducer and laryngeal pressure are often needed to clearly visualize the glottis</li> </ul>	<ul style="list-style-type: none"> <li>Storz V-MAC</li> <li>C-MAC</li> <li>McGrath MAC</li> </ul>
Hyperangulated videolaryngoscope (HA-VL)	<ul style="list-style-type: none"> <li>J-shaped or hyper-angulated blade able to visualize the glottis without flexing or extending the neck</li> <li>Offers an indirect view of the glottis, and intubation requires a pre-shaped stylet</li> <li>Upper airway trauma could occur when the tip is not visible</li> </ul>	<ul style="list-style-type: none"> <li>Gleescope</li> <li>McGrath Series 5</li> <li>TruView Devices</li> <li>Bullard laryngoscope</li> <li>Storz D-blade</li> <li>AP Venner scope</li> </ul>
Channelled videolaryngoscope (Channelled-VL)	<ul style="list-style-type: none"> <li>J-shaped or hyper-angulated blade with a canal that guides the tracheal tube to the glottis</li> <li>The tube tip is visible throughout tracheal intubation</li> </ul>	<ul style="list-style-type: none"> <li>Airtraq</li> <li>Pentax-Airway Scope</li> </ul>

### Videolaryngoscopy as a primary intubation modality in obstetrics: A narrative review of current evidence

Tatjana Stopar Pintarič

📄 FULL TEXT
📄 PROVISIONAL PDF
📄 Citations
0



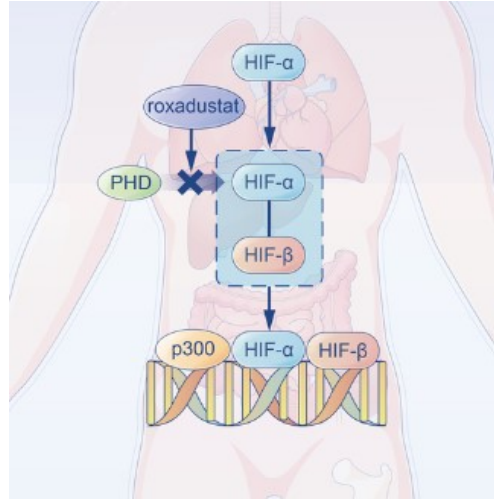
### A comprehensive review on the molecular basis and therapeutic targets in prostate cancer

Florentina Claudia Militaru, Valentin Militaru, Nicolae Crisan, Ioana Corina Bocsan, Anghel Adrian Udrea, Andreea Catana, Eniko Kutasi, Mariela Sanda Militaru

 FULL TEXT

 PROVISIONAL PDF

 Citations 0



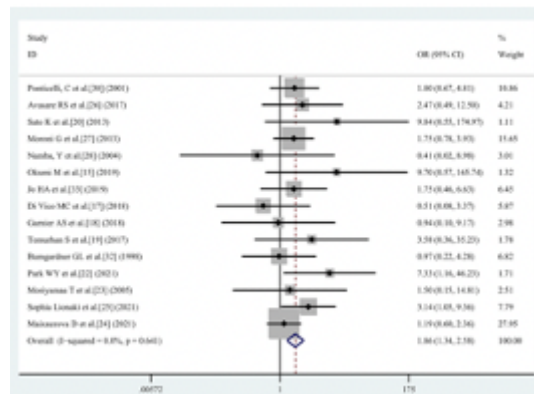
**Roxadustat: Do we know all the answers?**

Qiu-Yu Li, Qian-Wen Xiong, Xuefeng Yao, Fei Liu, Xiaoxiao Tang, Haidong Fu, Tong Tong, Jianhua Mao, Wan-Xin Peng

 FULL TEXT

 PDF

 Citations 0



**Risk factors for recurrent IgA nephropathy after renal transplantation: A meta-analysis**

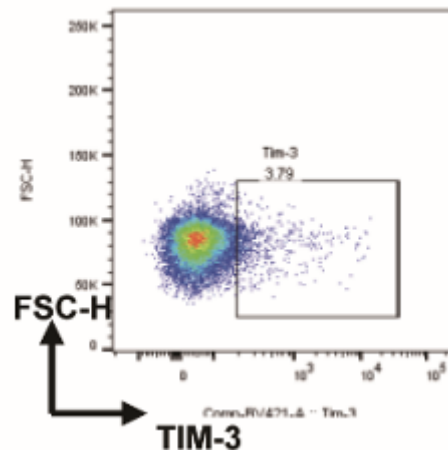
Jiang Bai, Qiong Wu, Jing Chen, Zhifang Zheng, Jiarong Chang, Liangliang Wang, Yun Zhou, Qiang Guo

 FULL TEXT

 PDF

 Citations 0

**IMMUNOLOGY**



### Increased co-expression of TIM-3 with TIGIT or 2B4 on CD8+ T cells is associated with poor prognosis in locally advanced nasopharyngeal carcinoma

Xiaolin Xie, Yaning Feng, Peiwen Fan, Danning Dong, Xuan Yao, Yanchun Peng, Ruo Zheng Wang

[FULL TEXT](#)

[PDF](#)

[Citations](#) 0



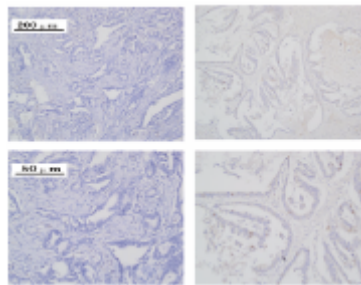
### Identification of RUNX1 and IFNGR2 as prognostic-related biomarkers correlated with immune infiltration and subtype differentiation of low-grade glioma

Xia Zhang, Hongyu Chu, Yuan Cheng, Jie Ren, Wei Wang, Xicheng Liu, Xiaodong Yan

[FULL TEXT](#)

[PDF](#)

[Citations](#) 0



### C5aR1 shapes a non-inflammatory tumor microenvironment and mediates immune evasion in gastric cancer

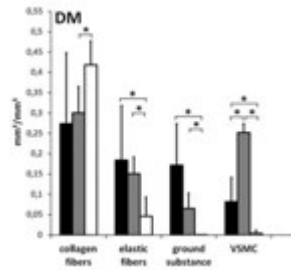
Honghong Shen, Xianhua Gu, Xinwei Li, Zheng Xiang, Rong Zhang, Fan Shi, Mingyue Tang, Huiyuan Li, Guangzheng Zhu, Fang Su, Zishu Wang

[FULL TEXT](#)

[PDF](#)

[Citations](#) 0

## PATHOLOGY



## Differences between inflammatory cells infiltrated into tunica intima, media, and adventitia of ascending aortic aneurysms within diabetic and hypertensive patients

Aleksandra Milutinović, Marko Zivin, Ruda Zorc-Pleskovič

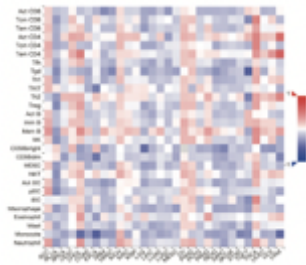
FULL TEXT

PDF

Citations 0

## MOLECULAR BIOLOGY

---



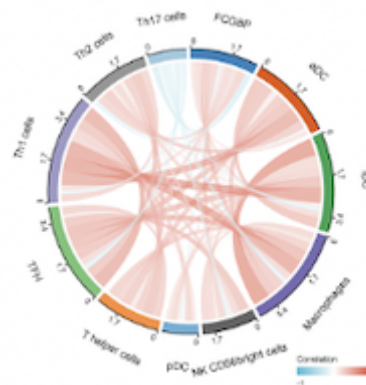
## Systemic analysis of the expression and prognostic significance of USP31 in endometrial cancer

Yuzhen Huang, Peng Jiang, Yuting Chen, Jinyu Wang, Rui Yuan

FULL TEXT

PDF

Citations 0



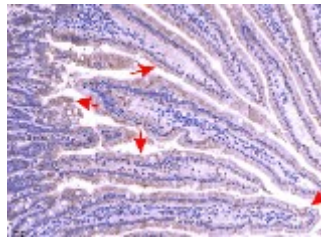
## Fc fragment of IgG binding protein is correlated with immune infiltration levels in hepatocellular carcinoma

Yuhong Suo, Chunyu Hou, Guang Yang, Hongfeng Yuan, Lina Zhao, Yufei Wang, Ningning Zhang, Xiaodong Zhang, Wei Lu

FULL TEXT

PDF

Citations 0



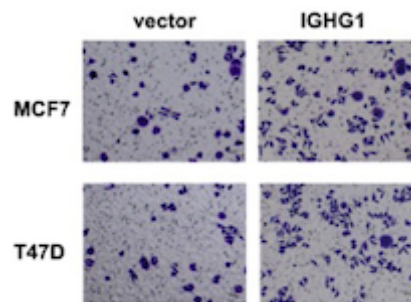
## Promotive role of IRF7 in ferroptosis of colonic epithelial cells in ulcerative colitis by the miR-375-3p/SLC11A2 axis

Zepeng Chen, Qinglong Gu, Ruichao Chen

[FULL TEXT](#)

[PDF](#)

Citations 0



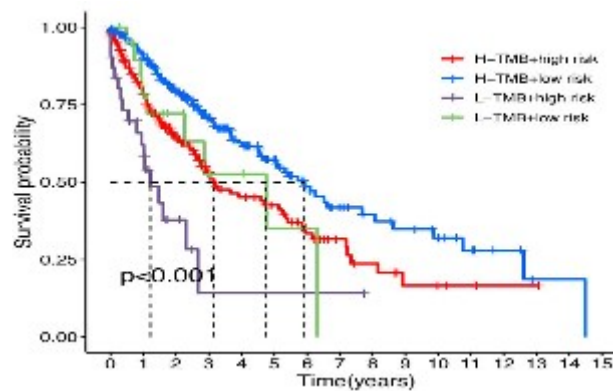
## IGHG1 promotes malignant progression in breast cancer cells through the regulation of AKT and VEGF signaling

Yong Zhang, Xueying Fang, Yan Sun

[FULL TEXT](#)

[PDF](#)

Citations 0



## A cuproptosis-associated long non-coding RNA signature for the prognosis and immunotherapy of lung squamous cell carcinoma

Chunlan Hou, Xiuping Wu, Caoyang Li, Chao Wang, Jinbo Liu, Qing Luo

[FULL TEXT](#)

[PDF](#)

Citations 0

Variables	DS with HT (n = 40)	Control (n = 50)	P value	OR (95% CI)
Genotypes, n (%)			0.031	
AA	6 (15)	20 (40)	<b>0.012*</b>	0.265 (0.094 - 0.746)
AG	22 (55)	21 (42)	0.221	1.688 (0.730 - 3.905)
GG	12 (30)	9 (18)	0.185	1.952 (0.726 - 5.248)
Allele, n (%)				
A	68 (42.5)	122 (61)	<b>0.0002*</b>	0.472 (0.309 - 0.721)
G	92 (57.5)	78 (39)	<b>0.0003*</b>	2.116 (1.386 - 3.23)

## Cytotoxic T-lymphocyte-associated protein 4 +49A/G polymorphism in Down syndrome children with Hashimoto's thyroiditis

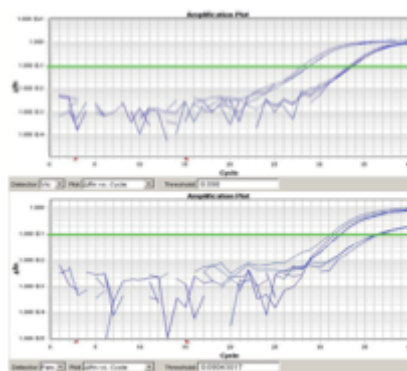
Muhammad Faizi, Nur Rochmah, Yuni Hisbiyah, Rayi Kurnia Perwitasari, Qurrota Ayuni Novia Putri, Sukmawati Basuki, Anang Endaryanto, Soetjipto Soetjipto

 FULL TEXT

 PDF

 Citations 0

## MICROBIOLOGY



## A novel approach for simultaneous detection of the most common food-borne pathogens by multiplex qPCR

Emir Hodzic, Aida Glavinic, Cara Wademan

 FULL TEXT

 PDF

 Citations 0

Table 1. Participants' demographics by sex and age.

		Sex		Age group (years)	
		Female n (%)	Male n (%)	18-36 n (%)	37-54 n (%)
Age group (years)	18-36	14 (87.5)	142 (81.6)	-	-
	37-54	2 (12.5)	32 (18.4)	-	-
Agwar residence	No	15 (93.8)	119 (68.4)	103 (66.0)	31 (91.2)
	Yes	1 (6.3)	55 (31.6)	53 (34.0)	3 (8.8)
Military serviceman	No	16 (100)	165 (94.8)	148 (94.9)	33 (97.1)
	Yes	0 (0)	9 (5.2)	8 (5.1)	1 (2.9)

## Transfusion transmitted virus and dengue virus among healthy blood donors: A prevalence report from Jordan



HOME / Editorial Team

## Editorial Team

### Editors-in-Chief

**Muzafer Mujic**, Association of Basic Medical Sciences, Bosnia and Herzegovina  
(muzafer.mujic@bjbms.org)

**Semir Vranic**, College of Medicine, Qatar University, Qatar (semir.vranic@bjbms.org)

### Editors

**Nicolai Wewer Albrechtsen**, University of Copenhagen, Copenhagen, Denmark

**Jurica Arapovic**, University of Mostar, Mostar, Bosnia and Herzegovina

**Zhe-Sheng Chen**, College of Pharmacy and Health Sciences, St. John's University, New York, USA

**Ivan Damjanov**, University of Kansas School of Medicine, Kansas City, USA

**Sijana Dzinic**, Wayne State University, Detroit, MI, USA

**Eyad Elkord**, University of Salford, Manchester, UK

**Emir Hodzic**, University of California, Davis School of Veterinary Medicine, USA

**Amina Kozaric**, Department of Pathology, University of Sarajevo Clinical Center, Bosnia and Herzegovina

**Gordan Srkalovic**, Sparrow Cancer Center, Sparrow Hospital System, Lansing, MI, United States

**Ljiljana Šerman**, University of Zagreb Medical School, Croatia

### Managing Editor

**Faruk Skenderi**, Department of Pathology, UniMed Clinic, Sarajevo School of Science and Technology, Bosnia and Herzegovina

### Editorial Board

**Saghir Akhtar**, College of Medicine, Qatar University, Doha, Qatar

**Kenan Arnautovic**, Semmes-Murphey Clinic, Memphis, TN, United States

**Mohammad Amin Sherifat Bafrouei**, Waisman Center, University of Wisconsin-Madison, WI,

United States

**Adnan Beganović**, University of Sarajevo Clinical Center, Bosnia and Herzegovina

**Margarida Bernardo**, Wayne State University School of Medicine-Karmanos Cancer Institute, Detroit, United States

**Semir Beslija**, University of Sarajevo Clinical Center, Sarajevo, Bosnia and Herzegovina

**Vanesa Bijol**, Northwell Health Physician Partners, New York, United States

**R. Daniel Bonfil**, Nova Southeastern University, Fort Lauderdale, FL, United States

**Mile Bosilkovski**, Clinic for Infectious Diseases and Febrile Conditions, Medical Faculty, Skopje, North Macedonia

**Giuseppe Broggi**, Department of Anatomic Pathology, University of Catania, Catania, Italy

**Juei-Tang Cheng**, Institute of Medical Research Chang Jung Christian University Tainan City, Taiwan, Province of China

**Adela Cemic**, Columbia University Irving Medical Center, New York, NY, USA

**Stefan Dojcinov**, Swansea University, United Kingdom

**Snježana Dotlić**, Clinical Hospital Center Rebro, Zagreb, Croatia

**Amel Dudakovic**, Mayo Clinic, Rochester, MN, United States

**Ammad Ahmad Farooqi**, Institute of Biomedical and Genetic Engineering (IBGE), Islamabad, Pakistan

**Ognjen Gajic**, Mayo Clinic, Rochester, Minnesota, United States

**Zoran Gatalica**, The University of Oklahoma Health Science Center, Oklahoma City, OK, USA

**Figen Govsa**, Department of Anatomy, Faculty of Medicine, Ege University, Izmir, Turkey

**Florian Grebien**, Ludwig Boltzmann Institute Cancer Research, Vienna, Austria

**Nermin Halkić**, Department of Visceral Surgery, Hospitalier Universitaire Vaudois, Lausanne, Switzerland

**Miroslav Harjacek**, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain, UAE

**Adnan Hasanovic**, Lenox Hill Hospital, New York, United States

**Stipan Jonjic**, Center for Proteomics, Faculty of Medicine, University of Rijeka, Croatia

**David Kachlik**, Department of Anatomy, Second and Third Faculty of Medicine, Charles University, Prague, Czechia

**Hulya Karatas-Kursun**, Institute of Neurological Sciences and Psychiatry, Hacettepe University, Ankara, Turkey

**Orsolya Kiss**, Semmelweis University, Budapest, Hungary

**Antonio Klasan**, Department for Orthopaedics and Traumatology, Kepler University Hospital Linz, Austria

**Senada Koljenovic**, Erasmus MC, Rotterdam, Netherlands

**Shuncong Wang**, Department of Imaging & Pathology, KU Leuven, Belgium

**Božo Krušlin**, Sestre milosrdnice University Hospital, Zagreb, Croatia

**Janina Kulka**, Semmelweis University, Budapest, Hungary

**Helena Kuivaniemi**, Faculty of Medicine and Health Sciences, Stellenbosch University, Tygerberg, South Africa

**Li Li**, School of Biomedical Engineering, Shanghai Jiao Tong University (SJTU), Shanghai, China  
**José I López**, Department of Pathology, Cruces University Hospital, University of the Basque Country, Bilbao, Spain  
**Jianfeng Lu**, School of Life Science and Technology at Tongji University, Shanghai, China  
**Boštjan Luzar**, Medical Faculty, University of Ljubljana, Ljubljana, Slovenia  
**Hideki Makishima**, Taussig Cancer Center, Cleveland Clinic, Cleveland, OH, United States  
**Sanjiv Manek**, Department of Cellular Pathology, John Radcliffe Hospital, Oxford, United Kingdom  
**Zaher K Otrock**, Washington University School of Medicine, St. Louis, MO, United States  
**Semir Ozdemir**, Department of Biophysics, Akdeniz University Medical Faculty, Antalya, Turkey  
**Mirza Pojskic**, Department of Neurosurgery, University Marburg, Germany  
**Yitao Qi**, Department of Cell Biology, College of Life Sciences, Shaanxi Normal University, China  
**Nermin N Salkic**, Department of Gastroenterology University Clinical Center Tuzla, Bosnia and Herzegovina  
**Mukesh Kumar Sriwastva**, University of Louisville, USA  
**Maja Šegvić Klarić**, Faculty of Pharmacy and Biochemistry, University of Zagreb, Croatia  
**Nirav R. Shah**, Carolinas Healthcare Systems, United States  
**Neil Shepherd**, Cheltenham General Hospital, Gloucester, United Kingdom  
**Zdenko Sonicki**, Andrija Stampar School of Public Health, University of Zagreb, Zagreb, Croatia  
**Valter Stemberga**, Department of Forensic Medicine, School of Medicine, Rijeka, Croatia  
**Mirza Suljagic**, International University of Sarajevo, Sarajevo, Bosnia and Herzegovina  
**Luca Testarelli**, Sapienza University of Rome, Rome, Italy  
**Kiril Trpkov**, Department of Pathology and Laboratory Medicine, University of Calgary, Calgary, Canada  
**Samra Turajlic**, Institute of Cancer Research, London, United Kingdom  
**Lejla Vajzovic**, Duke Eye Center, Durham, NC, United States  
**Tomaz Velnar**, University Medical Centre Maribor, Department of Neurosurgery, Slovenia  
**Edina Vranić**, University of Sarajevo Faculty of Pharmacy, Bosnia and Herzegovina  
**Gordan Vujanic**, Sidra Medical and Research Center, Sidra, Qatar  
**Enver Zerem**, Academy of Sciences and Arts of Bosnia and Herzegovina, Sarajevo, Bosnia and Herzegovina  
**Matjaž Zwitter**, Institute of Oncology, Ljubljana, Slovenia  
**Hongda Liu**, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China  
**Jinhui Liu**, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China

## Honorary Editorial Board

**Ondrej Hes**, Charles University in Prague, Faculty of Medicine in Plzen, Czechia  
**Muhidin Hamamdžić**, Academy of Sciences and Arts of Bosnia and Herzegovina, Bosnia and Herzegovina

## Founding Editor

Muzafer Mujić

## Previous Editors

Irfan Zulić (1998-2005)

[Muzafer Mujić](#) (2006-2008)

Denis Selimović (2009-2010)

Bakir Mehić (2011-2014)

## Production Editor

**Bakir Kudic**, Department of Pharmacology and Toxicology, Faculty of Medicine, University of Sarajevo, Bosnia and Herzegovina

## Editorial Assistants

**Ema Avdic**, Faculty of Medicine, University of Sarajevo, Bosnia and Herzegovina

**Nudžejma Dulic**, Faculty of Food Science and Agriculture, University of Sarajevo, Bosnia and Herzegovina

**Merima Bukva**, Institute of Pharmacology, Heidelberg University, Germany

**Nejra Selak**, Faculty of Medicine, University of Sarajevo, Bosnia and Herzegovina









**Ermina Vukalic**, Association of Basic Medical Sciences, Bosnia and Herzegovina

**Lamija Mlaco**, Department of Human Physiology, Faculty of Medicine, University of Sarajevo, Bosnia and Herzegovina

**Selma Susic**, Roanoke College, Salem, Virginia, USA

RESEARCH ARTICLE

# Cytotoxic T-lymphocyte-associated protein 4 +49A/G polymorphism in Down syndrome children with Hashimoto's thyroiditis

Muhammad Faizi <sup>1</sup>, Nur Rochmah <sup>1</sup>, Yuni Hisbiyah <sup>1</sup>, Rayi Kurnia Perwitasari <sup>1</sup>, Qurrota Ayuni Novia Putri <sup>2</sup>, Sukmawati Basuki <sup>3</sup>, Anang Endaryanto <sup>1</sup>, and Soetjipto Soetjipto <sup>4\*</sup>

Thyroid dysfunction is the most common endocrine disorder in Down syndrome (DS) children. Cytotoxic T-lymphocyte-associated antigen 4 (*CTLA-4*) is one of the immune regulatory genes that correlates with Hashimoto's thyroiditis (HT). However, studies on *CTLA-4* +49A/G in DS children with HT are still limited. We aimed to evaluate *CTLA-4* +49A/G gene polymorphism in DS children with HT. This case-control study, conducted from February 2020 to February 2022 at Dr. Soetomo General Hospital, Surabaya, enrolled 40 DS children with HT and 50 healthy children. The DNA sequencing was performed to identify the polymorphism (Sanger sequencing). Thyroid peroxidase antibodies (TPOAb), thyroglobulin antibodies (TgAb), thyroid-stimulating hormone (TSH), and free thyroxine (FT4) levels were analyzed by enzyme-linked immunosorbent assay (ELISA). The mean age of DS children with HT was 1.78 years. Males predominated in the study population. Subjects with GG genotype were diagnosed earliest with hypothyroidism (8 months) compared with other studies. The most common thyroid dysfunction was central hypothyroidism, with TgAb positivity present in all patients. The AA genotype (odds ratio [OR] 0.265, 95% confidence interval [CI] 0.094–0.746;  $P = 0.012$ ) and A allele (OR 0.472, 95% CI 0.309–0.721;  $P = 0.0002$ ) were significantly more frequent in the control group. The AG genotype (OR 2.65, 95% CI 0.094–0.746;  $P = 0.003$ ) and G allele (OR 2.116, 95% CI 1.386–3.23;  $P = 0.003$ ) were more frequent in the DS with HT group. The age of the subjects in this study was younger than in previous studies. The AG genotype and the G allele were more prevalent in the DS with HT group and may be a risk factor in HT development in DS children. Furthermore, the AA genotype may act as a protective factor against HT in DS children.

**Keywords:** Down syndrome (DS), Hashimoto's thyroiditis (HT), cytotoxic T-lymphocyte-associated protein 4 +49A/G (*CTLA-4* +49A/G).

## Introduction

Down syndrome (DS) is the most common chromosomal abnormality, occurring in 1 in over 800 births. It is mainly caused by the nondisjunction of chromosome 21, whereas the other cases are due to translocation or mosaicism [1–3]. According to the World Health Organization (WHO) report in 2018, approximately 3000–5000 children are born with chromosomal abnormalities each year [4]. There is an increased prevalence of DS in children from Indonesia, from 0.12% in 2010 to 0.21% in 2018, based on data obtained from the Ministry of Health in 2018 [5].

DS is associated with many abnormalities, such as pulmonary, gastrointestinal, cardiovascular, and endocrine problems, as well as developmental delay. Thyroid dysfunction is the most frequent endocrine disorder (4%–8%) in DS children [6]. Congenital hypothyroidism, subclinical hypothyroidism, hyperthyroidism, and acquired hypothyroidism (autoimmune and non-autoimmune) commonly occur in these children [7].

The prevalence of autoimmune thyroiditis increases in childhood due to genetic and environmental causes. Genetic factors are divided into two groups according to their functions: thyroid-specific genes and immune regulatory genes [8]. Cytotoxic T-lymphocyte-associated antigen 4 (*CTLA-4*) is one of the immune regulatory genes that are correlated with HT. Human *CTLA-4* is among the most researched loci (2q33) in terms of autoimmunity and its role in the development of Graves' disease (GD) and/or Hashimoto's thyroiditis (HT). *CTLA-4* is an immunoregulatory molecule found on the surface of activated T-cells that inhibits T-cell activation by interacting with the B7 molecule [9]. *CTLA-4* single nucleotide polymorphism is correlated to the T-effector activity [10]. In particular, *CTLA-4* adenine–guanine polymorphism exon 1 position 49 (a peptide with a Thr/Ala exchange) single nucleotide polymorphism leads to the development of a faulty receptor, which reduces the inhibitory effect of *CTLA-4* on

<sup>1</sup>Faculty of Medicine, Department of Child Health, Dr. Soetomo General Hospital, Universitas Airlangga, Surabaya, East Java, Indonesia; <sup>2</sup>Faculty of Medicine, Universitas Airlangga, Surabaya, East Java, Indonesia; <sup>3</sup>Institute of Tropical Disease, Universitas Airlangga, Surabaya, East Java, Indonesia; <sup>4</sup>Department of Medical Biochemistry, Faculty of Medicine, Universitas Airlangga, Surabaya, East Java, Indonesia.

\*Correspondence to Soetjipto Soetjipto: [Soetjipto1950@gmail.com](mailto:Soetjipto1950@gmail.com)

DOI: 10.17305/bb.2022.7869

© 2023 Faizi et al. This article is available under a Creative Commons License (Attribution 4.0 International, as described at <https://creativecommons.org/licenses/by/4.0/>).

T-cell activation and the important candidate for autoimmune thyroid diseases [11, 12, 52]. Antiperoxidase antibodies (TPOAb) and/or antithyroglobulin antibodies (TgAb) are the thyroid-specific antibodies observed in HT. The association between cytotoxic T-lymphocyte-associated protein 4 +49A/G (*CTLA-4* +49A/G) and HT has frequently been studied, whereas research on *CTLA-4* +49A/G in DS children with HT is still limited. This study aimed to evaluate the *CTLA-4* +49A/G in DS children with HT.

## Materials and methods

### Study design

Forty DS children with HT who visited the Pediatric Endocrine Clinic at Dr. Soetomo General Hospital Surabaya from February 2020 to February 2022 and met the inclusion and exclusion criteria were enrolled in this case-control study. As a control group, 50 healthy children without a history of DS and/or HT were included. The consecutive sampling was used for selecting the study participants. The inclusion criteria were as follows: (a) DS children with HT (proven by the positivity of TPOAb and/or TgAb) aged 1 month to 18 years treated as outpatients at Dr. Soetomo General Hospital Surabaya; (b) DS confirmed by a karyotyping test; and (c) the patients' parents signed the informed consent to study participation. DS children with HT who required Pediatric Intensive Care Unit care were excluded.

### DNA extraction and genotyping

The *CTLA-4* +49A/G (rs231775) was identified by DNA sequencing using the primer forward for *CTLA-4* +49A/G (rs231775) 5'-GCTCAGCTGAACCTGGCT-3', and primer reverse 5'-AAATCACTGCCCTTGACTGC-3'. DNA was extracted from peripheral blood mononuclear cells by using the QIAamp mini Kit as instruction (Qiagen®, Germany). In total, the reaction volume of 10 µL (a primer set of *CTLA-4* +49A/G, Go Tag DNA polymerase [Promega, USA], 25 mM MgCl<sub>2</sub>, H<sub>2</sub>O, and 2 µL template DNA) was amplified with the initial denaturation at 92 °C for 3 min, then followed by 92 °C for 45 s, 58 °C for 1 min, 70 °C for 1 min (35 cycles) with the final extension at 70 °C for 3 min in the PCR machine (Labycler Sensequest Biomedizinische, Germany). The PCR product was evaluated using agarose gel 2% that runs in the electrophoretic field and resulted in a single band at 165 bp. The PCR product was purified with ExoSAP-IT (USB, USA). PCR sequencing was performed on 2 µL purified amplicons with a dye terminator (BigDye™ Terminator v3.1, Applied Biosystems, USA) and forward or reverse primer of *CTLA-4* +49A/G (rs231775). The PCR sequencing product was precipitated by ethanol solution, then dissolved in H<sub>2</sub>O and placed into a well plate. The filled well plate was put into the sequencer machine (ABI/Hitachi 3500XL, USA). This DNA sequencing was applied at the Institute of Tropical Diseases, Universitas Airlangga, Surabaya, Indonesia. The *CTLA-4* +49A/G (rs231775) polymorphism was examined by using FinchTV (Geospiza Inc, USA) and NCBI softwares.

### Thyroid function and antibody markers

Thyroid functions (free thyroxine [FT4], thyroid-stimulating hormone [TSH], TPOAb, and TgAb) were assessed using blood

tests. The thyroid antibodies of HT were examined at the Institute of the Tropical Disease University of Airlangga, Surabaya. HT was postulated by the positivity of TPOAb and/or TgAb. The TPOAb values were classified as follows using a Demeditec kit (Demeditec Diagnostics GmbH, catalog number: DE7580): negative (<50 IU/mL); borderline (50–75 IU/mL), which was considered positive in this study; or positive (>75 IU/mL). The TgAb values were classified as follows using a Demeditec kit (Demeditec Diagnostics GmbH, catalog number: DE7590): negative (<100 IU/mL), borderline (100–150 IU/mL), which was considered positive in this study; or positive (≥150 IU/mL).

The thyroid function test results were analyzed at the Diagnostic Center of Soetomo General Hospital, Surabaya. The catalogue number for FT4 ELISA kit (Biochem Canada) was 'CAN-FT4-4340' and TSH Triiodothyronine was 'CAN-TSH-4080'. The reference range values for pediatric care by Sperling et al. (2020) [13] were used to determine the normal range of TSH and FT4. Central hypothyroidism was defined by the low TSH and FT4 levels, primary hypothyroidism by the high TSH and low FT4 levels, subclinical hypothyroidism by the elevated serum TSH levels with normal FT4 levels, and euthyroidism by the normal FT4 and TSH levels [7].

### Ethical statement

This study was approved by Dr. Soetomo General Hospital Ethics Committee (Ref. No. 1960/KEKP/IV/26020). The authors' institutional review board or comparable organization approved the present research for human subjects, which followed all relevant national rules, institutional procedures, and the precepts of the Helsinki Declaration.

### Statistical analysis

The SPSS 17 software was used to analyze all data. A descriptive analysis was performed on the distribution of the characteristics of participants and *CTLA-4* +49A/G (rs231775) polymorphism. The comparison of genetic polymorphism and allele distribution between DS patients with HT and the control group, thyroid dysfunction, TPO-Ab, and genetic polymorphism was analyzed with a chi-square test. Age at diagnosis of hypothyroidism and duration of therapy with levothyroxine compared with *CTLA-4* +49A/G (rs231775) polymorphism were analyzed with nonparametric Kruskal–Wallis H test. The regression test was analyzed with the logistic regression test. The significant result was defined as  $P < 0.05$ . The data are shown as mean and percentage (%).  $P$  value of Hardy–Weinberg Equilibrium (HWE) was also calculated.

## Results

### Characteristics of the study

The characteristics of the study participants are presented in Table 1. Forty DS children with HT and 50 healthy children voluntarily participated in this study. DS group with HT consisted predominantly of male participants, whereas the equal proportion of male and female participants was in the control group. Forty-five percent (18/40) of the DS children with HT had another congenital anomaly, such as congenital heart disease. FT4 and TSH levels in DS children with HT were

**Table 1.** Characteristics of the study participants

Characteristics	DS with HT (n = 40)	Control (n = 50)
Age (years), mean ± SD	1.78 ± 1.5	3.2 ± 2.27
Sex, n (%)		
Male	26 (65)	26 (52)
Female	14 (35)	24 (48)
Thyroid function test, n (%)		
Central hypothyroidism	31 (77.5)	–
Primary hypothyroidism	7 (14.6)	–
Subclinical hypothyroidism	2 (4.2)	–
Value, n (%)		
Positive for TgAb	40 (100)	–
Positive for both TPOAb and TgAb	26 (65)	–
Cumulative positive for autoimmune marker	40 (100)	–
Value, mean ± SD		
FT4 (ng/mL)	0.8 ± 0.5	–
TSH (μU/mL)	1.2 ± 6.6	–

Data presented as mean ± standard deviation (SD) or n (%). DS: Down syndrome; HT: Hashimoto’s thyroiditis; TPOAb: Thyroid peroxidase antibodies; TgAb: Thyroglobulin antibodies; FT4: Free thyroxine; TSH: Thyroid-stimulating hormone.

0.8 ± 0.5 ng/mL and 1.2 ± 6.6 μU/mL, respectively. All DS children with HT showed TgAb positivity.

#### CTLA-4 + 49A/G (rs231775) gene polymorphisms in DS children with HT

Three kinds of genotypes were noted in the study participants. The details of CTLA-4 + 49A/G (rs231775) gene polymorphism in DS patients with HT and controls are described in Table 3. The AG genotype was the most common in DS children with HT. DS participants with the AG genotype had a 1.688 times higher risk of developing HT than those with other genotypes, although the results did not show statistical significance (odds ratio [OR] 1.688, 95% confidence interval [CI] 0.730–3.905; *P* = 0.221). The AA genotype was the most common in the control group. The AA genotype had a protective effect on DS children with HT 0.265 times compared and showed statistical significance (OR 0.265, 95% CI 0.094–0.746; *P* = 0.012). The G allele had increased risk for DS with HT (OR 2.116, 95% CI 1.386–3.23; *P* = 0.003). Furthermore, the A allele had protective effect (OR 0.472, 95% CI 0.309–0.721; *P* = 0.0002) (Table 2). Distribution consistent to Hardy–Weinberg’s law at level of significance 0.05.

#### The correlation between thyroid function diagnoses and CTLA-4 + 49A/G (rs231775) gene polymorphism in DS children with HT

This study showed that AG was the most frequent genotype found. All subjects having an AG genotype were commonly diagnosed with central hypothyroidism (77.3%), followed by primary (18.2%) and subclinical hypothyroidism (4.6%) even though not significantly correlated (Table 3).

#### The correlation between TPOAb and CTLA-4 + 49A/G (rs231775) gene polymorphism in DS children with HT

The AG genotype was the most common genotype, and subjects with the AG genotype frequently had positive TPOAb. Also,

**Table 2.** CTLA-4 + 49A/G gene polymorphism in DS children with HT at Dr. Soetomo General Hospital

Variables	DS with HT (n = 40)	Control (n = 50)	<i>P</i> value	OR (95% CI)
Genotypes, n (%)			0.031	
AA	6 (15)	20 (40)	<b>0.012*</b>	0.265 (0.094–0.746)
AG	22 (55)	21 (42)	0.221	1.688 (0.730–3.905)
GG	12 (30)	9 (18)	0.185	1.952 (0.726–5.248)
Allele, n (%)				
A	68 (42.5)	122 (61)	<b>0.0002*</b>	0.472 (0.309–0.721)
G	92 (57.5)	78 (39)	<b>0.0003*</b>	2.116 (1.386–3.23)

Data presented as n (%). Logistic regression test, \**P* < 0.05 considered significant. DS: Down syndrome; HT: Hashimoto’s thyroiditis; OR: Odds ratio; CI: Confidence interval; CTLA-4: Cytotoxic T-lymphocyte-associated protein 4.

**Table 3.** Correlation between thyroid dysfunction diagnoses and CTLA-4 + 49 A/G in DS children with HT

Genotype	Central hypothyroidism	Primary hypothyroidism	Subclinical hypothyroidism	<i>P</i> value
AA, n (%)	4 (66.7)	2 (33.3)	0 (0)	0.667 <sup>a</sup>
AG, n (%)	17 (77.3)	4 (18.2)	1 (4.6)	
GG, n (%)	10 (83.2)	1 (8.4)	1 (8.4)	

Data presented as n (%). <sup>a</sup>Chi-square, *P* < 0.05 considered significant. DS: Down syndrome; HT: Hashimoto’s thyroiditis; CTLA-4: Cytotoxic T-lymphocyte-associated protein 4.

**Table 4.** The correlation between TPOAb and CTLA-4 + 49 A/G in DS children with HT

Genotype	TPOAb		<i>P</i> value
	Positive	Negative	
AA, n (%)	4 (66)	2 (34)	0.45 <sup>a</sup>
AG, n (%)	16 (73)	6 (27)	
GG, n (%)	6 (50)	6 (50)	

Data presented as n (%). <sup>a</sup>Chi-square test, *P* < 0.05 considered significant. DS: Down syndrome; HT: Hashimoto’s thyroiditis; TPOAb: Thyroid peroxidase antibodies; CTLA-4: Cytotoxic T-lymphocyte-associated protein 4.

the positivity of TPOAb showed dominance in other genotypes, even though TPOAb and CTLA-4 + 49A/G were not significantly correlated (*P* = 0.45). Otherwise, the correlation between TGAb and CTLA-4 + 49A/G gene polymorphism could not be done due to the constant result of TGAb positivity (Table 4).

#### Age at diagnosis of hypothyroidism and duration of therapy with levothyroxine compared with CTLA-4 + 49A/G polymorphism

There was no significant effect of age at diagnosis of hypothyroidism (*P* = 0.46), and duration of therapy with levothyroxine (*P* = 0.25) compared with CTLA-4 + 49A/G polymorphism (Table 5). Children with the GG genotype were diagnosed with hypothyroidism earlier than those with other genotypes.

**Table 5.** Age at diagnosis of hypothyroidism and duration of therapy with levothyroxine compared with *CTLA-4* +49A/G polymorphism

Genotype	Age at diagnosis of hypothyroidism (months)	<i>P</i> value	Duration of therapy with levothyroxine (months)	<i>P</i> value
AA, mean ± SD	18 ± 19.8	0.46*	6.7 ± 14.4	0.25*
AG, mean ± SD	11.4 ± 11.4		7.1 ± 14.8	
GG, mean ± SD	7.8 ± 7.7		17.3 ± 18.5	

Data presented as months, mean ± standard deviation (SD). \*Kruskall-Wallis H test,  $P < 0.05$  considered significant. *CTLA-4*: Cytotoxic T-lymphocyte-associated protein 4.

## Discussion

To the best of our knowledge, this is the first study that explored the *CTLA-4* +49A/G polymorphism in DS children with HT, as the studies on the association between *CTLA-4* and HT are limited. We showed that the *CTLA-4* +49A/G gene polymorphism in DS children with HT was dominated by the AG genotype (55%), although not significantly differed from the control group ( $P = 0.221$ ). Similar studies involving adult patients with HT but without DS in Poland and Slovakia (Caucasians), Turkey (Mixed), Japan and India (Asians) revealed that the most common genotype found in these patients was AG [15–22]. However, studies conducted in Thailand and Lebanon revealed that the most common genotype was AA [23, 24]. Meanwhile, Bicek et al. reported that GG is the most common genotype found in GD patients from Slovenia [25].

The AG genotype and the G allele had 1.688- and 2.116-fold higher risks, respectively, of developing DS with HT compared with other genotypes, similar to the results of some European and Asian studies [20, 22, 26, 27]. *CTLA-4* +49A/G increased the risk of HT among non-DS adults, particularly those with the AG genotype and G allele. The G49 allele was associated with HT, as confirmed by the higher thyroid autoantibody concentrations, including TgAb and TPOAb [28, 29]. According to previous studies, the G allele is associated with reduced control of T-cell proliferation, thereby contributing to the pathophysiology of autoimmune hypothyroidism, GD, and other autoimmune diseases [11].

Regarding chromosome 2q33, a susceptibility locus for thyroid autoantibody production was discovered around the marker D2S155, which is linked to the *CTLA4* and *CD28* genes [28]. Further research revealed that a significant gene for thyroid autoantibody production on chromosome 2q33 is most likely the *CTLA4* gene rather than the *CD28* gene [28].

Yung et al. reported that the G allele predominates in the Asian population compared to other ethnicities, whereas the A allele is commonly found in whites; moreover, the individuals with the AG genotype were reported to be more susceptible to developing HT [27, 30]. However, the patients in most case-control studies were ethnically matched, and both whites and Asians were found to have a link to the disease. As a result, ethnic and regional heterogeneities may only play a minor role in explaining discrepancies among the association studies [26]. Conflicting studies found no linkage between the *CTLA-4* gene and HT in Japan, Tunisia, and another Chinese pedigree [26, 31–33].

The AA genotype was significantly more prevalent in the control group in our study. The AA genotype as well as the A allele were significant protective factors against HT development. These results are consistent with those of European and Asian population-based studies [20, 26, 27].

The rate of GG genotypes in this study was not different between the DS patients with HT and controls. This result is contrary to Pastuszak et al.'s study involving GD patients from Poland and Patel et al.'s study involving non-DS patients from India; they reported that the GG genotype increased the risk for HT development in these patient groups [20, 22].

Congenital alteration in thyroid gland regulation among DS children is directly related to the trisomy of chromosome 21. The present study showed that central hypothyroidism was the most common diagnosis among DS children. This finding is different from those of other studies. Subclinical hypothyroidism was the most common thyroid dysfunction in DS children with HT, which is commonly asymptomatic and may not require treatment. However, subclinical hypothyroidism can progress to primary hypothyroidism associated with autoimmune factors [34–47]. TgAb positivity was more prevalent compared to TPOAb positivity, as observed in our study. A similar study by Gentile et al. demonstrated that TgAb was also more prevalent in HT patients [48]. Contrarily, Bjoro et al.'s study showed that TPOAb positivity was more frequently observed among HT patients, and it usually appears before thyroid dysfunction occurs, as compared to TgAb [48]. Thyroid autoimmunity affects DS children from an early age, but another study reported that thyroid autoantibodies start to appear at 5–8 years of age [34], indicating that autoimmune regression increases with age. Genetics has become a factor in autoimmune disorders, although the underlying reasons remain unknown [8].

The mean age of participants in this study was 1.78 years. Patients with GG genotype were diagnosed with hypothyroidism earliest compared with those with other genotypes and were younger compared with other studies, although there was no significant correlation. Studies by Aversa et al. stated that DS children develop thyroid dysfunction at 6.5 years, i.e., at a younger age than non-DS children, who develop thyroid dysfunction at 11 years old [49]. Similar to Sanyal and Bhat-tacharjee, the median age of DS children with hypothyroidism is younger than that non-DS children [50].

In our study population, the proportion of boys was greater than that of girls, which was similar to the findings of Musdalipa et al. and Kawanto et al. [46, 50]. However, Guaraldi



et al. pointed out that there is no difference in the rate of HT between sexes [51].

In the present study, another congenital anomaly besides thyroid dysfunction, i.e., congenital heart disease, was present in 18 (45%) children with DS. A previous study showed that cardiovascular diseases are observed in 58% of DS children [51].

The major limitation of our study is that it was performed in only one health center and with limited sample size. We suggest performing the multicenter study on this topic.

## Conclusion

Central hypothyroidism is the most common thyroid dysfunction observed among DS children with HT. AG genotype and G allele of *CTLA-4* +49A/G gene polymorphism were frequently found in the patients' group; these may be risk factors for the susceptibility to HT among DS children. Moreover, the AA genotype may play as a protective factor against HT in DS children.

## Acknowledgments

The authors would like to thank the patients, their families, the Universitas Airlangga Institute of Tropical Disease team, and the endocrine staffs of the Faculty of Medicine, Universitas Airlangga Dr. Soetomo General Hospital in Surabaya, Indonesia.

**Conflicts of interest:** Authors declare no conflicts of interest.

**Funding:** Authors received no specific funding for this work.

Submitted: 20 July 2022

Accepted: 06 January 2023

Published online: 30 January 2023

## References

- Wayne H. Examination paediatrics: a guide to paediatric training. 5<sup>th</sup> ed. Chatswood (Australia): Elsevier; 2019. p. 336-348.
- Real de Asua D, Quero M, Moldenhauer F, Suarez C. Clinical profile and main comorbidities of Spanish adults with Down syndrome. *Eur J Intern Med* 2015;26:385-91. <https://doi.org/10.1016/j.ejim.2015.05.003>.
- Weijerman ME, de Winter JP. Clinical practice. The care of children with Down syndrome. *Eur J Pediatr* 2010;169:1445-52. <https://doi.org/10.1007/s00431-010-1253-0>.
- World Health Organization Regional Office for Europe (2018). Births with Down's syndrome per 100 000 live births. 2018. Available from: [https://gateway.euro.who.int/en/indicators/hfa\\_603-7120-births-with-downs-syndrome-per-100-000-live-births](https://gateway.euro.who.int/en/indicators/hfa_603-7120-births-with-downs-syndrome-per-100-000-live-births)
- Kemenkes RI. Infodatin (Pusat data dan informasi kementerian kesehatan RI). Kementerian Kesehatan Republik Indonesia; 2018.
- Bull ML. Committee on Genetics. Health supervision for children with Down syndrome. *Pediatrics* 2011;128:393-406. <https://doi.org/10.1542/peds.2011-1605>.
- Pierce MJ, LaFranchi SH, Pinter JD. Characterization of thyroid abnormalities in a large cohort of children with Down syndrome. *Horm Res Paediatr* 2017;87:170-8. <https://doi.org/10.1159/000457952>.
- Iughetti L, Lucaccioni L, Fugetto F, Mason A, Barbara P. Thyroid function in Down syndrome. *Expert Rev Endocrinol Metab* 2015;10:525-32. <https://doi.org/10.1586/17446651.2015.1063995>.
- Brunner MC, Chambers CA, Chan FK, Hanke J, Winoto A, Allison JP. *CTLA-4*-mediated inhibition of early events of T cell proliferation. *J Immunol* 1999;162:5813-20. <https://doi.org/10.4049/jimmunol.162.10.5813>.
- Manzotti CN, Tipping H, Perry LC, Mead KI, Blair PJ, Zheng Y, et al. Inhibition of human T cell proliferation by *CTLA-4* utilizes CD80 and requires CD25<sup>+</sup> regulatory T cells. *Eur J Immunol* 2002;32:2888-96. [https://doi.org/10.1002/1521-4141\(200210\)32:10<2888::AID-IMMU2888>3.0.CO;2-F](https://doi.org/10.1002/1521-4141(200210)32:10<2888::AID-IMMU2888>3.0.CO;2-F).
- Kouki T, Sawai Y, Gardine CA, Fisfalen M-E, Alegre M-L, DeGroot LJ. *CTLA-4* gene polymorphism at position 49 in exon 1 reduces the inhibitory function of *CTLA-4* and contributes to the pathogenesis of Graves' disease. *J Immunol* 2000;165:6606-11. <https://doi.org/10.4049/jimmunol.165.11.6606>.
- Mäurer M, Loserth S, Kolb-Mäurer A, Ponath A, Wiese S, Kruse N, et al. A polymorphism in the human cytotoxic T-lymphocyte antigen 4 (*CTLA4*) gene (exon 1 +49) alters T-cell activation. *Immunogenetics* 2002;54:1-8. <https://doi.org/10.1007/s00251-002-0429-9>.
- Sperling, M. Pediatric endocrinology. Philadelphia (PA): ELSEVIER-HEALTH Science; 2020.
- Zemel BS, Pipan M, Stallings VA, Hall W, Schadt K, Freedman DS, et al. Growth charts for children with Down syndrome in the United States. *Pediatrics* 2015;136(5):e1204-11. <https://doi.org/10.1542/peds.2015-1652>.
- T. Dallos, M. Avbelj, E. Barak L, Zapletalová J, Pribilincová Z, Krajčířová M, et al. *CTLA-4* gene polymorphisms predispose to autoimmune endocrinopathies but not to celiac disease. *Neuro Endocrinol Lett* 2009;29(3):334-340.
- Kucharska AM, Gorska E, Wasik M, Pyrzak B, and Demkow U. Expression of CD152 (*CTLA-4*) in children with autoimmune thyroiditis and +49 A/G polymorphism of exon 1 of the *CTLA-4* gene. *J Physiol Pharmacol* 2009;60:77-80.
- Sahin M, Gursoy A, Erdogan MF. Cytotoxic T lymphocyte-associated molecule-4 polymorphism in Turkish patients with Hashimoto thyroiditis. *Int J Immunogenet* 2009;36(2):103-6. <https://doi.org/10.1111/j.1744-313X.2009.00831.x>.
- Guo Z, Chen X, Wu P, Wu G. Relationships between *CTLA-4* gene polymorphism and Graves' disease in Han population in Western region of Guangdong province. *J Guangdong Med Coll* 2010;28:1-3. <https://doi.org/10.1186/s12881-018-0665-y>.
- Veeramuthumari P, Isabel W, Kannan K. A study on the level of T(3), T(4), TSH and the association of A/G polymorphism with *CTLA-4* gene in graves' hyperthyroidism among south Indian population. *Indian J Clin Biochem* 2011;26(1):66-9. <https://doi.org/10.1007/s12291-010-0093-z>.
- Pastuszak-Lewandoska D, Sewerynek E, Domańska D, Gładys A, Skrzypczak R, Brzeźniańska E. *CTLA-4* gene polymorphism and their influence on predisposition to autoimmune thyroid diseases (Graves' disease and Hashimoto's thyroiditis). *Arch Med Sci* 2012;8(3):415-21. <https://doi.org/10.5114/aoms.2012.28593>.
- Tomoyose T, Komiya I, Takara M, Yabiku K, Kinjo Y, Shimajiri Y, et al. Cytotoxic T-lymphocyte antigen-4 gene polymorphisms and human T-cell lymphotropic virus-1 infection: their associations with Hashimoto's thyroiditis in Japanese patients. *Thyroid* 2002 Aug;12(8):673-7. <https://doi.org/10.1089/105072502760258640>.
- Patel H, Mansuri MS, Singh M, Begum R, Shastri M, Misra A. Association of cytotoxic T-lymphocyte antigen 4 (*CTLA4*) and thyroglobulin (TG) genetic variants with autoimmune hypothyroidism. *PLoS One* 2016;11(3):1-16. <https://doi.org/10.1371/journal.pone.0149441>.
- Kimkong I, Nakkuntod J, Sae-Ngow S, Snaboon T, Avhingsanon Y, Hirankarn N. Association between *CTLA-4* polymorphisms and the susceptibility to systemic lupus erythematosus and Graves' disease in Thai population. *Asian Pac J Allergy Immunol* 2011;29(3):229-35.
- Farra C, Awwad J, Fadlallah A, Sebaly G, Hage G, Souaid M, et al. Genetics of autoimmune thyroid disease in the Lebanese population. *J Community Genet* 2012;3(4):259-64. <https://doi.org/10.1007/s12687-012-0085-1>.
- Bicek A, Zaletel K, Gaberscek S, Pirnat E, Khirin B, Stopar TG et al. 49A/G and CT60 polymorphisms of the cytotoxic T-lymphocyte-associated antigen 4 gene associated with autoimmune thyroid disease. *Human Immunol* 2009;70(10):820-4. <https://doi.org/10.1016/j.humimm.2009.06.016>.
- Chistiakov DA, Turakulov RI. *CTLA-4* and its role in autoimmune thyroid disease. *J Mol Endocrinol* 2003;31:21-36. <https://doi.org/10.1677/jme.0.0310021>.
- Kavvoura FK, Akamizu T, Awata T, Ban Y, Chistiakov DA, Frydecka I, et al. Cytotoxic T-lymphocyte associated antigen 4 gene polymorphisms and autoimmune thyroid disease: a meta-analysis. *J Clin*

- Endocrinol Metab 2007;92:3162–70. <https://doi.org/10.1210/jc.2007-0147>.
- [28] Tomer Y, Greenberg DA, Barbesino G, Concepcion E, Davies TF. CTLA-4 and not CD28 is a susceptibility gene for thyroid autoantibody production. *J Clin Endocrinol Metab* 2001;86(4):1687–93. <https://doi.org/10.1210/jc.86.4.1687>.
- [29] Zaletel K, Krhin B, Gaberscek S, Pirnat E, Hojker S. The influence of the exon 1 polymorphism of the cytotoxic T lymphocyte antigen 4 gene on thyroid antibody production in patients with newly diagnosed Graves' disease. *Thyroid* 2002;12:373–6. <https://doi.org/10.1210/jcem.86.4.7372>.
- [30] Yung E, Cheng PS, Fok TF, Wong GWK. CTLA-4 gene A-G polymorphism and childhood Graves' disease. *Clin Endocrinol* 2002;56(5):649–53. <https://doi.org/10.1046/j.1365-2265.2002.01234.x>.
- [31] Akamizu T, Sale MM, Rich SS, Hiratani H, Noh JY, Kanamoto N, et al. Association of autoimmune thyroid disease with microsatellite markers for the thyrotropin receptor gene and CTLA-4 in Japanese patients. *Thyroid* 2000;10(10):851–8. <https://doi.org/10.1089/thy.2000.10.851>.
- [32] Maalej A, Bougacha N, Rebai A, Bellassouad M, Ayadi-Makni F, Abid M, et al. Lack of linkage and association between autoimmune thyroid diseases and the CTLA-4 gene in a large Tunisian family. *Human Immunol* 2001;62(11):1245–50. [https://doi.org/10.1016/S0198-8859\(01\)00316-0](https://doi.org/10.1016/S0198-8859(01)00316-0).
- [33] Villanueva R, Tomer Y, Greenberg DA, Mao C, Concepcion ES, Tucci S, et al. Autoimmune thyroid disease susceptibility loci in a large Chinese family. *Clin Endocrinol* 2002;56(1):45–51. <https://doi.org/10.1046/j0300-0664.2001.01429.x>.
- [34] Karlsson B, Gustafsson J, Hedov G, Ivarsson S-A, Annerén G. Thyroid dysfunction in Down's syndrome: relation to age and thyroid autoimmunity. *Arch Dis Childhood* 1998;79(3):242–5. <https://doi.org/10.1136/adc.79.3.242>.
- [35] Popova G, Paterson WF, Brown A, Donaldson MDC. Hashimoto's thyroiditis in Down's syndrome: clinical presentation and evolution. *Hormone Res* 2008;70(5):278–84. <https://doi.org/10.1159/000157874>.
- [36] AlAaraj N, Soliman AT, Itani M, Khalil A, De Sanctis V. Prevalence of thyroid dysfunctions in infants and children with Down Syndrome (DS) and the effect of thyroxine treatment on linear growth and weight gain in treated subjects versus DS subjects with normal thyroid function: a controlled study. *Acta Biomed* 2019;90(8-S):36–42. <https://doi.org/10.23750/abm.v90i8-S.8503>.
- [37] Zelazowska-Rutkowska B, Jakubiuk-Tomaszuk, Cylwik B. Thyroid function in children with Down syndrome in the polish population: a case-control study. *Arch Iran Med* 2020;23(6):386–90. <https://doi.org/10.34172/aim.2020.31>.
- [38] Gentile F, Conte M, Formisano S. Thyroglobulin as an autoantigen: What can we learn about immunopathogenicity from the correlation of antigenic properties with protein structure? *Immunology* 2004;112(1):13–25. <https://doi.org/10.1111/j.1365-2567.2004.01861.x>.
- [39] Awata T, Kurihara S, Iitaka M, Takei S-I, Inoue I, Ishii C, et al. Association of CTLA-4 gene A-G polymorphism (*ITIM2 locus*) with acute-onset and insulin-depleted IDDM as well as autoimmune thyroid disease (Graves' disease and Hashimoto's thyroiditis) in the Japanese population. *Am Diabetes Assoc* 1998;47(1):128–9. <https://doi.org/10.2337/diab.47.1.128>.
- [40] Zimmermann MB, Moretti D, Chaouki N, Torresani T. Introduction of iodized salt to severely iodine-deficient children does not provoke thyroid autoimmunity: a one year prospective trial in northern Morocco. *Thyroid* 2003;13:199–203. <https://doi.org/10.1089/105072503321319512>.
- [41] Hashemipour M, Amini M, Aminorroaya A, Dastjerdi MA, Rezvanian H, Kachoei A, et al. High prevalence of goiter in an iodine replete area: do thyroid auto-antibodies play a role? *Asia Pac J Clin Nutr* 2007;16:403–10.
- [42] Zois C, Stavrou I, Kalogera C, Svarna E, Dimoliatis I, Seferiadis K, et al. High prevalence of autoimmune thyroiditis in schoolchildren after elimination of iodine deficiency in Northwestern Greece. *Thyroid* 2003;13:485–9. <https://doi.org/10.1089/105072503322021151>.
- [43] Vasheghani M, Jalali R, Dabbaghmanesh MD, Sadeghalvad AS, Omrani GR. Thyroid autoimmunity role in the evolution of endemic goiter in rural area, Fars, Iran. *Arch Iran Med* 2011;14:164–6.
- [44] Jakić J, Dumić M, Filipović B, Ille J, Cvijetić M, Gjurij G. Thyroid diseases in a school population with thyromegaly. *Arch Dis Child* 1994;70:103–6. <https://doi.org/10.1136/adc.70.2.103>.
- [45] Ghorashian SM, Moghaddam SHH, Afkhami-Ardekani M. Relationship between anti-thyroid peroxidase antibody and thyroid function test. *Iran J Immunol* 2006;3:146–149.
- [46] Musdalipa M, Wulandari R, Maddeppungeng M. Maternal age distribution of Down-syndrome at pediatric growth and development clinic, 2015–2019. *Green Med J* 2021;3(2):73–80. <https://doi.org/10.33096/gmj.v3i2.84>.
- [47] Guaraldi F, Giaccherion RR, Lanfranco F, Motta G, Gori D, Arvat E, et al. Endocrine autoimmunity in Down's syndrome. *Endocrine Immunol Kanger* 2017;48:133–46. <https://doi.org/10.1159/000452912>.
- [48] Bjoro T, Holmen J, Krüger O, Midthjell K, Hunstad K, Scheriner K, et al. Prevalence of thyroid disease, thyroid dysfunction and thyroid peroxidase antibodies in a large, unselected population. The health study of Nord Trøndelag (HUNT). *Eur J Endocrinol* 2000;143:639–47. <https://doi.org/10.1530/eje.0.1430639>.
- [49] Aversa T, Salerno M, Radetti G, Faienza MF, Iughetti L, Corrias A, et al. Peculiarities of presentation and evolution over time of Hashimoto's thyroiditis in children and adolescents with Down's syndrome. *Hormones* 2015;14(3):410–6. <https://doi.org/10.14310/horm.2002.1574>.
- [50] Sanyal D, Bhattacharjee K. High thyroid-stimulating hormone level in Down's syndrome: a mere resetting of hypothalamic-pituitary axis in subclinical hypothyroidism? *Indian J Endocrinol Metab* 2020;24(5):406–9. [https://doi.org/10.4103/ijem.IJEM\\_422\\_20](https://doi.org/10.4103/ijem.IJEM_422_20).
- [51] Kawanto FH, Soedjatmiko S, Hendarto A. Factors associated with intelligence in young children with Down syndrome. *Paediatrica Indonesiana* 2012;52(4):194. <https://doi.org/10.14238/pi52.4.2012.194-9>.
- [52] Vieland VJ, Huang Y, Bartlett C, Davies TF, Tomer Y. A multilocus model of the genetic architecture of autoimmune thyroid disorder, with clinical implications. *Am J Hum Genet* 2008 Jun;82(6):1349–56. <https://doi.org/10.1016/j.ajhg.2008.04.017>.

## Related articles published in BJBMS

1. Vascular endothelial growth factor (VEGF)-related polymorphisms rs10738760 and rs6921438 are not risk factors for proliferative diabetic retinopathy (PDR) in patients with type 2 diabetes mellitus (T2DM)

Jana Sajovic et al., BJBMS, 2019

2. The joint effect of the endothelin receptor B gene (EDNRB) polymorphism rs10507875 and nitric oxide synthase 3 gene (NOS3) polymorphism rs869109213 in Slovenian patients with type 2 diabetes mellitus and diabetic retinopathy

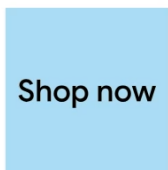
Editors' Choice. Dejan Bregar et al., BJBMS, 2018



Scimago Journal & Country Rank

Enter Journal Title, ISSN or Publisher Name

- Home
- Journal Rankings
- Country Rankings
- Viz Tools
- Help
- About Us



Same Day Delivery. T&Cs Apply  
HP Store Indonesia

-10%	-10%	-36%
-9%	-9%	-10%

# Bosnian Journal of Basic Medical Sciences

## COUNTRY

Bosnia and Herzegovina



Universities and research institutions in Bosnia and Herzegovina



Media Ranking in Bosnia and Herzegovina

## SUBJECT AREA AND CATEGORY

Medicine  
Medicine  
(miscellaneous)

## PUBLISHER

Association of Basic Medical Sciences  
Federation of Bosnia and Herzegovina

## H-INDEX

# 29

## PUBLICATION TYPE

Journals

## ISSN

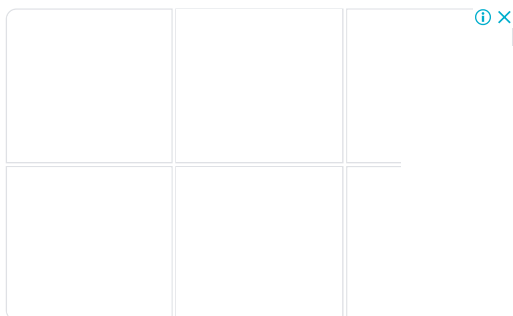
15128601,  
18404812

## COVERAGE

2004-2021

## INFORMATION

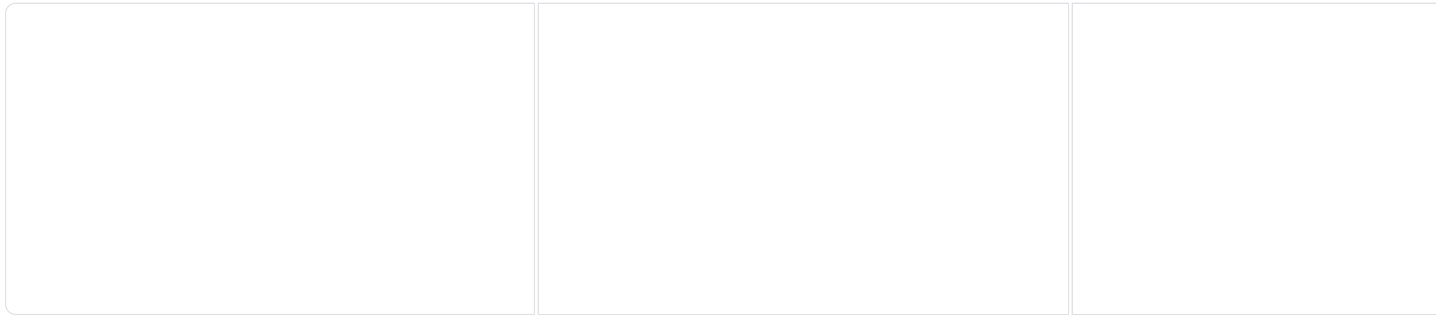
- [Homepage](#)
- [How to publish in this journal](#)
- [Contact](#)



Original HP Toner Cartridge  
HP Store Indonesia

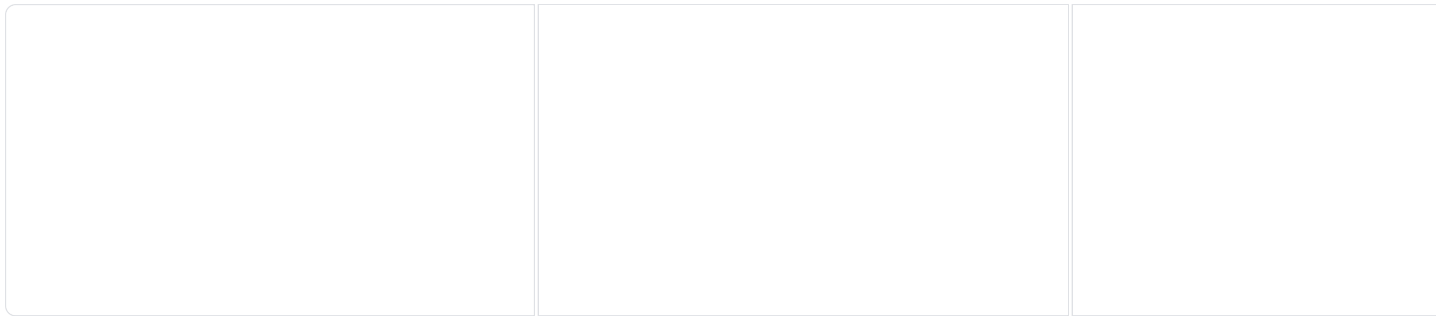
The Bosnian Journal of Basic Medical Sciences (BJBMS) is an international, English-language, peer reviewed journal, publishing original articles from different disciplines of basic medical sciences. BJBMS welcomes original research and comprehensive reviews as well as short research communications in the field of biochemistry, genetics, immunology, microbiology, pathology, pharmacology, pharmaceutical sciences and physiology.

Join the conversation about this journal



Original HP Toner Cartridge  
HP Store Indonesia

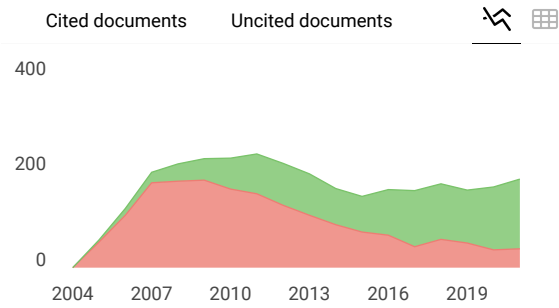
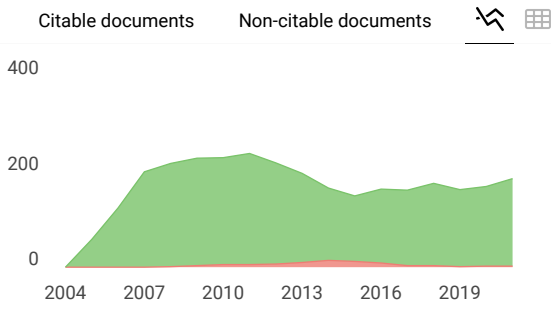
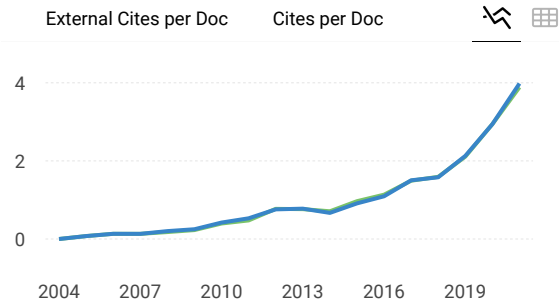
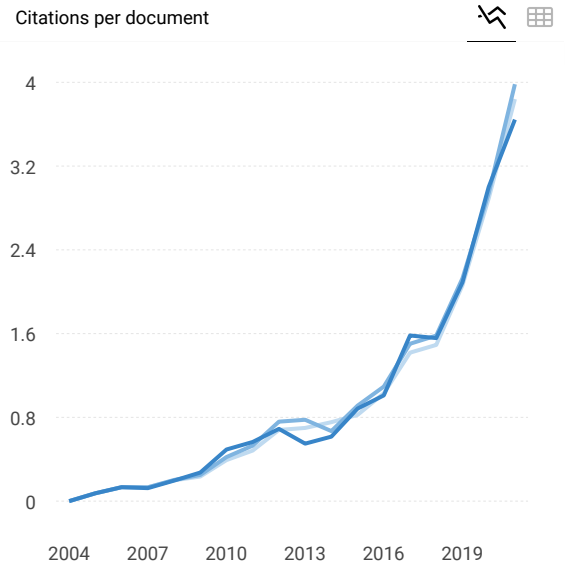
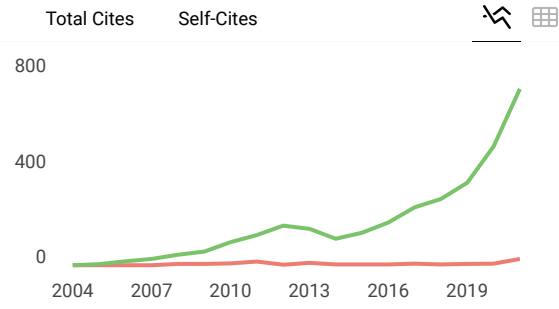
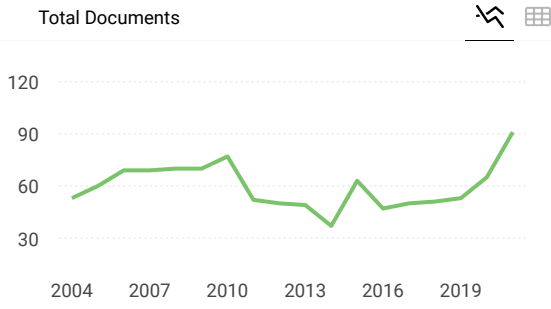
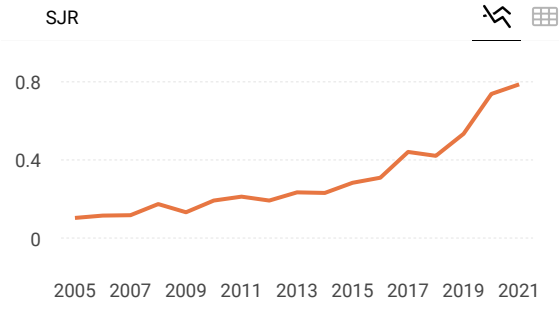
Quartiles



Original HP Toner Cartridge  
HP Store Indonesia

FIND SIMILAR JOURNALS ?

<p>1 <b>International Journal of Medical Sciences</b> AUS</p> <p>68%</p>	<p>2 <b>Experimental and Therapeutic Medicine</b> GRC</p> <p>68%</p>	<p>3 <b>Biomedical Reports</b> GRC</p> <p>66%</p>	<p>4 <b>Disease Ma</b> EGY</p> <p>6</p>
--	--	---	---



**Bosnian Journal of Basic Medical Sciences**

**Q2**    Medicine (miscellaneous)    best quartile

**SJR 2021**  
**0.79**

powered by scimagojr.com

← Show this widget in your own website

Just copy the code below and paste within your html code:

```
<a href="https://www.scimaç
```



new data visualization  
tool.



Metrics based on Scopus® data as of April 2022



Loading comments...

Developed by:



Powered by:



Follow us on @ScimagoJR

Scimago Lab, Copyright 2007-2022. Data Source: Scopus®

EST MODUS IN REBUS

Horatio (Satire 1, 1, 106)

[Cookie settings](#)

[Cookie policy](#)

