

# **Editorial system registration**

1 message

**Global Journal of Environmental Science and Management** <no-reply@sinaweb.net> To: sucipto-h@fst.unair.ac.id

Mon, Aug 2, 2021 at 12:30 PM



Dear Sucipto Hariyanto,

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Thank you for your registration in the Editorial System Online Subbmission and peer review tracking system. Below please find your username and confidential password, which you need to access the system at:

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Global Journal of Environmental Science and Management

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# Acknowledgement of Submission (#GJESM-2108-3815)

2 messages

Global Journal of Environmental Science and Management <no-reply@sinaweb.net>

Mon, Aug 2, 2021 at 2:59 PM

To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com, sivakumar.gjesm@gmail.com





Quarterly Publication

Author Resubmit:

Manuscript ID: GJESM-2108-3815

Manuscript Title: Characteristics assessment of dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality

Authors: Okik Hendriyanto Cahyonugroho, Sucipto Hariyanto, Ganden Supriyanto

Dear Dr. Sucipto Hariyanto

I wish to acknowledge receiving the above-mentioned manuscript. It should be noted that the manuscript will be reviewed for possible publication in the Global Journal of Environmental Science and Management (GJESM). The regular review process includes the following items:

- 1- Checking the manuscript English language
- 2- Format/Style check based on the GJESM Paper Template
- 3- Plagiarism check (less than 15% duplicates)

#### Airlangga University Mail - Acknowledgement of Submission (#GJESM-2108-3815)

10/20/23, 12:26 PM

4- Reaching at the blind peer reviews the manuscript at least by two potential reviewers in the field of the study, if it passes the three above-mentioned items.

Please be sure that the submitted manuscript has not been published or submitted elsewhere prior to GJESM decision.

The editor's decision will be brought to you for your attention and doing any possible further action in completing the peer review process, once the paper has been reviewed due to the reviewers' considerations.

GJESM Journal is mainly indexed in Web of Science and Scopus. The editorial initial screening of the submitted manuscript will immediately forward you within 1 to 2 days so that you can review it beforehand for the further peer review. The peer review is completed through a fast-tracked model in about 2 to 3 weeks up to the entire publication.

It should also be noted that according to the new GJESM editorial decision, all articles submitted after March 1, 2021 will apply the Article Processing Charge (APC). Therefore, to publish open access in the GJESM Journal, international authors are eligible to pay APC: USD500.00 (for internal authors: 1.000.000 Tomans), when it passes positively through the peer reviews. Otherwise, you must immediately withdraw your submitted paper.

Finally, the GJESM Publisher has decided to award an annual prize to the most innovative paper of the year as USD 1000.

I wish to take this opportunity to thank you for sharing your work with GJESM Journal.

Truly yours,

Professor J. Nouri

Editor in Chief

Global Journal of Environmental Science and Management

sucipto hariyanto <sucipto-h@fst.unair.ac.id>
To: okik hendriyanto <okik.hendriyanto.cahyonugroho-2019@fst.unair.ac.id>

Mon, Aug 2, 2021 at 6:45 PM

[Quoted text hidden]



UNIVERSITY	
Manuscript Needs Resubmission (#GJESM-2108-3815) message	
Wed, o: sucipto-h@fst.unair.ac.id c: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com	Aug 4, 2021 at 3:09 PM
Manuscript ID: GJESM-2108-3815	
Manuscript Title: Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality	
Authors: Okik Hendriyanto Cahyonugroho,Sucipto Hariyanto,Ganden Supriyanto	
Dear <b>Dr. Sucipto Hariyanto</b> ,	
Thank you very much for your interest in publishing your work in the <b>Global Journal of Environmental Science and Management</b> . Your manuscript recorded above cannot be considered for possible publipresented form.	ication in the
Therefore, we suggest you revise your attached manuscript CAREFULLY where you can find it through the website system which some editorial comment is proposed on the proposed file. Thus, after correct through your own dashboard on the journal website system. To resubmit your manuscript, log into <a href="https://www.gjesm.net/">https://www.gjesm.net/</a> and enter as "Author", where you will find your manuscript title listed under "Manuscript".	ction, resubmit it ript needs to be
Editor-in-Chief / Editor Note:	
Truly yours,	

Professor J. Nouri

Editor-in-chief

Global Journal of Environmental Science and Management,



# Acknowledgement of Submission (#GJESM-2108-3815)

4 messages

Global Journal of Environmental Science and Management <no-reply@sinaweb.net>

Wed, Aug 4, 2021 at 1:58 PM

To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com, sivakumar.gjesm@gmail.com





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Professor J. Nouri

Editor in Chief

Global Journal of Environmental Science and Management

#### Global Journal of Environmental Science and Management <no-reply@sinaweb.net>

To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com, sivakumar.gjesm@gmail.com

[Quoted text hidden]

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To: sucipto-h@fst.unair.ac.id

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Wed, Aug 4, 2021 at 3:08 PM

Wed, Aug 4, 2021 at 7:27 PM

Thu, Aug 5, 2021 at 1:59 AM



# RE: Submitted Manuscript #3815 Edited

1 message

GJESM Journal <gjesm.publication@gmail.com>

Thu, Aug 5, 2021 at 2:18 AM

To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id

# Dear Author(s),

Your attached file is just gone for the fast-tracked review. The reason we attach the primary final revision for you is because every time you change the name of the file as well as change the editorial format such as the Tables titles and Figures captions must be in font 10p.

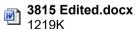
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Global Journal of Environmental Science and Management (GJESM)

Tel.: +9821- 26105110 Fax: +9821- 26105110





SMART UNIVERSITY	sucipto hariyanto <sucipto-h@fst.unair.ac.id></sucipto-h@fst.unair.ac.id>
RE: Your 3 proposed Reviewers  message	
Global Journal of Environmental Science and Management <no-reply@sinaweb.net> o: sucipto-h@fst.unair.ac.id</no-reply@sinaweb.net>	Sat, Aug 14, 2021 at 10:00 PM
GJESM-2108-3815	
Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality	
Dear Author(s),	
As the GJESM Journals are reviewers reviwed your paper positively, if one of your assigned reviewers responds as soon as possible, your the case will be forward	ed to you for your action.
Editorial Office	



# Manuscript Needs Major Revision (#GJESM-2108-3815 (R1))

1 message

Global Journal of Environmental Science and Management <no-reply@sinaweb.net>

Mon, Aug 16, 2021 at 1:55 PM

To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com

Manuscript ID: GJESM-2108-3815 (R1)

Manuscript Title: Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality

Authors: Okik Hendriyanto Cahyonugroho, Sucipto Hariyanto, Ganden Supriyanto

Dear Dr. Sucipto Hariyanto

Your manuscript has now been peer reviewed and virtually resulted as **Major Revision**. As numerous technical comments have pointed out by the reviewers, please revise your manuscript carefully according to the reviewers' comments as well as the editor notifications, within 10 days.

Should the reviewers and editor be satisfied with your amendments, you will be notified the acceptance of your manuscript for publication.

Please do not hesitate to contact us if you have any further inquiries regarding your manuscript revision.

Wishing you every success in your future endeavors.

Truly yours,

**Managing Editor** 

Global Journal of Environmental Science and Management

Dear Author(s),

- 1- Your manuscript review has now been performed as a fast-tacked peer reviewed exceptionally and virtually resulted as Minor Revision where the three reviewers' comments can be found through your own dashboard in the website system (#3815). Therefore you must satisfy the both reviewers by your accurate and completed revisions.
- 2- You will also find an "Author Query Form" that you have to add the manuscript reviewer' inquiries and then response each item into the "Author's response" column carefully and correctly in order to be recognized by the reviewers and editor.
- 3- REMEMBER, any alteration and corrections must be done JUST on the attached final modified manuscript file (#3815 Edited), highlighting with RED paint in order to be recognized by the Editor and Reviewers
- 4- After completion of your paper revisions, please resubmit your revised manuscript as well as the completed Author Query Form file through your own dashboard back (#3815) where you had already submitted your manuscript in the system as soon as possible to avoid any further delays for the latter processing as your paper may be included at the forthcoming issue publication as the last released package.
- 5- Please also finalize your manuscript English content, otherwise, the manuscript processing will be delayed.
- 6- Should the reviewers and editor be satisfied with your amendments, you will be notified to receive the Galley Proof and related copyright forms.
- 7- You will have 5 days' time to return your completed revised manuscript back. In case of not receiving your revised file after the deadline, we assume that you do not like or cannot revise your last stage of article process of publication and therefore your file will be closed with no further action.

Editorial Office

Reviewers Recommendation:

#### Reviewer 1:

Reviewer Comment For Author:

#### Comments:

- 1. The author should present in the Abstract the quantity of any units in terms of the resulting values or the resulting data of the four components of dissolved organic matter namely AP-like, HA-like, SMPs-like, and FA-like to make the findings more specific.
- 2. The Abstract mentions an acronym (FRI) that is not spelled out. According to the GJESM Authors Guideline for Manuscript Preparation, the Abstract should not contain any undefined abbreviation.
- 3. The manuscript title highlighted water quality where other parameters may be of important considerations such as total suspended solids, turbidity, dissolved oxygen, etc. Any justifications why these parameters are not considered in the study?
- 4. With "water quality" as one of the mean contents of the manuscript title, a specific review of literature about water quality is helpful to enhance the scientific basis of the study. The literature can be seen in the following link:

https://www.innspub.net/wp-content/uploads/2018/04/JBES-Vol-12-No-3-p-201-209.pdf

- 5. It is highly suggested to present a GIS-based generated map of the study site to make the presentation more comprehensible.
- 6. The reviewer assumes to have only two (2) replicates of samples being carried out in the study based on the number of data collected which was done twice per week from January to March 2021. Generally, in biology using one-way ANOVA needs at least three (3) replicates to make it acceptable. Any justification for this guery?
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- 8. The paper would be more robust if the author/s present a discussion about the other sources of dissolved organic matter which may be included in the correction analysis between the phytoplankton abundance and the amount of DOM in the surface water.
- 9. Some of the labels in Fig. 4: are too small. It is suggested to increase their font size to enhance readability.
- 10. The labels in Figures 6 and 7 are too small. They should be improved.
- 11. The discussions mention Figures 6a and 6d as well as Figures 7a and 7d. However, these labels are not found in the respective graphs.
- 12. The authors have to recheck carefully the language, grammar including the punctuation of the entire manuscript.
- 13. The conclusion is too generic. A conclusion should highlight the significant findings supported with specific values of the study results.

#### Reviewer 2:

Reviewer Comment For Author:

The manuscript is well written and the methodology was well developed.

Minor recommendations:

- 1. Improve the description in the legend of the figures. The reader should know that it is Sta. 1, Sta. 2, Sta. 3 and Sta. 4 without having to search in the text.
- 2. In Figures 4 and 5, improve the quality of the axis labels.

- 3. It would be interesting to have an image of the study area, since this could contextualize the readers about the characteristics and functioning of the ecosystem. Which could be modulating the behavior found
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#### Reviewer 3:

Reviewer Comment For Author:

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Global Journal of Environmental Science and Management <no-reply@sinaweb.net></no-reply@sinaweb.net>	Tue, Aug 17, 2021 at 7:46 PM
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To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com, sivakumar.gjesm@gmail.com

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Manuscript Title: Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality
Authors: Okik Hendriyanto Cahyonugroho,Sucipto Hariyanto,Ganden Supriyanto
Date: 2021-08-04
Dear <b>Dr. Sucipto Hariyanto</b>
Thank you for submitting the revised file of your manuscript to the Global Journal of Environmental Science and Management
The Editorial Office will proceed on your manuscript and inform you in the earliest time.
If there is anything else, please do not hesitate to contact us.
Truly yours.

Professor D. Sivakumar

Managing Editor

Global Journal of Environmental Science and Management



Managing Editor

sucipto hariyanto <sucipto-h@fst.unair.ac.id>

Request for Submit/Confirm	Galley Proof	(#GJESM-2108-3815	(R1))
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Manuscript ID: GJESM-2108-3815 (R1)	
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Authors: Okik Hendriyanto Cahyonugroho,Sucipto Hariyanto,Ganden Supriyanto	
Dear <b>Dr. Sucipto Hariyanto</b>	
We are pleased to inform you that your paper for the GJESM journal is ready for publication. The page proofs are available at:	
https://www.gjesm.net/	
Attached, you will find three files through the system. 1) Galley Proof, 2) Copyright Release, 3) Conflict of Interest Forms. Please read and revise the Galley Proof carefully, if required corrections must be highlighted with RED fonts to be recognized by the editor. After the final correction, complete and sign the two copyright forms and immediately return them into PDF for dashboard via the system. In addition, finalize English correction if required. The contents must be returned immediately for the entire online publication.	d to be corrected. Therefore, all new ormat for final steps through your own
Truly yours,	
Professor D. Sivakumar	

#### Global Journal of Environmental Science and Management

(URGENT PLEASE)!

Dear Author,

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Please read the Galley Proof carefully and correct any remaining minor corrections. Then, complete and sign the two forms and return them into PDF format through your dashboard (#3815) in the system immediately for the final processes.

In addition, finalize the English language correction, if required. The materials must be returned immediately for the entire publication as the forthcoming issue is going to be released.

**Editorial Office** 



# The Final Stage of Publication (APC payment for Paper # 3815)

1 message

Jafar Nouri <nourijafar@gmail.com>

Wed, Aug 18, 2021 at 8:33 PM

To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id

(URGENT MESSAGE)

#### Dear Author(s),

- 1- First it is necessary to mention that in the time of spiking the hyperlinks into the references, most of the reference parts were deleted by your mandatory action. You were lucky that we found out and recovered them. You may view from your first Galley Proof as well as the finalized file as attached!
- 2- As you may see from the website "Article in Press", at the moment there are 9 articles published and there is only one place remaining for one more article to be completed and releasing the forthcoming issue. In this regard, aside from your prepared article for publication at this issue, another accepted article belonging to professor Marwan Ramli from Universitas Syiah Kuala, Indonesia is forwarded to him for the APC payment today. Now, if anyone pays the APC earlier, it will have the priority to be included in the forthcoming issue which is going to be released and published on the website by next week.

\* \* \* \* \*

At this stage, **USD 525** (APC **500** + **25** subject to VAT) Article Processing Charge (APC) is required due to the acceptance of your manuscript for the publication to deposit to the below account. After the APC payment, you should email us your bank payment receipt within 3 days in order your paper can be included at the forthcoming issue publication. Then, your payment confirmation will be sent to you along with the article acceptance letter and subsequently your published article will immediately appear in the title Website "Press Article".

Besides, if there are some additional corrections, please revise the final version of the manuscript as it is also attached. In between, if you require any further inquiry, please feel free to contact us.

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# 2- PayPal

PayPal Account name: Global Journal of Environmental Science and Management

https://www.paypal.me/GJESM

(It must be delivered as personal, not as business delivery!)

# 3- Western Union Money Transfer

**Account Name:** 

Mr. Nima Nouri

Address: 19203 Ansel, Irvine, California 92618 USA

Phone: +1 (949) 397-0003

\_\_\_\_\_

#### J. Nouri

#### Founder and Editor in Chief

Global Journal of Environmental Science and Management (GJESM)

#### **Emeritus Professor**

Department of Environmental Health Engineering

Tehran University of Medical Sciences

Tehran, Iran

#### **Journal Adviser**

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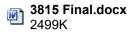
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# Acceptance of Manuscript (#GJESM-2108-3815 (R1))

1 message

Global Journal of Environmental Science and Management <no-reply@sinaweb.net>

Fri, Aug 20, 2021 at 2:36 PM

To: sucipto-h@fst.unair.ac.id

Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com



Manuscript ID: GJESM-2108-3815 (R1)

Manuscript Title: Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality

Authors: Okik Hendriyanto Cahyonugroho, Sucipto Hariyanto, Ganden Supriyanto

Dear Dr. Sucipto Hariyanto

#### **Acceptance Letter**

This is to confirm that after technical and in-house evaluation, the above mentioned manuscript is finalized and recommended by the Editorial Board Committee to be accepted for publication in the Global Journal of Environmental Science and Management (GJESM).

It is necessary to mention that GJESM is an open access, double-blind, peer reviewed quarterly publication, which is indexed and cited in the well-known world databases mainly at the Web of Science, Scopus, SJR (Q2), EBSCO, ProQuest, Ulrichsweb, Cabi, Agricola and Chemical Abstract. The title is committee on Publication Ethics (COPE) and meets the highest ethical standards in accordance with ethical rules (COPE).

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Editor in Chief

Global Journal of Environmental Science and Management





Fri, Aug 20, 2021 at 2:53 PM

# Manuscript Published Online (#GJESM-2108-3815 (R1))

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Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com, sivakumar.gjesm@gmail.com

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Authors: Okik Hendriyanto Cahyonugroho, Sucipto Hariyanto, Ganden Supriyanto

Dear Dr. Sucipto Hariyanto

Your article abstract is now published in the "Article in Press" for the upcoming issues of the Global Journal of Environmental Science and Management (GJESM). The PDF full-text will also be included shortly.

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Mon, Aug 23, 2021 at 12:53 AM

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Cc: okhecah@gmail.com, ganden-s@fst.unair.ac.id, gjesm.publication@gmail.com, sivakumar.gjesm@gmail.com

Manuscript ID: GJESM-2108-3815 (R1)

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Authors: Okik Hendriyanto Cahyonugroho, Sucipto Hariyanto, Ganden Supriyanto

Dear Dr. Sucipto Hariyanto

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Please note that from now on, in case of your current published article gets more citations, in different sources specially in the Scopus and Web of Science journal articles as well as the social media, your future submitted manuscripts will have a better positive impact on easier acceptance in the GJESM Journal. In this regard, We hope to receive your highly cited scientific articles in the future.

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Global Journal of Environmental Science and Management

# REVIEWER RECOMMENDATIONS AND COMMENTS & RESPONSES TO REVIEWER COMMENTS

#### REVIEWER COMMENTS

Dear Author(s),

- 1- Your manuscript review has now been performed as a fast-tacked peer reviewed exceptionally and virtually resulted as Minor Revision where the three reviewers' comments can be found through your own dashboard in the website system (#3815). Therefore you must satisfy the both reviewers by your accurate and completed revisions.
- 2- You will also find an "Author Query Form" that you have to add the manuscript reviewer' inquiries and then response each item into the "Author's response" column carefully and correctly in order to be recognized by the reviewers and editor.
- 3- REMEMBER, any alteration and corrections must be done JUST on the attached final modified manuscript file (#3815 Edited), highlighting with RED paint in order to be recognized by the Editor and Reviewers
- 4- After completion of your paper revisions, please resubmit your revised manuscript as well as the completed Author Query Form file through your own dashboard back (#3815) where you had already submitted your manuscript in the system as soon as possible to avoid any further delays for the latter processing as your paper may be included at the forthcoming issue publication as the last released package.
- 5- Please also finalize your manuscript English content, otherwise, the manuscript processing will be delayed.
- 6- Should the reviewers and editor be satisfied with your amendments, you will be notified to receive the Galley Proof and related copyright forms.
- 7- You will have 5 days' time to return your completed revised manuscript back. In case of not receiving your revised file after the deadline, we assume that you do not like or cannot revise your last stage of article process of publication and therefore your file will be closed with no further action.

**Editorial Office** 

Reviewers Recommendation:

#### Reviewer 1:

Reviewer Comment For Author:

## Comments:

- 1. The author should present in the Abstract the quantity of any units in terms of the resulting values or the resulting data of the four components of dissolved organic matter namely AP-like, HA-like, SMPs-like, and FA-like to make the findings more specific.
- 2. The Abstract mentions an acronym (FRI) that is not spelled out. According to the GJESM Authors Guideline for Manuscript Preparation, the Abstract should not contain any undefined abbreviation.

- 3. The manuscript title highlighted water quality where other parameters may be of important considerations such as total suspended solids, turbidity, dissolved oxygen, etc. Any justifications why these parameters are not considered in the study?
- 4. With "water quality" as one of the mean contents of the manuscript title, a specific review of literature about water quality is helpful to enhance the scientific basis of the study. The literature can be seen in the following link:

https://www.innspub.net/wp-content/uploads/2018/04/JBES-Vol-12-No-3-p-201-209.pdf

- 5. It is highly suggested to present a GIS-based generated map of the study site to make the presentation more comprehensible.
- 6. The reviewer assumes to have only two (2) replicates of samples being carried out in the study based on the number of data collected which was done twice per week from January to March 2021. Generally, in biology using one-way ANOVA needs at least three (3) replicates to make it acceptable. Any justification for this query?
- 7. The manuscript could be more enhanced if the author/s will present a precise description of the study site in terms of climatic, environmental, or bio-physical conditions.
- 8. The paper would be more robust if the author/s present a discussion about the other sources of dissolved organic matter which may be included in the correction analysis between the phytoplankton abundance and the amount of DOM in the surface water.
- 9. Some of the labels in Fig. 4: are too small. It is suggested to increase their font size to enhance readability.
- 10. The labels in Figures 6 and 7 are too small. They should be improved.
- 11. The discussions mention Figures 6a and 6d as well as Figures 7a and 7d. However, these labels are not found in the respective graphs.
- 12. The authors have to recheck carefully the language, grammar including the punctuation of the entire manuscript.
- 13. The conclusion is too generic. A conclusion should highlight the significant findings supported with specific values of the study results.

### **Reviewer 2:**

Reviewer Comment For Author:

The manuscript is well written and the methodology was well developed. Minor recommendations:

- 1. Improve the description in the legend of the figures. The reader should know that it is Sta. 1, Sta. 2, Sta. 3 and Sta. 4 without having to search in the text.
- 2. In Figures 4 and 5, improve the quality of the axis labels.
- 3. It would be interesting to have an image of the study area, since this could contextualize the readers about the characteristics and functioning of the ecosystem. Which could be modulating the behavior found.
- 4. In Table 1, where the results of the correlation are shown, not only the value of the correlation coefficient should be shown, but also the p-value.

# **Reviewer 3:**

**Reviewer Comment For Author:** 

1. Why does surface water have quite high TOC concentration, is it probably due to phytoplankton activities?

- 2. Figure 4, the author showed I, II, IV, it is confused. The reviewer suggested that I, II, IV should be written as name of region, such as AP-like, etc., as mentioned on the paragraph.
- 3. How do you define the area of spectrum Figure 4? which reference?
- 4. Did the author found the other species of phytoplankton? how is the abundance value of those phytoplankton?
- 5. This study should make a suggestion that further research in the lab scale will be conducted in order to identify the characteristic of organic matter has been been released by a kind of phytoplankton species.
- 6. Table 1 seems a miss typed or TOP, it should be TOC.

s

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# Global Journal of Environmental Science and Management

Quarterly Publication

# Manuscript # 3815

# **Author Query Form**

Add the manuscript reviews comments of each reviewer separately in the "Review Details Required" and then respond to each of the items in the "Author's Response" column to be recognize by the reviewers and editor

# Reviewer # 1:

Query	Review Details Required	Author's Response  (Author MUST show the place of performed corrections in the revised manuscript at this column)
1.	The author should present in the Abstract the quantity of any units in terms of the resulting values or the resulting data of the four components of dissolved organic matter namely AP-like, HA-like, SMPs-like, and FA-like to make the findings more specific.	The quantity of unit fluorescence spectra is arbitrary unit (A.U). It has been added on the abstract of the revised manuscript. (Page 1)
2.	The Abstract mentions an acronym (FRI) that is not spelled out. According to the GJESM Authors Guideline for Manuscript Preparation, the Abstract should not contain any undefined abbreviation.	The acronym of FRI is Fluorescence Regional Integration, it has been added on the abstract of the revised manuscript. (Page 1)
3.	The manuscript title highlighted water quality where other parameters may be of important considerations such as total suspended solids, turbidity, dissolved oxygen, etc. Any justifications why these parameters are not considered in the study?	The title highlighted water quality, however the parameter such as total suspended solids, turbidity, dissolved oxygen, etc. are not included in the manuscript because this manuscript focused on dissolved organic matter, since the author assumed that phytoplankton could give effect on the quantity and quality of organic matter in the aquatic. However, this study measured the parameters of water quality, such as total organic carbon.
4.	With "water quality" as one of the mean contents of the manuscript title, a specific review of literature about water quality is helpful to enhance the scientific basis of the study. The literature can be seen in the following link:  https://www.innspub.net/wp-content/uploads/2018/04/JBES-Vol-12-No-3-p-201-209.pdf	The literature suggestion has been added in the revised manuscript. Allochthonous and autochthonous with effluent organic matter are the source of dissolved organic matter (DOM) in the surface water, since allochthonous could be generated from the upstream, midstream and the downstream. The upstream was found to be covered with perennial vegetation; the midstream is used for agriculture and covered with least forest; the downstream was mainly used for residential and utilized for different forms of agriculture (Dumago et al., 2018) (Page 2)

5.	It is highly suggested to present a GIS-based generated map of the study site to make the presentation more comprehensible.	The reviewer suggestion is accepted. A GIS-based generated map of the study site has been added as Figure 1 of the revised manuscript.  Therefore, the existing figure has shifted into labels shifted into the following number. (Page 3)
6.	The reviewer assumes to have only two (2) replicates of samples being carried out in the study based on the number of data collected which was done twice per week from January to March 2021. Generally, in biology using one-way ANOVA needs at least three (3) replicates to make it acceptable. Any justification for this query?	The sample was collected twice per week means sampling time has been conducted two times per week. Each sampling time taken three samples or three replicates.  We made mistake in type two replications. That has been revised in the last paragraph of material and methods. (Page 4)
7.	The manuscript could be more enhanced if the author/s will present a precise description of the study site in terms of climatic, environmental, or bio-physical conditions.	A description of the study site in terms of climate, environmental condition has been added in Table 1 of the revised manuscript. Table 1 is a new table. (Page 3)
8.	The paper would be more robust if the author/s present a discussion about the other sources of dissolved organic matter which may be included in the correction analysis between the phytoplankton abundance and the amount of DOM in the surface water.	The correction analysis between DOM, which has been released by the phytoplaknton abundance, and the amount of DOM in the surface water did not include in this study. However, the reviewer suggestion will be implemented in the future work, as it has been added in the conclusion of the revised manuscript. (Page 12)
9.	Some of the labels in Fig. 4: are too small. It is suggested to increase their font size to enhance readability.	The labels in Figure 4 (rename as Figure 5) has been revised. (Page 7)
10.	The labels in Figures 6 and 7 are too small. They should be improved.	The labels in Figure 6 (rename as Figure 7) and Figure 7 (rename as Figure 8) has been improved. (Page 9 – 10)
11.	The discussions mention Figures 6a and 6d as well as Figures 7a and 7d. However, these labels are not found in the respective graphs.	The labels "a, b, c, d" has been added in the Figure 7 and Figure 8 of the revised manuscript. (Page 9 – 10)
12.	The authors have to recheck carefully the language, grammar including the punctuation of the entire manuscript.	The authors did recheck the language, grammar and the punctuation in the manuscript.
13.	The conclusion is too generic. A conclusion should highlight the significant findings supported with specific values of the study results.	The conclusion has been added significant findings supported with spesific values of the study results. (Page 12)

# Reviewer # 2:

Query	Review Details Required	Author's Response
1.	Improve the description in the legend of the figures. The reader should know that it is Sta. 1, Sta. 2, Sta. 3 and Sta. 4 without having to search in the text.	The description in the legend of the figure, such as Sta.1, Sta.2, Sta.3, and Sta.4 has been improved into Station 1, Station 2, and Station 3. It has been added in the revised manuscript. (Page 5, 6, 8, 9, 10)
2.	In Figures 4 and 5, improve the quality of the axis labels.	The quality of Figure 4 (rename as Figure 5) and Figure 5 (rename as Figure 6) has been improved. (Page 7 – 8)
3.	It would be interesting to have an image of the study area, since this could contextualize the readers about the characteristics and functioning of the ecosystem. Which could be modulating the behavior found.	An image of the study area has been added as Figure 1 in the revised manuscript. (Page 3)
4.	In Table 1, where the results of the correlation are shown, not only the value of the correlation coefficient should be shown, but also the p-value.	Table 1 (rename as Table 2) has shown the results of the correlation coefficient and the p-value. It has been added on the revised manuscript. (Page 11)
5.		
6.		
7.		
8.		
9.		
10.		

# Reviewer # 3:

Query	Review Details Required	Author's Response
1.	Why does surface water have quite high TOC concentration, is it probably due to phytoplankton activities?	Surface water has quite high TOC concentration, it is affected by allochthonous and autochthonous sources. Allochthonous could be generated through watershed discharge into surface water, while autochthonous was produced by microorganism activities in water bodies. This information has been explained in the introduction of the revised manuscript. (Page 2)
2.	Figure 4, the author showed I, II, II, IV, it is confused. The reviewer suggested that I, II, II, IV should be written as name of region, such as AP-like, etc., as mentioned on the paragraph.	The labels "I, II, III, IV" has been revised into "Aromatic protein-like; Fulvic acid -like, Soluble microbials products, humic acid-like" as shown in Figure 4 (rename as Figure 5) of the revised manuscript. (Page 7)
3.	How do you define the area of spectrum Figure 4? which reference?	The area of spectrum has been defined according to the reference of Chen et al., 2003. It has been mentioned in the revised manuscript. (Page 6)
4.	Did the author found the other species of phytoplankton? how is the abundance value of those phytoplankton?	This study was found the other phytoplankton species, such as Oscillatoria sp. (1002 cell/L), Scenesdemus sp. (390 cell/L), Spyrogyra sp. (238 cell/L), Synedra sp. (270 cell/L), Terpsione sp. (202 cell/L) and Mougeotya sp. (262 cell/L).
5.	This study should make a suggestion that further research in the lab scale will be conducted in order to identify the characteristic of organic matter has been been released by a kind of phytoplankton species.	The reviewer suggestion has been added in the conclusion of the revised manuscript. (Page 12)
6.	Table 1 seems a miss typed or TOP, it should be TOC.	"TOP" has been revised into "TOC" in the Table 1 (rename as Table 2) of the revised manuscript. (Page 11)
7.		
8.		
9.		
10.		

# Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality

#### **ABSTRACT:**

**BACKGROUND AND OBJECTIVES:** Dissolved organic matter has a fundamental role in supporting phytoplankton abundance and growth in aquatic environments. However, these organisms produce dissolved organic matter with varied quantities or characteristics depending on the nutrient availability and the species composition. Therefore, this study aims to assess the characteristic of dissolved organic matter on surface water and its correlation with phytoplankton abundance for monitoring water quality.

**METHODS:** The sample was obtained at four Kali Surabaya river stations for further dissolved organic matter analysis and phytoplankton species analysis. The analysis was presented through bulk parameters of total organic, ultraviolet at 254 nm wavelength, specific ultraviolet absorbance value, and fluorescence spectroscopy using excitation-emission matrices with fluorescence regional integration analysis.

**FINDINGS:** The results showed the bulk parameters of dissolved organic matter at all stations were significantly different, as Station 1 and 2 were higher, while 3 and 4 had a lower concentration. Furthermore, the fluorescence spectroscopy identified four components of dissolved organic matter at all stations, namely aromatic proteins-like, humic acid-like, soluble microbial by-products-like, and fulvic acid-like, which is the unit of fluorescence spectra in arbitrary unit. Also, stations 1 and 2 were grouped in the high percentage fluorescence regional integration of humic substance (fulvic acid-like and humic acid-like), while 3 and 4 were classified in the high percentage fluorescence regional integration of non-humic substances (aromatic proteins-like and soluble microbial by-products-like).

**CONCLUSION:** The main phytoplankton species, namely Plectonema sp., Pinularia sp., Nitzchia sp., Navicula sp., had the highest abundance at Stations 1, 3, and 4, respectively. A strong correlation between dissolved organic matter analysis and phytoplankton abundance led to the usage of these methods for monitoring surface water quality.

**KEYWORDS**: Correlation; Dissolved organic matter; Fluorescence spectroscopy; Phytoplankton.

NUMBER OF REFERENCES	NUMBER OF FIGURES	NUMBER OF TABLES		
55	8	2		

**RUNNING TITLE:** Assessing of characteristic dissolved organic matter and its correlation.

#### INTRODUCTION

Human, industrial and agricultural activities have significantly changed aquatic ecosystems due to high organic and inorganic wastewater discharge. This runoff has appeared in the eutrophication of rivers and tributary (Conley *et al.*, 2009; Bhattacharya and Osburn, 2017) causing blooming phytoplankton and consequently, and the environmental issues (Paerl *et al.*, 2008; Heisler *et al.*, 2008; Biggs, 2000). It is eminent that phytoplankton community dynamics (i.e., taxonomic composition, abundance, and biomass) regard the quantity of inorganic phosphorus and nitrogen in the aquatic surrounding (Cao *et al.*, 2016; Cuvin-Aralar *et al.*, 2004). Furthermore, the impact of the organic pollutants contributes to

the quantity or quality of dissolved organic matter in surface water. Allochthonous and autochthonous with effluent organic matter are the source of dissolved organic matter (DOM) in the surface water, since allochthonous could be generated from the upstream, midstream and the downstream. The upstream was found to be covered with perennial vegetation; the midstream is used for agriculture and covered with least forest; the downstream was mainly used for residential and utilized for different forms of agriculture (Dumago et al., 2018). In addition, biogeochemical cycles will affect the quality and quantity of DOM from the surrounding environment. Also, DOM has an essential role in supporting phytoplankton abundance and growth in aquatic surroundings (Kissman et al., 2017; Burpee et al., 2016) due to its usage as an organic nutrient source. It can be used by these micro-organisms as a source of nitrogen, phosphorus, and carbon when inorganic phosphorus and nitrogen are unavailable (Burpee et al., 2016). The primary producers were proposed as an important source that influences its composition in surface water (Biddanda and Benner, 1997). Conversely, DOM can be produced by phytoplankton (Thornton, 2014), with varied characteristics and quantity which are mostly dependent on nutrient availability (Myklestad, 1995), composition of phytoplankton type (Biddanda and Benner, 1997), and bacterial interaction (Ramanan et al., 2016). According to previous studies, various types of DOM have been found and released by different taxonomic groups of phytoplankton (Fukuzaki et al., 2014; Romera-Castillo et al., 2010). Phytoplankton production, microbial metabolism, residue from microbial degradation after their death and other processes, release protein-like materials as one of DOM components (Liu et al., 2019; Mangal et al., 2016). The fluorescence spectroscopy fingerprints, identified the signals of protein-like and humic-like materials released from extracellular Microcystis aeruginosa (Ziegmann et al., 2010). In addition, the DOM which is closely related to the phytoplankton community dynamics, mainly consist of humic-like and protein-like materials (Suksomjit et al., 2009; Zhang et al., 2014) and exhibits their blooming (Altman and Paerl, 2012; Hounshell et al., 2017). The qualitative and quantitative methods for characterizing organic matter analysis have been implemented to clarify the types of DOM transformation through the treatment process or in source water and their following removal. For example, using the bulk parameters of dissolved organic carbon (DOC) concentration, UV/vis at 254 nm wavelength to measure the aromaticity degree of organic matter and specific ultraviolet absorbance (SUVA) (Edzwald et al., 1985; Lai et al., 2015; Hidayah et al., 2017), high-performance size exclusion chromatography (HPSEC) with ultraviolet detector (UVD) or an on-line organic carbon detector (OCD) (Jiao et al., 2014; Lai et al., 2015), fluorescence spectroscopy as well as fluorescence excitation-emission matrices (FEEM) (Hidayah et al., 2017; Ho et al., 2019). These procedures have been previously applied in observing the contribution of phytoplankton degradation to DOM as chromophoric by using fluorescent spectroscopy (Zhang et al., 2009), to characterize DOM excreted by phytoplankton (Chari et al., 2013), and to reveal its relationship with the community (Liu et al., 2021). The use of bulk parameters and fluorescent spectroscopy methods, simultaneously for characterizing organic matter considering the phytoplankton abundance, have been rarely observed. Therefore, resulting in poor implementation of optimal water quality control measures. Furthermore, using these techniques to characterize organic matter and its correlation with phytoplankton abundance for monitoring surface water quality seems to urgently need implementation. Hence, this study aims to assess the characteristic of dissolved organic matter on surface water, as well as its correlation with phytoplankton abundance using the bulk parameters and fluorescence spectroscopy to monitor surface water quality. This study was conducted in the Kali Surabaya River, Surabaya, Indonesia, in 2021.

## **MATERIALS AND METHODS**

## Data collection

This study used water from the Kali Surabaya River in Surabaya city, a surface water source for public supply. The position of station 1 to station 4 is as shown in Table 1 and Fig. 1. The sample was collected twice per week from January to March 2021, and the DOM analysis, as well as phytoplankton abundance was measured through the bulk parameters and fluorescence spectroscopy. The parameters include TOC, UV<sub>254</sub>, SUVA value, while fluorescence spectroscopy identified aromatic proteins-like (AP-like), humic acid-like (HA-like), soluble microbial products-like (SMPs-like), and fulvic acid-like (FA-like). As this study targeted on dissolved organic matter in source water, 0.45 m filter

paper was used to filter the collected source water (Millipore Corporation, USA) to eliminate suspended particles before analysis the parameters. Furthermore, the ultraviolet absorbance at 254 nm (UV<sub>254</sub>) and total organic carbon (TOC) concentration of the water was measured for common physicochemical characteristics based on Standard Methods procedures (APHA *et al.*, 2012).

Table 1: The study sampling location characteristics

No.	Sampling station	Coordinate	Climate	Environmental condition
1	Rolag Telu dam	7°26′40″ S 112°27′25″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature 29°C</li> </ul>	<ul><li>Downstream of the Brantas river</li><li>Stagnant water</li><li>No residential</li></ul>
2	Wringin Anom district	7°24′21″ S 112°30′27″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature</li> <li>29°C</li> </ul>	<ul> <li>Agricultural land</li> <li>There are residential</li> <li>There are domestic activities (bathing, washing, latrine)</li> </ul>
3	Cangkir district	7°22′04″ S 112°37′47″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature</li> <li>29°C</li> </ul>	<ul><li>Industrial area</li><li>Densely populated</li><li>Temporary dump site</li></ul>
4	Karang Pilang drinking water company inlet	7°20′54″ S 112°40′51″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature 29°C</li> </ul>	<ul><li>Industrial area</li><li>There are residential</li><li>There are domestic activities (bathing, washing, latrine)</li></ul>



Fig. 1: Geographic location of the study area in the Kali Surabaya River, Indonesia

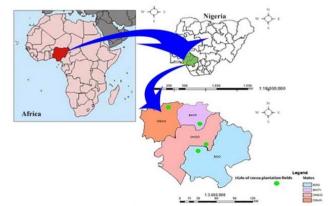
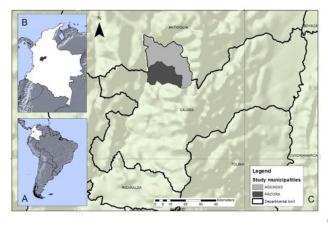
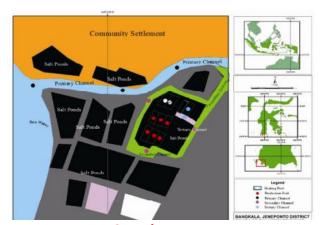


Fig. 1: Locations of the study area







**4 Sample Maps** 

TOC was quantified using TOC-500 Model (Shimadzu, Japan), while UV<sub>254</sub> was detected by UV/vis spectrophotometer Model U-2001 (Hitachi, Japan). The SUVA value showed the dissolved organics were contained in hydrophilic fraction as calculated from measurements of UV<sub>254</sub> and DOC samples. Perkin Elmer LS-55 spectrometer with excitation-emission wavelength pair was used to measure the fluorescence in the source water. Moreover, the excitation-emission matrix (EEM) were resulted for each sample by skimming overexcitation (Ex) wavelengths between 230 and 400 nm at an interval of 10 nm with emission (Em) wavelengths between 300 and 547.5 nm at 0.5 nm interval (Murphy *et al.*, 2013; Hidayah *et al.*, 2017). Counting of fluorescence regional integration (FRI) analysis was used to provide the cumulative fluorescence reaction of organic matter with identical characteristic in selected regions by integration beneath EEMs (Chen *et al.*, 2003). The phytoplankton sampling was conducted using a plankton net mesh size 60 mm as much as 100 liters. Meanwhile, its identification was carried out in the laboratory using a binocular microscope with 10 x 10 magnification (AmScope B100B-MS). Also, the abundance was calculated using Sedgewick-Rafter Counting Chamber for three replications (Marienfeld GmbH).

#### Analytical framework

The Kolmogorov-Smirnov, one-way Analysis of Variance (ANOVA), and Pearson correlations were applied utilizing SPSS Statistics 17.0 software (IBM, Armonk, NY, USA). The Kolmogorov-Smirnov test opposed the empirical cumulative distribution function of bulk parameters data and the results of FRI analysis with the distribution expected when the data were standard. When the observed difference is adequately significant, the test will reject the hypothesis of bulk parameters data, the results of FRI analysis data, and phytoplankton abundance normality. However, when the p-value of this test is less than 5%, it can be concluded that the bulk parameters data, the results of FRI analysis data, and phytoplankton abundance are non-normal. The one-way ANOVA was applied to determine whether any statistically significant differences between the means of bulk parameters and the results of FRI data. It was also used to determine at least two groups of the parameters data as the results of FRI analysis were different. In addition, The Pearson correlation coefficients measured the strength of the linear relationship variables among TOC, UV<sub>254</sub>, SUVA value, AP-like, FA-like, SMPs-like, HA-like, and phytoplankton abundance.

#### **RESULTS AND DISCUSSION**

The bulk parameters of dissolved organic matter in the river segment.

The distribution data for the bulk parameters of dissolved organic matter in the river segment as tested by Kolmogorov–Smirnov showed the TOC concentration (P > 0.15), UV<sub>254</sub> concentration (P > 0.15), and SUVA value (P > 0.15) was normal. Furthermore, the normal distribution data was performed using ANOVA testing to know the differences in mean concentrations of TOC and UV<sub>254</sub>, as well as SUVA value. ANOVA with the Tukey 95% confidence interval also determined whether there were statistically significant or non-significance differences. The results indicated statistically significant differences in the mean concentration of the bulk parameters among the river segment with a p-value of 0.011, 0.001, and 0.004 in TOC, UV<sub>254</sub>, and SUVA values, respectively. Moreover, enough evidence was provided, which concluded that the average of the bulk organic matter parameters at all stations was significantly different. The Tukey analysis classified the bulk parameters concentration at each station into two main groups. Station 1 and 2 were grouped in the high concentration, while 3 and 4 were classified in the bulk parameters' low concentration, which means the former had averages significantly different from the latter. The average TOC concentration for stations 1 and 2 was about a value 10.1-11.7 mg/L, while 3 and 4 were between 9.8-10.9 mg/L. The average UV<sub>254</sub> concentration for stations 1 and 2 was in the range of 10.1-11.7 mg/L, while 3 and 4 were in between 9.8-10.9 mg/L. The average UV<sub>254</sub> concentration for stations 1 and 2 was in the range of 0.65-0.8/cm, while 3 and 4 were 0.39-0.65/cm. The average SUVA concentration of stations 1 and 2 was 5.3-6.4 L/mg/m, while 3 and 4 were 4.0-5.3 L/mg/m. Furthermore, statistical box plot analysis presented the pattern for the bulk parameters of dissolved organic matter in the surface water. Figs. 2, 3, and 4 show a box plot of the average concentration of TOC, UV<sub>254</sub>, and average SUVA value respectively. Fig. 2 shows that the highest

average TOC concentration occurred at Station 2 with a varying range. In comparison, the lowest average TOC concentration with a low range occurred at station 4. In addition, the results showed the average concentration from the highest to the lowest was found at stations 2, 1, 3, and 4. The surface water used in this study contained 7.36 – 15.50 mg/L TOC concentration, which was typically associated with the DOC range. River water has a typical concentration about 2 to 10 mg/L of dissolved organic carbon, which was much higher than groundwater and seawater. Variation in average concentrations of TOC indicated various physical or ecological drivers, chemical processes, spatial changes, which can significantly affect on organic matters dynamics (Maie et al., 2006). The organic matter compositional changes could be induced by biophysical controls, such as changes in composition, which likely result in bioavailability, photoreactivity, nutrient cycling, or chelating capacity and can affect carbon fluxes consequentially ecological drivers not accounted for (Jaffe, 2008). In addition, the hydrology dynamics of surface runoff contributed to the surface water stream (Hood et al., 2006).

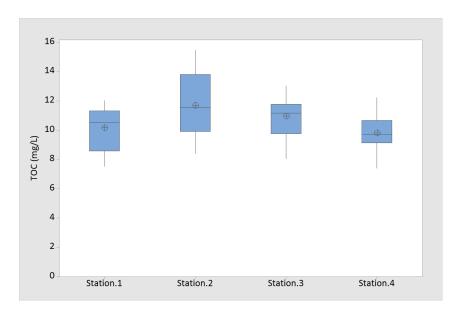


Fig. 2: The average TOC concentration in the river segment at various stations.

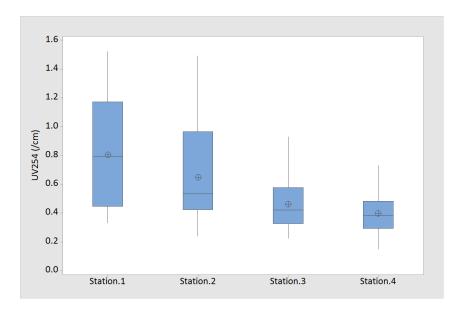


Fig. 3: The average UV<sub>254</sub> concentration in the river segment at various station

Fig. 3 describes the concentration of  $UV_{254}$ , which corresponded to the organic compounds with an aromatic structure, double bonds of C=C (Matilainen *et al.*, 2011). In this study, the concentration of  $UV_{254}$  for surface water was 0.148 - 1.524/cm, which was within the typical range of river (0.085 - 0.4/cm) (Edzwald *et al.*, 1985). The results showed that the average highest aromatic compound was detected at Station 1, while Station 4 had the average lowest concentration. Therefore, Station 1 contained higher humic matter with conjugated C=C double structural bonds than the others. Meanwhile, Station 4 contained lower humic matter than the others. As well known, organic compounds of humic matter contain unsaturated carbon bonds (double or triple) or aromatic rings in their molecular structure. Hence, it absorbs an amount of UV light through the water sample (Her *et al.*, 2002).

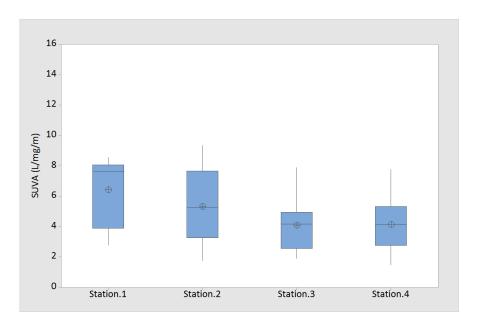


Fig. 4: The average SUVA value in the river segment at various station

Fig. 4. Shows the hydrophobicity of organic matter characteristic or specific UV-absorbance (SUVA) value. The results revealed a value between 1.45 – 9.36 L/mg/m. However, it was mostly higher than 4 along the river segment, which means that the organic matter is mainly consists of humic, hydrophobic, and high molar mass organic material. According to Edzwald and Tobiason (2011), SUVA is a parameter of the organic matter composition in water. Source water with SUVA values ≥ 4 indicated that natural organic matter composed mainly of humic or hydrophobic matter, while those < 2 contained mainly non-humic or hydrophilic natural organic matter. The results were consistent with the high concentration of  $UV_{254}$  (0.148 – 1.524/cm). The values typically ranged from 1.0 to 6.0 L/mg/m for surface water. However, values greater than 6.0 were revealed for interstitial waters dominated by a solid terrestrial signature (Jaffe et al., 2008). According to previous studies, these higher values can be as a result of the absorption at 254 nm from colloids, iron, or other components in the sample (Weishaar et al., 2003; Hudson et al., 2007). Combining the bulk parameters of TOC, UV<sub>254</sub>, and SUVA value led to characterize the organic matter in the river. Station 2 was mainly composed of the highest TOC with lower aromatic and hydrophobic than 1, and vice versa. Also, station 4 was mainly composed of lower bulk parameters than 3. Therefore, 2 contained more aliphatic organic matter that does not absorb at 254 nm than the others. The lower SUVA value among all stations indicated the mixtures of aquatic humics, hydrophobic and hydrophilic, and molecular weights of organic matter.

Characteristic of fluorescence dissolved organic matter in the river segment through volumetric fluorescence distribution.

Fig. 5. Illustrates the fluorescence excitation-emission matrices (FEEMs) for dissolved organic matter in the river segment at a different station, taken on the first week sampling time. Dissolved organic carbon was classified into four regions based on its excitation/emission wavelengths (Ex/Em), namely Region 1 indicated the aromatic proteins-like (AP-like), such as tyrosine and tryptophan, at Ex/Em <250 nm/<350 nm. Region 2 identified the fulvic acid-like (FA-like) substances at Ex/Em <250 nm/>380 nm, Region 3 was corresponded to the soluble microbial by products-like (SMPs-like) substances at Ex/Em 250-280 nm /<380 nm, while Region 4 was identified as the humic acid-like (HA-like) substances with Ex/Em >280 nm/>380 nm (Chen et al., 2003).

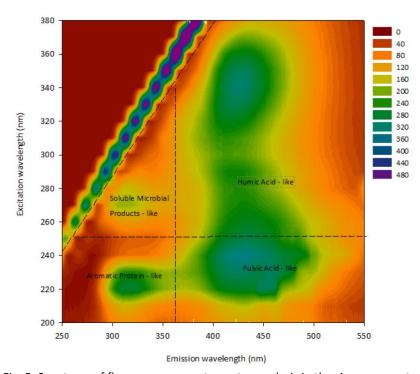


Fig. 5: Spectrum of fluorescence spectrometer analysis in the river segment

This study shows that the fluorescence component from FEEM analysis has consistent results with previous studies (Her et al., 2003; Yao et al., 2016; Moradi et al., 2018; Hidayah et al., 2020). Generally, HA-like and FA-like correlated with aromatic compounds. They mainly exist as carboxylic and phenolic functional groups in natural dissolved organic matter. These fluorescence structures are mostly present as a significant percentage of humic substances, which typically represent over 50% of natural organic matter (Shon et al., 2012). In addition, source water may contain protein-like materials which microbial activities can generate. The amount, characteristics, and properties of dissolved organic matter in the aquatic system depend on their origin and environmental biochemical cycles. Sources of organic matter are classified as allochthonous (generated from a terrestrial watershed) and autochthonous (produced by organism activities, such as phytoplankton activities) (Chari et al., 2013; Zhang et al., 2009; Haraguchi et al., 2019). Terrestrial watershed is mainly composed of humic substances such as fulvic and humic acids as well as humin, which are primarily hydrophobic and rich in aromatic carbon. The autochthonous source material is microbially derived organic, such as algalderived and effluent organic matter (Kelso and Baker, 2020). Fig. 6 showed the percentage fluorescence response, which was calculated by Fluorescence Regional Integration (FRI) method. The percentage of fluorescence distribution indicated the four fractions quantity of fluorescence organic matter. This study classified the fraction into humic and non-humic substances-like. The first was represented by Region 2 (FA-like) and 4 (HA-like), while the second one by Region 1 (AP-like) and 3 (SMPs-like). Firstly, the results showed the highest total percentage of FRI in Region 2 and 4 was at Station 1 (76.6%), and the lowest total percentage for humic substances-like was at Station 4 (69.2%). Both components are classified as humic substances and are mainly composed of aromatic compounds with high to medium molecular weight (Watson et al., 2018; Hua et al., 2020). Their total percentage

FRI showed a consistent UV<sub>254</sub> concentration and SUVA value. Furthermore, Station 1 had the highest bulk parameters, while 4 had the lowest. Secondly, the highest total percentage FRI of Region 1 and 3 (30.8%) was identified at Station 4, with the lowest at 1 (23.4%). This indicated that Station 4 contained abundant proteins substances and microbial-like fluorescence than the others and followed the lowest SUVA value of Station 4 with the highest for Station 1. Region 1 and 3 correlated with high molecular weight protein-like, which had chemical properties related to aromatic amino acids, tryptophan or tyrosine-like (Yamashita *et al.*, 2008; Hua *et al.*, 2020) and low molecular weight microbial humic-like as well as less conjugated double bond organic matter (Nguyen *et al.*, 2013; Hua *et al.*, 2020).

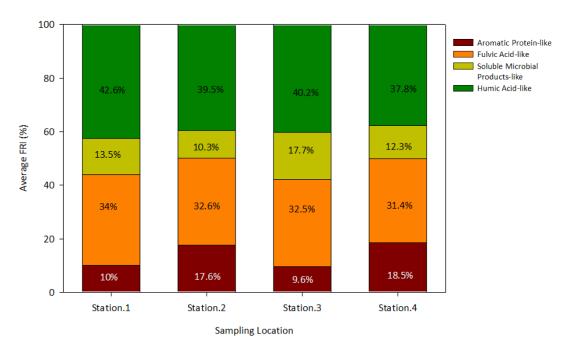


Fig. 6: FRI distribution of fractionated organic matter from the various river segment

The distribution data for the fluorescence of dissolved organic matter in the river segment was tested by Kolmogorov-Smirnov and the bulk parameters. The results showed distribution data for percentage FRI of Region 1 (AP-like), 2 (FA-like), 3 (SMPs-like), and Region 4 (HA-like) with P > 0.000, 0.007, 0.000 and 0.013 respectively were normal. Furthermore, Analysis of Variance (ANOVA) testing was carried out to determine the differences in mean percentage FRI for each region. The statistical analysis ANOVA One-Way with the Tukey 95% confidence interval also determined whether statistically significant or non-significance differences in percentage FRI of AP-like, FA-like, SMPs-like, and HA-like among all stations. The results showed statistically significant differences in the mean percentage FRI of all fluorescence organic fractions at all stations with p = 0.000, 0.007, 0.000, and 0.013 in AP-like, FA-like, SMPs-like, HA-like, respectively. The results provided enough evidence to conclude that the mean percentage FRI of all fluorescence organic fractions at all stations was significantly different. Moreover, the Tukey analysis classified their percentage FRI at each station into two main groups. Station 1 and 2 were grouped in the high percentage FRI of humic substance-like (FA-like and HA-like), while 3 and 4 were classified in the low percentage. This means the former had an average percentage FRI of FA-like and HA-like, which were significantly different from the latter. In addition, stations 3 and 4 were grouped in the high percentage FRI of non-humic substance-like (AP-like and SMPs-like), while stations 3 and 4 were grouped in the low percentage. This showed both had average percentage FRI of AP-like and SMPs-like, which were significantly different from stations 1 and 2. Moreover, statistical box plot analysis presented the pattern of the fluorescence organic matter in the river segments. Fig. 7a to 7d presented box plot with average percentage FRI of the organic matter. Firstly, a comparison among all fluoresces organic compounds showed the average FRI of HA-like was much higher and much lower for SMPs-like than the others. However, HA-like, located at Region 4 of the fluorescence spectra, had the most extensive range of excitation and emission wavelengths. Therefore, the humic acid substances-like region had the most extensive volume distribution of FRI when compared to others (Chen et al., 2003). Meanwhile, SMPs-like or Region 3 comprised a dominant percentage of the fluorescence in wastewater treatment plant effluent (Chen et al., 2003) and was closely related to the phytoplankton activities (Liu et al., 2021; Hua et al., 2020). Second, the average percentage FRI of the organic matter indicated different quantities and quality at each station. The non-humic substanceslike fluorescence as presented by AP-like, SMPs-like, with statistical analysis, had a higher percentage FRI at stations 3 and 4 than the others. It was likely that Station 3 and 4 had a higher percentage of extracellular biological organic matter fraction than the other river segments. The fraction was supposed to contain soluble microbial products of amino acids and carbohydrates. Tryptophan and tyrosine which are aromatic amino acids, were confirmed as biological activity products in natural systems and exhibited a distribution of fluorescence response similar to AP-like and SMPs-like of this study (Coble 2007; Determann et al., 1998). The humic substances-like fluorescence as presented by FA-like and HA-like had a higher percentage FRI at stations 1 and 2 than others and were tested by ANOVA One-Way. Combining the bulk parameters of TOC, UV254, SUVA value, and fluorescence spectroscopy convinced the characteristic of organic matter in the river. Station 1 and 2 had high UV<sub>254</sub> concentration, SUVA value, high percentage FRI of FA-like and HA-like substances. It was conjectured that stations 1 and 2 were mainly composed of aromatic, hydrophobic, humic substances organic matter, which may be generated from terrestrial systems.

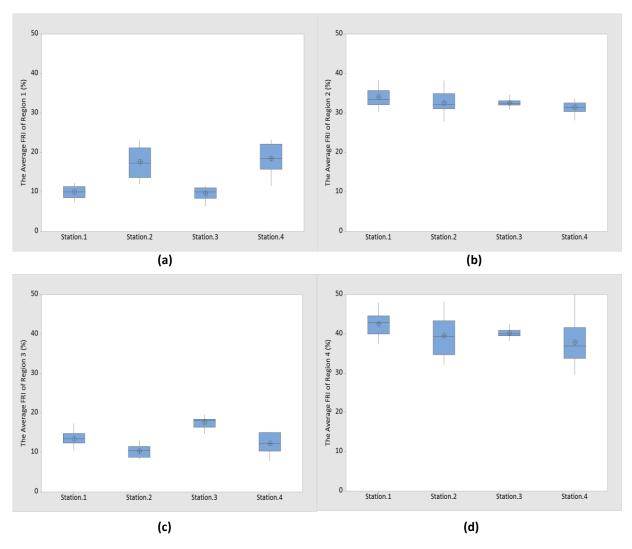


Fig. 7: The average percentage FRI of fluorescence organic matter in the river segment at various stations

Station 3 and 4 had lower UV concentration and SUVA values, with a high percentage FRI of AP-like and SMPs-like than the others. There was a lower SUVA value among all stations indicates in the mixtures of aquatic humics, hydrophobic and hydrophilic, as well as molecular weights of organic

matter. This showed that Station 3 and 4 comprised more autochthonous and sources of organic matter from anthropogenic activities. The river ecosystem, which source is terrestrial, autochthonous, and anthropogenic, provided hotspots for storing, transporting, and transforming organic matter. The sources proportions were primarily and terrestrially derived with increased autochthonous inputs from macrophytes. In addition, the sources of dissolved organic matter are a mixture of terrestrial, autochthonous, or primarily from wastewater effluent (Kelso and Baker, 2020).

Contribution of phytoplankton abundance to fluorescence dissolved organic matter in the river segment.

This study discovered four main phytoplankton species with various abundance in the river segments, namely Plectonema sp., Nitzchia sp., Navicula sp., and Pinularia sp. The distribution data of the phytoplankton abundance in this segment was tested by Kolmogorov-Smirnov. The results showed a usual distribution data for Plectonema sp., Nitzchia sp., Navicular sp., and Pinularia sp. as abundance P > 0.000, 0.007, 0.000, and 0.013, respectively. Furthermore, ANOVA testing was carried out to determine the differences in the mean phytoplankton abundance of the river segments. The statistical analysis ANOVA One-Way with the Tukey 95% confidence interval determined whether there were statistically significant or non-significance differences in the abundance of the species among all stations. According to the results, there were statistically significant differences in the mean abundance of phytoplankton at all stations with p-value = 0.006 and 0.01 in Plectonema sp. and Nitzchia sp. abundance, respectively. Meanwhile, the analysis generated p-value = 0.156 and 0.412 for Navicula sp. and Pinularia sp. abundance, respectively, therefore, classified as only one group of phytoplankton abundance. This showed that there were non-significantly differences in both species abundance among all stations. The Tukey analysis classified Plectonema sp. and Nitzchia sp. abundance at each station into two main groups. Station 1 and 2 were grouped in the high Plectonema sp. and low Nitzchia sp. abundance, while Station 3 and 4 were classified in the low Plectonema sp. and high Nitzchia sp. abundance. Furthermore, the statistical box plot analysis presented the pattern of the phytoplankton abundance in the river segments.

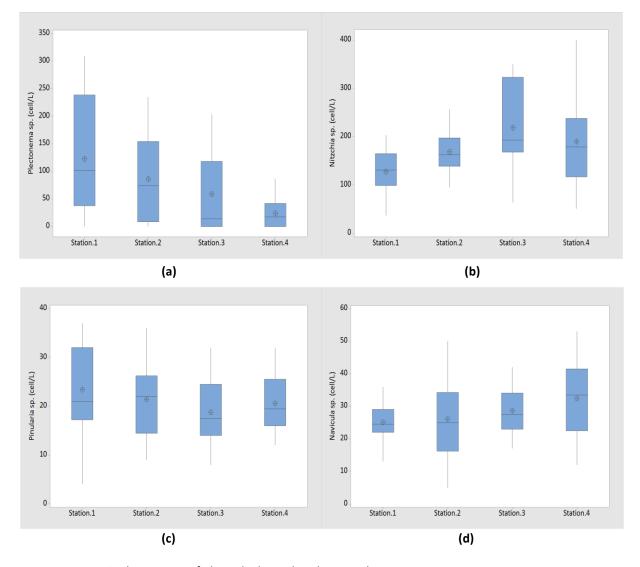


Fig. 8: The average of phytoplankton abundance in the river segment at various station

Fig. 8a to 8d present box plots of their average abundance. Firstly, a comparison among the species at all stations conjectured that Nitzchia sp. had a higher abundance, and Pinularia sp. was lower than the others. Meanwhile, Plectonema sp. had the highest at Station 1 and the lowest at 4. Nitzchia sp. had a higher abundance at Station 3 and lower at 1. Moreover, Navicula sp. had the highest abundance at Station 4 and the lowest at Station 1. Pinnularia sp. gave the highest at Station 1, with the lowest at Station 3. This phytoplankton abundance was strongly influenced by migration, which can occur due to population density and physical environmental conditions, such as changes in temperature and currents (Basu and Mackey, 2018). Secondly, Station 1 was likely to contain a similar abundance in Plectonema sp. and Nitzchia sp., and the same for Navicula sp. and Pinularia sp. Stations 2 and 3 showed that the abundance of Nitzchia sp. was primarily dominant than others. However, Navicular sp. was similar to Pinularia sp. Station 4 identified a similar abundance of Plectonema sp., Navicular sp., and Pinularia sp. There is competition in several phytoplankton species that use the same resource lacking in availability, or even regardless of sufficient availability, and competition still occurs when they take advantage of the resource, with one attacking the other or vice versa (Burson et al., 2018).

The relationship among the bulk parameters, organic fluorescence parameters, and phytoplankton abundance

The degree correlation between the bulk parameters, fluorescence organic matter, and phytoplankton abundance was examined, as shown in Table 2. Correlation analysis was carried out using TOC and  $UV_{254}$  concentrations, SUVA value with percentage FRI of AP-like, FA-like, SMPs-like, or HA-like, as well

as the abundance of Plectonema sp., Nitzchia sp., Navicula sp., and Pinularia sp. Firstly, based on the correlations of the bulk parameters, TOC concentration was positively higher with Region 1 (AP-like) and Region 2 (FA-like). In addition,  $UV_{254}$  concentration and SUVA value were significantly correlated with Region 1(AP-like) and Region 4 (HA-like). The results showed fluorescence spectroscopy, which fractionated AP-like, FA-like, SMPs-like, and HA-like could be used to identify the quantity and quality of organic matter in the source water.

Table 2: The degree correlation among the bulk parameters, fluorescence organic matter, and phytoplankton abundance

Parameters	ТОС	UV <sub>254</sub>	SUVA	AP- like	FA- like	SMPs -like	HA- like	Navicula sp.	Plectone ma sp.	Pinnularia sp.
UV <sub>254</sub>	0.085 0.502	•	•	•	•		•			•
SUVA	-0.044 0.729	0.887 0.000								
AP-like	-0.287 0.022	-0.440 0.000	-0.373 0.002							
FA-like	0.254 0.042	0.105 0.411	0.047 0.710	-0.249 0.048						
SMPs-like	-0.038 0.764	-0.198 0.116	-0.228 0.070	0.638 0.000	-0.085 0.505					
HA-like	-0.035 0.786	0.344 0.005	0.344 0.005	-0.674 0.000	-0.022 0.862	-0.348 0.005				
Navicula sp.	-0.109 0.392	-0.331 0.007	-0.289 0.021	0.193 0.126	-0.102 0.422	0.090 0.480	-0.082 0.521			
Plectonema sp.	0.271 0.030	0.137 0.281	0.131 0.303	-0.346 0.005	0.293 0.019	-0.057 0.652	0.110 0.386	0.166 0.189		
Pinnularia sp.	-0.097 0.448	-0.239 0.058	-0.292 0.470	-0.245 0.051	0.142 0.263	-0.268 0.032	0.220 0.080	0.137 0.279	0.320 0.010	
Nitzchia sp.	-0.243 0.053	-0.283 0.023	-0.203 0.108	0.160 0.205	-0.203 0.107	0.176 0.164	0.070 0.585	0.168 0.184	0.035 0.785	0.174 0.170

Cell Contents description; Pearson correlation (the first row of the number of correlation between parameters); P-value (the second row of the number of correlation between parameters)

This result was expected since TOC measured all organic carbon, including humic and non-humic substances, as presented by AP-like and FA-like. Secondly, a strong positive correlation between UV<sub>254</sub> concentration and SUVA value indicated that higher aromatic conjugated double bond corresponded to higher molecular weight organic, more hydrophobic, and content of humic substances. These results are consistent with the Pearson correlation between bulk parameters of UV<sub>254</sub> correlation, SUVA, and fluorescence organic matters of AP-like and HA-like. Furthermore, it was conjectured that fluorescence spectroscopy could be used to assess the properties of organic matter existing in the source water. Thirdly, the results showed that TOC had a stronger correlation with AP-like than HA-like. This was probably because the humic structure may incorporate protein-like-fluorophores due to weak interactions based on x-x or van der Waals forces between the dissolved organic matter components. Previous studies indicated that proteins and humic supramolecules containing specific structures attained from phenol or aniline might contribute to the fluorescence. Fourth, this study discovered a strong correlation between DOM and phytoplankton abundance. Plectonema sp. correlated with TOC, AP-like, and FA-like, while Navicula sp. and Nitzchia sp. correlated with UV<sub>254</sub>, and Pinularia sp. with SMPs-like. The existence of phytoplankton was likely to enhance the quantity and characteristics of DOM in the aquatic environment. The production of marine-like fluorophores accompanied phytoplankton degradation as a significant source of autochthonous DOM (Wada et al., 2007). In addition, higher molecular weight compounds such as protein (tryptophan)-like fluorescence were presented in exudates when phytoplankton grows (Chari et al., 2013). The combination of the bulk parameters (TOC, UV<sub>254</sub>, and SUVA value), fluorescence spectroscopy, and phytoplankton abundance convinced the quality of organic matter in the surface water. However, it could be eventually used to monitor the water's quality.

#### **CONCLUSION**

This study showed that the quality and quantity of DOM at all stations were significantly different, as classified into two groups with higher bulk parameters at stations 1 and 2 and a lower concentration at 3 and 4. The average TOC concentration for stations 1 and 2 was about a value 10.1-11.7 mg/L, while 3 and 4 were in between 9.8-10.9 mg/L. The average  $UV_{254}$  concentration for stations 1 and 2 was in the range of 10.1-11.7 mg/L, while 3 and 4 were between 9.8-10.9 mg/L. The average  $UV_{254}$ concentration for stations 1 and 2 was 0.65-0.8/cm, while 3 and 4 were 0.39-0.65/cm. The average SUVA concentration of stations 1 and 2 was in the range 5.3-6.4 L/mg/m, while 3 and 4 were 4.0-5.3 L/mg/m. In addition, fluorescence spectroscopy with FRI analysis showed stations 1 and 2 were grouped in the high percentage FRI of humic substance-like (FA-like and HA-like) about 74.35%. It was conjectured that stations 1 and 2 were mainly composed of aromatic, hydrophobic, humic substances organic matter, which may be generated from terrestrial systems, while stations 3 and 4 were classified in high percentages non-humic substances-like (AP-like and SMPs-like) about 29.05%. This showed that Station 3 and 4 comprised more autochthonous and sources of organic matter from anthropogenic activities. According to phytoplankton abundance, Station 1 had a high abundance of Plectonema sp. (238.5 cell/L) and Pinularia sp. (32 cell/L), while stations 2 and 3 mainly consisted of Nitzchia sp. (197.5 cell/L and 322.75 cell/L), and Navicula sp. (41.5 cell/L) was dominant at Station 4. The Pearson correlation showed a strong relationship between DOM and phytoplankton abundance. Therefore, Plectonema sp. was in correlation with TOC (0.271), AP-like (-0.346), and FA-like (0.293), while Navicula sp. and Nitzchia sp. correlated with UV<sub>254</sub> (-0.331 and -0.283), and Pinularia sp. correlated with SMPslike (-0.268). This study conjectured that the bulk parameters of DOM, fluorescence spectroscopy, and phytoplankton abundance could be used to assess the characteristic of DOM, while the combination of these methods could be used to monitor the surface water quality. Future work should be conducted on the laboratory scale for phytoplankton observation in order to identify the characteristic of organic matter that a kind of phytoplankton species has released. Therefore, it could be used to predict the amount of DOM derived by phytoplankton, DOM derived in the aquatic, and DOM from the terrestrial watershed.

# **AUTHOR CONTRIBUTIONS**

Okik Hendriyanto Cahyonugroho performed the experimental design, analyzed the data, and prepared the manuscript text as well as the literature review. Sucipto Hariyanto and Ganden Supriyanto interpreted the data and helped in manuscript preparation.

# **ACKNOWLEDGEMENT**

This study was supported by the Directorate of Research and Community Service, Deputy for Strengthening Research and Development of the Ministry of Research and Technology/National Research and Innovation Agency through Postgraduate Research — Doctoral Dissertation Research Scheme for multi-year, with Contract No. 489/UN3.15/PT/2021.

## **CONFLICT OF INTEREST**

The authors declare no potential conflict of interest regarding the publication of this work. Also, ethical issues, including plagiarism, informed consent, misconduct, data fabrication and/or falsification, double publication and submission, as well as redundancy, have been entirely witnessed by the authors.

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#### **ABBREVIATIONS**

% Percent
/cm Per centimeter

ANOVA Analysis of variance
AP-like Aromatic proteins-like
C=C Carbon chain double bonds

cell/L The number of phytoplankton cell per liter

DOCDissolved organic carbonDOMDissolved organic matterEmEmission wavelengthExExcitation wavelength

FA-like Fulvic acid-like

FEEM Fluorescence spectroscopy using excitation-emission matrices

FRI Fluorescence regional integration

HA-like Humic acid-like

*L/mg/m* Liter per miligrams per meter

mg/L Miligrams per liter

mm Milimeter
 μm Micrometer
 Navicula sp. Navicula species
 Nitzchia sp. Nitzchia species

NOM Natural organic matter

*nm* Nanometer

OCD Organic carbon detector
 P > Probability value more than
 P = Probability value equal
 Pinnularia sp.
 Plectonema sp.
 Plectonema species
 P-value
 Probability value

SMPs-like Soluble microbial by products-like SUVA Specific ultraviolet absorbance

TOC Total organic carbon

UV<sub>254</sub> Ultraviolet at 254 nm wavelength

UVD Ultraviolet detector UV/vis Ultraviolet visible

#### **REFERENCES**

Altman, J.C.; Paerl, H.W., (2012). Composition of inorganic and organic nutrient sources influences phytoplankton community structure in the New River Estuary, North Carolina. Aquat. Ecol., 46(3): 269-282 (14 pages).

https://link.springer.com/article/10.1007/s10452-012-9398-8

APHA, (2012). Standard Methods for the Examination of Water and Wastewaters, 21<sup>th</sup> ed. American Public Health Association: Washington, DC.

http://sricstaff.santarosa.edu/~oraola/Assets/APHA SM 20.pdf

Basu, S.; Mackey, K.R.M., (2018). Phytoplankton as key mediators of the biological carbon pump: their responses to a changing climate. J. Sustainability. 10(3): 1-18 (18 pages).

https://www.mdpi.com/2071-1050/10/3/869/htm

Bhattacharya, R.; Osburn, C.L., (2017). Multivariate analyses of phytoplankton pigment fluorescence from a freshwater river network. Environ. Sci. Technol., 51(12): 6683-6690 (8 pages).

https://pubs.acs.org/doi/pdf/10.1021/acs.est.6b05880?rand=lld0z3bx&

Biddanda, B.; Benner, R., (1997). Carbon, nitrogen, and carbohydrate fluxes during the production of particulate and dissolved organic matter by marine phytoplankton. Limnol. Ocean., 42: 506–518 (13 pages).

https://aslopubs.onlinelibrary.wiley.com/doi/abs/10.4319/lo.1997.42.3.0506

Biggs, B.J.F., (2000). Eutrophication of streams and rivers: dissolved nutrient chlorophyll relationships for benthic algae. J. North Am. Benthol. Soc., 19(1): 17-31 (15 pages).

https://www.jstor.org/stable/1468279

Burpee, B.; Saros, JE; Northington, R.M.; Simon, K.S., (2016). Microbial nutrient limitation in Arctic lakes in a permafrost landscape of southwest Greenland. Biogeosci., 13(2): 365-374 (10 pages). https://bg.copernicus.org/articles/13/365/2016/

Burson A.; Stomp M.; Greenwell E.; Grosse J.; Huisman J. (2018). Competition for nutrients and light: testing advances in resource competition with a natural phytoplankton community. J. Ecol. 99(5): 1108 – 1118 (11 pages).

https://esajournals.onlinelibrary.wiley.com/doi/full/10.1002/ecy.2187

Cao, X.; Wang, J.; Liao, J.; Sun, J.; Huang, Y., (2016). The threshold responses of phytoplankton community to nutrient gradient in a shallow eutrophic Chinese lake. Ecol. Indicat., 61: 258-267 (10 pages).

https://daneshyari.com/article/preview/6293585.pdf

Chari, N.V.H.K.; Keerthi, S.; Sarma, N.S.; Pandi, S.R.; Chiranjeevulu, G.; Kiran, R.; Koduru, U., (2013). Fluorescence and absorption characteristics of dissolved organic matter excreted by phytoplankton species of western Bay of Bengal under axenic laboratory condition. J. Exp. Mar. Biol. Ecol., 445: 148-155 (8 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0022098113001238?via%3Dihub

Chen, W.; Westerhoff, P.; Leenheer, J.A.; Booksh, K., (2003). Fluorescence excitation-emission matrix regional integration to quantify spectra for dissolved organic matter. J. Environ. Sci. Technol., 37: 5701-5710 (10 pages).

https://pubs.acs.org/doi/10.1021/es034354c

Coble, P.G., (2007). Marine chemical biogeochemistry: the chemistry of ocean color. Chem. Rev., 107: 402-418 (17 pages).

https://pubs.acs.org/doi/abs/10.1021/cr050350+

Conley, D.J.; Paerl, H.W.; Howarth, R.W.; Boesch, D.F.; Seitzinger, S.P.; Havens, K.E.; Lancelot, C.; Likens, G.E., (2009). Controlling eutrophication: nitrogen and phosphorus. J. Sci, 323(5917): 1014-1015 (2 pages).

https://science.sciencemag.org/content/323/5917/1014

- Cuvin-Aralar, M.L.; Focken, U.; Becker, K.; Aralar, E.V., (2004). Effects of low nitrogen phosphorus ratios in the phytoplankton community in Laguna de Bay, a shallow eutrophic lake in the Philippines. Aquat. Ecol. 38(3): 387-401 **(15 pages)**.
- https://link.springer.com/article/10.1023/B:AECO.0000035174.35091.0b
- Determann, S.; Lobbes, J.M.; Reuter, R.; Rullko tter, J., (1998). Ultraviolet fluorescence excitation and emission spectroscopy of marine algae andbacteria. Mar. Chem., 62(1–2): 137–156 (19 pages)
- Dumago, S.W.L.; Puno, G.R.; Ingotan, S.S., (2018). Water quality assessment in various land use and land cover of Muleta Watershed Bukidnon, Philippines. J. Bio. Environ. Sci., 112(3): 201-209 (8 pages)
- https://www.innspub.net/wp-content/uploads/2018/04/JBES-Vol-12-No-3-p-201-209.pdf
- Edzwald, J.K.; Becker, W.C.; Wattier, K.L., (1985). Surrogate parameters for monitoring organic matter and THM precursors. J. Am. Water Works Assn., 77(4): 122–131 (10 pages).
- https://awwa.onlinelibrary.wiley.com/doi/abs/10.1002/j.1551-8833.1985.tb05521.x
- Edzwald, J.K.; Tobiason, J.E., (2011). Chemical principles, source water composition, and watershed protection. In Water Quality & Treatment: A handbook on drinking water; Edzwald, J.K., Eds. New York: AWWA McGraw-Hill.
- https://www.worldcat.org/title/water-quality-treatment-a-handbook-on-drinking-water/oclc
- Fukuzaki, K.; Imai, I.; Fukushima, K.; Ishii, K.I.; Sawayama, S.; Yoshioka, T., (2014). Fluorescent characteristics of dissolved organic matter produced by bloom-forming coastal phytoplankton. J. Plankton. Res., 36: 685–694 **(10 pages)**.
- https://core.ac.uk/download/pdf/39322149.pdf
- Haraguchi, L.; Asmala, E.; Jakobsen, H.H.; Carstensen, J., (2019). Composition of natural phytoplankton community has minor effects on autochthonous dissolved organic matter characteristics. Mar. Biol. Res., 1-19 (19 pages).
- https://www.tandfonline.com/doi/abs/10.1080/17451000.2019.1662449?journalCode=smar20
- Heisler, J.; Glibert, P.M.; Burkholder, J.M.; Anderson, D.M.; Cochlan, W.; Dennison, W.C.; Dortch, Q.; Gobler, C.J.; Heil, C.A.; Humphries, E.; Lewitus, A.; Magnien, R.; Marshall, H.G.; Sellner, K.; Stockwell, D.A.; Stoecker, D.K.; Suddleson, M., (2008). Eutrophication and harmful algal blooms: a scientific consensus. Harmful Algae, 8(1): 3-13 (11 pages).
- https://www.sciencedirect.com/science/article/abs/pii/S1568988308001066?via=ihub
- Her, N.; Amy, G.; Foss, D.; Cho, J.; Yoon, Y.; Kosenka, P., (2002). Optimization of method for detecting and characterizing NOM by HPLC-size exclusion chromatography with UV and on-line DOC detection. Environ. Sci. Technol., 36: 1069–1076 (8 pages).
- https://pubs.acs.org/doi/10.1021/es015505j
- Her, N.; Amy, G.; McKnight, D.; Sohn, J.; Yoon, Y., (2003). Characterization of DOM as a function of MW by fluorescence EEM and HPLC-SEC using UVA, DOC, and fluorescence detection. Water Res., 37: 4295-4303 (9 pages).
- https://www.sciencedirect.com/science/article/abs/pii/S0043135403003178?via=ihub
- Hidayah, E.N.; Chen, Y.C.; Yeh, H.H., (2017). Comparison between HPSEC-OCD and F-EEMs for assessing DBPs formation in water. J. Environ. Sci. Health. Part A., 52(4): 391-402 (12 pages).
- $\underline{https://www.tandfonline.com/doi/abs/10.1080/10934529.2016.1262607?journalCode=lesa20}\\$

Hidayah, E.N.; Pachwarya, R.B.; Cahyonugroho, O.H.; Ramanathan, A.L., (2020). Characterization of molecular weight-based fluorescent organic matter and its removal in combination of constructed wetland with activated sludge process. Water Air Soil Pollut., 231(41): 1-12 (12 pages).

https://link.springer.com/article/10.1007/s11270-020-4405-5

Ho, H.H.; Cao, J.W.; Kao, C.M.; Lai, W.L., (2019). Characterization of released metabolic organics during AOC analyses by P17 and NOX strains using 3-D fluorescent signals. Chemosphere. 222: 205-213 (9 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0045653519301201?via=ihub

Hood, E.; Gooseff, M.N.; Johnson, S.L., (2006). Changes in the character of streamwater dissolved organic carbon during flushing in three small watersheds, Oregon. J. Geophys. Res., 111 (G01007): 1-8 (8 pages).

https://andrewsforest.oregonstate.edu/sites/default/files/lter/pubs/pdf/pub4000.pdf

Hounshell, A.G.; Peierls, B.L.; Osburn, C.L.; Paerl, H.W., (2017). Stimulation of phytoplankton production by anthropogenic dissolved organic nitrogen in a coastal plain estuary. Environ. Sci. Technol., 51(22): 13104-13112 (9 pages).

https://pubs.acs.org/doi/10.1021/acs.est.7b03538

Hua, L.C.; Chao, S.J.; Huang, K.; Huang, C.P., (2020). Characteristics of low and high SUVA precursors: Relationships among molecular weight, fluorescence, and chemical composition with DBP formation. Sci. Total Environ., 727: 1-10 (10 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0048969720321549?via=ihub

Hudson, N.; Baker, A.; Reynolds, D., (2007). Fluorescence analysis of dissolved organic matter in natural, waste and polluted waters- a review. River Res. Appl., 23: 631-649 (19 pages). https://onlinelibrary.wiley.com/doi/abs/10.1002/rra.1005

Jaffe, R.; McKnight, D.; Maie, N.; Cory, R.; McDowell, W.H.; Campbell, J.L., (2008). Spatial and temporal variations in DOM composition in ecosystems: The importance of long-term monitoring of optical properties. J. Geo. Res., 113(G04032): 1-15 (15 pages).

https://www.nrs.fs.fed.us/pubs/53685

Jiao, R.; Chow, C.W.K.; Xu, H.; Yang, X.; Wang, D., (2014). Organic removal assessment at full-scale treatment facilities using advanced organic characterization tools. Environ. Sci. Processes Impacts, 16: 2451-2459 (9 pages).

https://pubs.rsc.org/en/content/articlelanding/2014/EM/c4em00227j - !divAbstract

Kelso, J.E.; Baker, M.A., (2020). Organic matter is a mixture of terrestrial, autochthonous, and Wastewater effluent in an urban river. Front. in Environ. Sci., 7(202): 1-16 (16 pages). https://www.frontiersin.org/articles/10.3389/fenvs.2019.00202/full

Kissman, C.E.H.; Williamson, C.E.; Rose, K.C.; Saros, J.E., (2017). Nutrients associated with terrestrial dissolved organic matter drive changes in zooplankton: phytoplankton biomass ratios in an alpine lake. Freshwater Biol., 62(1): 40-51 (12 pages).

https://onlinelibrary.wiley.com/doi/10.1111/fwb.12847

Lai, C.H.; Chou, Y.C.; Yeh, H.H., (2015). Assessing the interaction effects of coagulation pretreatment and membrane material on UF fouling control using HPSEC combined with peak-fitting. J. Member. Sci., 474: 207-214 (8 pages).

https://www.infona.pl/resource/bwmeta1.element.elsevier-6817faf9-8391-3a9f-90a6-8643ec573c04

Liu, Q.; Jiang, Y.; Tian, Y.; Hou, Z.; He, K.; Fu, L.; Xu, H., (2019). Impact of land use on the DOM composition in different seasons in a subtropical river flowing through a region undergoing rapid urbanization. J. Clean. Prod., 212: 1224-1231 (8 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0959652618337326?via%3Dihub

Liu, Q.; Tian, Y.; Liu, Y.; Yu, M.; Hou, Z.; He, K.; Xu, H.; Cui, B.; Jiang, Y., (2021). Relationship between dissolved organic matter and phytoplankton community dynamics in a human-impacted subtropical river. J. Clean. Prod., 289: 1-10 (10 pages).

https://www.sciencedirect.com/science/article/abs/pii/S095965262035188X?via%3Dihub

Mangal, V.; Stock, N.L.; Gueguen, C., (2016). Molecular characterization of phytoplankton dissolved organic matter (DOM) and sulfur components using high resolution orbitrap mass spectrometry. Anal. Bioanal. Chem., 408(7): 1891-1900 (10 pages).

https://link.springer.com/article/10.1007%2Fs00216-015-9295-9

Maie, N.; Boyer, J. N.; Yang, C.; Jaffe, R., (2006). Sp.atial, geomorphological and seasonal variability of CDOM in estuaries of the Florida Coastal Everglades, Hydrobiol., 569: 135-150 (16 pages). <a href="https://link.springer.com/article/10.1007/s10750-006-0128-x">https://link.springer.com/article/10.1007/s10750-006-0128-x</a>

Matilainen, A.; Gjessing, E.T.; Lahtinen, T.; Hed, L.; Bhatnagar, A.; Sillanpää, M., (2011). An overview of the methods used in the characterization of natural organic matter (NOM) in relation to drinking water treatment. Chemosphere. 83(11): 1431-1442 (12 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0045653511000208?via%3Dihub

Moradi, S.; Sawade, E.; Aryal, R.; Chow, C.W.K.; van Leeuwen, J.; Drikas, M.; Cook, D.; Amal, R., (2018). Tracking changes in organic matter during nitrification using fluorescence excitation-emission matrix spectroscopy coupled with parallel factor analysis (feem/parafac). J. Environ. Chem. Eng., 6(1): 1522-1528 (7 pages).

https://www.sciencedirect.com/science/article/abs/pii/S2213343718300757?via%3Dihub

Murphy, K.R.; Stedmon, C.A.; Graeber, D.; Bro, R., (2013). Fluorescence spectroscopy and multi-way techniques. PARAFAC. Anal. Methods, 5(23): 6557-6566 (10 pages).

https://pubs.rsc.org/en/content/articlelanding/2013/ay/c3ay41160e - !divAbstract

Myklestad, S.M., (1995). Release of extracellular products by phytoplankton with special emphasis on polysaccharides. Sci. Total Environ., 165: 155–164 **(10 pages)**.

https://www.sciencedirect.com/science/article/abs/pii/004896979504549G

Nguyen, K.A.; Zhang, H.; Stewart, R.A., (2013). An intelligent pattern recognition model to automate the categorisation of residential water end-use events, J. Environ. Soft., 47: 108-127 (19 pages). https://www.sciencedirect.com/science/article/abs/pii/S1364815213001084?via%3Dihub

Paerl, H.W.; Huisman, J., (2008). Blooms like it hot. Science. 320(5872): 57-58 (2 pages). https://science.sciencemag.org/content/320/5872/57.full

Ramanan, R.; Kim, B.H.; Cho, D.H.; Oh, H.M.; Kim, H.S., (2016). Algae-bacteria interactions: evolution, ecology and emerging applications. Biotech. Adv. 34: 14–29 (16 pages). https://www.sciencedirect.com/science/article/pii/S0734975015300586

Romera-Castillo, C.; Sarmento, H.; Álvarez-Salgado, X.A.; Gasol, J.M.; Marrasé, C., (2011). Net production and consumption of fluorescent colored dissolved organic matter by natural

bacterial assemblages growing on marine phytoplankton exudates. Appl. Environ. Microbiol., 77: 7490–7498 (9 pages).

https://journals.asm.org/doi/10.1128/AEM.00200-11

- Shon, H.K.; Vigneswaran, S.; Snyder, S.A., (2012). Effluent organic matter (EfOM) in wastewater: constituents, effect, and treatment. Crit. Rev. Environ. Sci. Technol., 36(4): 327-374 (48 pages). <a href="https://www.tandfonline.com/doi/abs/10.1080/10643380600580011">https://www.tandfonline.com/doi/abs/10.1080/10643380600580011</a>
- Suksomjit, M.; Nagao, S.; Ichimi, K.; Yamada, T.; Tada, K., (2009). Variation of dissolved organic matter and fluorescence characteristics before, during and after phytoplankton bloom. J. Oceanogr. 65(6): 835-846 (12 pages).

https://link.springer.com/article/10.1007/s10872-009-0069-x

Thornton, D.C.O., (2014). Dissolved organic matter (DOM) released by phytoplankton in the contemporary and future ocean. European. J. Phys., 49: 20–46 **(27 pages)**.

https://www.tandfonline.com/doi/full/10.1080/09670262.2013.875596

Wada, S.; Aoki, M.N.; Tsuchiya, Y.; Sato, T.; Shinagawa, H.; Hama, T., (2007). Quantitative and qualitative analyses of dissolved organic matter released from Ecklonia cava Kjellman, in Oura Bay, Shimoda, Izu Peninsula, Japan. J. Exp. Mar. Biol. Ecol., 349:344–358 (14 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0022098107002699?via%3Dihub

Watson, K.; Farre, M.J.; Leusch, F.D.L.; Knight, N., (2018). Using fluorescence-parallel factor analysis for assessing disinfection by-product formation and natural organic matter removal efficiency in secondary treated synthetic drinking waters. Sci. Total Environ., 640-641: 31-40 (10 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0048969718319338?via=ihub

Weishaar, J.L.; Aiken, G.R.; Bergamaschi, B.A.; Fram, M.S.; Fuji, R.; Mopper, K., (2003). Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. Environ. Sci. Technol., 37(20): 4702–4708 (7 pages).

https://pubs.acs.org/doi/10.1021/es030360x

Yamashita, Y.; Jaffé, R.; Maie, N.; Tanoue, E., (2008). Assessing the dynamics of dissolved organic matter (DOM) in coastal environments by excitation-emission matrix fluorescence and parallel factor analysis (EEM-PARAFAC). Limnol. Ocean., 53: 1900–1908 (9 pages).

https://aslopubs.onlinelibrary.wiley.com/doi/abs/10.4319/lo.2008.53.5.1900

Yao, Y.; Li, Y. Z.; Zhang, Y. P.; Guo, X. J.; Yuan, F., (2016). Changes and characteristics of dissolved organic matter in a constructed wetland system using fluorescence spectroscopy. Environ. Sci. Pollut. Res., 23: 12237–12245 (9 pages).

https://link.springer.com/article/10.1007%2Fs11356-016-6435-5

Zhang, F.; Harir, M.; Moritz, F.; Zhang, J.; Witting, M.; Wu, Y.; Schmitt-Kopplin, P.; Fekete, A.; Gaspar, A.; Hertkorn, N., (2014). Molecular and structural characterization of dissolved organic matter during and post cyanobacterial bloom in Taihu by combination of NMR spectroscopy and FTICR mass spectrometry. Water Res., 57: 280-294 (15 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0043135414001791?via%3Dihub

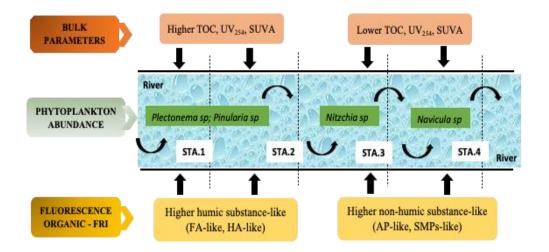
Zhang, Y.; van Dijk, MA; Liu, M.; Zhu, G.; Qin, B., (2009). The contribution of phytoplankton degradation to chromophoric dissolved organic matter (CDOM) in eutrophic shallow lakes: field and experimental evidence. Water Res., 43: 4685-4697 (13 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0043135409004928?via%3Dihub

Ziegman, M.; Abert, M.; Muler, M.; Frimmel, F.H., (2010). Use of fluorescence for the estimation of bloom formation and toxin production of Microcystis aeruginosa. Water Res., 44: 195-204 (10 pages).

https://www.sciencedirect.com/science/article/abs/pii/S0043135409006186?via%3Dihub

## **GRAPHICAL ABSTRACT**



## **HIGHLIGHTS**

- ➤ Characteristic of organic matter and the phytoplankton abundance is different at each segment of surface water;
- There is a substantial correlation among the bulk parameters, fluorescence spectroscopy of DOM and phytoplankton abundance;
- Phytoplankton abundance combined with DOM analysis could be used to evaluate the quantity and quality of organic matter for monitoring surface water quality.

# Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality

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Received: 04 April 2021 Revised: 17 July 2021 Accepted:+ Published: 20 August 2021

#### ABSTRACT:

**BACKGROUND AND OBJECTIVES:** Dissolved organic matter has a fundamental role in supporting phytoplankton abundance and growth in aquatic environments. However, these organisms produce dissolved organic matter with varied quantities or characteristics depending on the nutrient availability and the species composition. Therefore, this study aims to assess the characteristic of dissolved organic matter on surface water and its correlation with phytoplankton abundance for monitoring water quality.

**METHODS:** The sample was obtained at four Kali Surabaya river stations for further dissolved organic matter analysis and phytoplankton species analysis. The analysis was presented through bulk parameters of total organic, ultraviolet at 254 nm wavelength, specific ultraviolet absorbance value, and fluorescence spectroscopy using excitation-emission matrices with fluorescence regional integration analysis.

**FINDINGS:** The results showed the bulk parameters of dissolved organic matter at all stations were significantly different, as Station 1 and 2 were higher, while 3 and 4 had a lower concentration. Furthermore, the fluorescence spectroscopy identified four components of dissolved organic matter at all stations, namely aromatic proteins-like, humic acid-like, soluble microbial byproducts-like, and fulvic acid-like, which is the unit of fluorescence spectra in arbitrary unit. Also, stations 1 and 2 were grouped in the high percentage fluorescence regional integration of humic substance (fulvic acid-like and humic acid-like), while 3 and 4 were classified in the high percentage fluorescence regional integration of non-humic substances (aromatic proteins-like and soluble microbial by-products-like).

**CONCLUSION:** The main phytoplankton species, namely Plectonema sp., Pinularia sp., Nitzchia sp., Navicula sp., had the highest abundance at Stations 1, 3, and 4, respectively. A strong correlation between dissolved organic matter analysis and phytoplankton abundance led to the usage of these methods for monitoring surface water quality.

**KEYWORDS**: Correlation; Dissolved organic matter; Fluorescence spectroscopy; Phytoplankton.

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1

NUMBER OF REFERENCES	NUMBER OF FIGURES	NUMBER OF TABLES
55	8	2

**RUNNING TITLE:** Assessing of characteristic dissolved organic matter and its correlation.

#### INTRODUCTION

Human, industrial and agricultural activities have significantly changed aquatic ecosystems due to high organic and inorganic wastewater discharge. This runoff has appeared in the eutrophication of rivers and tributary (Conley et al., 2009; Bhattacharya and Osburn, 2017) causing blooming phytoplankton and consequently, and the environmental issues (Paerl et al., 2008; Heisler et al., 2008; Biggs, 2000). It is eminent that phytoplankton community dynamics (i.e., taxonomic composition, abundance, and biomass) regard the quantity of inorganic phosphorus and nitrogen in the aquatic surrounding (Cao et al., 2016; Cuvin-Aralar et al., 2004). Furthermore, the impact of the organic pollutants contributes to the quantity or quality of dissolved organic matter in surface water. Allochthonous and autochthonous with effluent organic matter are the source of dissolved organic matter (DOM) in the surface water, since allochthonous could be generated from the upstream, midstream and the downstream. The upstream was found to be covered with perennial vegetation; the midstream is used for agriculture and covered with least forest; the downstream was mainly used for residential and utilized for different forms of agriculture (Dumago et al., 2018). In addition, biogeochemical cycles will affect the quality and quantity of DOM from the surrounding environment. Also, DOM has an essential role in supporting phytoplankton abundance and growth in aquatic surroundings (Kissman et al., 2017; Burpee et al., 2016) due to its usage as an organic nutrient source. It can be used by these micro-organisms as a source of nitrogen, phosphorus, and carbon when inorganic phosphorus and nitrogen are unavailable (Burpee et al., 2016). The primary producers were proposed as an important source that influences its composition in surface water (Biddanda and Benner, 1997). Conversely, DOM can be produced by phytoplankton (Thornton, 2014), with varied characteristics and quantity which are mostly dependent on nutrient availability (Myklestad, 1995), composition of phytoplankton type (Biddanda and Benner, 1997), and bacterial interaction (Ramanan et al., 2016). According to previous studies, various types of DOM have been found and released by different taxonomic groups of phytoplankton (Fukuzaki et al., 2014; Romera-Castillo et al., 2010). Phytoplankton production, microbial metabolism, residue from microbial degradation after their death and other processes, release protein-like materials as one of DOM components (Liu et al., 2019; Mangal et al., 2016). The fluorescence spectroscopy fingerprints, identified the signals of protein-like and humic-like materials released from extracellular Microcystis aeruginosa (Ziegmann et al., 2010). In addition, the DOM which is closely related to the phytoplankton community dynamics, mainly consist of humic-like and protein-like materials (Suksomjit et al., 2009; Zhang et al., 2014) and exhibits their blooming (Altman and Paerl, 2012; Hounshell et al., 2017). The qualitative and quantitative methods for characterizing organic matter analysis have been implemented to clarify the types of DOM transformation through the treatment process or in source water and their following removal. For example, using the bulk parameters of dissolved organic carbon (DOC) concentration, UV/vis at 254 nm wavelength to measure the aromaticity degree of organic matter and specific ultraviolet absorbance (SUVA) (Edzwald et al., 1985; Lai et al., 2015; Hidayah et al., 2017), high-performance size exclusion chromatography (HPSEC) with ultraviolet detector (UVD) or an on-line organic carbon detector (OCD) (Jiao et al., 2014; Lai et al., 2015), fluorescence spectroscopy as well as fluorescence excitation-emission matrices (FEEM) (Hidayah et al., 2017; Ho et al., 2019). These procedures have been previously applied in observing the contribution of phytoplankton degradation to DOM as chromophoric by using fluorescent spectroscopy (Zhang et al., 2009), to characterize DOM excreted by phytoplankton (Chari et al., 2013), and to reveal its relationship with the community (Liu et al., 2021). The use of bulk parameters and fluorescent spectroscopy methods, simultaneously for characterizing organic matter considering the phytoplankton abundance, have been rarely observed. Therefore, resulting in poor implementation of optimal water quality control measures. Furthermore, using these techniques to characterize organic matter and its correlation with phytoplankton abundance for monitoring surface water quality seems to urgently need implementation. Hence, this study aims to assess the characteristic of dissolved organic matter on surface water, as well as its correlation with phytoplankton abundance using the bulk parameters and fluorescence spectroscopy to monitor surface water quality. This study was conducted in the Kali Surabaya River, Surabaya, Indonesia, in 2021.

## **MATERIALS AND METHODS**

#### Data collection

This study used water from the Kali Surabaya River in Surabaya city, a surface water source for public supply. The position of station 1 to station 4 is as shown in Table 1 and Fig. 1. The sample was collected twice per week from January to March 2021, and the DOM analysis, as well as phytoplankton abundance was measured through the bulk parameters and fluorescence spectroscopy. The parameters include TOC,  $UV_{254}$ , SUVA value, while fluorescence spectroscopy identified aromatic proteins-like (AP-like), humic acid-like (HA-like), soluble microbial products-like (SMPs-like), and fulvic acid-like (FA-like). As this study targeted on dissolved organic matter in source water, 0.45 m filter paper was used to filter the collected source water (Millipore Corporation, USA) to eliminate suspended particles before analysis the parameters. Furthermore, the ultraviolet absorbance at 254 nm ( $UV_{254}$ ) and total organic carbon (TOC) concentration of the water was measured for common physicochemical characteristics based on Standard Methods procedures (APHA *et al.*, 2012).

Table 1: The study sampling location characteristics

No.	Sampling station	Coordinate	Climate	<b>Environmental condition</b>
1	Rolag Telu dam	7°26′40″ S 112°27′25″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature</li> <li>29°C</li> </ul>	<ul><li>Downstream of the Brantas river</li><li>Stagnant water</li><li>No residential</li></ul>
2	Wringin Anom district	7°24′21″ S 112°30′27″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature</li> <li>29°C</li> </ul>	<ul><li>Agricultural land</li><li>There are residential</li><li>There are domestic activities (bathing, washing, latrine)</li></ul>
3	Cangkir district	7°22′04″ S 112°37′47″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature</li> <li>29°C</li> </ul>	<ul><li>Industrial area</li><li>Densely populated</li><li>Temporary dump site</li></ul>
4	Karang Pilang drinking water company inlet	7°20′54″ S 112°40′51″ E	<ul> <li>Tropical</li> <li>Sunny weather</li> <li>Temperature</li> <li>29°C</li> </ul>	<ul><li>Industrial area</li><li>There are residential</li><li>There are domestic activities (bathing, washing, latrine)</li></ul>



Fig. 1: Geographic location of the study area in the Kali Surabaya River, Indonesia

TOC was quantified using TOC-500 Model (Shimadzu, Japan), while UV<sub>254</sub> was detected by UV/vis spectrophotometer Model U-2001 (Hitachi, Japan). The SUVA value showed the dissolved organics were contained in hydrophilic fraction as calculated from measurements of UV<sub>254</sub> and DOC samples. Perkin Elmer LS-55 spectrometer with excitation-emission wavelength pair was used to measure the fluorescence in the source water. Moreover, the excitation-emission matrix (EEM) were resulted for each sample by skimming overexcitation (Ex) wavelengths between 230 and 400 nm at an interval of 10 nm with emission (Em) wavelengths between 300 and 547.5 nm at 0.5 nm interval (Murphy *et al.*, 2013; Hidayah *et al.*, 2017). Counting of fluorescence regional integration (FRI) analysis was used to provide the cumulative fluorescence reaction of organic matter with identical characteristic in selected regions by integration beneath EEMs (Chen *et al.*, 2003). The phytoplankton sampling was conducted using a plankton net mesh size 60 mm as much as 100 liters. Meanwhile, its identification was carried out in the laboratory using a binocular microscope with 10 x 10 magnification (AmScope B100B-MS). Also, the abundance was calculated using Sedgewick-Rafter Counting Chamber for three replications (Marienfeld GmbH).

#### Analytical framework

The Kolmogorov-Smirnov, one-way Analysis of Variance (ANOVA), and Pearson correlations were applied utilizing SPSS Statistics 17.0 software (IBM, Armonk, NY, USA). The Kolmogorov-Smirnov test opposed the empirical cumulative distribution function of bulk parameters data and the results of FRI analysis with the distribution expected when the data were standard. When the observed difference is adequately significant, the test will reject the hypothesis of bulk parameters data, the results of FRI analysis data, and phytoplankton abundance normality. However, when the p-value of this test is less than 5%, it can be concluded that the bulk parameters data, the results of FRI analysis data, and phytoplankton abundance are non-normal. The one-way ANOVA was applied to determine whether any statistically significant differences between the means of bulk parameters and the results of FRI data. It was also used to determine at least two groups of the parameters data as the results of FRI analysis were different. In addition, The Pearson correlation coefficients measured the strength of the

linear relationship variables among TOC, UV<sub>254</sub>, SUVA value, AP-like, FA-like, SMPs-like, HA-like, and phytoplankton abundance.

#### **RESULTS AND DISCUSSION**

The bulk parameters of dissolved organic matter in the river segment.

The distribution data for the bulk parameters of dissolved organic matter in the river segment as tested by Kolmogorov–Smirnov showed the TOC concentration (P > 0.15), UV<sub>254</sub> concentration (P > 0.15), and SUVA value (P > 0.15) was normal. Furthermore, the normal distribution data was performed using ANOVA testing to know the differences in mean concentrations of TOC and UV254, as well as SUVA value. ANOVA with the Tukey 95% confidence interval also determined whether there were statistically significant or non-significance differences. The results indicated statistically significant differences in the mean concentration of the bulk parameters among the river segment with a p-value of 0.011, 0.001, and 0.004 in TOC, UV<sub>254</sub>, and SUVA values, respectively. Moreover, enough evidence was provided, which concluded that the average of the bulk organic matter parameters at all stations was significantly different. The Tukey analysis classified the bulk parameters concentration at each station into two main groups. Station 1 and 2 were grouped in the high concentration, while 3 and 4 were classified in the bulk parameters' low concentration, which means the former had averages significantly different from the latter. The average TOC concentration for stations 1 and 2 was about a value 10.1-11.7 mg/L, while 3 and 4 were between 9.8-10.9 mg/L. The average UV<sub>254</sub> concentration for stations 1 and 2 was in the range of 10.1-11.7 mg/L, while 3 and 4 were in between 9.8-10.9 mg/L. The average UV<sub>254</sub> concentration for stations 1 and 2 was in the range of 0.65-0.8/cm, while 3 and 4 were 0.39-0.65/cm. The average SUVA concentration of stations 1 and 2 was 5.3-6.4 L/mg/m, while 3 and 4 were 4.0-5.3 L/mg/m. Furthermore, statistical box plot analysis presented the pattern for the bulk parameters of dissolved organic matter in the surface water. Figs. 2, 3, and 4 show a box plot of the average concentration of TOC, UV<sub>254</sub>, and average SUVA value respectively. Fig. 2 shows that the highest average TOC concentration occurred at Station 2 with a varying range. In comparison, the lowest average TOC concentration with a low range occurred at station 4. In addition, the results showed the average concentration from the highest to the lowest was found at stations 2, 1, 3, and 4. The surface water used in this study contained 7.36 - 15.50 mg/L TOC concentration, which was typically associated with the DOC range. River water has a typical concentration about 2 to 10 mg/L of dissolved organic carbon, which was much higher than groundwater and seawater. Variation in average concentrations of TOC indicated various physical or ecological drivers, chemical processes, spatial changes, which can significantly affect on organic matters dynamics (Maie et al., 2006). The organic matter compositional changes could be induced by biophysical controls, such as changes in composition, which likely result in bioavailability, photoreactivity, nutrient cycling, or chelating capacity and can affect carbon fluxes consequentially ecological drivers not accounted for (Jaffe, 2008). In addition, the hydrology dynamics of surface runoff contributed to the surface water stream (Hood et al., 2006).

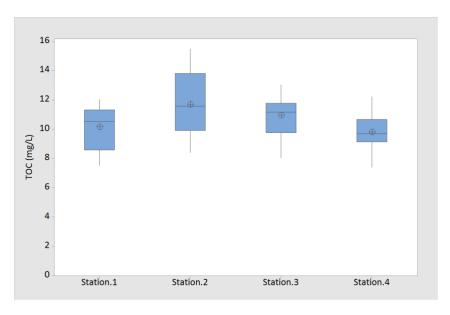


Fig. 2: The average TOC concentration in the river segment at various stations.

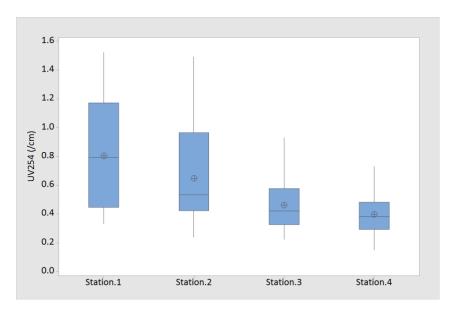


Fig. 3: The average UV<sub>254</sub> concentration in the river segment at various station

Fig. 3 describes the concentration of  $UV_{254}$ , which corresponded to the organic compounds with an aromatic structure, double bonds of C=C (Matilainen *et al.*, 2011). In this study, the concentration of  $UV_{254}$  for surface water was 0.148 - 1.524/cm, which was within the typical range of river (0.085 - 0.4/cm) (Edzwald *et al.*, 1985). The results showed that the average highest aromatic compound was detected at Station 1, while Station 4 had the average lowest concentration. Therefore, Station 1 contained higher humic matter with conjugated C=C double structural bonds than the others. Meanwhile, Station 4 contained lower humic matter than the others. As well known, organic compounds of humic matter contain unsaturated carbon bonds (double or triple) or aromatic rings in their molecular structure. Hence, it absorbs an amount of UV light through the water sample (Her *et al.*, 2002).

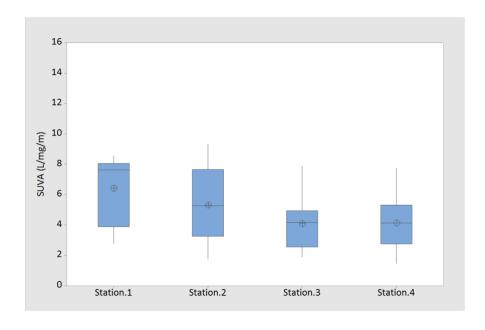


Fig. 4: The average SUVA value in the river segment at various station

Fig. 4. Shows the hydrophobicity of organic matter characteristic or specific UV-absorbance (SUVA) value. The results revealed a value between 1.45 – 9.36 L/mg/m. However, it was mostly higher than 4 along the river segment, which means that the organic matter is mainly consists of humic, hydrophobic, and high molar mass organic material. According to Edzwald and Tobiason (2011), SUVA is a parameter of the organic matter composition in water. Source water with SUVA values ≥ 4 indicated that natural organic matter composed mainly of humic or hydrophobic matter, while those < 2 contained mainly non-humic or hydrophilic natural organic matter. The results were consistent with the high concentration of  $UV_{254}$  (0.148 – 1.524/cm). The values typically ranged from 1.0 to 6.0 L/mg/m for surface water. However, values greater than 6.0 were revealed for interstitial waters dominated by a solid terrestrial signature (Jaffe et al., 2008). According to previous studies, these higher values can be as a result of the absorption at 254 nm from colloids, iron, or other components in the sample (Weishaar et al., 2003; Hudson et al., 2007). Combining the bulk parameters of TOC, UV<sub>254</sub>, and SUVA value led to characterize the organic matter in the river. Station 2 was mainly composed of the highest TOC with lower aromatic and hydrophobic than 1, and vice versa. Also, station 4 was mainly composed of lower bulk parameters than 3. Therefore, 2 contained more aliphatic organic matter that does not absorb at 254 nm than the others. The lower SUVA value among all stations indicated the mixtures of aquatic humics, hydrophobic and hydrophilic, and molecular weights of organic matter.

Characteristic of fluorescence dissolved organic matter in the river segment through volumetric fluorescence distribution.

Fig. 5. Illustrates the fluorescence excitation-emission matrices (FEEMs) for dissolved organic matter in the river segment at a different station, taken on the first week sampling time. Dissolved organic carbon was classified into four regions based on its excitation/emission wavelengths (Ex/Em), namely Region 1 indicated the aromatic proteins-like (AP-like), such as tyrosine and tryptophan, at Ex/Em <250 nm/<350 nm. Region 2 identified the fulvic acid-like (FA-like) substances at Ex/Em <250 nm/>380 nm, Region 3 was corresponded to the soluble microbial by products-like (SMPs-like) substances at Ex/Em 250-280 nm /<380 nm, while Region 4 was identified as the humic acid-like (HA-like) substances with Ex/Em >280 nm/>380 nm (Chen et al., 2003).

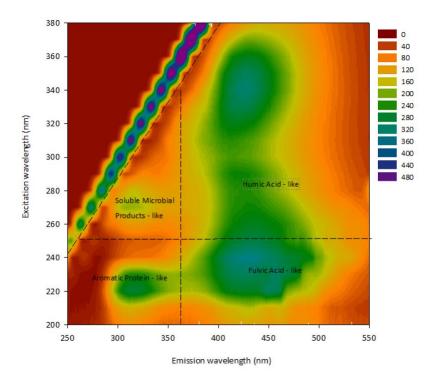


Fig. 5: Spectrum of fluorescence spectrometer analysis in the river segment

This study shows that the fluorescence component from FEEM analysis has consistent results with previous studies (Her et al., 2003; Yao et al., 2016; Moradi et al., 2018; Hidayah et al., 2020). Generally, HA-like and FA-like correlated with aromatic compounds. They mainly exist as carboxylic and phenolic functional groups in natural dissolved organic matter. These fluorescence structures are mostly present as a significant percentage of humic substances, which typically represent over 50% of natural organic matter (Shon et al., 2012). In addition, source water may contain protein-like materials which microbial activities can generate. The amount, characteristics, and properties of dissolved organic matter in the aquatic system depend on their origin and environmental biochemical cycles. Sources of organic matter are classified as allochthonous (generated from a terrestrial watershed) and autochthonous (produced by organism activities, such as phytoplankton activities) (Chari et al., 2013; Zhang et al., 2009; Haraguchi et al., 2019). Terrestrial watershed is mainly composed of humic substances such as fulvic and humic acids as well as humin, which are primarily hydrophobic and rich in aromatic carbon. The autochthonous source material is microbially derived organic, such as algalderived and effluent organic matter (Kelso and Baker, 2020). Fig. 6 showed the percentage fluorescence response, which was calculated by Fluorescence Regional Integration (FRI) method. The percentage of fluorescence distribution indicated the four fractions quantity of fluorescence organic matter. This study classified the fraction into humic and non-humic substances-like. The first was represented by Region 2 (FA-like) and 4 (HA-like), while the second one by Region 1 (AP-like) and 3 (SMPs-like). Firstly, the results showed the highest total percentage of FRI in Region 2 and 4 was at Station 1 (76.6%), and the lowest total percentage for humic substances-like was at Station 4 (69.2%). Both components are classified as humic substances and are mainly composed of aromatic compounds with high to medium molecular weight (Watson et al., 2018; Hua et al., 2020). Their total percentage FRI showed a consistent UV<sub>254</sub> concentration and SUVA value. Furthermore, Station 1 had the highest bulk parameters, while 4 had the lowest. Secondly, the highest total percentage FRI of Region 1 and 3 (30.8%) was identified at Station 4, with the lowest at 1 (23.4%). This indicated that Station 4 contained abundant proteins substances and microbial-like fluorescence than the others and followed the lowest SUVA value of Station 4 with the highest for Station 1. Region 1 and 3 correlated with high molecular weight protein-like, which had chemical properties related to aromatic amino acids, tryptophan or tyrosine-like (Yamashita et al., 2008; Hua et al., 2020) and low molecular weight microbial humic-like as well as less conjugated double bond organic matter (Nguyen et al., 2013; Hua et al., 2020).

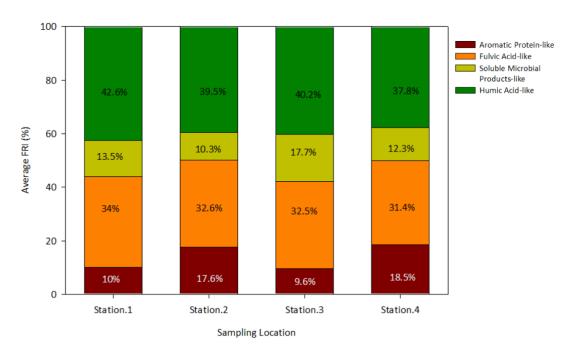


Fig. 6: FRI distribution of fractionated organic matter from the various river segment

The distribution data for the fluorescence of dissolved organic matter in the river segment was tested by Kolmogorov-Smirnov and the bulk parameters. The results showed distribution data for percentage FRI of Region 1 (AP-like), 2 (FA-like), 3 (SMPs-like), and Region 4 (HA-like) with P > 0.000, 0.007, 0.000 and 0.013 respectively were normal. Furthermore, Analysis of Variance (ANOVA) testing was carried out to determine the differences in mean percentage FRI for each region. The statistical analysis ANOVA One-Way with the Tukey 95% confidence interval also determined whether statistically significant or non-significance differences in percentage FRI of AP-like, FA-like, SMPs-like, and HA-like among all stations. The results showed statistically significant differences in the mean percentage FRI of all fluorescence organic fractions at all stations with p = 0.000, 0.007, 0.000, and 0.013 in AP-like, FA-like, SMPs-like, HA-like, respectively. The results provided enough evidence to conclude that the mean percentage FRI of all fluorescence organic fractions at all stations was significantly different. Moreover, the Tukey analysis classified their percentage FRI at each station into two main groups. Station 1 and 2 were grouped in the high percentage FRI of humic substance-like (FA-like and HA-like), while 3 and 4 were classified in the low percentage. This means the former had an average percentage FRI of FA-like and HA-like, which were significantly different from the latter. In addition, stations 3 and 4 were grouped in the high percentage FRI of non-humic substance-like (AP-like and SMPs-like), while stations 3 and 4 were grouped in the low percentage. This showed both had average percentage FRI of AP-like and SMPs-like, which were significantly different from stations 1 and 2. Moreover, statistical box plot analysis presented the pattern of the fluorescence organic matter in the river segments. Fig. 7a to 7d presented box plot with average percentage FRI of the organic matter. Firstly, a comparison among all fluoresces organic compounds showed the average FRI of HA-like was much higher and much lower for SMPs-like than the others. However, HA-like, located at Region 4 of the fluorescence spectra, had the most extensive range of excitation and emission wavelengths. Therefore, the humic acid substances-like region had the most extensive volume distribution of FRI when compared to others (Chen et al., 2003). Meanwhile, SMPs-like or Region 3 comprised a dominant percentage of the fluorescence in wastewater treatment plant effluent (Chen et al., 2003) and was closely related to the phytoplankton activities (Liu et al., 2021; Hua et al., 2020). Second, the average percentage FRI of the organic matter indicated different quantities and quality at each station. The non-humic substanceslike fluorescence as presented by AP-like, SMPs-like, with statistical analysis, had a higher percentage FRI at stations 3 and 4 than the others. It was likely that Station 3 and 4 had a higher percentage of extracellular biological organic matter fraction than the other river segments. The fraction was supposed to contain soluble microbial products of amino acids and carbohydrates. Tryptophan and tyrosine which are aromatic amino acids, were confirmed as biological activity products in natural systems and exhibited a distribution of fluorescence response similar to AP-like and SMPs-like of this study (Coble 2007; Determann et al., 1998). The humic substances-like fluorescence as presented by FA-like and HA-like had a higher percentage FRI at stations 1 and 2 than others and were tested by ANOVA One-Way. Combining the bulk parameters of TOC, UV254, SUVA value, and fluorescence spectroscopy convinced the characteristic of organic matter in the river. Station 1 and 2 had high UV<sub>254</sub> concentration, SUVA value, high percentage FRI of FA-like and HA-like substances. It was conjectured that stations 1 and 2 were mainly composed of aromatic, hydrophobic, humic substances organic matter, which may be generated from terrestrial systems.

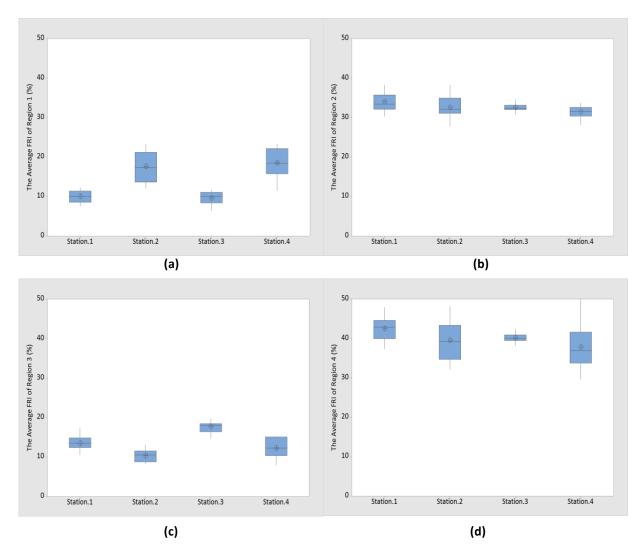


Fig. 7: The average percentage FRI of fluorescence organic matter in the river segment at various stations

Station 3 and 4 had lower UV concentration and SUVA values, with a high percentage FRI of AP-like and SMPs-like than the others. There was a lower SUVA value among all stations indicates in the mixtures of aquatic humics, hydrophobic and hydrophilic, as well as molecular weights of organic matter. This showed that Station 3 and 4 comprised more autochthonous and sources of organic matter from anthropogenic activities. The river ecosystem, which source is terrestrial, autochthonous, and anthropogenic, provided hotspots for storing, transporting, and transforming organic matter. The sources proportions were primarily and terrestrially derived with increased autochthonous inputs from macrophytes. In addition, the sources of dissolved organic matter are a mixture of terrestrial, autochthonous, or primarily from wastewater effluent (Kelso and Baker, 2020).

Contribution of phytoplankton abundance to fluorescence dissolved organic matter in the river segment.

This study discovered four main phytoplankton species with various abundance in the river segments, namely Plectonema sp., Nitzchia sp., Navicula sp., and Pinularia sp. The distribution data of the phytoplankton abundance in this segment was tested by Kolmogorov–Smirnov. The results showed a usual distribution data for Plectonema sp., Nitzchia sp., Navicular sp., and Pinularia sp. as abundance P > 0.000, 0.007, 0.000, and 0.013, respectively. Furthermore, ANOVA testing was carried out to determine the differences in the mean phytoplankton abundance of the river segments. The statistical analysis ANOVA One-Way with the Tukey 95% confidence interval determined whether there were

statistically significant or non-significance differences in the abundance of the species among all stations. According to the results, there were statistically significant differences in the mean abundance of phytoplankton at all stations with p-value = 0.006 and 0.01 in Plectonema sp. and Nitzchia sp. abundance, respectively. Meanwhile, the analysis generated p-value = 0.156 and 0.412 for Navicula sp. and Pinularia sp. abundance, respectively, therefore, classified as only one group of phytoplankton abundance. This showed that there were non-significantly differences in both species abundance among all stations. The Tukey analysis classified Plectonema sp. and Nitzchia sp. abundance at each station into two main groups. Station 1 and 2 were grouped in the high Plectonema sp. and low Nitzchia sp. abundance, while Station 3 and 4 were classified in the low Plectonema sp. and high Nitzchia sp. abundance. Furthermore, the statistical box plot analysis presented the pattern of the phytoplankton abundance in the river segments.

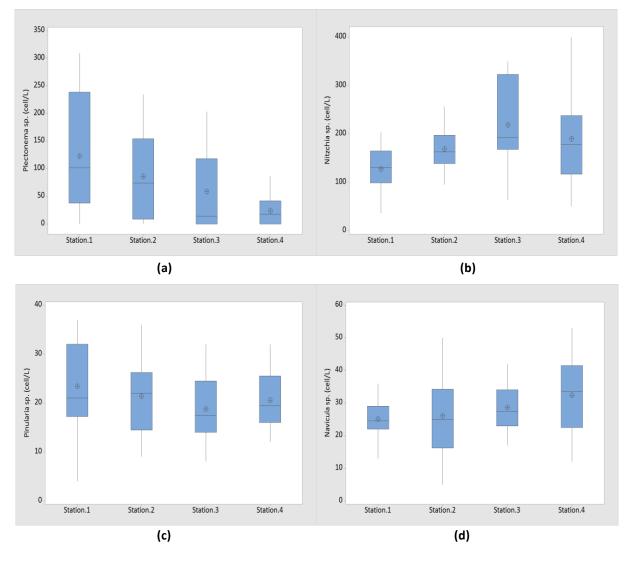


Fig. 8: The average of phytoplankton abundance in the river segment at various station

Fig. 8a to 8d present box plots of their average abundance. Firstly, a comparison among the species at all stations conjectured that Nitzchia sp. had a higher abundance, and Pinularia sp. was lower than the others. Meanwhile, Plectonema sp. had the highest at Station 1 and the lowest at 4. Nitzchia sp. had a higher abundance at Station 3 and lower at 1. Moreover, Navicula sp. had the highest abundance at Station 4 and the lowest at Station 1. Pinnularia sp. gave the highest at Station 1, with the lowest at Station 3. This phytoplankton abundance was strongly influenced by migration, which can occur due to population density and physical environmental conditions, such as changes in temperature and currents (Basu and Mackey, 2018). Secondly, Station 1 was likely to contain a similar abundance in Plectonema sp. and Nitzchia sp., and the same for Navicula sp. and Pinularia sp. Stations 2 and 3 showed that the abundance of Nitzchia sp. was primarily dominant than others. However, Navicular sp. was similar to Pinularia sp. Station 4 identified a similar abundance of Plectonema sp., Navicular sp., and Pinularia sp. There is competition in several phytoplankton species that use the same resource lacking in availability, or even regardless of sufficient availability, and competition still occurs when they take advantage of the resource, with one attacking the other or vice versa (Burson *et al.*, 2018).

The relationship among the bulk parameters, organic fluorescence parameters, and phytoplankton abundance

The degree correlation between the bulk parameters, fluorescence organic matter, and phytoplankton abundance was examined, as shown in Table 2. Correlation analysis was carried out using TOC and

 $UV_{254}$  concentrations, SUVA value with percentage FRI of AP-like, FA-like, SMPs-like, or HA-like, as well as the abundance of Plectonema sp., Nitzchia sp., Navicula sp., and Pinularia sp. Firstly, based on the correlations of the bulk parameters, TOC concentration was positively higher with Region 1 (AP-like) and Region 2 (FA-like). In addition,  $UV_{254}$  concentration and SUVA value were significantly correlated with Region 1(AP-like) and Region 4 (HA-like). The results showed fluorescence spectroscopy, which fractionated AP-like, FA-like, SMPs-like, and HA-like could be used to identify the quantity and quality of organic matter in the source water.

Table 2: The degree correlation among the bulk parameters, fluorescence organic matter, and phytoplankton abundance\*

Parameters	тос	UV <sub>254</sub>	SUVA	AP- like	FA- like	SMPs -like	HA- like	Navicula sp.	Plectone ma sp.	Pinnularia sp.
UV <sub>254</sub>	0.085 0.502			•	•	•		· · ·	·	·
SUVA	-0.044 0.729	0.887 0.000								
AP-like	-0.287 0.022	-0.440 0.000	-0.373 0.002							
FA-like	0.254 0.042	0.105 0.411	0.047 0.710	-0.249 0.048						
SMPs-like	-0.038 0.764	-0.198 0.116	-0.228 0.070	0.638 0.000	-0.085 0.505					
HA-like	-0.035 0.786	0.344 0.005	0.344 0.005	-0.674 0.000	-0.022 0.862	-0.348 0.005				
Navicula sp.	-0.109 0.392	-0.331 0.007	-0.289 0.021	0.193 0.126	-0.102 0.422	0.090 0.480	-0.082 0.521			
Plectonema sp.	0.271 0.030	0.137 0.281	0.131 0.303	-0.346 0.005	0.293 0.019	-0.057 0.652	0.110 0.386	0.166 0.189		
Pinnularia sp.	-0.097 0.448	-0.239 0.058	-0.292 0.470	-0.245 0.051	0.142 0.263	-0.268 0.032	0.220 0.080	0.137 0.279	0.320 0.010	
Nitzchia sp.	-0.243 0.053	-0.283 0.023	-0.203 0.108	0.160 0.205	-0.203 0.107	0.176 0.164	0.070 0.585	0.168 0.184	0.035 0.785	0.174 0.170

\*Cell Contents description; Pearson correlation (the first row of the number of correlation between parameters); P-value (the second row of the number of correlation between parameters)

This result was expected since TOC measured all organic carbon, including humic and non-humic substances, as presented by AP-like and FA-like. Secondly, a strong positive correlation between UV<sub>254</sub> concentration and SUVA value indicated that higher aromatic conjugated double bond corresponded to higher molecular weight organic, more hydrophobic, and content of humic substances. These results are consistent with the Pearson correlation between bulk parameters of UV<sub>254</sub> correlation, SUVA, and fluorescence organic matters of AP-like and HA-like. Furthermore, it was conjectured that fluorescence spectroscopy could be used to assess the properties of organic matter existing in the source water. Thirdly, the results showed that TOC had a stronger correlation with AP-like than HA-like. This was probably because the humic structure may incorporate protein-like-fluorophores due to weak interactions based on x-x or van der Waals forces between the dissolved organic matter components. Previous studies indicated that proteins and humic supramolecules containing specific structures attained from phenol or aniline might contribute to the fluorescence. Fourth, this study discovered a strong correlation between DOM and phytoplankton abundance. Plectonema sp. correlated with TOC, AP-like, and FA-like, while Navicula sp. and Nitzchia sp. correlated with UV<sub>254</sub>, and Pinularia sp. with SMPs-like. The existence of phytoplankton was likely to enhance the quantity and characteristics of DOM in the aquatic environment. The production of marine-like fluorophores accompanied phytoplankton degradation as a significant source of autochthonous DOM (Wada et al., 2007). In addition, higher molecular weight compounds such as protein (tryptophan)-like fluorescence were presented in exudates when phytoplankton grows (Chari et al., 2013). The combination of the bulk parameters (TOC, UV<sub>254</sub>, and SUVA value), fluorescence spectroscopy, and phytoplankton abundance convinced the quality of organic matter in the surface water. However, it could be eventually used to monitor the water's quality.

#### **CONCLUSION**

This study showed that the quality and quantity of DOM at all stations were significantly different, as classified into two groups with higher bulk parameters at stations 1 and 2 and a lower concentration at 3 and 4. The average TOC concentration for stations 1 and 2 was about a value 10.1-11.7 mg/L, while 3 and 4 were in between 9.8-10.9 mg/L. The average UV<sub>254</sub> concentration for stations 1 and 2 was in the range of 10.1-11.7 mg/L, while 3 and 4 were between 9.8-10.9 mg/L. The average UV<sub>254</sub> concentration for stations 1 and 2 was 0.65-0.8/cm, while 3 and 4 were 0.39-0.65/cm. The average SUVA concentration of stations 1 and 2 was in the range 5.3-6.4 L/mg/m, while 3 and 4 were 4.0-5.3 L/mg/m. In addition, fluorescence spectroscopy with FRI analysis showed stations 1 and 2 were grouped in the high percentage FRI of humic substance-like (FA-like and HA-like) about 74.35%. It was conjectured that stations 1 and 2 were mainly composed of aromatic, hydrophobic, humic substances organic matter, which may be generated from terrestrial systems, while stations 3 and 4 were classified in high percentages non-humic substances-like (AP-like and SMPs-like) about 29.05%. This showed that Station 3 and 4 comprised more autochthonous and sources of organic matter from anthropogenic activities. According to phytoplankton abundance, Station 1 had a high abundance of Plectonema sp. (238.5 cell/L) and Pinularia sp. (32 cell/L), while stations 2 and 3 mainly consisted of Nitzchia sp. (197.5 cell/L and 322.75 cell/L), and Navicula sp. (41.5 cell/L) was dominant at Station 4. The Pearson correlation showed a strong relationship between DOM and phytoplankton abundance. Therefore, Plectonema sp. was in correlation with TOC (0.271), AP-like (-0.346), and FA-like (0.293), while Navicula sp. and Nitzchia sp. correlated with UV<sub>254</sub> (-0.331 and -0.283), and Pinularia sp. correlated with SMPslike (-0.268). This study conjectured that the bulk parameters of DOM, fluorescence spectroscopy, and phytoplankton abundance could be used to assess the characteristic of DOM, while the combination of these methods could be used to monitor the surface water quality. Future work should be conducted on the laboratory scale for phytoplankton observation in order to identify the characteristic of organic matter that a kind of phytoplankton species has released. Therefore, it could be used to predict the amount of DOM derived by phytoplankton, DOM derived in the aquatic, and DOM from the terrestrial watershed.

#### **AUTHOR CONTRIBUTIONS**

O.H. Cahyonugroho performed the experimental design, analyzed the data, and prepared the manuscript text as well as the literature review. S. Hariyanto and G. Supriyanto interpreted the data and helped in manuscript preparation.

#### **ACKNOWLEDGEMENT**

This study was supported by the Directorate of Research and Community Service, Deputy for Strengthening Research and Development of the Ministry of Research and Technology/National Research and Innovation Agency through Postgraduate Research – Doctoral Dissertation Research Scheme for multi-year, with Contract No. [489/UN3.15/PT/2021].

#### CONFLICT OF INTEREST

The authors declare no potential conflict of interest regarding the publication of this work. Also, ethical issues, including plagiarism, informed consent, misconduct, data fabrication and/or falsification, double publication and submission, as well as redundancy, have been entirely witnessed by the authors.

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## **ABBREVIATIONS**

% Percent
/cm Per centimeter

ANOVA Analysis of variance
AP-like Aromatic proteins-like
C=C Carbon chain double bonds

cell/L The number of phytoplankton cell per liter

DOCDissolved organic carbonDOMDissolved organic matterEmEmission wavelengthExExcitation wavelength

FA-like Fulvic acid-like

FEEM Fluorescence spectroscopy using excitation-emission matrices

FRI Fluorescence regional integration

*HA-like* Humic acid-like

*L/mg/m* Liter per miligrams per meter

mg/L Miligrams per liter

mm Milimeter
 μm Micrometer
 Navicula sp. Navicula species
 Nitzchia sp. Nitzchia species

NOM Natural organic matter

*nm* Nanometer

OCD Organic carbon detector
 P > Probability value more than
 P = Probability value equal
 Pinnularia sp.
 Plectonema sp.
 Plectonema species
 P-value
 Probability value

SMPs-like Soluble microbial by products-like SUVA Specific ultraviolet absorbance

TOC Total organic carbon

*UV*<sub>254</sub> Ultraviolet at 254 nm wavelength

UVD Ultraviolet detector UV/vis Ultraviolet visible

#### **REFERENCES**

- Altman, J.C.; Paerl, H.W., (2012). Composition of inorganic and organic nutrient sources influences phytoplankton community structure in the New River Estuary, North Carolina. Aquat. Ecol., 46(3): 269-282 (14 pages).
- APHA, (2012). Standard Methods for the Examination of Water and Wastewaters, 21<sup>th</sup> ed. American Public Health Association: Washington, DC.
- Basu, S.; Mackey, K.R.M., (2018). Phytoplankton as key mediators of the biological carbon pump: their responses to a changing climate. J. Sustainability. 10(3): 1-18 (18 pages).
- Bhattacharya, R.; Osburn, C.L., (2017). Multivariate analyses of phytoplankton pigment fluorescence from a freshwater river network. Environ. Sci. Technol., 51(12): 6683-6690 (8 pages).
- Biddanda, B.; Benner, R., (1997). Carbon, nitrogen, and carbohydrate fluxes during the production of particulate and dissolved organic matter by marine phytoplankton. Limnol. Ocean., 42: 506–518 (13 pages).
- Biggs, B.J.F., (2000). Eutrophication of streams and rivers: dissolved nutrient chlorophyll relationships for benthic algae. J. North Am. Benthol. Soc., 19(1): 17-31 (15 pages).
- Burpee, B.; Saros, JE; Northington, R.M.; Simon, K.S., (2016). Microbial nutrient limitation in Arctic lakes in a permafrost landscape of southwest Greenland. Biogeosci., 13(2): 365-374 (10 pages).
- Burson A.; Stomp M.; Greenwell E.; Grosse J.; Huisman J. (2018). Competition for nutrients and light: testing advances in resource competition with a natural phytoplankton community. J. Ecol. 99(5): 1108 1118 (11 pages).
- Cao, X.; Wang, J.; Liao, J.; Sun, J.; Huang, Y., (2016). The threshold responses of phytoplankton community to nutrient gradient in a shallow eutrophic Chinese lake. Ecol. Indicat., 61: 258-267 (10 pages).
- Chari, N.V.H.K.; Keerthi, S.; Sarma, N.S.; Pandi, S.R.; Chiranjeevulu, G.; Kiran, R.; Koduru, U., (2013). Fluorescence and absorption characteristics of dissolved organic matter excreted by phytoplankton species of western Bay of Bengal under axenic under axenic laboratory condition. J. Exp. Mar. Biol. Ecol., 445: 148-155 (8 pages).
- Chen, W.; Westerhoff, P.; Leenheer, J.A.; Booksh, K., (2003). Fluorescence excitation-emission matrix regional integration to quantify spectra for dissolved organic matter. J. Environ. Sci. Technol., 37: 5701-5710 (10 pages).
- Coble, P.G., (2007). Marine chemical biogeochemistry: the chemistry of ocean color. Chem. Rev., 107: 402-418 (17 pages).
- Conley, D.J.; Paerl, H.W.; Howarth, R.W.; Boesch, D.F.; Seitzinger, S.P.; Havens, K.E.; Lancelot, C.; Likens, G.E., (2009). Controlling eutrophication: nitrogen and phosphorus. J. Sci, 323(5917): 1014-1015 (2 pages).
- Cuvin-Aralar, M.L.; Focken, U.; Becker, K.; Aralar, E.V., (2004). Effects of low nitrogen phosphorus ratios in the phytoplankton community in Laguna de Bay, a shallow eutrophic lake in the Philippines. Aquat. Ecol. 38(3): 387-401 (15 pages).
- Determann, S.; Lobbes, J.M.; Reuter, R.; Rullko tter, J., (1998). Ultraviolet fluorescence excitation and emission spectroscopy of marine algae and bacteria. Mar. Chem., 62(1–2): 137–156 (19 pages).
- Dumago, S.W.L.; Puno, G.R.; Ingotan, S.S., (2018). Water quality assessment in various land use and land cover of Muleta Watershed Bukidnon, Philippines. J. Bio. Environ. Sci., 112(3): 201-209 (8 pages).
- Edzwald, J.K.; Becker, W.C.; Wattier, K.L., (1985). Surrogate parameters for monitoring organic matter and THM precursors. J. Am. Water Works Assn., 77(4): 122–131 (10 pages).
- Edzwald, J.K.; Tobiason, J.E., (2011). Chemical principles, source water composition, and watershed protection. In Water Quality & Treatment: A handbook on drinking water; Edzwald, J.K., Eds. New York: AWWA McGraw-Hill.
- Fukuzaki, K.; Imai, I.; Fukushima, K.; Ishii, K.I.; Sawayama, S.; Yoshioka, T., (2014). Fluorescent characteristics of dissolved organic matter produced by bloom-forming coastal phytoplankton. J. Plankton. Res., 36: 685–694 **(10 pages)**.

- Haraguchi, L.; Asmala, E.; Jakobsen, H.H.; Carstensen, J., (2019). Composition of natural phytoplankton community has minor effects on autochthonous dissolved organic matter characteristics. Mar. Biol. Res., 1-19 (19 pages).
- Heisler, J.; Glibert, P.M.; Burkholder, J.M.; Anderson, D.M.; Cochlan, W.; Dennison, W.C.; Dortch, Q.; Gobler, C.J.; Heil, C.A.; Humphries, E.; Lewitus, A.; Magnien, R.; Marshall, H.G.; Sellner, K.; Stockwell, D.A.; Stoecker, D.K.; Suddleson, M., (2008). Eutrophication and harmful algal blooms: a scientific consensus. Harmful Algae. 8(1): 3-13 (11 pages).
- Her, N.; Amy, G.; Foss, D.; Cho, J.; Yoon, Y.; Kosenka, P., (2002). Optimization of method for detecting and characterizing NOM by HPLC-size exclusion chromatography with UV and on-line DOC detection. Environ. Sci. Technol., 36: 1069–1076 (8 pages).
- Her, N.; Amy, G.; McKnight, D.; Sohn, J.; Yoon, Y., (2003). Characterization of DOM as a function of MW by fluorescence EEM and HPLC-SEC using UVA, DOC, and fluorescence detection. Water Res., 37: 4295-4303 (9 pages).
- Hidayah, E.N.; Chen, Y.C.; Yeh, H.H., (2017). Comparison between HPSEC-OCD and F-EEMs for assessing DBPs formation in water. J. Environ. Sci. Health. Part A., 52(4): 391-402 (12 pages).
- Hidayah, E.N.; Pachwarya, R.B.; Cahyonugroho, O.H.; Ramanathan, A.L., (2020). Characterization of molecular weight-based fluorescent organic matter and its removal in combination of constructed wetland with activated sludge process. Water Air Soil Pollut., 231(41): 1-12 (12 pages).
- Ho, H.H.; Cao, J.W.; Kao, C.M.; Lai, W.L., (2019). Characterization of released metabolic organics during AOC analyses by P17 and NOX strains using 3-D fluorescent signals. Chemosphere. 222: 205-213 (9 pages).
- Hood, E.; Gooseff, M.N.; Johnson, S.L., (2006). Changes in the character of streamwater dissolved organic carbon during flushing in three small watersheds, Oregon. J. Geophys. Res., 111 (G01007): 1-8 (8 pages).
- Hounshell, A.G.; Peierls, B.L.; Osburn, C.L.; Paerl, H.W., (2017). Stimulation of phytoplankton production by anthropogenic dissolved organic nitrogen in a coastal plain estuary. Environ. Sci. Technol., 51(22): 13104-13112 (9 pages).
- Hua, L.C.; Chao, S.J.; Huang, K.; Huang, C.P., (2020). Characteristics of low and high SUVA precursors: Relationships among molecular weight, fluorescence, and chemical composition with DBP formation. Sci. Total Environ., 727: 1-10 (10 pages).
- Hudson, N.; Baker, A.; Reynolds, D., (2007). Fluorescence analysis of dissolved organic matter in natural, waste and polluted waters- a review. River Res. Appl., 23: 631-649 (19 pages).
- Jaffe, R.; McKnight, D.; Maie, N.; Cory, R.; McDowell, W.H.; Campbell, J.L., (2008). Spatial and temporal variations in DOM composition in ecosystems: The importance of long-term monitoring of optical properties. J. Geo. Res., 113(G04032): 1-15 (15 pages).
- Jiao, R.; Chow, C.W.K.; Xu, H.; Yang, X.; Wang, D., (2014). Organic removal assessment at full-scale treatment facilities using advanced organic characterization tools. Environ. Sci. Processes Impacts, 16: 2451-2459 (9 pages).
- Kelso, J.E.; Baker, M.A., (2020). Organic matter is a mixture of terrestrial, autochthonous, and Wastewater effluent in an urban river. Front. in Environ. Sci., 7(202): 1-16 (16 pages).
- Kissman, C.E.H.; Williamson, C.E.; Rose, K.C.; Saros, J.E., (2017). Nutrients associated with terrestrial dissolved organic matter drive changes in zooplankton: phytoplankton biomass ratios in an alpine lake. Freshwater Biol., 62(1): 40-51 (12 pages).
- Lai, C.H.; Chou, Y.C.; Yeh, H.H., (2015). Assessing the interaction effects of coagulation pretreatment and membrane material on UF fouling control using HPSEC combined with peak-fitting. J. Member. Sci., 474: 207-214 (8 pages).
- Liu, Q.; Jiang, Y.; Tian, Y.; Hou, Z.; He, K.; Fu, L.; Xu, H., (2019). Impact of land use on the DOM composition in different seasons in a subtropical river flowing through a region undergoing rapid urbanization. J. Clean. Prod., 212: 1224-1231 (8 pages).
- Liu, Q.; Tian, Y.; Liu, Y.; Yu, M.; Hou, Z.; He, K.; Xu, H.; Cui, B.; Jiang, Y., (2021). Relationship between dissolved organic matter and phytoplankton community dynamics in a human-impacted subtropical river. J. Clean. Prod., 289: 1-10 (10 pages).

- Mangal, V.; Stock, N.L.; Gueguen, C., (2016). Molecular characterization of phytoplankton dissolved organic matter (DOM) and sulfur components using high resolution orbitrap mass spectrometry. Anal. Bioanal. Chem., 408(7): 1891-1900 (10 pages).
- Maie, N.; Boyer, J. N.; Yang, C.; Jaffe, R., (2006). Sp.atial, geomorphological and seasonal variability of CDOM in estuaries of the Florida Coastal Everglades, Hydrobiol., 569: 135-150 (16 pages).
- Matilainen, A.; Gjessing, E.T.; Lahtinen, T.; Hed, L.; Bhatnagar, A.; Sillanpää, M., (2011). An overview of the methods used in the characterization of natural organic matter (NOM) in relation to drinking water treatment. Chemosphere. 83(11): 1431-1442 (12 pages).
- Moradi, S.; Sawade, E.; Aryal, R.; Chow, C.W.K.; van Leeuwen, J.; Drikas, M.; Cook, D.; Amal, R., (2018). Tracking changes in organic matter during nitrification using fluorescence excitation-emission matrix spectroscopy coupled with parallel factor analysis (feem/parafac). J. Environ. Chem. Eng., 6(1): 1522-1528 (7 pages).
- Murphy, K.R.; Stedmon, C.A.; Graeber, D.; Bro, R., (2013). Fluorescence spectroscopy and multi-way techniques. PARAFAC. Anal. Methods, 5(23): 6557-6566 (10 pages).
- Myklestad, S.M., (1995). Release of extracellular products by phytoplankton with special emphasis on polysaccharides. Sci. Total Environ., 165: 155–164 **(10 pages)**.
- Nguyen, K.A.; Zhang, H.; Stewart, R.A., (2013). An intelligent pattern recognition model to automate the categorisation of residential water end-use events, J. Environ. Soft., 47: 108-127 (19 pages).
- Paerl, H.W.; Huisman, J., (2008). Blooms like it hot. Science. 320(5872): 57-58 (2 pages).
- Ramanan, R.; Kim, B.H.; Cho, D.H.; Oh, H.M.; Kim, H.S., (2016). Algae-bacteria interactions: evolution, ecology and emerging applications. Biotech. Adv. 34: 14–29 (16 pages).
- Romera-Castillo, C.; Sarmento, H.; Álvarez-Salgado, X.A.; Gasol, J.M.; Marrasé, C., (2011). Net production and consumption of fluorescent colored dissolved organic matter by natural bacterial assemblages growing on marine phytoplankton exudates. Appl. Environ. Microbiol., 77: 7490–7498 (9 pages).
- Shon, H.K.; Vigneswaran, S.; Snyder, S.A., (2012). Effluent organic matter (EfOM) in wastewater: constituents, effect, and treatment. Crit. Rev. Environ. Sci. Technol., 36(4): 327-374 (48 pages).
- Suksomjit, M.; Nagao, S.; Ichimi, K.; Yamada, T.; Tada, K., (2009). Variation of dissolved organic matter and fluorescence characteristics before, during and after phytoplankton bloom. J. Oceanogr. 65(6): 835-846 (12 pages).
- Thornton, D.C.O., (2014). Dissolved organic matter (DOM) released by phytoplankton in the contemporary and future ocean. European. J. Phys., 49: 20–46 (27 pages).
- Wada, S.; Aoki, M.N.; Tsuchiya, Y.; Sato, T.; Shinagawa, H.; Hama, T., (2007). Quantitative and qualitative analyses of dissolved organic matter released from Ecklonia cava Kjellman, in Oura Bay, Shimoda, Izu Peninsula, Japan. J. Exp. Mar. Biol. Ecol., 349:344–358 (14 pages).
- Watson, K.; Farre, M.J.; Leusch, F.D.L.; Knight, N., (2018). Using fluorescence-parallel factor analysis for assessing disinfection by-product formation and natural organic matter removal efficiency in secondary treated synthetic drinking waters. Sci. Total Environ., 640-641: 31-40 (10 pages).
- Weishaar, J.L.; Aiken, G.R.; Bergamaschi, B.A.; Fram, M.S.; Fuji, R.; Mopper, K., (2003). Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. Environ. Sci. Technol., 37(20): 4702–4708 (7 pages).
- Yamashita, Y.; Jaffé, R.; Maie, N.; Tanoue, E., (2008). Assessing the dynamics of dissolved organic matter (DOM) in coastal environments by excitation-emission matrix fluorescence and parallel factor analysis (EEM-PARAFAC). Limnol. Ocean., 53: 1900–1908 (9 pages).
- Yao, Y.; Li, Y. Z.; Zhang, Y. P.; Guo, X. J.; Yuan, F., (2016). Changes and characteristics of dissolved organic matter in a constructed wetland system using fluorescence spectroscopy. Environ. Sci. Pollut. Res., 23: 12237–12245 (9 pages).
- Zhang, F.; Harir, M.; Moritz, F.; Zhang, J.; Witting, M.; Wu, Y.; Schmitt-Kopplin, P.; Fekete, A.; Gaspar, A.; Hertkorn, N., (2014). Molecular and structural characterization of dissolved organic matter during and post cyanobacterial bloom in Taihu by combination of NMR spectroscopy and FTICR mass spectrometry. Water Res., 57: 280-294 (15 pages).

Zhang, Y.; van Dijk, MA; Liu, M.; Zhu, G.; Qin, B., (2009). The contribution of phytoplankton degradation to chromophoric dissolved organic matter (CDOM) in eutrophic shallow lakes: field and experimental evidence. Water Res., 43: 4685-4697 (13 pages).

Ziegman, M.; Abert, M.; Muler, M.; Frimmel, F.H., (2010). Use of fluorescence for the estimation of bloom formation and toxin production of Microcystis aeruginosa. Water Res., 44: 195-204 (10 pages).

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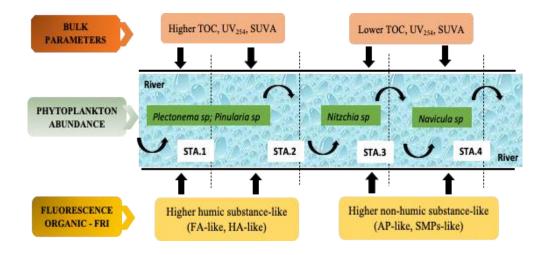
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#### **HOW TO CITE THIS ARTICLE**

Cahyonugroho, O.H.; Hariyanto, S.; Supriyanto, G., (2022). Dissolved organic matter and its correlation with phytoplankton abundance for monitoring surface water quality. Global J. Environ. Sci. Manage., 8(1): ...,...

## **GRAPHICAL ABSTRACT**



#### **HIGHLIGHTS**

- Characteristic of organic matter and the phytoplankton abundance is different at each segment of surface water;
- There is a substantial correlation among the bulk parameters, fluorescence spectroscopy of DOM and phytoplankton abundance;
- ➤ Phytoplankton abundance combined with DOM analysis could be used to evaluate the quantity and quality of organic matter for monitoring surface water quality.