medical archives

JOURNAL OF THE ACADEMY OF MEDICAL SCIENCES IN BOSNIA AND HERZEGOVINA









SUBMIT ARTICLE

MEDICAL ARCHIVES

Journal of the Academy of Medical Sciences of Bosnia and Herzegovina

We use cookies to ensure that we give you the best experience on our website. If you continue to use this site we will assume that you are happy with it.

Issue: 72-5

Giving Birth After Fertility Sparing Treatment of Embrional Carcinoma Figo III C: Case Report and Literature Review

Anis Cerovac, Dzenita Ljuca, Enida Nevacinovic, Azur Tulumovic, Ermina Iljazovic

MEDARCH:2018; 72-5: 371-373

PDF - FULL TEXT | DOI: 10.5455/medarh.2018.72.371-373

One Hundred Fifty Years of Organized Health Care Services in Bosnia and Herzegovina Izet Masic

MEDARCH:2018; 72-5: 374-388

PDF - FULL TEXT | DOI: 10.5455/medarh.2018.72.374-388

Incidence of Cardiac Dysfunction After Brain Injury

Selma Sijercic, Alisa Krdzalic, Goran Krdzalic

MEDARCH:2018; 72-5: 316-318

PDF - FULL TEXT | DOI: 10.5455/medarh.2018.72.316-318

Impact of Clopidogrel Loading for Coronarography on Bleeding After Urgent First Time CABG

Aleksander Hoxha, Sokol Shehu, Rezar Deveja, Thoma Qirjazi

MEDARCH:2018; 72-5: 319-324

PDF - FULL TEXT | DOI: 10.5455/medarh.2018.72.319-324

Hepatitis C Treatment in Patients with Drug Addiction Is Effective or Not Effective?

Seyed Amineh Hojati, Elham Maserat, Mohammad Ghorbani, Alireza Safarpour, Mohammad Reza Fattehi

MEDARCH:2018; 72-5: 325-329

PDF - FULL TEXT | DOI: 10.5455/medarh.2018.72.325-329

The Importance of Acinetobacter Species in the Hospital Environment

Velma Rebic, Nejra Masic, Sanela Teskeredzic, Mufida Aljicevic, Amila Abduzaimovic, Damir Rebic

MEDARCH:2018; 72-5: 330-334

PDF - FULL TEXT | DOI: 10.5455/medarh.2018.72.330-334

We use cookies to ensure that we give you the best experience on our website. If you continue to use this site we will assume that you are happy with it.

Ok







Medical Archives is official journal of Academy of Medical Sciences in Bosnia and Herzegovina

We use cookies to ensure that we give you the best experience on our website. If you continue to use this site we will assume that you are happy with it.

Ok







Medical Archives is official journal of Academy of Medical Sciences in Bosnia and Herzegovina

We use cookies to ensure that we give you the best experience on our website. If you continue to use this site we will assume that you are happy with it.

MEMBERS OF THE BOARD

Prof. Kenan Arnautovic, MD, PhD. (University of Tennessee, USA), Prof. Jacob Bergsland, PhD (The Intervention Centre, Oslo University Hospital, Oslo, Norway), Prof. Marko Buksa, PhD (Clinic for Internal diseases, Faculty of medicine, Sarajevo, BiH), Prof. Benjamin Djulbegovic, PhD (University of Tampa, Florida, USA), Bahare Fazeli (Inflammation and Inflammatory Diseases Research Center, Medical school, Mashhad, Iran, ORCID ID), Prof Armen Yuri Gasparyan, MD, PhD. (Department of Rheumatology, Clinical Research Unit, Russell's Hall Hospital, Dudley, UK, ORCID ID), Prof. Vjekoslav Gerc, PhD (University of Sarajevo, BiH), Prof. Mehmed Gribajcevic, PhD (University of Sarajevo, BiH), Prof. Mirko Grujic, PhD (University of Mostar and Sarajevo, BiH), Prof. Zoran Hadziahmetovic, PhD (Clinic for Emeregency medicine, Clinical center of University of Sarajevo, BiH), Prof. Ahmad-Reza Jamshidi (Rheumatology Research Center Tehran University of Medical Sciences: Tehran, Tehran, Iran, ORCID ID), Prof. Jasenko Karamehic, PhD (Clinical center of University of Sarajevo, BiH), Prof. Abdulah Kucukalic, PhD (Psychiatric clinic, Clinical center of University of Sarajevo, BiH), Prof. Asim Kurjak, PhD (Dubrovnik International University, Dubrovnik, Croatia), Prof. Pavle Milenkovic, PhD (Faculty of medicine, University of Belgrade, Serbia), Prof Resia Pretorius, PhD (Faculty of Health Sciences, University of Pretoria, ORCID ID), (Prof. Zeljko Reiner, PhD (Clinical center "Rebro", Faculty of medicine, University of Zagreb, Croatia), Prof. Osman Sinanovic, PhD (Neurology clinic, University clinical center Tuzla, BiH), Murat Ugurlucan (Istanbul, Turkey, ORCID ID), Prof Muharem Zildzic, MD, PhD (Academy of the medical sciences of Bosnia and Herzegovina, Sarajevo, Bosnia and Herzegovina-AMNuBiH, ORCID ID), Prof. Sukrija Zvizdic, PhD (Department for microbiology, Faculty of medicine, University of Sarajevo, BiH)

ADDRESS OF THE BOARD

Sarajevo, Mis Irbina 11, E-mail: www.amn.ba

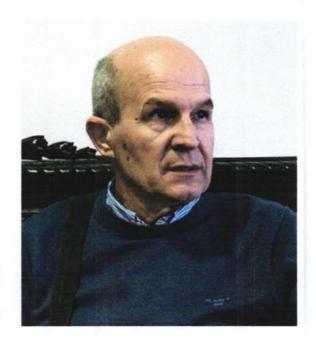
PUBLISHED BY

Academy of Medical Informatics in Bosnia and Herzegovina, Mis Irbina 11, 71000 Sarajevo, Bosnia and Herzegovina, amnubih@gmail.ccom

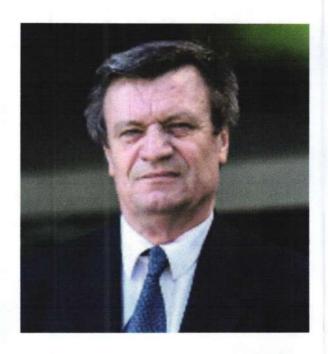
We use cookies to ensure that we give you the best experience on our website. If you continue to use this site we will assume that you are happy with it.

Ok

Editorial Board



IZET HOZO Editor-In-Chief ORCID ID



IZET MASIC Co-Editor-In-Chief ORCID ID





We use cookies to ensure that we give you the best experience on our website. If you continue to use this site we will assume that you are happy with it.

ORIGINAL PAPER

doi: 10.5455/medarh.2018.72.348-351
MED ARCH. 2018 OCT; 72(5): 348-351
RECEIVED: AUG 28, 2018 | ACCEPTED: OCT 05, 2018

¹Anatomy Department, Faculty of Medicine, Wijaya Kusuma Surabaya University

²Farmacology Department, Faculty of Medicine, Wijaya Kusuma Surabaya University

³Anatomy Department, Faculty of Medicine, Airlangga University, Surabaya

Proffessor of Internal Medicine at Doctoral Programme, Faculty of Medicine, Brawijaya University, Malang

⁵Proffessor of Clinical Pathology Doctoral Programme, Faculty of Medicine, Brawijaya University, Malang

Research Group of Smart Molecule of Natural Genetics Resources UB, and Department of Biology, Faculty of Sciences, Brawijaya University

Corresponding author: Prof. Fatchiyah F. PhD. Head of Research Group of Smart Molecule of Natural Genetics Resources, Brawijaya University, Jl. Veteran, Malang 65145, East Java, Indonesia. Email: fatchiya@ub.ac.id; Telp./Fax: +62-341-575841

© 2018 Ibrahim Njoto, Ayly Soekanto, Ernawati Ernawati, Abdurrachman Abdurrachman, Handono Kalim, Kusworini Handono, Djoko W. Soeatmadji, Fatchiyah Fatchiyah

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Chondrocyte Intracellular Matrix Strain Fields of Articular Cartilage Surface in Hyperglycemia Model of Rat: Cellular Morphological Study

Ibrahim Njoto¹, Ayly Soekanto¹, Ernawati Ernawati², Abdurrachman Abdurrachman³, Handono Kalim⁴, Kusworini Handono⁵, Djoko W. Soeatmadji⁴, Fatchiyah Fatchiyah⁵

ABSTRACT

Introduction: Chondrocyte is one cell in articular cartilage was products many proteins, molecules, and other factors. The external influence of cartilage, such as: hyperglycemia was entering joint capsule and impact to the chondrocytes and the cartilage. Hyperglycemia caused modification of heparan sulfate proteoglycan 2 (perlecan) proteins through glycation process. Aim: The aim of this study was to analyze morphological changing of chondrocytes pericellular matrix by the influence of hyperglycemia. Material and Methods: Eighteen adult male rats were divided into six groups: control, rat treated with sugar intake was 0.5 mg/kg, 0.75 mg/ kg, 1mg/kg, 1.5 g/kg and 2 mg/kg of body weight. The animal model was dislocated and left knee was taken to observe changing of chondrocytes pericellular matrix strain fields by Scanning Electron Microscope (SEM) from perpendicular to femoral condylus cartilage. Results: Changing of chondrocytes intracellular matrix strain fields as changing of cell diameters and cell distances at group control and group I to group V, which cell diameters was lower level and cell distances was the highest level at over diet 2. This changing of intracellular matrix strain fields was corresponding to changing chondrocytes morphology in hyperglycemia condition. due to hypertrophic stage as adaptive responses. This research as based on next research for accomplish of hyperglycemia influence to morphology articular cartilage changing to prevent degeneration of cartilage towards osteoarthritis. Conclusions: Present study concludes that hyperglycemia influence to chondrocyte intracellular matrix strain fields changing.

Keywords: cartilages, chondrocytes, heparan sulfate proteoglycan 2 (perlecan) proteins, hyperglycemia,

1. INTRODUCTION

Research on the effect of excessive carbohydrate intake on joint health, is still not explained yet in detail about changes in the morphology of chondrocytes in the articular cartilage. Previous studies have suggested that enlargement of chondrocyte size due to the influence of hyperglycemia in diabetes patients (1). The chondrocytes in the hypertrophy phase is not beneficial for cartilage health, because according to previous studies, it was found that hypertrophic chondrocytes secrete catabolic factors over anabolic factors, thus risking the integrity of the articular cartilage matrix (2,3). In addition to other previous studies, it was found that type 2 diabetes mellitus was a common predictor of severity progression of osteoarthritis disease (4).

This study is expected to fulfill the research gap before diabetes

emerged, it starts hyperglycemia condition, there has been a change in chondrocytes which at the risk of causing articular cartilage degeneration. Other literature states that osteoarthritis also increases the risk of type 2 diabetes mellitus (5). Various results from previous studies have not been explained about changes at the cellular level, when hypertrophy of chondrocytes happens, it will have an impact on the increase in secretion of catabolic factors over catabolic factors. The matrix degradation was starting at the superficial layer of articular cartilage due to mechanical compression during joint movement (6). This condition lasts will have an impact on articular cartilage erosion. The entire process has not been carried out research on the length of the chondrocyte diameter and the distance between chondrocytes in the

	С	SI 0,5	SI 0,75	SI 1	SI 1,5	SI 2
Blood Sugar level (mg/dl)	86.33±0,58	143,33±1,53	160.67±0,58	127.83±1,04	201,77±1,97*	202±2,64*

Table 1, Blood sugar level of control group and sugar intake treatment group, C = control; SI: Sugar intake (0,5mg/kg BW; 0,75mg/kg BW; 1mg/kg BW; 1,5mg/kg BW; 2mg/kg BW). * = significant different level (p<0,05)

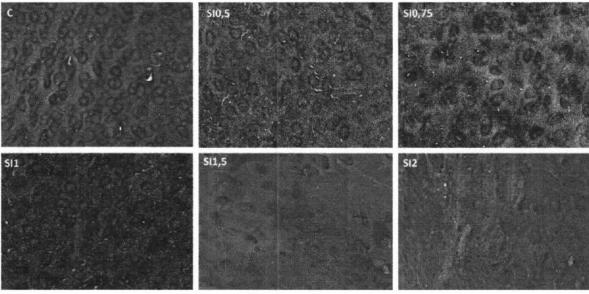


Figure 1. Morphology of chondrocyte and pericellular matrix (1000x) by Scanning Electron Microscope (SEM), C = control; SI: Sugar intake (0,5mg/kg BW; 0,75mg/kg BW; 1,5mg/kg BW; 2mg/kg BW).

surface layer of chondrocytes in hyperglycemia condi-

The parameters are expected to represent the chondrocytes intracellular matrix strain fields, which are needed to reflect the integrity of the articular cartilage matrix. The results of morphological changes in chondrocytes include superficial cell diameters and cell distance at the superficial layer of articular cartilage giving answers to the morphological changes in chondrocytes when cartilage degeneration begins, which is the starting point for osteoarthritis.

2. AIM

The aim of this study to analyze occurrence of hyperglycemia will increase the risk of osteoarthritis due to degeneration of the superficial layer of the articular cartilage caused by chondrocytes during the hypertrophy phase secreting catabolic factors over anabolic secretion.

3. MATERIAL AND METHODS

The experimental animal model was eighteen of male rats (*Rattus norvegicus* strain Wistar), 1.5 to 2 months old and 100 to 150 g body weight, divided into six groups of control group and rat was oral administration with sugar intake 0.5 mg/ kg BW (SI0,5); 0.75 mg/ kg BW (SI0,75), 1mg/kg BW (SI1), 1.5 g/kg BW (SI1,5) and 2 mg/kg BW (SI2). Sugar intake treatment for three times daily (in the morning, afternoon and evening). All groups were fed once a day in the afternoon was given 30 g of the standard diet.

All treatment was maintained for two months, after that all animal models were taking a blood sample to check the blood glucose level before being sacrificed. All treatment above was doing at standard above and sugar intake treatment until hyperglycemia condition. The sample was right condylar of femur bone, and processing to observe at the highest point of condylar with the perpendicular laser shooting of Scanning Electron Microscope device. Each condylar was shot for three times. All data of morphological chondrocytes changing was processed by Olympus TM 1000. Then the data continue to be analyzed with statistical analysis by one-way ANOVA. This study had approved by the research Ethics committee of Brawijaya University, Malang, East Java, Indonesia

4. RESULTS

The blood sugar level of SI2 group highest level significantly among of all treatment was 202±2,64* (Table 1). Whereas, SI 1,5 group was higher blood glucose level compared with control group, low sugar intake and standard. Control group and lower sugar intake (S10,5; SI0,75 and SI1) have still range 86 mg/kg BW until 160.67 mg/kg BW.

The morphological changing of chondrocytes at the superficial layer of articular cartilage by Scanning Electron Microscope for measure cell diameters and distance can show in Figure 1 and 2.

The group of SI1, 5 and SI2 shows that the cell is not compact and rarely (Figure 1 and 2). Vice versa, Control group and the lower sugar and standard sugar intake still show compact cell. This condition also supported with the cell diameter of chondrocyte show that SI1 and SI2 was shortest diameter significantly among of all groups (Figure 3A). Whereas, the cell distances of chondrocyte

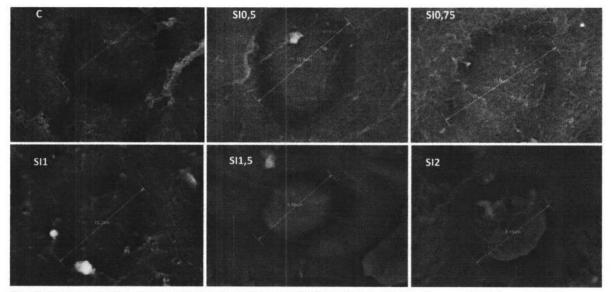


Figure 2. Morphology and size cell of chondrocyte and pericellular matrix (9000x) by Scanning Electron Microscope (SEM), C = control; SI: Sugar intake (0,5mg/kg BW; 0,75mg/kg BW; 1,5mg/kg BW; 2,5mg/kg BW).

of group SI1,5 and SI2 was increasing significantly that show the cell is rarely along with increasing blood sugar level (Figure 3B).

The result showed that blood sugar level was increased, according to carbohydrate diet in the SI 0.5 group with BSL: 142 mg/dl; SI 0.75 group with BSL: 160.5 mg/dl and SI 1.0 group with BSL: 127.0; SI 1.5 group with BSL: 200.3 mg/dl and SI 2 group with BSL: 184.0 mg/dl, all of five groups has BSL above the control group. The result of cell diameters was decreasing according to BSL increasing, but cell distances were increasing. The most significant change of the longest diameter at SI 1.5 group: 9.69 μ M and at SI 2 group: 8.51 μ M comparable to control group: 11.40 μ M. The most significant of cell distances changing was increased in SI 1.5 group: 5.25 μ M and 10.29 μ M. This condition is shown that increasing of blood sugar level of animal model influence to chondrocytes shape and formation.

5. DISCUSSION

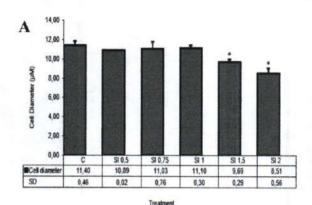
The previous studies result that was a link between hyperglycemia in type 2 diabetes with osteoarthritis disease (7). Hyperglycemia induce chondrocytes to produce pro-inflammatory mediators, such as: advanced glycation end products (AGEs); known as chondrocytes activation (8). This condition increasing local toxicity to joint tissue which caused chondrocytes dedifferentiation and goes apoptotic (9). Above condition has not explained cellular and morphological changes of chondrocytes formation at articular cartilage. Previous literature stated that chondrocytes will become hypertrophic (10) during osteoarthritis. Current conditions occur in the articular cartilage and have not been explained how the morphology chondrocytes changing, especially at the superficial layer of cartilage. The present study has an answer about this morphological changing of chondrocytes at the superficial layer of articular cartilage.

The result of this research was representative of the morphological condition of chondrocytes, which being

influenced by hyperglycemia at five groups of treatment. The impact of excess carbohydrate diet was increasing of blood sugar level of animal models which caused the changing of cell diameters and cell distances. Cell diameters at this study were measurement of the longest diameter of each chondrocyte, cell distances was representative of chondrocytes anatomical form. It represents that excess carbohydrate diets influence to chondrocytes at the superficial layer of articular cartilage of animal model. The long diameters of chondrocytes were increasing, it means that chondrocyte has rounder shape and less flat form, at normal condition the chondrocyte form of superficial layer must be flat.

Shape changing of decreasing of long diameters of chondrocytes at this study as a sign which chondrocytes entering a hypertrophic phase from proliferation phase. This condition will give no good impact to cartilage health in the future, because hypertrophic chondrocytes produce more catabolic factor such as: IL-1ß (Interleukin 1 beta), FGF-2 (Fibroblast Growth Factor 2) which increasing of cartilage matrix degradation (11). Changing of chondrocytes distance as a sign that hyperglycemia of animal model at present study showed the negative impact to the chondrocytes population at the superficial layer of cartilage layer. It means that hyperglycemia influence to chondrocytes formation, more rarely than normal condition. Changing of chondrocytes diameter and cell distances as representing the condition of chondrocytes formation, which two parameters indicate of changing of intracellular matrix strain fields changing at the superficial layer of articular cartilage.

When chondrocytes formation at superficial layer was changed, it can be followed by changing of chondrocytes intracellular matrix strain fields. Chondrocytes are the only permanent residence at articular cartilage (12). It produces many major and minor components of articular cartilage, such as: collagen type II, glicosaminoglican and hialuronan, woven around condrosit as a major component (13),(14), and collagen type V, VI, IX, X, XI,



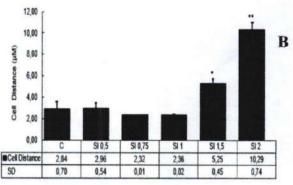


Figure 3. A) Cell diameters and B) Cell distances of chondrocyte and pericellular matrix. C = control; SI: Sugar intake (0,5mg/kg BW; 0,75mg/kg BW; 1mg/kg BW; 1,5mg/kg BW; 2mg/kg BW). *= significantly different level (p<0,05)

XII, XIV, decorin, biglican, fibromodulin, perlecan; non colagenus protein: matrilins, trombospondin-5/ COMP (15),(16),(17). All of the components above need to support matrix integrity, if chondrocytes distance was increased by the influence of hyperglycemia at animal model, it increases risk to lowering of matrix density, which can decrease intracellular matrix strain fields by the dynamical movement of the joint (18). It can increase the risk of articular cartilage damage, beginning at superficial layer.

6. CONCLUSION

This study concludes that hyperglycemia can be act as a trigger factor of osteoarthritis disease, which it can be worse by chondrocytes senescence as a pathogenic factor. Present as basic to continue with next research which can measure of cartilage pericellular matrix strain fields and counting of the chondrocyte's population at the surface, middle and deep layer of articular cartilage of animal model, as complement to the pathophysiology of correlation between diabetic with osteoarthritis diseases. Present study shown that hyperglycemia by excess carbohydrate diets already influence to articular cartilage, so the suggestion was controlling carbohydrate diets need to support articular cartilage in the future.

- Acknowledgments: This study was supported by DGHE, Ministry
 of Science, Technology and Higher Education research grant No. 3/E/
 KPT/III/2018 and thanks to Biosains Institute staff and analyst, Brawijaya University for providing the laboratory facilities.
- Authors' contributions: All the authors were involved during the investigation process in all stages of this study including a primary data collection, analysis and the documentation of the collection.
- · Conflict of interest: none declared

REFERENCES

- Salinas D, Minor CA, Carlson RP, McCutchen CN, Mumey BM, June RK. Combining targeted metabolomic data with a model of glucose metabolism: Toward progress in chondrocyte mechanotransduction. PLoS One. 2017; 12(1).
- Loeser RF, Gandhi U, Long DL, Yin W, Chubinskaya S. Aging and oxidative stress reduce the response of human articular chondrocytes to insulin-like growth factor-1 and osteogenic protein-1. Arthritis Rheumatol (Hoboken, NJ). 2014; 66(8): 2201-2209.
- Loeser R. Age-related changes in the muscoskeletal system and the devel-

opment of osteoarthritis. Clin Geriatr Med. 2010; 26(3): 371-386.

Treatment

- Schett G, Kleyer A, Perricone C, Sahinbegovic F, Jagnocco A, Zwerina J, et al. Diabetes is an independent predictor for severe osteoarthritis: Results from a longitudinal cohort study. Diabetes Care. 2013; 36(2): 403-409.
- Rahman MM, Cibere J, Anis AH, Goldsmith CH, Kopec JA. Risk of Type 2 Diabetes among Osteoarthritis Patients in a Prospective Longitudinal Study. Int J Rheumatol. 2014; 2014; 1-8.
- Sakai N, Hashimoto C, Yarimitsu S, Sawae Y, Komori M, Murakami T. A functional effect of the superficial mechanical properties of articular cartilage as a load bearing system in a sliding condition. Biosurface and Biotribology. 2016; 2(1): 26-39.
- Oren TW, Botolin S, Williams A, Bucknell A, King KB. Arthroplasty in veterans: analysis of cartilage, bone, serum, and synovial fluid reveals differences and similarities in osteoarthritis with and without comorbid diabetes. J Rehabil Res Dev. 2011; 48: 1195-1210.
- Laiguillon MC, Courties A, Houard X, Auclair M, Sautet A, Capeau J, et al. Characterization of diabetic osteoarthritic cartilage and role of high glucose environment on chondrocyte activation: toward pathophysiological delineation of diabetes mellitus-related osteoarthritis. Osteoarthr Cartil 2015; 23: 1513-1522. doi:10.1016/j.joca.2015.04.026.
- Courties A, Sellam J. Osteoarthritis and type 2 diabetes mellitus: what are the links?. HAL ld 2016: hal-01396521.
- Willett TL, Kandel R, De Croos JNA, Avery NC, Grynpas MD. Enhanced levels of non-enzymatic glycation and pentosidine crosslinking in spontaneous osteoarthritis progression. Osteoarthr Cartil. 2012; 20(7): 736-744.
- Wang J., Kramer W.C., Schroeppel J.P. Transcriptional regulation of articular chondrocyte function and its implication in osteoarthritis. Principles of Osteoarthritis Its Definition, Character, Derivation and Modality-Related Recognition. 2012; 474-488.
- Baugé C, Duval E, Ollitrault D, Girard N, Leclercq S, Galéra P, et al. Type II TGF? receptor modulates chondrocyte phenotype. Age (Omaha). 2013; 35(4): 1105-1116.
- Heinegard D., Saxne T. The Role of Cartilage Matrix in Osteoarthritis. Abstract Nature Reviews Rheumatology. 2011; 7: 50-56.
- Shen J, Li J, Wang B, Jin H, Wang M, Zhang Y, et al. Deletion of the transforming growth factor β Receptor type II gene in articular chondrocytes leads to a progressive osteoarthritis-like phenotype in mice. Arthritis Rheum. 2013; 65(12): 3107-3119.
- Goepfert C, Lutz V, Linse S, Kittel S, Wiegandt K, Kammal M, et al. Evaluation of cartilage specific matrix synthesis of human articular chondrocytes after extended propagation on microcarriers by image analysis. Int J Artif Organs. 2010; 33(4): 204-218.
- Mackie EJ, Tatarczuch L, Mirams M. The skeleton: A multi-functional complex organ. The growth plate chondrocyte and endochondral ossification. Journal of Endocrinology. 2011; 109-121.
- Shen J, Wang M, Jin H, Sampson E, Che D. Genetic Mouse Models for Osteoarthritis Research. Princ Osteoarthritis- Its Defin Character, Deriv Modality-Related Recognit [Internet]. 2012; 321-334.
- Weber JF. The sensitivity of articular chondrocytes to dynamic mechanical stimulation By. 2015; (December)...