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Bioanalytical method development and validation for quantification of morachalcone A in rabbit plasma using high performance liquid chromatography

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Abstract: Artocarpus champeden (A. champeden) ethand 2 extract has been reported as antimalarial activity and prospective to be developed as phytomedicine products. The active marker compound was identical with known prenylated chalcone compound, Morachalcone A. To further develop phytomedicine products from A. champeden especially in aspects of bioavailability and pharmacok 13 tic, a valid, selective and sensitive analytical method becomes important to determine morachalcone A in plasma. The aim of study was to develop and validate selectivity and sensitivity of High Performance 21 uid Chromatography (HPLC) method to determine morachal 10 to A in rabbit plasma. This method was used a RP-18 Column (250 x 4.6 mm i.d, 5 µm), under isocratic elution and acetonitrile:water (50:50 v/v) was used as mobile phase with flow rate of 1.0ml/min. Detection was carried out at 368 nm, 4-hydroxychalcone and methanol were used as internal standard and precipitant. Results showed that this HPLC method was selective with good linearity in range of 3096.774 to 154.839ng/ml. LO 12 nd LLOQ were 89.384 and 154.839ng/ml, respectively. The mean %different was found between 2.79 to 14.33%. Intra and inter-day precision were $\leq 15\%$ and recovery from this extraction method of morachalcone A and Internal Standard were 80-120%.

Keywords: HPLC, Morachalcone A, bioavailability, pharmacokinetic, 4-hydroxychalcone, rabbit plasma.

INTRODUCTION

Artocarpus champeden (A. champeden) is one of plant species in fa2 ily of Moraceae and locally known as "cempedak". It is widely spread in Indonesia and has been traditionally used for treatment of malaria (Hakim et al., 2006). In previous studies 201. champeden extract have been reported for its antimalarial 2 ctivity against Plasmodium falciparum 3D7 strain and several prenylated flavonoids isolated from A. champeden exhibited in vitro antime 40 al activity against *P. falciparum* 3D7 strain as well (Widyawaruyanti et al., 2007). One of prenylated flavonoids from A. champeden is mora 20 cone A which was isolated from the ethanolic extract 2 A. champeden stem bark. It was exhibited antimalarial activity against P. *falciparum* 3D7 strain with 39 C₅₀ value of 0.18µg/ml. Regarding to its antimalaria 2 ctivity, the ethanolic extract of A. champeden stem bark is prospective to be developed as antimalarial phytomedicine product with morachalcone A as an active marker compound (Hafid et al., 2012).

Method development and validation plays a significant role in evaluation and interpretation of bioavailability, bioequivalence, pharmacokinetic and toxicokinetic studies. These studies were important in the development

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of antimalarial phytomedicine product from A. champeden ethanolic extract (Ma et al., 2015). Such study is required to obtain a reliable analytical and sensitive method to analyze marker compound of A. champeden ethanolic extract, especially at a number of tracer (Harahap et al., 2016). One of these methods is High Performance Liquid Chromatography (HPLC).

Despite morachalcone A can be quantified in the et an of extract of *A. champeden* stem bark using High Performance Liquid Chromatography Photo Diode Array (HPLC-PDA) (Hafid *et al.*, 2012). However, there are still no reported methods for determination of morachalcone A in plasma. Therefore, in this present work, we wanted to develop an accurate, selective and validated analytic 35 method to determine morachalcone A in rabbit plasma. In this work, we report a development and validation of HPLC method to determine morachalcone A in rabbit plasma.

MATERIALS AND METHOD

Chemicals and reagents

Morachalcone A (98% on assay) was purchased from Chemfaces (China). The internal standard 4-Hydroxychalcone (97% on assay) was obtained from Organic Chemistry Laboratory, Yogyakarta State

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University (Indonesia). Acetonitrile, methanol and water were HPLC grade. All solvent was purchased from Merck (Singapore). Lithium heparin was purchased from BD (USA). Six rabbits (New Zealand rabbit, male, 4-5 months old and with 2-3 kg body weight) were used for the study and were purchased from rabbit farms in Malang (Indonesia).

Apparatus

Bath sonicator (Sonica EP series), microbalance (Radwag), centrifuge (Thermo Legend Micro 17 e), micropipettes (Soccorex), vortex mixture (Maxi mix II Barnstead Thermolyne) were used for the study.

HPLC system and conditions

System consisted of a pump (LC-06 AD) equipped with SPD-M20A and connected (1) CBM-20A Shimadzu (Japan) with computer (HP). Manual Rheodyne injector model 7725i (with 20 µl loop size). The software used was LC Workstation. Chromatographic separations were performed 32 Lichrosphere 100 RP18 5µm LichoCART (2) 250-4 (250 x 4,6 mm i.d.) column and attached to a guard column (octad 19 silane guard column) and at ambient temperature. Mobile phase consists of mixture acetonitrile and water (50:50 v/v), which w 6 pumped 1.0ml/min and in isocratic mode. Before use, the mobile phase was filtered with 0.22µm cellulose membrane (Whatman) and degassed. Wavelength to analysis was detected at 368 nm.

Blank rabbit plasma sample preparation

Six male healthy New Zealand rabbits weighing 2-3 kg body were acclimatized for one weeks in order to observe their good health and 36 tability. After one week, approximately 5ml rabbit blood samples were taken and 16 ected into heparinized tubes via vena auricuralis then centrifuged (3,000 rpm, 10 min) to separate the plasma. The plasma samples were st 17 at -20^oC prior to analysis. This research protocol was approved by Animal Care and Use Committee (ACUC), Faculty of Veterinary Medicine, Universitas Airlangga with reference number 660-KE for notice of approval.

Preparation of stock, calibrations solutions and quality control samples

Stock solution of morachalcone A (0.96mg/ml) and 4hydroxychalcone as Internal Standard (0.96mg/ml) were diluted in methanol. Calibration and controls was made by dilute stock solution with methanol until certain concentration. Solution of Inte 3 ll Standard (IS) (300.387ng/ml) was made by dilute the IS stock solution. All solutions were kept at 4° C and brought to room temperature before use. For calibrations, rabbit plasma contain morachalcone A was prepared in concentration range of 3096.774; 1548.387; 774.194; 387.097; 193.548ng/mL; and 154.839ng/ml. After that, into the calibrations plasma was added 10.0μ l a certain of morachalcone A working solution into 90.0μ l of blank rabbit plasma and shortly vortexed. Quality control (QC) samples were made by the same method as making calibration standards using three concentrations: low (464.516ng/ml), medium (1470.968ng/ml) and high (2477.419ng/ml).

Sample preparation

 5.0μ l of IS solution (300.387ng/ml) was added into 50.0μ l aliquot of rabbit plasma containing certain concentrations of morachalcone A. The solution then vortexed (30 seconds) and 100 μ l of methanol was added to precipitate protein in plasma. Samples were vortexed (1 minute), sonicated (3 minutes) then centrifuged at 14,000 rpm (10 min) at 4^oC, 20.0 μ l of supernatant was taken and inject to the HPLC system.

System suitability test

System suitability test performed on morachalcone A (619.355ng/mL) and IS (300.387ng/ml) in rabbit plasma then extracted up g protein precipitation method. 20.0μ l of supernatant injected into the HPLC equipment in optimum condition, done six times a repetition then record the number of the retical plate, selectivity factor and asymmetry factor. The coefficient of variation of retention time and area ratio chromatogram also calculated too.

Method validation

Selectivity 31

Selectivity was determined by analyzing blank plasma from six different rabbits, blank plasma was spiking with morachalcone A (at LLOQ concentration) and the IS. The %different of morachalcone A also was calculated and its value should be less than 20% (Harahap *et al.*, 2016).

Linearity, limit of detection (LOD) and lower limit of quantification (LLOQ) 18

Linearity was determined by plotting peak area ratio (y) of morachalcone A to internal standard (morachalcone A/IS) versus the concentration (x) of morachalcone A. Linearity was made by analyzing spiked samples on five different days. The corresponding slope (b) and residual standard deviation (Sy) values were used to calculate limit of detection (LOD) using following equation (Hafid *et al.*, 2015):

$$LOD = \frac{3 \text{ x Sy}}{\text{slope (b)}}$$

The $\overline{\text{LLOQ}}$ was defined as lowest concentration with accoefficient variation (%CV) and accuracy (%(15) ifferent) should be- $\leq 20\%$. The response signal of the LLOQ sample should be at least 5 times response signal of blank sample (FDA, 2013).

Precision and accuracy

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Intra-day precision and accuracy were performed by analyzing samples (n=5) in replicates at three concentrations (464.516ng/ml, 1470.968ng/ml) and 2477.419ng/ml) on one day and the time gap was eight hours. Inter-day precision and accuracy were analyzed by

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analyzing same \overline{QC} sample 11 three consecutive days in samples replicates (n=5). Intra and inter-day precision were estimated by calculate the coefficients of variation (% CV) from QC samples." %different" or "%bias" was used to calculate intra and inter-day accuracy, which can be calculated using the following equation (Kumar *et al.*, 2006):

% different = <u>Observed concentration - nominal concentration</u> <u>nominal concentration</u>

Extraction recovery

The extraction recovery from clean up sample from plasma was determined at 464.516, 1470.968 and 2477.419ng/ml by spiking morachalcone A into drug-free plasma 34 prachalcone A extraction recovery from plasma sample was determined by comparing response from peak area ratio of QC samples with peak area response from non-extracted control samples which prepared at the same concentration level (Singh *et al.*, 2012). This step was repeated to determine the extraction recovery from IS.

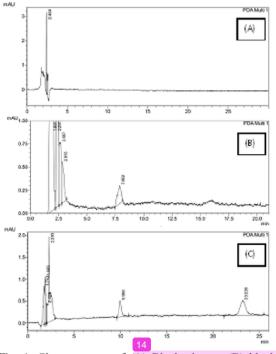


Fig. 1: Chromatogram of (A) Blank plasma, (B) blank plasma spiked with IS, (C) blank plasma spiked with morachalcone A (154.839ng/ml) and IS (300.387ng/ml).

Stability

Stability of morachalcone A in rabbit plasma was determined by analyze the QC plasma samples containing 3 prachalcone A at 464.516 and 2477.419ng/ml (n=3). These results were compared with freshly prepared plasma samples. Freeze/thawstability was 3 termined after 3 complete cycles (-20^oC) on three days. Short-term stability was determined following incubation of sat 9 les had been spiking with morachalcone A at room

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temperature (24h). Long-term stability was determined following keep of spiked plasma samples (30 days) at -20°C.

RESULTS

stem suitability test

The results of system suitability test repeatability demonstrated by injection, indicating that the analytical methods used have met the system suitability criteria. Coefficient variation value of retention time was 0.25 and 0.27% for morachalcone A and IS, respectively, the 110 of the chromatogram area was 6.01% which was <10% for the analysis of biological fluid samples (Harmita, 2006). The value of number of theoretical plate from morachalcone A 2167.445 and for IS was 2531.573 which were more than 2000, selectivity factor from morachalcone A was 2.650 and 11.2275 for IS which were more than 1.0 and asymmetry factor morachalcone A 0.940 and 0.984 for IS which less or equal than 2.0 (USP, 2015).

Selectivity

The aim of selectivity test was to ensure originality of the peak in sam 30 analysis. The selectivity was determined by analyze blank rabbit plasma sample, blank rabbit plasma that was spiked with internal standard (4-hydroxychalcone) and both of morachalcone A and internal standard (4-hydroxychalcone). fig. 1 showed that there was no interference of endogenous compounds from blank plasma from the six different rabbits. %different of morachalcone A was 13.66% which was <20% (Harahap *et al.*, 2016). Retention time of morachalcone A and IS were 23.228 and 9.996 minutes, respectively.

Linearity, LOD and LLOQ

The correlation between morachalcone A standard concentration versus response area showed linier correlation at concentration range of 3096.774 to 154.839ng/mL with $R^2 = 0.9994$ (r = 0.9997). LOD and LLOQ for morachalcone A in plasma were 89.384 and 154.839ng/ml, respectively. The precision (%CV) and accuracy (%different) from LLOQ were 7.59 and 10.87%, respectively which was <20% (FDA, 2013).

Precision and accuracy

QC samples plasma contain morachalcone A at 3 concentration levels of 464 9 6, 1470.968 and 2477.419ng/ml were analyzed for precision and accuracy. The inter- at 27 ntra-day precision and accuracy values of this method are presented in table 1. The range for intra-day precision (%CV) was 3.88-7.08% and range for inter-day precision (%CV) was 4.98-10.80%. The range for intra-day accuracy (%different) for morachalcone A was 3.35 to 14.33%, and range for inter-day accuracy (%different) was 2.79 to 10.07%. These results showed that this method fulfilled acceptance criteria of FDA because these values were <15% (FDA, 2013).

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Concentration (ng/ml) Precision Accuracy Assay Observed (mean ±SD) Nominal CV (%) Different (%) 464.516 397.947 ± 25.935 6.52 14.33 1470.968 1265.608 ± 49.166 13.96 Intra-day 3.88 2477.419 2560.525 ± 181.292 7.08 3.35 464.516 417.741 ± 20.819 4.98 10.07 Inter-day 1470.968 1429.971 ± 154.417 10.80 2.79 2477.419 2305.863 ± 221.884 9.62 6.92

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Table 1: Intra and inter-day precision and accuracy of morachalcone A in rabbit plasma (n=5)

Table 2: Stability of morachalcone A in rabbit plasma at different storage conditio

Storage condition	Concentration (ng/ml)		Different (%)
Storage condition	Nominal	Observed	
Three freeze-thaw cycles at -20 ⁰ C	2477.419	2088.691±272.723	8.03
	464.516	452.351±41.641	7.09
Room temperature for 24 hours	2477.419	2609.952±358.123	8.30
	464.516	500.505±60.746	14.23
Frozen (-20 [°] C) for 30 days	2477.419	2482.221±1462.859	2.05
	464.516	461.880±23.564	6.77

Extraction recovery

The aim of extraction recovery was to ensure pertains to the extraction efficiency of an analytical method within the limits of variability. Extraction recovery from morachalcone A in plasma at 464.516, 1470.968, and 2477.419ng/ml were found to be 95.38 ± 1.60 ; 84.45 ± 3.25 ; and $80.291\pm1.55\%$, respectively. Extraction recovery from IS was $99.54\pm1.31\%$. Extraction recovery for both morachalcone A and IS were within 80-120% and fulfilled the acceptance criteria (Caufield and Steward, 2012).

Stability

Stability of morachalcone A in plasma (464.516 and 2477.419ng/ml) under different temperature and storage condition is showed in table 2. The result of stability test showed that morachalcone A in 26 sma were stable for all conditions, i.e. 24h stored at room temperature; 3 freeze/thaw cycles and 30 days stored at -20°C. The %different value was <15% and fulfilled the acceptance criteria from FDA (FDA 2013).

DISCUSSION

The developed method provided a specific assay and sensitive for morachalcone A in rabbit plasma. 4-hydroxychalcone was selected as internal standard because its core structure is similar with morachalcone A, and it can be separated from morachalcone A. Another study also used 4-hydroxychalcone as internal standard to develop analytical method to determine pharmacokinetic parameter of ezetimibe in human plasma (B4 *et al.*, 2012). Morachalcone A was detected at 368 nm using PDA detector to increase the signal of compound and reduce the signal of plasma interferents. Ratio and flow rate of mobile phases were optimized by several trials to obtain good resolution and symmetric peak for

morachalcone A. This method 16 s optimized by observe 4 chromatographic parameters: retention time, resolution, number of theoretica 11 ate (N) and tailing factor of various compositions. Acetonitrile-water (50:50 v/v) and flow rate 1.0ml/min were selected as mobile phase with isocratic eluation because it is simple, easy to use and produces parameter which fulfilled the chromatography acceptance criteria (Kumar and Sunandamma, 2012). Method for sample clean up (to remove protein and interference) before sample was injected into HPLC was important also in the development of this method. A simple protein precipitation using methanol was employed in this study (Singh et al., 2012). Methanol was used as solvent to precipitate protein in plasma because its efficiency in precipitating protein in human plasma has been reported >90% and morachalcone A was dissolve in methanol as well (Bueno et al., 2011). This method also fulfills the acceptance criteria from FDA for bioanalytical method validation (FDA, 2013).

CONCLUSION

The HPLC method was valid for determination of morachalcone A in rabbit plasma (in vitro) and showed good chromatographic parameters including selectivity, linearity, sensitivity, accuracy, precision, stability and %recovery of extraction. The developed method also can be used to determine morachalcone A in rabbit plasma, thereby enabling to determine the bioavailability and pharmacokinetic parameter of *A. champeden* ethanolic extract using morachalcone A as active marker compound.

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