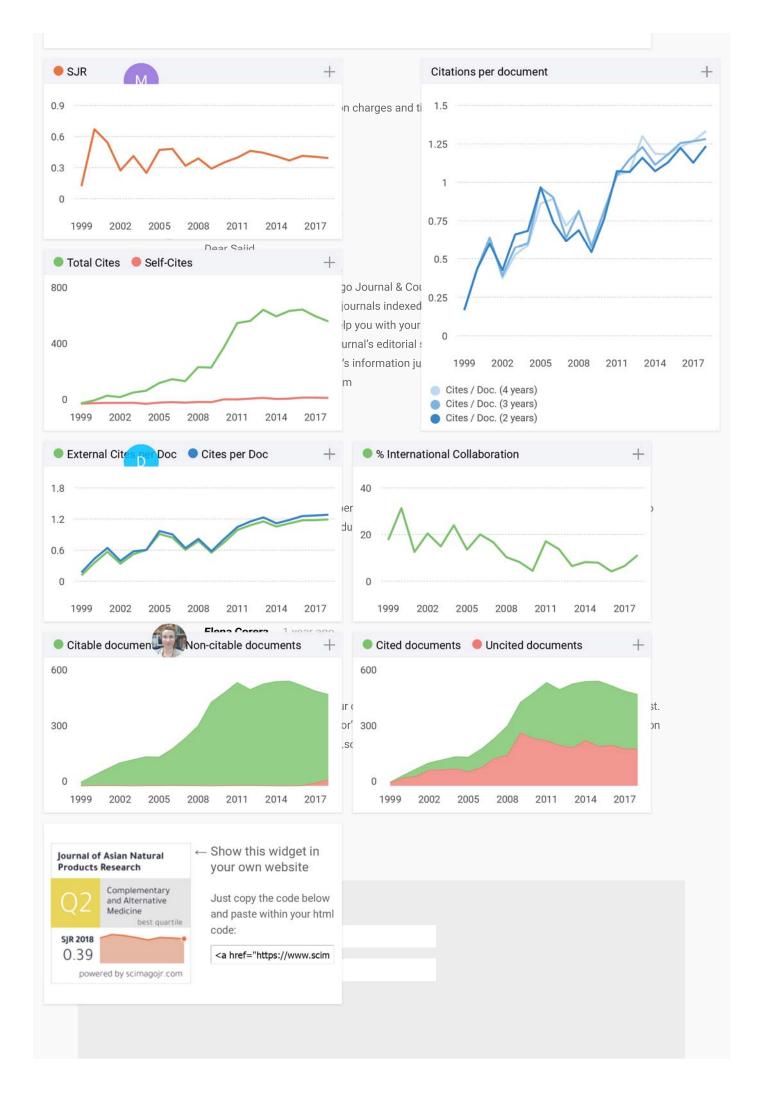
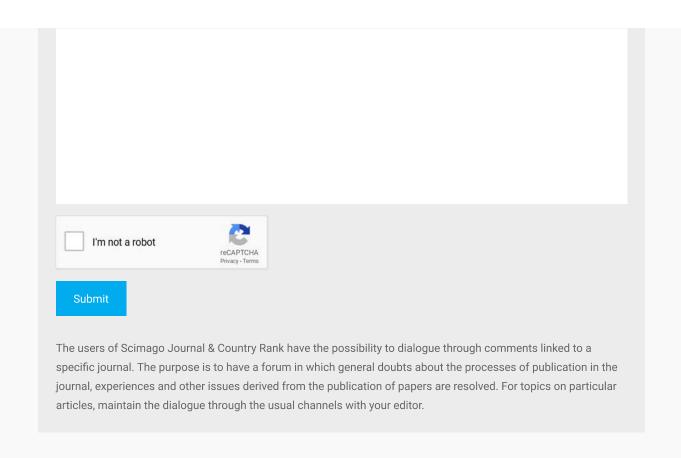


1 of 3 11/6/2019, 10:09 PM



2 of 3 11/6/2019, 10:09 PM





3 of 3 11/6/2019, 10:09 PM



browse our exclusive earlor's choice concedion for free

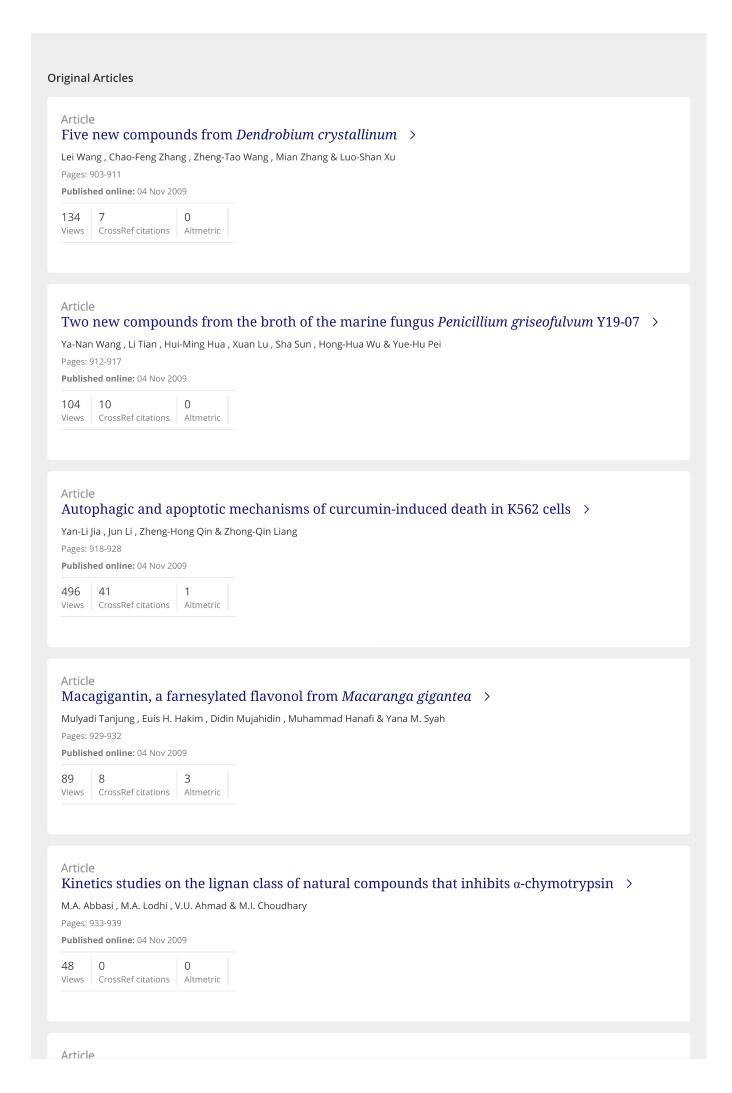
Latest articles

Article Article Article Article A new highly Two new 2-(2-A new single-stage Furanone oxygenated derivative and Hydroxymethod for pregnane and two sesquiterpene 2-phenylethyl)chr obtaining of new from Antarctic omens from betulin 5-hydroxymethylfurf phenylcarbamates marine-derived agarwood ural derivatives from originating from fungus Penicillium the water decoction of *Poria cocos* > sp. S-1-18 > Aquilaria crassna Wang et al. Mamaeva et al. Published online: 6 Oct 2017 Published online: 9 Oct 2017 Published online: 9 Oct 2017 Published online: 5 Oct 2017

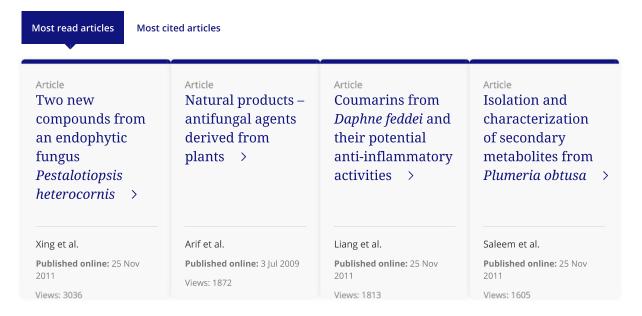
View more>

See all volumes and issues

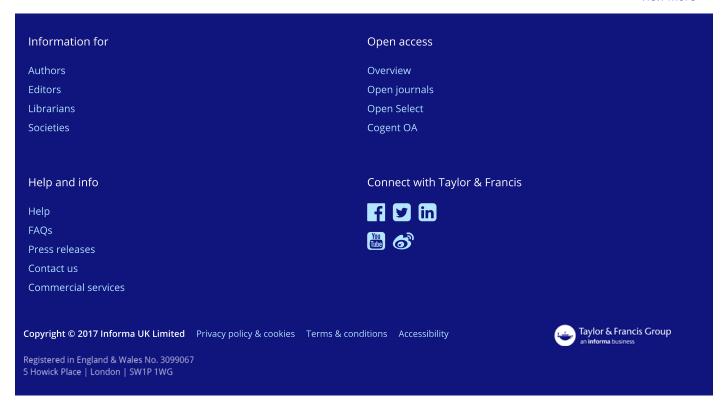




Explore



View more>





Editorial board

Editor-in-Chief:

De-Quan Yu - Institute of Materia Medica, Chinese Academy of Medical Sciences, 1 Xian Nong Tan Street, Beijing 100050, China

Vice Editors-in-Chief:

Li-Xin Dai - Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, Shanghai, 200032, China

Shi-Shan Yu - Institute of Materia Medica, Chinese Academy of Medical Sciences, 1 Xian Nong Tan Street, Beijing, 100050, China

Zhuo-Wei Hu - Institute of Materia Medica, Chinese Academy of Medical Sciences, 1 Xian Nong Tan Street, Beijing, 100050, China

Regional Editors:

Noguchi Hiroshi - School of Pharmaceutical Sciences, University of Shizuoka, Shizuoka 422-8526, Japan

Ching-jer Chang - Department of Medicinal Chemistry & Molecular Pharmacology, Purdue University, West Lafayette, Indiana 47907, USA

Editorial Board:

Zeper Abliz - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Atta-ur-Rahman - FRS, H.E.J. Research Institute of Chemistry, Karachi, Pakistan

Chun-Tao Che - The Chinese University of Hong Kong, Hong Kong

Xiao-Guang Chen - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Gui-Fang Cheng - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Guan-Hua Du - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Qi-Cheng Fang - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

De-An Guo - Chinese Academy of Sciences, Shanghai, China

Yue-Wei Guo - Chinese Academy of Sciences, Shanghai, China

Xiao-Jiang Hao - Chinese Academy of Sciences, Kunming, China

Prasat Kittakoop - Chulabhorn Graduate Institute and Chemical Biology Program, Thailand

Ling-Yi Kong - China Pharmaceutical University, Nanjing, China

Yueh-Hsiung Kuo - National Taiwan University, Taipei, Taiwan

Kuo-Hsiung Lee - University of North Carolina, USA

Lian-Niang Li - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Wen-Han Lin - Peking University, Beijing, China

Yue-Hu Pei - Shenyang Pharmaceutical University, Shenyang, China

Guo-Wei Qin - Chinese Academy of Sciences, Shanghai, China

Jian-Gong Shi - Chinese Academy of Medical Sciences, Beijing, China

Yan-Ping Shi - Chinese Academy of Sciences, Lanzhou, China

JongHeon Shin - Natural Products Research Institute, Seoul National University, Seoul, Korea

Han-Dong Sun - Chinese Academy of Sciences, Kunming, China

Ren-Xiang Tan - Nanjing University, Nanjing, China

Peng-Fei Tu - Peking University, Beijing, China

Yong-Qiang Tu - Lanzhou University, Lanzhou, China

Feng-Peng Wang - Sichuan University, Chengdu, China

Yu-Lin Wu - Chinese Academy of Sciences, Shanghai, China

Xiu-Wei Yang - Peking University, Beijing, China

Zhen Yang - Peking University, Beijing, China

Xin-Sheng Yao - Shenyang Pharmaceutical University, Shenyang, China

Wen-Cai Ye - Jinan University, Guangzhou, China

Yang Ye - Chinese Academy of Sciences, Shanghai, China

Da-Li Yin - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Biao Yu - Chinese Academy of Sciences, Shanghai, China

Jian-Min Yue - Chinese Academy of Sciences, Shanghai, China

Guo-Lin Zhang - Chinese Academy of Sciences, Chengdu, China

Li-He Zhang - Peking University, Beijing, China

Jun Zhou - Chinese Academy of Sciences, Kunming, China

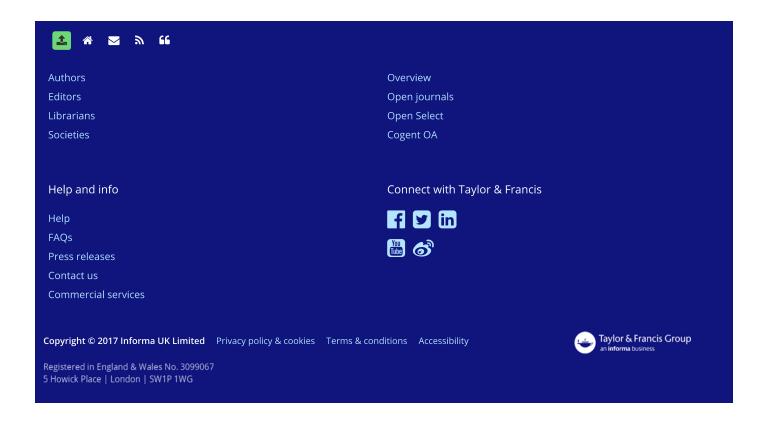
Wei-Dong Zhang - Second Military Medical University, Shanghai, China

Zhong-Mei Zou - Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China









This article was downloaded by: [Syah, Yana M.]

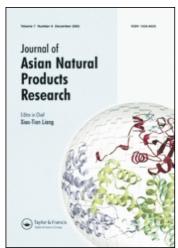
On: 5 November 2009

Access details: Access Details: [subscription number 916566229]

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House,

37-41 Mortimer Street, London W1T 3JH, UK



Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713454007

Macagigantin, a farnesylated flavonol from Macaranga gigantea

Mulyadi Tanjung ^a; Euis H. Hakim ^a; Didin Mujahidin ^a; Muhammad Hanafi ^b; Yana M. Syah ^a ^a Natural Products Chemistry Research Group, Organic Chemistry Division, Institut Teknologi Bandung, Bandung, Indonesia ^b Indonesian Institute of Science, Research Center for Chemistry, Tangerang, Indonesia

Online Publication Date: 01 November 2009

To cite this Article Tanjung, Mulyadi, Hakim, Euis H., Mujahidin, Didin, Hanafi, Muhammad and Syah, Yana M.(2009)'Macagigantin, a farnesylated flavonol from Macaranga gigantea', Journal of Asian Natural Products Research, 11:11,929 — 932

To link to this Article: DOI: 10.1080/10286020903302315 URL: http://dx.doi.org/10.1080/10286020903302315

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



Macagigantin, a farnesylated flavonol from Macaranga gigantea

Mulyadi Tanjung^a, Euis H. Hakim^a, Didin Mujahidin^a, Muhammad Hanafi^b and Yana M. Syah^a*

^aNatural Products Chemistry Research Group, Organic Chemistry Division, Institut Teknologi Bandung, Jalan Ganesha 10, Bandung 40132, Indonesia; ^bIndonesian Institute of Science, Research Center for Chemistry, Serpong, 15310 Tangerang, Indonesia

(Received 29 April 2009; final version received 1 September 2009)

A new farnesylated flavonol derivative, macagigantin (1), together with two known flavonoids, glyasperin A (2) and apigenin (3), had been isolated from the acetone extract of the leaves of *Macaranga gigantea*. The structure of the new compound was elucidated as 6-farnesylkaempferol based on its spectroscopic data, including UV, IR, 1D and 2D NMR, and HR-EI-MS spectra. Compounds 1-3 were evaluated for their cytotoxic properties against P-388 cells, their IC₅₀ values being 11.3, 6.0, and 5.1 μ M, respectively.

Keywords: macagigantin; farnesylated flavonol; *Macaranga gigantea*; Euphorbiaceae; cytotoxicity; P-388 cells

1. Introduction

The genus *Macaranga* (Euphorbiaceae) contains about 250 species distributed very widely from Africa and Madagascar in the west to tropical Asia, North Australia, and Pacific Islands in the east [1]. This genus has been shown to produce a number of phenolic compounds, particularly flavonoids and stilbenoids [2]. Recently, we have reported the first dihydrochalcone derivatives, along with flavanones, from an endemic *Macaranga* species in Indonesia, Macaranga trichocarpa [3]. In continuation of our phytochemical work on Indonesian tropical plants aiming to find new cytotoxic compounds [4-9], we had examined another *Macaranga* species, Macaranga gigantea (Reichb.f. & Zoll.) Müll. Arg. In this paper, we report the isolation of a farnesylated flavonol, 6farnesylkaempferol (1), together with two known compounds 2 and 3 (Figure 1), from the acetone extract of the title plant leaves. The cytotoxic properties of compounds 1-3 against murine leukemia P-388 cells are also briefly described.

2. Results and discussion

Compound 1 was isolated as a yellow solid and the molecular formula $C_{30}H_{34}O_6$ was deduced from its HR-EI-MS data. The UV spectrum of 1 exhibited maxima typical for a flavonol structure (λ_{max} 271, 348, and 283 nm), and showed bathochromic shifts on addition of AlCl₃ and NaOAc [10]. The IR spectrum indicated absorptions for hydroxyl (3419 cm⁻¹), conjugated carbonyl (1649 cm⁻¹), and aromatic (1608 and 1560 cm⁻¹) groups. In the ¹³C NMR spectrum (APT experiment; Table 1), 28 carbon signals representing 30 carbon atoms were observed. Two of them,

Figure 1. Structures of flavonoids 1-3.

namely the signals at $\delta_{\rm C}$ 136.5 and 176.5, are characteristic for C-3 and C-4 of a flavonol structure [11,12]. These spectroscopic data, therefore, suggested that 1 is a flavonol containing a C-15 side chain. Moreover, the presence of other five oxyaryl signals ($\delta_{\rm C}$ 146.6, 155.6, 158.1, 160.0, and 162.6) indicated that the flavonol is a derivative of kaempferol.

The side chain was deduced to be a farnesyl group from the observation in the ¹H NMR spectrum (Table 1) of four methyl singlets at $\delta_{\rm H}$ 1.51, 1.54, 1.57, and 1.79, five methylene signals at $\delta_{\rm H}$ 1.8–3.5, and three methine vinyl signals of three substituted alkenes at $\delta_{\rm H}$ 5.00, 5.06, and 5.29. The presence of the proton signals of a pair of doublets $(J = 9.0 \,\mathrm{Hz})$ in the aromatic region at $\delta_{\rm H}$ 7.00 and 8.12 (each 2H), assignable to the signals of a phydroxyphenyl group, and a singlet at $\delta_{\rm H}$ 6.59, suggested that the farnesyl group is either at C-6 or C-8 of the kaempferol structure. The presence of long-range correlations in the HMBC spectrum of 1 between the proton signal of a chelated -OH group at $\delta_{\rm H}$ 12.41 and three quaternary carbon signals at $\delta_{\rm C}$ 104.0 (C-4), 158.1 (C-5), and 111.7 (C-6) unambiguously placed the farnesyl group at C-6.

Therefore, compound **1**, trivially named macagigantin, was elucidated as 6-farnesylkaempferol or 6-[(2*E*,6*E*)-3",7",11"-trimethyldodeca-2",6",10"-trienyl]-kaempferol. Other HMBC correlations consistent with structure **1** are shown in Table 1. Further support for structure **1** was also obtained from the comparison of the NMR spectral data with those reported for 6-geranylkaempferol (macarangin) [11,12]. To our knowledge, compound **1** was the first example of flavonoid in the genus *Macaranga* with a farnesyl side chain.

On cytotoxic evaluation against P-388 cells, compounds 1-3 exhibited IC₅₀ values of 11.3 ± 0.4 , 6.0 ± 0.9 , and $5.1 \pm 0.7 \,\mu\text{M}$, respectively (artonin E as a positive control, IC₅₀ 1.38 ± 0.2). These cytotoxic data suggested that the presence of a terpenoid (farnesyl) substituent at C-6 (ring A) of the kaempferol structure reduces cytotoxic properties, while a similar substituent (isoprenyl) in the ring B enhances cytotoxicity.

3. Experimental

3.1 General experimental procedures

UV and IR spectra were measured with a Varian 100 Conc and an FT-IR Spectrum

Table 1. The NMR spectral data (d_6 -acetone) of compound 1.

No.	δ_{H} (multiplicity, J in Hz)	$\delta_{ m C}$	$HMBC (H \Leftrightarrow C)$
2	_	146.6	_
3	_	136.5	_
4	_	176.5	_
4a	_	104.0	-
5	_	158.1	-
6	_	111.7	_
7	_	162.6	-
8	6.59 (s)	93.8	C-4, C-4a, C-6, C-7, C-8a
8a	_	155.6	_
1'	_	123.4	-
2'/6'	8.12 (d, 9.0)	130.3	C-2, C-4', C-6'/2'
3'/5'	7.00 (d, 9.0)	116.2	C-4', C-1', C-5'/3'
4′	_	160.0	_
1"	3.37 (d, 6.7)	21.9	C-5, C-6, C-7, C-2", C-3"
2"	5.29 (tm, 6.7)	123.2	C-1", C-4", C-15"
3"	_	135.1	_
4"	1.97 (t, 7.3)	40.4	C-2", C-5", C-3", C-6", C-15"
5"	2.06 (br q, 7.3)	27.0	C-4", C-6", C-7"
6"	5.06 (br t, 7.3)	124.8	C-5", C-8", C-14"
7"	_	135.4	_
8"	1.85 (br t, 7.3)	40.4	C-6", C-7", C-9", C-10", C-14"
9"	1.93 (br q, 7.3)	27.3	C-8", C-10", C-11"
10"	5.00 (br t, 7.3)	125.0	C-9", C-12", C-13"
11"	_	131.5	-
12"	1.57 (br s)	25.8	C-10", C-11", C-13"
13"	1.51 (br s)	17.6	C-10", C-11", C-12"
14"	1.54 (br s)	16.2 ^a	C-6", C-7", C-8"
15"	1.79 (br s)	16.1 ^a	C-2", C-3", C-4"
5-OH	12.41 (s)		C-4a, C-5, C-6

^aAssignments could be interchanged.

One Perkin-Elmer instrument, respectively. 1H and 13C NMR spectra were recorded with a JEOL ECA500 spectrometer operating at 500 (¹H) and 125 (13C) MHz, using residual and deuterated solvent peaks ($\delta_{\rm H}$ 2.04 and $\delta_{\rm C}$ 29.8, respectively) as reference standards. Mass spectra were obtained with a VG Autospec mass spectrometer (EI mode). Vacuum liquid chromatography (VLC) and radial chromatography were carried out using Si gel 60 G (article no. 1.07731.1000, Merck KgaA, 64271 Darmstadt, Germany) and Si gel 60 PF₂₅₄ (article no. 1.07749.1000, Merck KgaA, 64271), respectively, and for TLC analysis, precoated Si gel 60 F₂₅₄ plates (article no. 1.05554.0001, Merck KgaA, 64271) were used. Solvents used for extraction and separation were of technical grades that were distilled before use.

3.2 Plant material

The leaves of *M. gigantea* were collected in November 2007 from Lungkut Layang Village, District Kapuas, Kalimantan, Indonesia. The specimen was identified by Mr Ismail, Herbarium Bogoriense, Center of Biological Research and Development, National Institute of Science, Bogor, Indonesia, and the voucher specimen has been deposited at the herbarium.

3.3 Extraction and isolation

The dried and powdered leaves of *M. gigantea* (1.2 kg) were macerated in

acetone at room temprature $(3 \times , 7.5 \text{ liters})$, and the acetone extract was evaporated under reduced pressure to give a crude acetone extract (50 g). The extract was fractionated by VLC on silica gel eluted with *n*-hexane–EtOAc mixtures [17:3 (4 \times) and $4:1 (4\times)$] to give five major fractions A-E. On TLC analysis, fraction D (1g) showed two major spots ($R_{\rm f}$ 0.71 and 0.43, eluent $CHCl_3-EtOAc = 9:1$) and on purification of this fraction using centrifugal planar chromatography eluted with nhexane-CHCl₃ (7:3 \rightarrow 1:9), CHCl₃, and CHCl₃-EtOAc (9:1) afforded macagigantin (1) (23 mg) and glyasperin A (2) (64 mg) [13]. The same procedure was applied to fraction E (330 mg) to give apigenin (3) (25 mg) [14].

3.3.1 Macagigantin (1)

A yellow solid. UV (MeOH) λ_{max} (log ε): 204 (4.58), 271 (4.20), 333 (4.13), 368 (4.18) nm, (MeOH+NaOH) 210 (4.90), 240 (sh, 4.32), 279 (4.22), 322 (4.09), 417 (4.27) nm, $(MeOH + AlCl_3)$ 203 (4.59), 273 (4.27), 347 (3.85), 431 (4.25) nm, (MeOH + AlCl₃/HCl) 203 (4.58), 272 (4.26), 348 (3.86), 431 (4.24) nm; (MeOH + NaOAc) 203 (4.85), (4.21), 333 (4.07), 368 (4.06) nm; IR (KBr) ν_{max} : 3419 (OH), 2922, 2852 (CHalkyl), 1649 (conj. C=O), 1608, 1560 (C=C aromatic) cm⁻¹; ¹H NMR (500 MHz) spectral data, see Table 1; ¹³C NMR (125 MHz) spectral data, see Table 1; HR-EI-MS m/z: 490.2357 [M]⁺ (calcd for $C_{30}H_{34}O_6$, 490.2355); EI-MS *m*/*z* (% rel.): 490 (17), 421 (24), 353 (56), 337 (15), 311 (12), 299 (100), 121 (10), 69 (25).

3.4 Cytotoxic evaluation

Cytotoxic properties of the isolated compounds 1-3 against murine leukemia P-388 cells were evaluated according to the method of MTT assay as described previously [15].

Acknowledgements

The authors are grateful for the financial support from the office of the Ministry of National Education, Republic of Indonesia (Hibah Pasca Grant VII 2009). We also thank Associate Prof. E.L. Ghisalberti, the University of Western Australia, for mass spectra measurements.

References

- [1] F.R. Blattner, K. Weising, G. Banfer, U. Maschwitz, and B. Fiala, Mol. Phyl. Evol. 19, 331 (2001).
- [2] B.J. Yoder, S. Cao, A. Norris, J.S. Miller, F. Ratooson, J. Razafitsalama, R. Andriantsiferana, V.E. Rasamison, and D.G.I. Kingston, J. Nat. Prod. 70, 342 (2007).
- [3] Y.M. Syah, E.H. Hakim, S.A. Achmad, M. Hanafi, and E.L. Ghisalberti, *Nat. Prod. Commun.* 4, 63 (2009).
- [4] E.H. Hakim, S.A. Achmad, L.D. Juliawaty, L. Makmur, Y.M. Syah, N. Aimi, M. Kitajima, H. Hiromitsu, and E.L. Ghisalberti, J. Nat. Med. 60, 161 (2006).
- [5] Y.M. Syah, L.D. Juliawaty, E.H. Hakim, S.A. Achmad, and E.L. Ghisalberti, *J. Nat. Med.* **60**, 308 (2006).
- [6] Muhtadi, E.H. Hakim, L.D. Juliawaty, Y.M. Syah, S.A. Achmad, J. Latif, and E.L. Ghisalberti, *Fitoterapia* 77, 550 (2006).
- [7] H. Saroyobudiono, L.D. Juliawaty, Y.M. Syah, S.A. Achmad, and E.H. Hakim, J. Nat. Med. 62, 195 (2008).
- [8] Ferlinahayati, E.H. Hakim, Y.M. Syah, L.D. Juliawaty, H. Takayama, I.M. Said, and J. Latip, Z. Naturforsch. 63c, 35 (2008).
- [9] I. Musthapa, L.D. Juliawaty, Y.M. Syah, E.H. Hakim, J. Latip, and E.L. Ghisalberti, Arch. Pharm. Res. 32, 191 (2009).
- [10] T.J. Mabry, K.R. Markham, and M.B. Thomas, *The Systematic Identification of Flavonoids* (Springer-Verlag, New York, 1970), pp. 41–164.
- [11] E. Hnawia, O. Thoison, F. Gueritte-Voegelein, D. Bourret, and T. Sevenet, *Phytochemistry* 29, 2367 (1990).
- [12] S. Sutthivaiyakit, S. Unganont, and P. Sutthivaiyakit, *Tetrahedron* 58, 3619 (2002).
- [13] L. Zeng, T. Fukai, T. Nomura, R.Y. Zhang, and Z.C. Lou, *Heterocycles* 34, 575 (1992).
- [14] T.J. Batterham and R.J. Highet, Aust. J. Chem. 17, 428 (1964).
- [15] Sahidin, E.H. Hakim, L.D. Juliawaty, Y.M. Syah, L.B. Din, E.L. Ghisalberti, J. Latip, I.M. Said, and S.A. Achmad, Z. Naturforsch. 60c, 723 (2005).