Bioprospecting thermostable enzymes-producing thermophiles from Indonesia

by Sucipto Hariyanto

Submission date: 09-Mar-2020 06:43PM (UTC+0800)

Submission ID: 1272185308

File name: g-thermostable-enzymes-producing-thermophiles-from-Indonesia.pdf (81.34K)

Word count: 2621

Character count: 15128

Eco. Env. & Cons. 25 (July Suppl. Issue) : 2019; pp. (S57-S61) Copyright@ EM International ISSN 0917–765X

Bioprospecting thermostable enzymes-producing thermophiles from Indonesia

Almando Geraldia, Aken Puti Wanguyun and Sucipto Hariyanto

¹Department of Biology, Faculty of Science and Technology, UniversitasAirlangga, Surabaya, Indonesia

(Received 27 March, 2019; Accepted 30 May, 2019)

ABSTRACT

Thermostable enzymes-producing thermophilic microorganisms have been of industrial interest due to their ability to work optimally in high temperature which ensures high reaction efficiency and productivity, as well as reduces contamination risks. Efforts on isolating thermophilic microorganism with industrial potential have long been conducted in Indonesia, where there is large number of hot springs and geothermal area. Bacterial strains with the ability to produce thermostable industrial enzymes such as amylases, chitinase, lipase, protease, and xylanase have been isolated from various regions in Indonesia. This review is the first endeavor to collectively look into the researches on thermophilic microorganisms with industrial potential in Indonesia, their outcomes and proposed future directions based on the limitation of current studies.

Key words: Thermophilic Microorganisms, Thermostable Enzymes, Hot Springs, Indonesia, Bioindustry

Introduction

Thermophilic microorganisms which grow optimally at temperature above 50 °C, have been of great interest in bioindustry. Thermostable enzymes produced by those microorganisms can be utilized in bioprocess involving high temperature (60-105 °C) such as starch liquefaction and lignocellulosic materials saccharification (Elleuche et al., 2014). Furthermore, thermophilic microorganisms are promising hosts for fermentation offering a number potential of advantage over mesophilic hosts. Thermophilic hosts work optimally at high temperature comparable to chemical refineries ensuring high efficiency and productivity as shown in the production of fructose, methane, and hydrogen (Zeldes et al., 2015). Moreover, high-temperature bioprocessing offers reduced risks of contamination from unwanted microorganisms and phage infection, reduced cooling costs of the fermenter, as well

as reduced water and energy consumption for sterilization (Chen and Jiang, 2018).

Due to the numerous advantages on bioindustry, the diversity and potential of thermophilic microorganisms have been widely explored. Survey and exploration to hot springs, geothermal fields, hydrothermal vents, desert, and even man-made environments such as compost facilities resulted in the discovery of novel thermophilic microorganism strains and species with industrial potential (Urbieta *et al.*, 2015)

One of the best areas to explore the biodiversity thermophilic microorganisms with industrial potential is Indonesia. As one of the most tectonically active areas in the world, with over 70 volcanoes, Indonesia has substantial number of hot springs and geothermal regions which harborlarge communities of thermophilic bacteria (Yohandini *et al.*, 2015). Efforts to isolate novel strain and species of thermophilic microorganisms from those ecosystems and

Corresponding author email: almando.geraldi@fst.unair.ac.id

screen the ability of isolated microorganisms to produce thermostable industrial enzymes have long been conducted.

In this review, the recent discovery of thermophilic microorganisms with the ability to produce thermostable industrial enzymes in Indonesia will be discussed. This review is the first that focusing the current status of exploration of thermophilic microorganisms with industrial potential in Indonesia.

Area and methods of exploration

Based on Table 1, most of thermophilic microorganism explorations in the last 10 years in Indonesia were conducted in Java, Sumatera, and Sulawesi islands. Interestingly some of the studied environments are suitable for hyperthermophiles where temperature reached >80°C and thermophiles-acidophiles where pH was below 4 (Urbieta *et al.*, 2015). The biodiversity of thermophiles in those ecosystems were mainly explored using culture-dependent methods such as traditional microorganism culture and culture-independent methods using molecular biology techniques.

Although, traditional microorganism culture methods have been important in characterizing the biochemical and physiological properties, as well as potential application of thermophiles isolates, however, this method is only able to reveal limited number of thermophiles. This is due to the commonly understood fact that only small fraction of microorganism in a given environment is culturable (Boteva and Kambourova, 2018; Chaudhary *et al.*, 2019). Most of the isolates obtained using this method are from order Bacilliales family Paenibacillaceae and

Bacillaceae. The member of family Paenibacillaceae isolated from Tanjung (Brevibacillusthermoruber) and Cisolok (Paenibacilluscisolokensis) (Yohandini et al., 2015; Yokota et al., 2016) While the isolated member of Bacillaceae were Bacillus sp from West Sumatera, as well as Anoxybacillusrupiensis, Anoxybacillus flavithermus, and Geobacilluspallidus, from South Sumatera (Harnentis et al., 2013; Yohandini et al., 2015). Even isolates from the same genus (Bacillusspp) were obtained from the explorationsin Sumatera (Jambi) and Sulawesi (North Sulawesi) islands, where the sampling area have different characteristics and are distantly apart (more than 4000 km) (Arzita et al., 2017; Simanjuntak and Samuel, 2018).

A more inclusive approach on studying biodiversity of thermophiles was represented by the use of molecular biology techniques such as 16S rRNA gene sequencing of environmental DNA and denaturing gradient gel electrophoresis (DGGE). The use of those methods successfully revealed the abundance of unique and previously unculturable microorganisms such as members of genus *Ralstonia, Delftia*, and *Thermus* in Gedongsongo and members of phylum Crenarchaeota in Kamojang (Aditiawati *et al.*, 2009; Aminin *et al.*, 2008).

Thermophilic microorganisms with industrial potential

Thermophilic microorganism explorations in Indonesia have been conducted mainly for these 3 purposes: [1]Bioprospectingthermophilic microorganisms with industrial potential, [2] Profiling the microbial communities in hot ecosystems, and [3] Dis-

Table 1. Selected exploration area for thermophiles in Indonesia

Name	Area	Temperature (°C)	рН	References
Tanjung Sakti hot spring	South Sumatera	80-91	7-8	(Yohandini et al., 2015)
Hot spring at South Solok District	West Sumatera	75-95	8	(Harnentis et al., 2013)
Sungai Tutunghot spring at Kerinci Seblat National Park	Jambi (Sumatera)	70-85	8.4	(Arzita <i>et al.</i> , 2017)
Kalianda Island coastal hot springs	Lampung (Sumater	a) 58.5-68.5	6.0-6.5	(Xu et al., 2013)
Geyser at Cisolok	West Java	70-80	7	(Yokota et al., 2016)
Kawah Hujan B hot spring, Kamojang	West Java	90-92	1.8-1.9	(Aditiawati et al., 2009)
GS-2 hot spring at Gedongsongo field	Central Java	50	4	(Aminin et al., 2008)
Lake Linow hot mud	North Sulawesi	90-110	7.08-8.35	(Simanjuntak and Samuel, 2018)

covering of novel. Among them, bioprospecting of thermophilic microorganisms with thermostable industrial enzymes activities has become the main goal of the most of the explorations.

As shown in table 2, a wide variety of industrially important enzymes were produced by thermophilic bacteria isolated from hot springs in Indonesia. Thermostable agarose is an important enzyme for agar hydrolysis in microbiological media industry and extract biological substances, such as unsaturated fatty acids, vitamins, and carotenoids from algae (Li *et al.*, 2014). Thermostable amylase can be utilized for starch gelatinization and liquefication in the food and bioethanol industries (Ibrahim *et al.*, 2013). Xylanase and mannase with high stability would increase the efficiency of biobleaching, food and feed processing and the solubilization of ligno-

cellulose for the production of second generation bioethanol (Elleuche *et al.*, 2015), while thermostable protease and lipase mainly used in food and detergent industries (Rigoldi *et al.*, 2018).

Future Perspectives

The efforts on discovering thermophilic microorganisms with industrial potential in Indonesia have been conducted for around 20 years (Huber *et al.*, 1996; Lee *et al.*, 1999; Nam *et al.*, 2002). Since then, numerous explorations have been conducted in various hot ecosystems in Indonesia with a small number mentioned in table 2. However, although volcanic and geothermal area are spread across Indonesia, current explorations are still limited to Java, Sumatera, and Sulawesi islands. Future exploration efforts in another area (i.e. Lesser Sunda Is-

Table 2. Selected Thermostable Enzymes-producing Thermophilic Microorganisms isolated from various area in Indonesia

Enzymes	Source	Associated ecosystem	Properties	References
Agarase	Bacillus sp. BI-3	Kalianda Island coastal hot spring, Lampung	• Work optimally at 70°C, pH 6.4	(Li et al., 2014)
Amylase	<i>Bacillus</i> sp. RSII-1b	Lejja hot sprin, South Sulawesi	 Work optimally at 55°C-60°C, pH 6.0 Activity of 0.165U/mL crude extract 	(Arfah <i>et al.</i> , 2015)
	Bacillus licheniformisBT5.9	Cangar hot spring, East Java	 α-amylase with optimum expression at 50°C, pH 5.0 Activity of 0.327 U/ml crude extract 	(Ibrahim <i>et al.</i> , 2013)
Chitinase	Bacillus licheniformis	Cangar hot spring, East Java	• Expressed at 52°C, pH 7.0	(Chrisnasari et al., 2018)
Lipase	DMS-3 isolate (rod-shaped, Gram positive bacteria)	Domas hot spring, West Java	 Alkaline lipase with optimum expression at 70°C, pH 9.0 	(Febriani <i>et al.</i> , 2010)
Mannase	Bacillus sp	Hot springs at South Solok District, West Sumatera	 Work optimally at 55°C-60°C, pH 6.0 Activity of 0.165U/mL crude extract 	(Harnentis et al., 2013)
Protease	Fictibacillusgelatini	Sungai Tutung hot spring, Jambi	 Alkaline protease expressed at 60°C, pH 8.0 Proteolytic index of 6.15 	(Arzita <i>et al.,</i> 2017)
	<i>Brevibacillus</i> sp PLI-1	Kalianda Island coastal hot spring, Lampung	 Alkaline protease expressed at 70°C, pH 8.0-9.0 Proteolytic index of 10.3 	(Wang <i>et al.,</i> 2012)
Xylanase	Bacillussp	Sapan Sungai Aro hot spring, West Sumatera	 Expressed at 60°C Xylanolytic index of 0.74 	(Irdawati <i>et al.,</i> 2018)
	Paenibacillussp. XJ18	TNBD Forest, Jambi	 The highest activity showed at 90°C, pH 5.0. 	(Kurrataa'yun and Meryandini, 2015)

lands and Molucca Archipelago) would provide valuable information regarding the biodiversity of beneficial thermophiles in Indonesia.

Furthermore, a major limitation of current study on thermophiles with industrial potential in Indonesia is that very few or even almost no attempts have been made towards development to commercial scale. Thus, efforts to bridge the gap between research on thermophilic microorganism potential and commercialization need to be carried out.

Acknowledgement

The authors would like to express our gratitude to the Faculty of Science and Technology, Universitas Airlangga through the scheme of RKAT 2019 No. 2419/UN3.1.8/LT/2019 for funding this research and sponsoring this publication.

References

- Aditiawati, P., Yohandini, H., Madayanti, F. and Akhmaloka. 2009. Microbial Diversity of Acidic Hot Spring (Kawah Hujan B) in Geothermal Field of Kamojang Area, West Java-Indonesia. *The Open Mi*crobiology Journal. 3: 58–66.
- Aminin, A. L. N., Warganegara, F. M. and Aditiawati, P. 2008. Culture-independent and Culture-dependent Approaches on Microbial Community Analysis at Gedongsongo (GS-2) Hot Spring. *International Journal of Integrative Biology*. 2 (3): 145-152.
- Arfah, R. A., Ahmad, A., Djide, M. N., Anis, M. and Zakir, M. 2015. Production Optimization and Characterization of Amylase Enzyme Isolated from Termofil Bacteria *Bacillus* sp RSAII-1b from Lejja Hot Spring South Sulawesi. *American Journal of Biomedical and Life Sciences*. 3 (6): 115–119.
- Arzita, S., Agustien, A. and Rilda, Y. 2017. The Diversity of the Alkaline Protease Producers, Thermophilic Obligate *Bacillus* spp., from Sungai Tutung Hot Spring, Kerinci, Jambi, Indonesia. *Journal of Pure and Applied Microbiology*. 11 (4): 1789–1797.
- Boteva, N. and Kambourova, M. 2018. Thermophiles and Their Exploration for Thermostable Enzyme Production, In *Extremophiles in Eurasian Ecosystems: Ecol*ogy, Diversity, and Applications, Springer, pp. 167– 186.
- Chaudhary, D. K., Khulan, A. and Kim, J. 2019. Development of a Novel Cultivation Technique for Uncultured Soil Bacteria. Scientific Reports 9 (1): 6666.
- Chen, G. Q. and Jiang, X. R. 2018. Next Generation Industrial Biotechnology Based on Extremophilic Bacteria. Current Opinion in Biotechnology. 50:94–100.

- Chrisnasari, R., Verina, D., Tapatfeto, A. C., Pranata, S., Patjajani, T., Wahjudi, M. and Purwanto, M. G. M. 2018. Isolating and Characterising Chitinolytic Thermophilic Bacteria from Cangar Hot Spring, East Java. Pertanika Journal of Tropical Agricultural Science 41(3): 1437-1448.
- Elleuche, S., Schäfers, C., Blank, S., Schröder, C. and Antranikian, G. 2015. Exploration of Extremophiles for High Temperature Biotechnological Processes. *Current Opinion in Microbiology*. 25: 113–119.
- Elleuche, S., Schroder, C., Sahm, K. and Antranikian, G. 2014. Extremozymes—Biocatalysts with Unique Properties from Extremophilic Microorganisms. *Current Opinion in Biotechnology*. 29: 116–123.
- Febriani, I., Hertadi, R., Kahar, P. and Akhmaloka, M. F. 2010. Isolation and Purification of Novel Thermostable Alkaline Lipase from Local Thermophilic Microorganism. *Biosciences Biotechnology Research* Asia. 7(2): 617–622.
- Harnentis, H., Marlida, Y., Rizal, Y. and Mahata, M. E. 2013. Isolation, Characterization and Production of Mannanase from Thermophilic Bacteria to Increase the Feed Quality. *Pakistan Journal of Nutrition*. 12(4): 360–364.
- Huber, R., Rossnagel, P., Woese, C. R., Rachel, R., Langworthy, T. A. and Stetter, K. O. 1996. Formation of Ammonium from Nitrate during Chemolithoautotrophic Growth of the Extremely Thermophilic Bacterium Ammonifex degensii gen. nov. sp. nov. Systematic and Applied Microbiology. 19(1): 40–49.
- Ibrahim, D., Zhu, H. L., Yusof, N., Isnaeni and Hong, L. S. 2013. Bacillus licheniformis BT5.9 Isolated from Changar Hot Spring, Malang, Indonesia as a Potential Producer of Thermostable α-amylase. Tropical life Sciences Research. 24 (1): 71–84.
- Irdawati, I., Syamsuardi, S., Agustien, A. and Rilda, Y. 2018. Screening of Thermophilic Bacteria Produce Xylanase from Sapan Sungai Aro Hot Spring South Solok, In *IOP Conference Series: Materials Science and Engineering*, Vol. 335, IOP Publishing, p. 12021.
- Kurrataa'yun and Meryadini, A. 2015. Characterization of Xylanase Activity Produced by Paenibacillus sp. XJ18 from TNBD Jambi, Indonesia. HAYATI Journal of Biosciences. 22(1): 20–26.
- Lee, D. W., Koh, Y. S., Kim, K. J., Kim, B. C., Choi, H. J., Kim, D. S., Suhartono, M. T. and Pyun, Y. R. 1999. Isolation and Characterization of a Thermophilic Lipase from *Bacillus thermoleovorans* ID-1, *FEMS Microbiology Letters*. 179 (2): 393–400.
- Li, J., Sha, Y., Seswita-Zilda, D., Hu, Q. and He, P. 2014. Purification and Characterization of Thermostable Agarase from *Bacillus* sp. BI-3, a Thermophilic Bacterium Isolated from Hot Spring. *Journal of Microbiology and Biotechnology*. 24 (1): 19–25.
- Nam, G. W., Lee, D. W., Lee, H. S., Lee, N. J., Kim, B. C.,

- Choe, E. A., Hwang, J.-K., Suhartono, M. T. and Pyun, Y. R. 2002. Native-feather Degradation by Fervidobacterium islandicum AW-1, a Newly Isolated Keratinase-producing Thermophilic Anaerobe. Archives of Microbiology. 178 (6): 538–547.
- Rigoldi, F., Donini, S., Redaelli, A., Parisini, E. and Gautieri, A. 2018. Engineering of Thermostable Enzymes for Industrial Applications. APL bioengineering. 2(1): 11501.
- Simanjuntak, S. and Samuel, M. Y. 2018. Isolation and Identification of Thermophilic Bacteria, Producer of Amylase Enzyme, from Lake Linow, North Sulawesi. *Journal of Pure and Applied Microbiology*. 12(2): 543–554.
- Urbieta, M. S., Donati, E. R., Chan, K. G., Shahar, S., Sin, L. L. and Goh, K. M. 2015. Thermophiles in The Genomic Era: Biodiversity, Science, and Applications. *Biotechnology Advances*. 33 (6): 633–647.
- Wang, S., Lin, X., Huang, X., Zheng, L. and Zilda, D. S. 2012. Screening and characterization of the alkaline protease isolated from PLI-1, a strain of *Brevibacillus* sp. collected from Indonesia's hot springs. *Journal of*

- ocean University of China. 11(2): 213-218.
- Xu, S. Y., He, P. Q., Dewi, S. Z., Zhang, X. L., Ekowati, C., Liu, T. J. and Huang, X. H. 2013. Hydrogen-producing microflora and Fe–Fe hydrogenase diversities in seaweed bed associated with marine hot springs of Kalianda, Indonesia. Current Microbiology. 66 (5): 499–506
- Yohandini, H., Julinar and Muharni. 2015. Isolation and Phylogenetic Analysis of Thermophile Community Within Tanjung Sakti Hot Spring, South Sumatera, Indonesia. *HAYATI Journal of Biosciences*. 22(3): 143– 148
- Yokota, A., Ningsih, F., Nurlaili, D. G., Sakai, Y., Yabe, S., Oetari, A., Santoso, I. and Sjamsuridzal, W. 2016. Paenibacillus cisolokensis sp. nov., isolated from litter of a geyser. International Journal of Systematic and Evolutionary Microbiology. 66 (8): 3088–3094.
- Zeldes, B. M., Keller, M. W., Loder, A. J., Straub, C. T., Adams, M. W. W. and Kelly, R. M. 2015. Extremely thermophilic microorganisms as metabolic engineering platforms for production of fuels and industrial chemicals. Frontiers in Microbiology. 6: 1209.

Bioprospecting thermostable enzymes-producing thermophiles from Indonesia

ther	mophiles from Indonesia			
ORIGINA	ALITY REPORT			
SIMILA	3% 5% INTERNET SOURCES	11% PUBLICATIONS	3% STUDENT PAPERS	
PRIMAR	Y SOURCES			_
1	Submitted to Universita Student Paper	s Airlangga	3	6
2	Muhammad Fauzul Imr Kurniawan, Agoes Soeg "Characterization of me potential bacteria isolat active sanitary landfill le Indonesia under differe Journal of Environment	ianto. ercury-reducing ed from Keput eachate, Surab nt saline cond	zih non- aya, itions",	_
3	"Microbial Diversity in E Sustainability and Biote Applications", Springer Media LLC, 2019	chnological		%
4	Jujjavarapu Satya Eswar Ramkrishna Sen. "Therr Industry", Springer Scie	mophiles for B	iotech	6

Publication

Media LLC, 2019

5	Skander Elleuche, Christian Schäfers, Saskia Blank, Carola Schröder, Garabed Antranikian. "Exploration of extremophiles for high temperature biotechnological processes", Current Opinion in Microbiology, 2015 Publication	1 %
6	M. Sofía Urbieta, Edgardo R. Donati, Kok-Gan Chan, Saleha Shahar, Lee Li Sin, Kian Mau Goh. "Thermophiles in the genomic era: Biodiversity, science, and applications", Biotechnology Advances, 2015 Publication	1 %
7	Karl-Heinz Schleifer. "Phylum XIII. Firmicutes Gibbons and Murray 1978, 5 (Firmacutes [sic] Gibbons and Murray 1978, 5)", Systematic Bacteriology, 2009 Publication	1 %
8	Riffel, A "Purification and characterization of a keratinolytic metalloprotease from Chryseobacterium sp. kr6", Journal of Biotechnology, 20070220 Publication	1 %
9	repository.up.ac.za Internet Source	1 %
10	www.science.gov Internet Source	1 %

11	and Phylogenetic Analysis of Thermophile Community Within Tanjung Sakti Hot Spring, South Sumatera, Indonesia", HAYATI Journal of Biosciences, 2015	<1%
12	journal.ipb.ac.id Internet Source	<1%
13	"Extremophiles in Eurasian Ecosystems: Ecology, Diversity, and Applications", Springer Science and Business Media LLC, 2018 Publication	<1%
14	Almando Geraldi, Ni'matuzahroh, Fatimah, Chang-Hao Cui, Thi Thuy Nguyen, Sun Chang Kim. "Enzymatic biotransformation of ginsenoside Rb1 by recombinant β-glucosidase of bacterial isolates from Indonesia", Biocatalysis and Agricultural Biotechnology, 2020	<1%

Bioprospecting thermostable enzymes-producing thermophiles from Indonesia

GRADEMARK REPORT	
FINAL GRADE	GENERAL COMMENTS
/0	
PAGE 1	
PAGE 2	
PAGE 3	
PAGE 4	
PAGE 5	