African Journal of Infectious Diseases (AJID)

Vol.12 No 1S March 2018 ISSN 2006-0165 Journal articles safe health "santé est la richesse!" a life is today wealth



- HIV, HEPATITIS B Virus and Siphilis
- Incidence of dengue virus infections . . .
- Rapid evolution of hemorrhagic fever . . .

EDITORIAL BOARD

EDITORIAL OFFICE

COJA VILLA, No 7 Road 1, Otun Maye Square, Ajebamidele, Ile-Ife, Osun State, Nigeria

Managing Editor:

Mr. Babatunde O. OLAGUNJU; MSc (COJA VILLA, No 7 Road 1, Otun Maye Square, Ajebamidele, Ile-Ife, Osun State, Nigeria). E-mail: bolagunju@athmsi.org

Editorial Assistants:

Ms Omolade OLABOYE and Mr. Hammed IBRAHEEM, (COJA VILLA, No 7 Road 1, Otun Maye Square, Ajebamidele, Ile-Ife, Osun State, Nigeria).

Editors-in-Chief:

Prof. Anthony O. ONIPEDE, B.Sc; MBChB; MSc; FWACP (Lab. Med.) Deoartment of Medical Microbiology, Obafemi, Awolowo University, Ile-Ife, Nigeria. E-mail: anthony_onipede@yahoo.ca, editor@athmsi.org

Dr. Gbola OLAYIWOLA, BPharm; MSc; PhD.

Department of Clinical Pharmacy and Administration, Obafemi Awolowo University, Ile-Ife, Nigeria. E-mail: gbolaolayiwola@yahoo.com, editor@athmsi.org

Associate Editors:

Prof. Ademola OLANIRAN; PhD (University of KwaZulu-Natal, Durban, South Africa). E-mail: olanirana@ukzn.ac.za

Prof. Francesca MANCIANTI; DVM (Faculty of Medicine, University of Pisa, Italy). E-mail: francesca.mancianti@unipi.it

Dr. Emel SONMEZ; PhD (Anadolu University, Eskischir, Turkey). E-mail: emls222224@gmail.com

Prof. Hui WANG; DVM (Chinese Academy of Agricultural Sciences, Gansu, China). E-mail: wanghui01@caas.cn

Editorial Board:

Dr. Saajida MOHAMED; MMed, FPHM (University of KwaZulu-Natal, Durban, South Africa).

E-mail: mahomeds@ukzn.ac.za

Dr. Frank ONYAMBU; PhD (Kenya Institute of Applied Sciences, Moi University, Kenya). E-mail: frank.onyambu@iscb.org Dr. Ezekiel AKINKUNMI; PhD (Obafemi Awolowo University, Ile-Ife, Nigeria). E-mail: eoakinmi@oauife.edu.ng

Dr. Josyline KABURI; PhD (University of Nairobi, Kenya). E-mail: jeirindi@kemri.org

Dr. Balram OMAR; MD (King George's Medical Unieristy, Lucknow, India). E-mail: @gmail.com

Prof. Megbaru ABATE; PhD (Bahir Dar University, Ethiopia). E-mail: mgbeyney@gmail.com

Prof. Chrispinus MULAMBALAH; PhD (Moi University, School of Medicine, Kitale, Kenya). E-mail: csmulambalah@gmail.com

Dr. Alok KUMAR; MD (University of the West Indies, Cave Hill, Jamaica). E-mail: alokkumar.uwichill@gmail.com

Dr. Sharlene GOVENDER; PhD (Nelson Mandele Metropolitan University, Port Elizabeth, South Africa). E-mail: sharlene.govender@nmmu.ac.za

Dr. Babajide SADIQ; DrPH (Florida A&M University, Tallhassee, USA). E-mail: babajidesadiq@yahoo.com

Dr. Celsus SENTE; PhD (Makerere University, Kampala, Uganda). E-mail: csenhte@covab.mak.ac.ug

Prof. Phyllis KANKI; DVM, DSc. (Harvard School of Public Health, Boston, USA). E-mail: pkanki@hsph.harvard.edu

Prof. Vincent TITANJI; PhD (University of Buea, Cameroon). E-mail: vpk.titanji@yahoo.com

Prof. Ahmed ADU-OPPONG; PhD (Jiann-Ping Hsu College of Public Health, States Boro, USA). E-mail: aduoppong@georgiasouthern.edu

Dr. Suresh JOSHI; MD, PhD (Thomas Jefferson University Medical College, Phiadelphia, USA).

E-mail: surejoshi@yahoo.com

AFRICAN JOURNAL OF INFECTIOUS DISEASES

ISSN: 2006-0165,	EISSN: 2505-0419,	http://journals.sfu.ca/africanem/index.php/AJID
------------------	-------------------	---

Volume-12

No. 1S (Special Issue)

March 2018

Table of Contents

- IN VITRO STUDIES ON HEME OXYGENASE-1 AND P24 ANTIGEN HIV-1 LEVEL AFTERHYPERBARIC OXYGEN TREATMENTOFHIV-1 INFECTED ON PERIPHERAL BLOOD MONONUCLEAR CELLS (PBMCS) Retno Budiarti, Kuntaman Kuntaman, Muhammad Nasronudin, Muhammad Guritno Suryokusumo, Siti Qamariyah Khairunisa AJID, 12 (1S): 1-6
- GENOTYPING OF HUMAN PAPPILOMAVIRUS IN CERVICAL PRECANCEROUS LESION AND SQUAMOUS CELL CARCINOMA AT DR. SOETOMO HOSPITAL, SURABAYA, INDONESIA Gondo Mastutik, Rahmi Alia, Alphania Rahniayu, Anny Setijo Rahaju, Nila Kurniasari, Suhartono Taat Putra AJID, 12 (1S): 7-12
- DETERMINATION OF ENVIRONMENTAL FACTORS AFFECTING DENGUE INCIDENCE IN SLEMAN DISTRICT, YOGYAKARTA, INDONESIA Tri Wulandari Kesetyaningsih, Sri Andarini, Sudarto Sudarto, Henny Pramoedyo AJID, 12 (1S): 13-35
- ANTIVIRAL ACTIVITY OF Justicia gendarussa Burm.f. LEAVES AGAINST HIV-INFECTED MT-4 CELLS Agustinus Widodo, Prihartini Widiyanti, Bambang Prajogo AJID, 12 (1S): 36-43
- ACANTHAMOEBA SP.S-11 PHAGOCYTOTIC ACTIVITY ON MYCOBACTERIUM LEPRAE IN DIFFERENT NUTRIENT CONDITIONS Sepling Paling, Ratna Wahyuni, DEA Ni'matuzahroh, Dwi Winarni, M.KES, Iswahyudi, Linda Astari, Dinar Adriaty, Indropo Agusni, Shinzo Izumi AJID, 12 (1S): 44-48
- CD4+ AND CD8+ T-CELLS EXPRESSING INTERFERON GAMMA IN ACTIVE PULMONARY TUBERCULOSIS PATIENTS Betty Agustina Tambunan, Hery Priyanto, Jusak Nugraha, Soedarsono Soedarsono AJID, 12 (1S): 49-53

- THE ROLE OF PSYCHOLOGICAL WELL-BEING IN BOOSTING IMMUNE RESPONSE: AN OPTIMAL EFFORT FOR TACKLING INFECTION Abdurachman Latief, Netty Herawati AJID, 12 (1S): 54-61
- ANTIBACTERIAL ACTIVITY OF DRACONTOMELON DAO EXTRACTS ON METHICILLIN-RESISTANT S. AUREUS (MRSA) AND E. COLI MULTIPLE DRUG RESISTANCE (MDR) Yuniati Yuniati, Nurul Hasanah, Sjarif Ismail, Silvia Anitasari, Swandari Paramita AJID, 12 (1S): 62-67
- 9. INCREASED APOPTOSIS SKULL OF PUPS BORN TO TOXOPLASMA GONDII-INFECTED MICE ASSOCIATED WITH INCREASED EXPRESSION OF INTERFERON GAMMA, BUT NOT TUMOR NECROSIS FACTOR ALFA Lucia Tri Suwanti, Mufasirin Mufasirin AJID, 12 (1S): 68-71
- ADDITION OF ANTI- Toxoplasma gondii MEMBRANE IMMUNOGLOBULIN Y TO REDUCE NECROTIC INDEX IN MICE'S LIVER Heni Puspitasari, Lucia T. Suwanti, Mufasirin Djaeri AJID, 12 (1S): 72-75
- 11. SEROPREVALENCE AND RISK FACTOR OF TOXOPLASMOSIS IN SCHIZOPHRENIA PATIENTS REFERRED TO GRHASIA PSYCHIATRIC HOSPITAL, YOGYAKARTA, INDONESIA Nina Difla Muflikhah, Supargiyono Supargiyono, Wayan Tunas Artama AJID, 12 (1S): 76-82
- 12. CONCOMITANT SEXUALLY TRANSMITTED DISEASES IN PATIENTS WITH DIAGNOSED HIV/AIDS: A RETROSPECTIVE STUDY Densy Violina Harnanti, Afif Nurul Hidayati, Muhammad Miftahussurur AJID, 12 (1S): 83-89
- 13. RISK FACTORS OF VULVOVAGINAL CANDIDIASIS IN DERMATO-VENEREOLOGY OUTPATIENTS CLINIC OF SOETOMO GENERAL HOSPITAL, SURABAYA, INDONESIA Dharin Serebrina Arfiputri, Afif Nurul Hidayati, Samsriyaningsih Handayani, Evy Ervianti AJID, 12 (1S): 90-94

- 14. COMPARISON OF ANTI BACTERIAL EFFICACY OF PHOTODYNAMIC THERAPY AND DOXYCYCLINE ON AGGREGATIBACTER ACTINOMYCETEMCOMITANS Ernie Maduratna Setiawatie, Vina Puji Lestari, Suryani Dyah Astuti AJID, 12 (1S): 95-103
- 15. EVALUATION OF THE ANTIGENICITY AND IMMUNOGENICITY OF Eimeria tenella BY REPRODUCTIVE INDEX AND HISTOPATHOLOGICAL CHANGES OF CECAL COCCIDIOSIS VIRULENT LIVE VACCINE IN BROILER CHICKENS Endang Suprihati, Muchammad Yunus AJID, 12 (1S): 104-110
- 16. DETERMINATION OF EFFECTIVE DOSE OF ANTIMALARIAL FROM CASSIA SPECTABILIS LEAF ETHANOL EXTRACT IN PLASMODIUM BERGHEI-INFECTED MICE Wiwied Ekasari, Tutik Sri Wahyuni, Heny Arwaty, Nindya T. Putri AJID, 12 (1S): 111-115

17. A NEW COPPER (II)-IMIDAZOLE DERIVATIVE EFFECTIVELY INHIBITS REPLICATION OF DENV-2 IN VERO CELL Teguh Hari Sucipto, Siti Churrotin, Harsasi Setyawati, Fahimah Martak, Kris Cahyo Mulyatno, Ilham Harlan Amarullah, Tomohiro Kotaki, Masanori Kameoka, Masanori Kameoka, Subagyo Yotopranoto, Soegeng Soegijanto AJID, 12 (1S): 116-119

18. COMPARISON OF MULTIPLEX SINGLE ROUND PCR AND MICROSCOPY IN DIAGNOSIS OF AMOEBIASIS

BS Sri-Hidajati, Sukmawati Basuki, Suhintam Pusarawati, Kusmartisnawati Kusmartisnawati, Lynda Rossyanti, Sri Wijayanti Sulistyowati, Dwi Peni Kartikasari, Heny Arwati, Indah Tantular, Alpha Fardah, Andy Darma, Retno Handajani, Subijanto Marto Soedarmo

AJID, 12 (1S): 120-126

- 19. CLONING AND EXPRESSION OF MCE1A GENE FROM MYCOBACTERIUM TUBERCULOSIS BEIJING AND H37RV STRAIN FOR VACCINE CANDIDATE DEVELOPMENT Desi Indriarini, Andriansjah Rukmana, Andi Yasmon AJID, 12 (1S): 127-132
- 20. EFFECT OF VARYING INCUBATION PERIODS ON CYTOTOXICITY AND VIRUCIDAL ACTIVITIES OF Justicia gendarussa Burm.f. LEAF EXTRACT ON HIV-INFECTED MOLT-4 CELLS Prihartini Widiyanti, Bambang Prajogo, Agustinus Widodo AJID, 12 (1S): 133-139

- 21. IN SILICO SCREENING AND BIOLOGICAL EVALUATION OF THE COMPOUNDS OF Justicia gendarussa LEAVES EXTRACT AS INTERFERON GAMMA INDUCER: A STUDY OF ANTI HUMAN IMMUNODEFICIENCY VIRUS (HIV) DEVELOPMENT Restry Sinansari, Bambang EW Prajogo, Prihartini Widiyanti AJID, 12 (1S): 140-147
- 22. ISOLATION AND IDENTIFICATION OF BRUCELLA SUIS IN PIGS AS ZOONOTIC DISEASE IN ENDEMIC AREAS OF EAST JAVA, INDONESIA Emy S Koestanti, Wiwik Misaco, Sri Chusniati, Lilik Maslachah AJID, 12 (1S): 148-151
- 23. INSTRUCTIONS FOR AUTHORS Babatunde O Olagunju AJID, 12 (1S): 152-158

Koestanti et al., Afr., J. Infect. Dis. (2018) 12(S): 148-151

https://doi.org/10.2101/Ajid.12v1S.22

ISOLATION AND IDENTIFICATION OF *BRUCELLA SUIS* IN PIGS AS ZOONOTIC DISEASE IN ENDEMIC AREAS OF EAST JAVA, INDONESIA

Emy Koestanti S^{1*}, Wiwik Misaco², Sri Chusniati³, Lilik Maslachah⁴

¹Departmen of Animal Husbandry, ²Departement Clinic Veteriner, ³Department of Microbiology Veterinary, ⁴Department of Basic Medicine Veterinary, Faculty of Veterinary Medicine, Universitas Airlangga, C Campus UNAIR, Jl. Mulyorejo Surabaya 60115 Indonesia

*Corresponding Author Email: emykoestanti@yahoo.co.id

Article History

Received: March. 13, 2017 Revised Received: Sept. 09, 2017 Accepted: Sept. 21, 2017 Published Online: March. 07, 2018

Abstract

Background: Brucellosis in pigs at East Java Indonesia has not only cause great economic losses due to a decrease in productivity of livestock but also are zoonotic. Infection on free brucelosis pigs were initially begun with the infected pigs both male and female, or the use of superior male pigs together. The elimination of the disease either on a group or population is considered as the most effective way to prevent the spread of the disease in pigs. Prevention efforts mainly addressed to vaccination, sanitary maintenace and government policy. The purpose of this study was to isolated and identified *Brucella suis* as the causative agent.

Material and Methods: The survey area were the pig farm owned by breeder farmers in the area of East Java Indonesia, at Kediri, Malang, Blitar and Probolinggo district. Blood samples obtained were tested with RBT. Pigs are suspected of being infected with Brucella if the RBT was positive that characterized with agglutination in the test results. If RBT was positive, bacteriological examination will be performed, with samples of visceral foetus organ, ie liver, spleen, placenta and amniotic fluid. Isolation and identification of *Brucella suis* were used Brucella Broth and Brucella Agar, and if the bacteri growthwill be continued with biochemical test ie H2S, urease, citrate, catalase and oxidase test. The positive results of *Brucella suis* showed positive urease, catalase andoxidase,but negative for citrate and H2S.

Results: RBT and bacteriolgical examination showed that 1 sample was positive *Brucella suis*, and 19 negative. The positive results showed positive urease, catalase and oxidase,but negative for citrate and H2S

Conclusion: Based on RBT test and bacteriological examination, there was 1 positive sample of brucellla suis, that is sample coming from Kediri district.

Key words: Brucela suis, pig, isolation, identification, zoonotic

Introduction

Brucellosis is an infectious disease that can affect humans and animals (Alton et al., 1991). The cause of this disease is the bacterial genus *Brucella* which is an intracellular microorganism and can cause abortions and infertility (*orchitis* and *epididymitis*) in sheep, cattle, goats and pigs (Christi et al., 1968). This disease in humans is characterized with faint, fever, chills, sweating, pain in the joints, headaches and pain in the whole body (Priadi et al., 1992). Reservoirs of brucellosis due to *Brucella*. *suis* are wild animals and pigs (Corbel, 1985; Sudibyo, 1997). Infection on free brucelosis pigs were initially begun with the infected pigs both male or female, or the use of superior male pigs together. The spread of *Brucella suis* in pigs that bred by artificial insemination are also common (Corbel, 1997; Madkour, 1989). Piglets usually get the infection from their sow. The infection occurred at birth or fed up to infectious sow (Enright, F.M. 1990).

Screening tests or rapid test performed today in East Java, Indonesia still using *B. abortus* isolates of cattle species, that is S19, thus often occurs inaccurate results for the diagnosis of Brucellosis in pigs (Nicoletti, 1990).

Materials and Methods

The entire research was conducted appropriately following the ethics in using experimental animals and has been approved by the ethics commission of the Faculty of Veterinary Medicine, Universitas Airlangga.

The survey area were the pig farm owned by breeder farmers in the area of East Java Indonesia, at Kediri, Malang, Blitar and Probolinggo district. Blood serum samples and fetus originated from pigs who have abortus were collected. Blood samples were taken from pig that have experienced abortion, through the sow's ear veins to obtained the serum.

Serum samples were tested by RBT method (Corner and Alton, 1982) using artificial antigen commercial RBT made by Pusvetma Surabaya. The serum sample was mixed with antigen RBT. Both solutions placed on the glass object then stirred by rotating clockwise and then anti-clockwise direction gently for 2 minutes. The results obtained was negative : no agglutination occurred, a mixture of antigen antiserum looked pink homogeneous colored, (+1): seen a smooth agglutination and in perimeter seen as dotted line. (++): seen a clear smooth agglutination with wide margins in perimeter and a little bit clear of fluid around, (+++): seen a coarse / large agglutination and clear fluid around, dubious: seen agglutination in pink homogeneous colored. Pigs stated infected if RBT result was positive.

Isolation and identification of *Brucella suis* used Brucella Broth and Brucella Agar (Oxoid, England) with the addition of supplements. When the bacteria growth, it will be followed by a biochemical test namely H2S, urease, citrate, catalase and oxidase test. The positive results of *Brucella suis* showed positive urease, catalase and oxidase but negative for H2S and citrate.

Results

The survey showed that pig farms in Kediri, Malang, Blitar and Probolinggo showed the incidence of abortion. Twenty 20 sows showing symptoms of abortion. From Kediri, Malang Probolinggo and Blitar were obtained 10, 3, 5 and 2 respectively. The RBT showed that 1 sample from Kediri was positive and 19 other were negative (Table1)

Table 1: Rose Bengal Test Results

No.	Area		Result		
		Samples Amount	Positive	Negative	
1	Kediri	10	1	9	
2	Malang	3	0	3	
3	Probolinggo	5	0	5	
4	Blitar	2	0	2	

Table 2: Bacteriological Test

Table 2: Dact	0		TIOC	TT	Citerate	C + 1	0.11
Area	Number	Brucella agar	H2S	Urease	Citrate	Catalase	Oxidase
	of	media					
	Sample						
Kediri	1	negative	0	0	0	0	0
	2	negative	0	0	0	0	0
	3	growth	negative	Positive	negative	positive	positive
	4	negative	0	0	0	0	0
	5	negative	0	0	0	0	0
	6	negative	0	0	0	0	0
	7	negative	0	0	0	0	0
	8	negative	0	0	0	0	0
	9	negative	0	0	0	0	0
	10	negative	0	0	0	0	0
Malang	1	negative	0	0	0	0	0
	2	negative	0	0	0	0	0
	3	negative	0	0	0	0	0
Probolinggo	1	negative	0	0	0	0	0
	2	negative	0	0	0	0	0

One sample from Kediri showed growth in Brucella agar media, thus further processed for purification, and incubated for 3 days before the biochemical tests were done. Biochemical test showed positive urease, catalase and oxidase test but negative for H2S and citrate test. The results of this test ensure that isolated bacteria was *Brucella suis*.

Discussion

The test results using *Rose Bengal Test* (RBT) towards blood serum of pigs showed that there was a positive reaction in 1 sample. Positive results of RBT were characterized by agglutination, means that the antigen and antibody was homolog and eventually agglutinated. Negative results of RBT proved that antigen and antibody was not homolog thus agglutination not occurred (McCughey, 1972; Mylrea, 1972, Rolfe and Sykes, 1987).

Negative H_2S tests mean that bacteria did not break sulfides to H_2S , where it was characterized by the absence of black color on TSIA media. The negative urease means that the bacteria did not own an urease enzyme that have capability hydrolyzing urea which changed yellow-colored alkaline become pink acid by using methyl red indicator. The negative citrate means that bacteria could not break the citrate so that there was no carbon element for cell metabolism which may changed green color to blue with bromine thymol blue as an indicator.

Catalase is produced by certain bacteria, which acts as a catalyst in breakdown of hydrogen peroxide into water and oxygen. If bubbles are produced, the organism is catalase positive and if bubbles are not produced, the organism is catalase negative.

The oxidase test is used to identify bacteria that produce cytochrome c oxidase, an enzyme of the bacterial electron transport chain. When present, the cytochrome c oxidase oxidizes the reagent (tetramethyl-p-phenylenediamine) to (indophenols) **purple** color end product. When the enzyme is not present, the reagent remains reduced and is colorless (Sulaiman, et al., 1993; Sapardi, et al., 2004).

Brucella suis affected gestation pigs aged 2-3 months. *Brucella suis* affected both male and female pigs. In gestation female pigs would cause miscarriage, while in male pigs would cause orchitis. The Brucella's germs outside the sow body could survive in a variety of environmental conditions within a certain time. The ability of Brucella bacteria living on dry land is four days outside the room temperature, in the moist soil can survive for 66 days and on a muddy soil to survive for 151-185 days (Gray and Martin,1980). According to Sudibyo (1998), *Brucella* bacteria can survive for 2 days in dirt or cages waste with relatively high temperatures. In the livestock drinking water germs can survive for 5-144 days and in the waste water for 30-150 days (Heck et al., 1980). This disease still remains a problem in many countries in the world because of its economic impact as well as the health impacts of its veterineries (Darwesh and Benkirane, 2001; Lumb, 2003).

Serological diagnosis accuracy is a very important factor for the success of Brucellosis control and eradication program. Serological examination is the most widely used for the diagnosis of Brucellosis, because of the simple serology test, fast and has a high accuracy (Oliver and Cooper, 1981). The Rose Bengal Test (RBT) method is widely used in many countries as a screening test against Brucellosis. After the screening tests, confirmation of the diagnosis by bacteriological test for bacterial isolates were used for screening tests (Herr et al., 1982; Manickam and Mohan, 1987, Martin, et al., 1987, Wrathall, et al., 1993, Paulo, et al., 2000).

Conclusion

Based on RBT test and bacteriological examination, there was 1 positive sample of *Brucellla suis*, that sample coming from Kediri district.

Conflict of interest: The authors declare that they have no conflict interest.

Acknowledgements

We thank *Ditjen DIKTI* (Directorate General of Higher Education) for donating this study through scheme *Program Penelitian Unggulan Perguruan Tinggi (PUPT)* / Universities Leading Research Programme, Decentralization.

References

- 1. Alton, G.G., L.M., Jones, R.D., Angus, and J.M., Verger. (1991). Techniques for the Brucellosis Laboratory. Institut Nasional De La Recherche Agronomique. Paris.
- Christi, T.E., W.R., Kerr, W.J., McCaughey. (1968). Brucellosis Eradication in Northern Ireland. Veterinary Record. 83: 176-183.
- 3. Corbel, M.J., (1985). Recent advances in the study of Brucella antigens and their serological cross reaction. Veterinary Bulletin. 55 (12): 927-942.
- 4. Corbel, M.J., (1997). Brucellosis an overview. Emerging Infectious Diseases. 3(2):221.
- 5. Corner L.A, G.G Alton. 1982. Bovine Brucellosis Standard Bacteriological Techniques. Standing Committee Agriculture -Animal Health Committee. Australian Bureau of Animal Health, Canberra.
- 6. Darwesh, M.A., A., Benkirane. (2001). Field investigations of brucellosis in cattle and small ruminants in Syria, 1990-1996. Revue Scientifique et Technique. International Office of Epizootics . 20 (3): 769-775.

- 7. Enright, F.M., (1990). The pathogenesis and pathobiology of Brucella infection in domestic animals. In: Animal Brucellosis. Edited by: Nielsen, K. and Duncan J.R. Chemical Rubber Company Press, Boca Raton, Ann Arbor, Boston.
- 8. Gray, M.D., S.W., Martin. (1980). An evaluation of screening programs for the detection of Brucellosis in dairy herds. Canadian Journal of Comparative Medicine. 44: 52-60.
- Heck, F.C., J.D., Williams, J., Preutt, R., Sanders, D.L., Zink. (1980). Enzyme Linked Immunosorbent Assay for detecting antibodies to Brucella abortus in bovine milk and serum. American Journal of Veterinary Research. 41: 2082-2084.
- 10. Herr, F.C., D., Roux, P.M., Pieterson. (1982). The reproducibility of result in bovine brucellosis serology and their correlation with the isolation of Brucella abortus. Journal of Veterinary Research. 49: 79-83.
- 11. Lumb, S. (2003). International pig Topics. Vol 18. East Yorkshire England.
- 12. Madkour, M.M. (1989) Brucellosis. Butterworths, London, 294.
- 13. Manickam, R., M., Mohan. (1987). Epidemiology studies on Brucella abortus infection in Miltch cattle. Indian Veterinary Journal. 64: 546-549.
- 14. Martin, S.W, A.H., Meek, P., Willeberg. (1987). Vetrinary Epidemiology, Principle and Methods. Iowa State University Press/Ames. pages. 22 40.
- McCaughey, W.J. (1972). Brucella Milk Ring Test on churd samples: A three year study. Veterinary Record. 90: 6-10. Mylrea, P.J. (1972). The diagnosis of brucellosis in dairy herds. Australian Veterinary Journal. 48: 369-375.
- 16. Nicoletti, P. (1990). Vaccination. In: Animal Brucellosis. Edited by: Nielsen K. and Duncan J.R. Chemical Rubber Company Press, Boca Raton, Ann Arbor, Boston.
- Oliver, D.G., R.S., Cooper. (1981). Comperative study of ELISA (IgG and IgM) with standard serological tests for diagnosis of brucellosis in cattle. 24th Annual Proceeding American Association of Veterinary Laboratory Diagnosticians. pages. 187-202.
- Paulo, P.S., A.M., Vigliocco, R.F., Ramondino, D., Marticorena, E., Bissi, G., Briones, C., Gorch, D.Gall and K. Nielsen. 2000. Evaluation of primary binding assay for presumptive serodiagnosis of swine brucellosis in Argentina. Clinical and Diagnostic Laboratory. Immunology. 7(5):828-831.
- 19. Priadi, A. (1992). Brucella suis infection as a zoonosis in Java. Penyakit Hewan 24(44):110112.
- 20. Rolfe, D.C., W.E., Sykes. (1987). Monitoring of dairy herds for Brucella abortus infection when prevalence is low. Australian Veterinary Journal. 64: 97-100.
- 21. Sapardi, M, B., Poermadjaja, T.B., Usman dan I. Sulaiman. (2004). Monitoring *Brucella suis* pada Babi di Jawa Tahun 2002-2003. *Bulletin Veteriner* Vol. III, No. 1 Edisi Januari-Maret, 1-6.
- 22. Sudibyo, A. (1998). Studi patogenisitas *Brucella suis* isolat lapang dan kemampuan penularannya dari babi ke manusia. *Jurnal Ilmu Ternak dan Veteriner* / Indonesian Journal of Animal and Veterinary Science. 3(4):257-263.
- 23. Sulaiman, I., Patten, B.E.; Darmadi, P. (1993). The evaluation of serology and bacteriology for the diagnosis of bovine brucellosis in south Sulawesi. Penyakit Hewan
- 24. Wrathall, A.E., E.S. Broughton, K.P., Gill and G.P., Goldsmith. (1993). Serological reactions to Brucella species in British Pigs. Veterinary Record. 132:449-454.