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LIST OF SYMBOLS AND ABBREVIATIONS

 $^{1}O_{2}$ Singlet oxygen

The Antioxidant, Antidiabetic, Anticancer Sin War Naw **THESIS**

A549 Alveolar carcinoma

ABTS 2,2-azinobis-3-ethylbenzo thizoline-6-sulfonate

ADA Americans with Disabilities Act

APC Adenomatous polyposis coli

APOP Apoptosis

BCL-2 B-cell lymphoma-2

BCL-xL B-cell lymphoma-extra large

BER Base excision repair

BHA 2-*t*-butyl-4-methoxyphenol

BHT 2,6-di-*t*-butyl-4-methyl phenol

CHCL₃ Chloroform

CIN Chromosomal instability

DMSO Dimethly sulfoxide

DNA Deoxyribonucleic acid

DNase Deoxyribonuclease

DPP-4 Dipeptidyl peptidase-4 enzymes

DPPH 2,2-diphenyl-1-picrylhydrazyl

DW Dry weight

FADD Fas-associated death domain

G1 phase Gap 1 phase

GI Glycaemia index

GLI Zinc finger protein

H₂O₂ Hydrogen peroxide

H₂SO₄ Sulphuric acid

HbA 1c Hemoglobin A1c

Hela Cervical cancer

HepG2 Heptacellular carcinoma

HIF1 Hypoxia inducible factor 1

HO₂• Hydroperoxyl radical

Huh7 Hepatocarcinoma cell line

IARC International Agency for Research on Cancer

IC₅₀ The Half Maximal Inhibitory Concentration

IL-6 Interleukin-6

IR Insulin resistance

LMWF Lower molecular weight fucoidan

MCF-7 Breast cancer

MDA-MB- Human mammary adenocarcinoma

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MGC-803 Human gastric adenocarcinoma cancer cells

MMR DNA mismatch repair

Mn Manganses

MTT 3-(4,5-dimethyliazol-2yl)-2,5-diphenyltetrazolium bromide

Na₂CO₃ Sodium carbonate

NER Nucleotide excision repair

O₂ Superoxide radical

P53 Tumor suppression protein

PARP Poly (ADP-ribose) polymerase

PI3K Phosphoinositide 3-kinase

pNPG p-nirophenyl α-glucopyranoside

PUFAs Polyunsaturated fatty acids

RB Retinoblastoma tumor suppressor gene

ROS Reactive oxygen species

RPMI-7951 Human malignant melanoma obtained

RTK Receptor tyrosine kinase

SK-ML-28 Human malignant melanoma

SK-ML-5 Human malignant melanoma

SMAD Intracellular mediator

SOD Superoxide dismutase

SP Sulfated polysaccharide

T2D Type 2 diabetes

T47D Breast cancer cell line
TNF Tumor necrosis factor

TNFR Tumor necrosis factor receptor

UV Ultraviolet

SUMMARY

Indonesia, an archipelagic country with 81,000 km long coastline, has a great potential for seaweed production: the most commonly found species there are red and brown seaweeds. The increased cases of oxidative stress have attracted the scholar's attention to explore the importance of antioxidant to resolve various public health concerns. The seaweeds contain antioxidants including carotenoids, vitamins E, chlorophylls, and polyphenol of ascorbic acid which prevent the oxidative stress stimulated by reactive oxygen species (ROS) such as hydroxyl radical, hydrogen peroxide, superoxide anions and nitric oxide that were reacted with biomolecules such as DNA, proteins, lipids and change the normal cellular functions which cause tissue damage and cell death.

Diabetes is considered a major global health threat, which can affect the people of all ages from different demographic regions. It is commonly known to result from the defects of beta cells that produced insulin or when the body is not able to use the secreted insulin. Cancer has become the second most life-threatening disease and one of the important health problems in worldwide that caused by an abnormal growth of cells and tissues. Internal causes of cancer may be attributed to lack of apoptotic function, genetic mutation, oxidative stress, and hypoxia, while the external cause of cancer may be linked to excessive exposure to ultraviolet rays, radiation, pollution, smoking, and stress

The current study involved the investigation of crude extracts and three fractions of S. duplicatum and P. tetrastromatica from two different sites in phytochemical content, total phenolic content, antioxidant, antidiabetic, anticancer activities and toxicity. by using Folin-Ciocalteus method, the 2,2-diphernyl-1 picrylhydrazyl (DPPH), α-glucosidase enzyme, 3-(4,5-dimethylthiazol-2-yl)-2,5diphenyltetrazolium bromide (MTT) and brine shrimp assay, respectively. S. duplicatum and P. tetrastromatica were extracted with methanol solvent and the obtained crude extracts were partitioned with three solvents (normal hexane, ethyl acetate, and distilled water).

The result of total phenolic content in ethyl acetate fraction of S. duplicatum and P. tetrastromatica from non-oil extraction site had the highest contents (105.17±5.12^e and 589.79±7.14^g mg GAE/g) than those that are obtained from oil extraction site (66.20±6.54^{a,b} and 112.35±4.51^e mg GAE/g). Therefore, ethyl acetate fractions of S. duplicatum and P. tetrastromatica from non-oil extractions had the best potent of DPPH inhibitory activity (IC₅₀ 214.06±16.46^e and 25.25±5.15^{a,b} μg/mL) and the highest inhibitory effect against on α-glucosidase enzyme (IC₅₀ 712.51±9.44^h and 249.12±1.77^b µg/mL) as compared to those that are collected from oil extraction site (IC₅₀ 954.65 \pm 17.02¹ and 419.32 \pm 9.91^c μ g/mL). These fractions equally demonstrated the highest total phenol contents, which might be a major contributor to the antioxidant activities of the two seaweed species.

The inhibitory activity against on A549 (lung cancer cell line), crude extract and all fractions of P. tetrastromatica from non-oil extraction site demonstrated the strongest activities with IC₅₀ values of 80.44±12.88^b, 165.46±0.66^e, 70.56±2.56^a, 77.50±0.43^{a,b} µg/mL when compared with those that are from oil extraction site with IC_{50} values of 136.43 ± 7.12^d , 169.94 ± 1.19^e , 125.10 ± 0.51^c , 134.30 ± 1.14^d µg/mL. Meanwhile, the lowest activity was found in crude extract and all fractions of S. duplicatum from oil extraction site (IC₅₀ 241.49±3.83ⁱ, 361.22±3.20^j and 236.24±7.24ⁱ µg/mL) as a compared to those that are from non-oil extraction site $(IC_{50} 182.41\pm13.27^f, 214.98\pm1.33^g, 178.98\pm1.15^f, and 227.78\pm2.36^h \mu g/mL).$ According to exposures for toxicity test between 24 h and 48 h incubation time,

there was no 100% mortality rate found at a different concentration in the crude extracts and all fractions of *S. duplicatum* and *P. tetrastromatica* from two different sites. However, *S. duplicatum* and *P. tetrastromatica* from oil extraction site were considered to be mild toxic while those from non-oil extraction had nontoxic after 48 h of incubation.